Review Article MicroRNAs: New Insights into Chronic Childhood Diseases

Ahmed Omran,^{1,2} Dalia Elimam,¹ and Fei Yin²

¹ Department of Pediatrics and Neonatology, Suez Canal University, Ismailia 41522, Egypt

² Department of Pediatrics, Xiangya Hospital of Central South University, 87 Xiangya Road, Changsha, Hunan 410008, China

Correspondence should be addressed to Fei Yin; yf_2323@yahoo.com

Received 16 April 2013; Accepted 7 June 2013

Academic Editor: Glen Jickling

Copyright © 2013 Ahmed Omran et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chronic diseases are the major cause of morbidity and mortality worldwide and have shown increasing incidence rates among children in the last decades. Chronic illnesses in the pediatric population, even if well managed, affect social, psychological, and physical development and often limit education and active participation and increase the risk for health complications. The significant pediatric morbidity and mortality rates caused by chronic illnesses call for serious efforts toward better understanding of the pathogenesis of these disorders. Recent studies have shown the involvement of microRNAs (miRNAs) in various aspects of major pediatric chronic non-neoplastic diseases. This review focuses on the role of miRNAs in four major pediatric chronic diseases including bronchial asthma, diabetes mellitus, epilepsy and cystic fibrosis. We intend to emphasize the importance of miRNA-based research in combating these major disorders, as we believe this approach will result in novel therapies to aid securing normal development and to prevent disabilities in the pediatric population.

1. Introduction

The prevalence of children with chronic illnesses varies widely with an overall rate of 10% to 20% [1] and is expected to increase further. Childhood chronic illnesses represent a major challenge and burden for affected families and the health care system. There is evidence that chronically ill children and their families are at greater risk for developing psychological and emotional difficulties than healthy children and their families. Many chronically ill children grow up in hospitals and live a life far from normal due to recurrent hospitalizations. They often show growth retardation as a result of the illness itself or its pharmacological treatment options. The long-term requirement for medical and social care of these children can be extremely complex and expensive.

The mandate for the child to adopt many self-care skills for monitoring and safety represents a major part of the challenge during the disease course.

The main goal for pediatricians is to maximize the children's functional abilities and sense of well-being, their health-related quality of life, and their development into healthy and productive adults. Chronic diseases in children and adolescents are far from rare and are today more likely described as an epidemic, which calls for major efforts to understand causation and improve prevention and treatment protocols.

There is a wealth of evidence on the diverse role of miR-NAs in many biological processes, including proliferation, differentiation, apoptosis, and development. The list of diseases in which dysregulation of miRNAs has been implicated is constantly growing and includes major pediatric chronic non-neoplastic diseases. We recently reviewed the role of miRNAs in pediatric central nervous system and cardiovascular diseases including congenital heart diseases [2, 3]. This review summarizes recent progress in edge-cutting research about the involvement of miRNAs in bronchial asthma, diabetes mellitus, epilepsy, and cystic fibrosis (Table 1).

2. miRNAs and Bronchial Asthma

Bronchial asthma is a chronic disorder of the airways that is characterized by variable and recurring airflow obstruction, chronic airway inflammation, bronchial hyperresponsiveness, and tissue remodeling [4, 5]. Three hundred million people are suffering from asthma worldwide, over 22 million

Disease	miRNAs	Mechanism	Reference
Bronchial asthma			
(1) Risk	MiR-148a, miR-148b, and miR-152	Interacting with HLA-G	[9]
	pre-miRNAs	rs2910164G/C and rs2292832C/T SNP	[10]
	MiR-155	Decreased expression increase asthma severity	[11]
Disease Bronchial asthma (1) Risk (2) Pathogenesis (3) Therapeutic targets Diabetes mellitus (1) Physiological aspects (a) Pancreas development (b) Insulin biosynthesis (c) Insulin biosynthesis (c) Insulin secretion (d) Insulin actions (2) Type 1 diabetes (3) Type 2 diabetes	MiR-146b, miR-223, miR-29b, miR-29c, miR-483, miR-574, miR-5p, miR-672, and miR-690	Abnormally expressed in asthma models	[12–19]
	MiR-221	Regulate mast cell functions	[20, 21]
	MiR-21	Polarize Th cells toward Th2	[12]
	MiR-126	Its blockage diminished Th2 responses	[13]
Disease Bronchial asthma (1) Risk (2) Pathogenesis (3) Therapeutic targets Diabetes mellitus (1) Physiological aspects (a) Pancreas development (b) Insulin biosynthesis (c) Insulin biosynthesis (2) Type 1 diabