High K⁺-Induced Relaxation by Nitric Oxide in Human Gastric Fundus

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This study was designed to elucidate high K⁺-induced relaxation in the human gastric fundus. Circular smooth muscle from the human gastric fundus greater curvature showed stretch-dependent high K⁺ (50 mM)-induced contractions. However, longitudinal smooth muscle produced stretch-dependent high K⁺-induced relaxation. We investigated several relaxation mechanisms to understand the reason for the discrepancy. Protein kinase inhibitors such as KT 5823 (1 μ M) and KT 5720 (1 μ M) which block protein kinases (PKG and PKA) had no effect on high K⁺-induced relaxation. K⁺ channel blockers except 4-aminopyridine (4-AP), a voltage-dependent K⁺ channel (K_V) blocker, did not affect high K⁺-induced relaxation. However, N(G)-nitro-L-arginine and 1H-(1,2,4)oxadiazolo (4,3-A)quino-xalin-1-one, an inhibitors of soluble guanylate cyclase (sGC) and 4-AP inhibited relaxation and reversed relaxation to contraction. High K⁺-induced relaxation of the human gastric fundus was observed only in the longitudinal muscles from the greater curvature. These data suggest that the longitudinal muscle of the human gastric fundus greater curvature produced high K⁺-induced relaxation that was activated by the nitric oxide/sGC pathway through a K_V channel-dependent mechanism.

Key Words: Fundus, High K⁺, Human stomach, Longitudinal smooth muscle, Nitric oxide, Relaxation

INTRODUCTION

The stomach performs complex function such as storage of food, breaking up of food particles, and emptying of its contents at a controlled rate [1,2]. Ingested food is stored in the gastric fundus and then triturated into the antral region of the stomach. Liquids and chime are passed into the duodenum through sieving and decanting, whereas solid particles are retained for longer periods. How these complicating mechanisms and processes are coordinated is still incompletely understood, except for several reflexes [3]. To date, the gastric fundus has been known to accommodate

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food via a reflex called receptive relaxation [1,2,4]. This reflex is characterized by graded wall distension with a small increase in intragastric pressure [4,5]. As receptive relaxation has been reported only in the fundus, we evaluated each part of the stomach according to gastric muscle layer as well as gastric region to understand the exact location of this phenomenon.

A non-adrenergic non-cholinergic (NANC) inhibitory mechanism is the final effectors of gastric receptive relaxation in the gastric fundus [5]. Nitric oxide (NO), whose formation is catalyzed by nitric oxide synthase (NOS) from L-arginine, is an inhibitory neurotransmitter that mediates smooth muscle relaxation in the mammalian gastrointestinal (GI) tract [4]. Neurogenic relaxation of the gastric fundus depends on a combination of NO and vasoactive intestinal peptide release [6,7]. In addition, NO plays an important role in the relaxation process of the gastric fundus

ABBREVIATIONS: PK, protein kinase; sGC, an inhibitor of soluble guanylate cyclase; NANC, non-adrenergic noncholinergic; NO, nitric oxide; NOS, nitric oxide synthase; GI tract, gastrointestinal tract; VDCC_L, voltage-dependent L-type $\mathrm{Ca^{2+}}$ channels; GC, greater curvature; LC, lesser curvature; ACh, acetylcholine; SNP, sodium nitroprusside; TEA, tetraethylammonium; $\mathrm{K_{ca}}$ channel, $\mathrm{Ca^{2+}}$ -activated $\mathrm{K^{+}}$ channel; KBC, $\mathrm{K^{+}}$ channel blockers cocktail; L-NNA, N(G)-nitro-L-arginine; ODQ, 1H-(1,2,4)oxadiazolo (4,3-A)quinoxalin-1-one; 4-AP, 4-aminopyridine; $\mathrm{K_{V}}$ channel, voltage-dependent $\mathrm{K^{+}}$ channel; NBC, nerve blockers cocktail; $\mathrm{[Ca^{2+}]_{i,}}$ intracellular $\mathrm{Ca^{2+}}$; ICC, interstitial cells of Cajal.

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as reported in intact animals [8], and in isolated stomachs from guinea-pigs [4], mice [9], and humans [10]. To date, only a few studies have reported on the nature of neurogenic NANC relaxation in the human stomach.

Voltage dependent "L-type" Ca^{2^+} channels (VDCC_L) play a central role in the regulation of $[\operatorname{Ca}^{2^+}]_i$ in smooth muscle [11]. Ca^{2^+} is a key factor in the regulation of smooth muscle contraction [11]. Among various conditions, high K^+ stimulation produces contraction through membrane depolarization, which activates VDCC_L in gastric smooth muscle [12]. This mechanism is true except for the longitudinal muscles of the fundus, where high K^+ produces relaxation [13]. We recently found high K^+ -induced relaxation of human corporal longitudinal smooth muscle is NO mediated. However, we could not evaluate the precise mechanisms of the opposed response to high K^+ stimulation such as the NO source in these muscles. In addition, we still have no information on whether the high K^+ -mediated response shows regional differences in the human stomach.

Thus, this study focused on the region of the stomach that shows high K^+ -mediated relaxation and the main mechanisms responsible for the relaxation effect.

METHODS

Tissue preparation for isometric contraction

Human gastric tissues from both greater and lesser curvature were obtained from patients who underwent total gastrectomy. Some tissue sample obtained by total gastrectomy who already underwent subtotal gastrectomy. This specific sample was obtained from patient underwent repetitive gastrectomy for recurrence of gastric cancer. All patients gave written informed consent and this experimental protocol for using human stomach was also approved by the Institutional Review Board for Clinical Research of Chungbuk National University. Specimens from macroscopically normal tissue of neoplastic area were removed immediately after surgical resection of stomach. In Krebs (KRB) solution, specimens were pinned down on sylgard plate. After removal of mucosa and submucosa, muscle strips (0.5×2 cm, 0.5 cm thickness) were prepared from the fundus according to muscle direction (circular and longitudinal direction) and mounted to organ bath (25 ml and 75 ml) of isometric contractile measuring system. For confirmation pathologist identified smooth muscles of human stomach using HE staining later. In vertical chamber, one end of strip was tied tightly to the holder and the other side was linked to force transducer by hook type holder (Harvard, USA). Force transducer was connected to PowerLab-Data Acquisition System, which was linked to IBM compatible computer operated by Charter v5.5 software (ADinstruments, Colorado, USA) for measuring isometric contraction. Each strip was stretched passively to resting tension after $1 \sim 1.5$ hours equilibration. Then contractile responses of the strip to the high K⁺ (50 mM, 10 min) was repeated two or three times until the responses were reproducible.

Solution and drugs

KRB solution (CO₂/bicarbonate-buffered Tyrode) contained (in mM): NaCl 122, KCl 4.7, MgCl₂ 1, CaCl₂ 2, NaHCO₃ 15, KH₂PO₄ 0.93, and glucose 11 (pH $7.3 \sim 7.4$,

bubbled with 5% CO₂/95% O₂). Equimolar concentration of Na⁺ was replaced by K⁺ to make high K⁺ (50 mM) solution. The external solution was changed by solutions which had previously been incubated (bubbled with 5% CO₂/95% O₂, 36°C) in water bath before the application. Sustained response by high K⁺ stimulus in the presence and absence of various blockers in this study was recorded for 10 min and measured at end of recording at 10 min. And pretreatment of various blockers was applied for 12 min before the application of high K⁺. K⁺ channel blockers cocktail except 4-AP (KBC) was applied (tetraethylammonium, (TEA, 10 mM); apamin (APA, 300 nM); glibenclamide (Glib, 20 μM); (1 mM) before application of stimulators to block each K+ channel responses. To rule out nerve mediated response, sometimes nerve blockers cocktail (NBC; tetrodotoxin (TTX, 0.4 μ M), guanethidine (1 μ M), atropine (ATR, 1 μ M) was also used. Sometimes phentolamine (Phento, 5 μ M) and proparnonol (Propra, 5 μ M) was added to NBC for blocking adrenergic response too. All drugs used in this study were purchased from Sigma.

Statistics

Data were expressed as means \pm standard errors (means \pm SEM). The Student's t-test was used to measure the statistical significance. p value less than 0.05 was regarded as statistically significant.

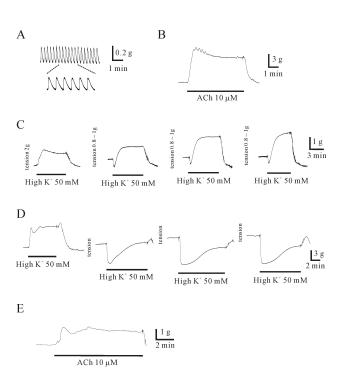


Fig. 1. Contractions of circular and longitudinal smooth muscle of the fundus greater curvature (GC). Circular muscle of the fundus GC showed spontaneous contraction (A). In (C) high-K $^+$ produced an initial and a tonic contraction and it reached peak by stepwise stretch (0.8 ~ 1 g). ACh (10 $\,\mu$ M) produced triphasic contractions in panel (B). Longitudinal muscle of the fundus lesser curvature (LC) obtained from a patient who underwent repetitive gastrectomy showed large high K $^+$ -induced relaxation (D). ACh produced contraction in the longitudinal muscle (E).

Acetylcholine (ACh, 10 mM) High K+-induced response Spontaneous contraction Phasic (g, cycles/min) Initial Initial peak Tonic Sustained (g, cycles/min) Fundus GC 0.2±0.06 g (n=17) 5.1±1.66 g (n=13) 3.7±0.97 g (n=14) 1.1±0.25 g (n=14) 0.7±0.22 g (n=12) 5.3±1.54 g (n=7) CircM 4.8±0.26 (n=17) Fundus GC 0.15±0.03 g (n=9) 0.5 ± 0.09 g (n=14) -1.4 ± 0.23 g (n=18) 1.34 ± 0.13 g (n=10) 1.0 ± 0.18 g (n=12) Long M 3.5 ± 0.17 (n=9) 0.03±0.01 g (n=4) 4.7 ± 0.13 g (n=4) 0.7 ± 0.27 g (n=2) Fundus LC 6.1±1.14 g (n=4) 5.0 ± 1.28 g (n=4) 2.5 ± 0.67 g (n=4) 1.5±0.21 (n=2) CircM1.7±0.24 (n=4) Fundus LC L: 1.7±0.20 g (n=4) 4.2±0.63 g (n=4) 0.7 ± 0.66 g (n=4) 6.0±1.0 g (n=4) 3.7 ± 2.2 g (n=4) 3.2±0.9 g (n=4) LongM 0.5 ± 0.05 (n=4) 1.1 ± 0.23 (n=4) S: 0.3±0.03 g (n=3)

Table 1. Analyzed data from circular and longitudinal smooth muscle in various regions of human stomach

GC, greater curvature; CircM, circular muscle; LongM, longitudinal muscle; L, large; S, small.

RESULTS

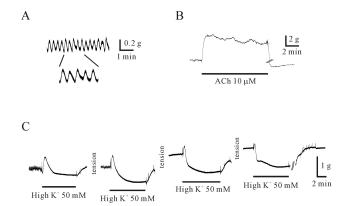
Isometric contraction of circular smooth muscle in the human gastric fundus greater curvature (GC)

Human gastric circular muscle from the fundus GC showed spontaneous contractions (0.2 g, n=17, Fig. 1A, Table 1). However, spontaneous contractility generally disappeared, except some cases due to stepwise stretch (Fig. 1C). High K⁺ (50 mM) also produced an initial peak and sustained tonic contraction of 5.3 g and 5.1 g (Fig. 1C, Table 1). Contraction of the GC reached a peak level by applying a stepwise stretch (0.8~1 g) from normal length (Fig. 1C). Initial transient, phasic and sustained tonic contractions were produced by acetylcholine (ACh) (10 μ M) (Fig. 1B; Table 1).

Isometric contraction of longitudinal smooth muscle in the human gastric fundus GC

Isometric contractions of the longitudinal muscle in the human gastric fundus were investigated (Fig. 1D and 1E). This sample was obtained from a patient who underwent repetitive gastrectomy for a gastric cancer recurrence. In contrast to circular muscle of the fundus GC (Fig. 1C), longitudinal smooth muscles showed high K⁺-induced relaxation repetitively with stepwise stretches (1.5 $\sim\!2$ g) (Fig. 1D and 1E). However, ACh (10 μ M) elicited contraction in these tissues (Fig. 1E). We investigated contractile properties of longitudinal muscle to verify these discrepancies between circular and longitudinal muscle. As shown in Fig. 2A, isometric contraction of longitudinal muscle resulted in spontaneous rhythmic contractions of 0.15 g (n=9; Fig. 2A; Table 1), which were attenuated and nearly abolished after applying a 1g load 3-5 times (Fig. 2C and 2D). High K and ACh also produced contractions (Fig. 2A and 2B; Table

To test the stretch-dependency of high K^+ -induced relaxation, we applied increasing tension to the fundus GC longitudinal muscle strip in a stepwise fashion. As shown in Fig. 2C, high K^+ -induced relaxation increased: initial contraction and subsequent relaxation $(0.5\pm0.09~g$ and of $-1.4\pm0.23~g$; n=14 and n=18; Fig. 2Ea). The response was $-61\pm9.67\%$ of total relaxation to basal tone (n=18, Fig. 2Eb). High K^+ -induced relaxation was compared to the effect of sodium nitroprusside (SNP), a nitric oxide (NO) donor. SNP



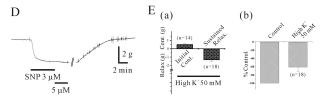


Fig. 2. High K⁺-induced relaxation of longitudinal smooth muscle in the fundus GC. Longitudinal muscle showed spontaneous (A) and ACh-induced contraction (B) however high K⁺ produced relaxation (C). High K⁺-induced relaxation was gradually increased by application of stepwise stretch (C). In (D) SNP (3 and 5 μ M) produced relaxation of -3.0 g. High K⁺ produced relaxation (-1.4 g; Ea) and it was -61% of total relaxation to basal tone (Eb).

(3 and 5 μ M) produced relaxation of -3.0 ± 0.39 g and -3.0 ± 0.41 g, which was equivalent to $-95\pm12.5\%$ and $-99\pm13.7\%$ from basal tone, respectively (n=4; Fig. 2D). Therefore, stretch-dependent relaxation (-1.5 ± 0.47 g) was equivalent to $-41\pm14.6\%$ of SNP (5 μ M)-induced relaxation (n=3, respectively).

Effects of K^{\dagger} channel blockers on stretch-dependent relaxation of longitudinal muscle from the human gastric fundus GC

Because high K^+ produced relaxation in longitudinal muscle, we studied whether activation of K^+ channels is

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associated with this stimulation using K⁺ channel blockers. Applying tetraetylammonium (TEA, 10 mM), a blocker of +-activated K⁺ channels (K_{Ca} channel), decreased phasic contractions (0.03±0.02 g, n=3 from 0.06±0.02 g, n=4), increased frequency (4.8±0.17 cycles/min, n=3 from 3.4±1.12 cycles/min, n=4) and produced slight tonic contractions (0.4 ±0.13 g, n=6). However, high K⁺-induced relaxation (-0.8 ± 0.15 g, n=6) was not inhibited but rather potentiated (-1.4 ±0.23 g, n=6) by TEA pre-treatment (Fig. 3A), as TEA (10 mM) produced tonic contractions (0.4 g). High K⁺-induced relaxation was also investigated in the presence of a K⁺ channel blocker cocktail (KBC, except 4-AP, see Methods). As shown in Fig. 3B and 3Ca, the KBC produced a tonic contraction of 0.6±0.24 g (n=6), and high K⁺-induced relaxation in the presence of the KBC was -1.2±0.28 g (n=6; $p \le 0.05$). The percent relaxation of high K^+ -induced relaxation with the KBC was comparable with -92±33.4 % from the initial basal tension level (n=6, Fig. 3Cb).

Effects of protein kinase (PK) inhibitors on high K^+ -induced relaxation of longitudinal smooth muscle in the human gastric fundus GC

The involvement of activated PKG and PKA on high K⁺-induced relaxation was evaluated. As shown in Fig. 3D, high K⁺-induced relaxation was produced in longitudinal

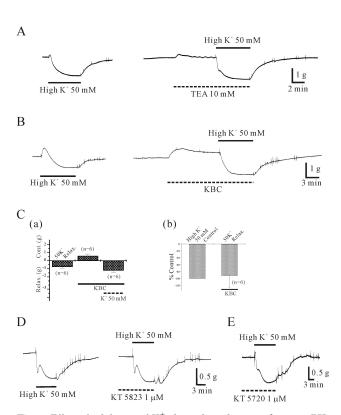


Fig. 3. Effect of inhibitors of K^+ channels and protein kinases (PK) on high K^+ -induced relaxation of longitudinal smooth muscle in the fundus GC. High K^+ -induced relaxation was not inhibited by tetraethylammonium (TEA, 10 mM) which is known inhibitor of Ca^{2+} -activated K^+ channel (K_{Ca} channel) (A) and also not by K^+ channel blockers cocktail (KBC except 4-AP, see methods; (B), (Ca, Cb). Furthermore, it was not affected by KT5823 and KT5720, blockers of PKG and PKA, respectively (D, E).

smooth muscle of the human fundus GC. High K⁺-induced relaxation was not inhibited by KT 5823 (1 μ M) or KT 5720 (1 μ M), blockers of PKG and PKA, respectively. High K⁺-induced relaxation was -1.8 ± 0.81 g and -2.0 ± 0.81 g in the absence and presence of KT 5823 (1 μ M, n=4), and was -1.5 ± 0.57 g and -1.3 ± 0.59 g in the absence and presence of KT 5720 (1 μ M, n=5), respectively (p>0.05; Fig. 3D and 3E). KT 5823 (1 μ M) and KT 5720 (1 μ M) weakly inhibited basal tone (-0.03 to -0.053 g; n=4 and 5; data not shown). Thus, the percent relaxation of high K⁺-induced relaxation was calculated as $-131.1\pm31.2\%$ and $-102.3\pm3.7\%$ in the presence of KT 5823 and KT 5720, respectively (n=4 and 5, p>0.05).

Effects of NO inhibitors and soluble guanylyl cyclase (sGC) on high K^{\dagger} -induced relaxation of longitudinal smooth muscle in the human gastric fundus GC

The involvement of NO and sGC on high K^+ -induced relaxation was investigated. High K^+ -induced relaxation was significantly antagonized and reversed to contraction by NG-nitro-L-arginine (L-NNA), an NO biosynthesis inhibitor. As shown in Fig. 4A, high K^+ -induced relaxation was -1.4 ± 0.34 g in the absence of L-NNA (n=14). However, high K^+ produced initial peak contraction (1.4 ±0.34 g, n=13) and sustained tonic contraction (1.01 ±0.29 g, n=14) after L-NNA treatment (100 μ M), which slightly increased basal tone to 0.02 ±0.13 g (n=14) (Fig. 4A and 4C). The effect

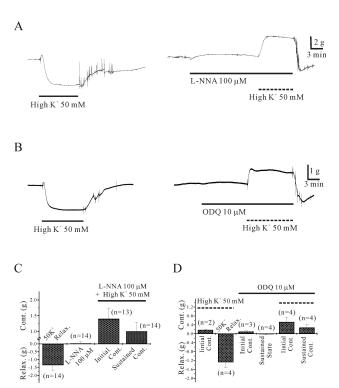


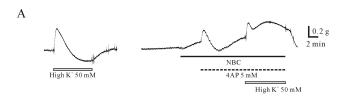
Fig. 4. Involvement of nitric oxide (NO) and soluble guanylyl cyclase (sGC) on high K^+ -induced relaxation of longitudinal smooth muscle in the fundus GC. In (A, B) high K^+ produced relaxation in longitudinal muscle in the fundus GC. However, L-NNA (100 $\mu\,\mathrm{M})$ and ODQ (10 $\mu\,\mathrm{M})$ blocked and reversed high K^+ -induced relaxation to contraction (A, B). The data were summarized in (C, D).

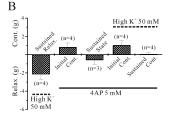
of ODQ, an sGC inhibitor, on high K $^+$ -induced relaxation was also studied. Applying ODQ (10 μ M) produced an initial contraction (0.1±0.04 g) and a slowly decaying contraction to -0.02 ± 0.02 g, respectively (n=3 and 4 respectively). In the absence of ODQ, high K $^+$ -induced relaxation (-1.3 ± 0.3 g, n=4) was reversed to initial and sustained contractions of 0.5±0.22 g and 0.3±0.16 g (n=4, respectively; Fig. 4B and 4D).

Effects of 4-aminopyridine (4-AP) on high K^+ -induced relaxation of longitudinal smooth muscle in the human gastric fundus GC

The involvement of K_V channels on high $K^+\text{-}induced$ relaxation was also investigated, using 4-AP and a nerve blocker cocktail (NBC, see methods). As shown in Fig. 5B, high $K^+\text{-}induced$ relaxation was -2.2 ± 0.59 g in the absence of 4-AP (n=4). 4-AP (5 mM) produced initial contractions, transient, and sustained relaxation of 0.8 ± 0.45 g, -1.3 ± 0.86 g, and -0.6 ± 0.49 g (n=4, 3 and 3, respectively). However, high $K^+\text{-}induced$ relaxation was reversed in the presence of 4-AP to initial (1.0±0.48 g, n=4) and following sustained contraction (0.1±0.15 g, n=4) (p<0.05).

NBC itself did not show any significant effect on basal contractility. High K^+ -induced relaxation in the absence and presence of the NBC was -2.6 ± 0.33 g and -2.8 ± 0.53 g, respectively (n=6, respectively; p>0.05). To study the





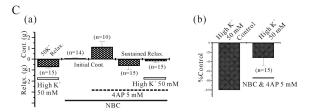


Fig. 5. Effects of 4-aminopyridine (4-AP) on high K^+ -induced relaxation of longitudinal smooth muscle in the fundus GC. Most high K^+ -induced relaxation were blocked or reversed to contraction (A), though some were not, by pretreatment of 4-AP with nerve blockers cocktail (NBC, see methods; C). In (B) effect of 4-AP (5 mM) on high K^+ -induced relaxation was summarized. In the presence of 4-AP (5 mM), high K^+ -induced relaxation was inhibited. In the presence of NBC and 4-AP, the percent relaxation of high K^+ -induced relaxation was suppressed (Ca) to -32% of the control (Cb).

possible involvement of neurotransmitters in the 4-AP effect, we also studied the effect of 4-AP in the presence of the NBC, as a neurotransmitter releasing effect by 4-AP, a Ky channel blocker, has been reported. As shown in Fig. 5C, high K⁺-induced relaxation in the control was -0.7 ± 0.1 g (n=15). Applying the NBC slightly increased basal tone by 0.1±0.04 g (n=14) and adding 4-AP produced an initial contraction of 1.1±0.48 g (n=10), followed by relaxation of -0.6±0.35 g (n=15). Most high K⁺-induced relaxation was inhibited or reversed to contraction in the presence of NBC and 4-AP, although some were not (Fig. 5A). Therefore, high K⁺-induced relaxation in the presence of 4-AP and the NBC significantly attenuated the value to -0.18 ± 0.14 g (n=15) compared to that of high K+-induced relaxation without 4-AP (-0.7 ± 0.1 g, n=15) (p<0.05; Fig. 5Ca). The percent relaxation of high K⁺-induced relaxation in the presence of the NBC with 4-AP was -32±16.2% from the initial basal tension level (n=15) (Fig. 5Cb).

Response of circular and longitudinal smooth muscle of the human stomach fundus lesser curvature (LC) to high K^{\dagger}

Circular smooth muscle from the fundus LC showed phasic contractions of 0.03 g. However, longitudinal muscle of the fundus LC showed large phasic contractions of 1.7 g, and small phasic contractions of 0.3 g were superimposed on top of these large contractions (n=4 and 3; Table 1). High $\rm K^+$ produced initial and tonic contractions of 6.1±1.14 g and 5.0±1.28 g in circular muscle (n=4, respectively; Fig. 6A). Longitudinal muscle also produced an initial contraction of 4.2±0.63 g followed by a tonic contraction of 0.7±0.66 g due to high $\rm K^+$ (n=4, Fig. 6B).

DISCUSSION

The stomach elicits various motor activities to perform its physiological functions such as acting as a reservoir and grinding and so on [14,15]. Among them, receptive relaxation is the best characterized function of the proximal stomach. Gastric relaxation of the fundus is key to accommodate ingested material with a minimal increase in intraluminal pressure. To date, gastric accommodation has been explained by various mechanisms. One of them is receptive relaxation known as the vago-vagal reflex [5]. The second mechanism is called adaptive relaxation by the short vagal

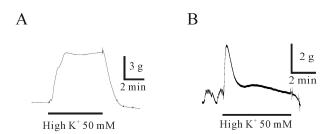


Fig. 6. High K⁺-induced response in the fundus LC. Contractile properties of circular and longitudinal smooth muscles of the fundus LC were investigated too. Circular muscle of the fundus LC showed high K⁺-induced contraction (A). Meanwhile, high K⁺ produced an initial contraction followed by an weak contraction in longitudinal muscle of the fundus LC (B).

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reflex. In particular, NO is a principal candidate for fundus relaxation [16]. We recently discovered NO-dependent relaxation in the corpus [13]. We also found that L-NNA inhibited high K^+ -induced relaxation in longitudinal smooth muscle of the fundus GC, as shown in Figs. 2C, 2E and $3 \sim 5$. Furthermore, ODQ blocked high K^+ -induced relaxation and reversed relaxation to contraction (Fig. 4B and 4D).

We also investigated the involvement of PKG on high K⁺-induced relaxation, as NO is known to activate guanylate cyclase (Fig. 3D and 3E). However, the PKG inhibitor KT5823 and the PKA inhibitor KT5720 (and S22536, not shown) did not affect this relaxation effect. Thus, we suggest that NO/cGMP-mediated the high K⁺-induced relaxation of longitudinal muscle in the human proximal stomach. Direct NO/cGMP-dependent and PKG-independent pathways for regulation of K channels are also present [17]. Interestingly, high K⁺-induced contraction of some circular muscle was followed by small transient relaxation, which was blocked by L-NNA (Fig. 1C). This observation suggests that NO mediated relaxing mechanisms are also present in circular muscle to a lesser degree.

NO is localized in myenteric neurons of the human gastric fundus and mediates relaxation of the fundus [10]. Relaxation by graded mechanical distention has been partly ascribed to a NO-mediated neurogenic component. Although it remains controversial, NO is believed to mediate gastric accommodation in the human stomach [10]. Because high K^+ -induced relaxation in human fundus longitudinal muscle was completely inhibited by L-NNA and converted to contraction (Fig. 4A and 4C), we believe that NO is a mediator for high K^+ -induced relaxation [18].

Several layers of muscle are found in the human stomach [19,20]. Circular and longitudinal muscle is regulated by the coordination of slow waves [21]. In this study, we found a functional dissociation between these muscles in the human stomach. Stimulation with high K^+ concentrations resulted in contraction of circular muscles but relaxation of longitudinal muscles. The mechanism underlying the responses of each muscle layer seems very complicated, indicating different regulation of intracellular Ca^{2+} ([Ca^{2+}];) by K^+ -stimulation and/or regional differences in electrical characteristics of each type of gastric muscle [12,22,23]. Dissociation of motility between both muscles might come from different characteristics of the muscles and/or neural responses [12].

The relaxation phenomenon of the human gastric fundus after gastrectomy is clinically interesting. We found that this relaxation was highly enhanced in a patient who underwent repetitive gastrectomy (Fig. 1D and 1E). A direct complication of gastrectomy is loss of gastric reservoir function. However, gastric volume after a subtotal gastrectomy is partly recovered by proper alimentotherapy. As a subtotal gastrectomy saves the gastric proximal region, its relaxation seems very natural. Further study is required to understand this phenomenon.

All visceral smooth muscles require Ca^{2+} for contraction, and the amplitude of the contraction depends on $[Ca^{2+}]_i$ [24]. To date, VDCC_L have been described in most excitable tissues, including the human stomach [25], and VDCC_L play an essential role in the regulation of $[Ca^{2+}]_i$ [11]. Spontaneous contractility of GI muscles is closely related to slow waves, and activation of VDCC_L on the slow wave plateau is necessary for excitation-contraction (E-C) coupling [26]. In general, phasic contraction of the GI tract

produces peristaltic contractions [28]. Therefore, an investigation of Ca2+ current regulation is essential to understand gastric functions [27]. We studied the effect of high K⁺-induced contraction in the human stomach (Fig. 1C). High K⁺ generally produces contractions via activation of VDCC_L in gastric smooth muscle [12]. However, we found relaxation due to high K+ stimulation in longitudinal muscle of the fundus from the GC. Relaxation of longitudinal smooth muscle in contrast to circular muscle occurs due to various reasons such as different spontaneous electrical activity in each muscle [23], dissociation of motility and regulation of [Ca²⁺]_i, including the different properties of the muscle fibers [12]. Differences in the cytoskeleton and/or some unknown regulatory mechanism related to mechanical stress might underlie this relaxation response [28,29]. These results suggest that longitudinal muscle might also have special directionality of its own like circular muscle.

As shown in Figs. 4A, 4C, and 5, high K⁺-induced relaxation of longitudinal muscle was transited to contraction by 4-AP and ODQ but not by KT5823 and KT5720 (Fig. 3D and 3E). These findings suggest that the NO/sGC pathway is selectively involved (Fig. 3D and 3E). In fact, K⁺ channels such as the K_V channel [17,30] and K_{Ca} channel [31] are directly regulated by the NO/sGC pathway in smooth muscle [13]. In addition, nNOS neurons are found in the myenteric region of the human stomach [32]. Interstitial cells of Cajal (ICC) express NO-sensitive sGC and are a primary target of NO released from nNOS neurons; thus, NO signals are transmitted to smooth muscle in the GI tract [33,34]. From these observations, we suggest that high K⁺-induced relaxation is produced by activating VDCC_L and by the NO/sGC-dependent pathway of the ICC in the longitudinal muscle layer.

NOS neurons are found in circular and longitudinal muscle layers, including the myenteric border of the GI tract [32]. However, we found that NO produced relaxation only in longitudinal smooth muscle of the human gastric greater curvature. Unfortunately, we could not clarify exactly where the NO originated. Although we used a NBC to block the nerve effect when applying high K^{\pm} stimulation, there still remained a possibility of NO release via activation of Ca^{2^+} channels in the NANC nerve. Therefore, the NANC nerve or other NO releasing cells could be involved in this relaxation response. In addition to an NO source, the reason why only longitudinal muscles respond with relaxation to high K^+ stimulation in the greater curvature side and why an enhanced response occurs in patients with recurrent gastric cancer must be investigated.

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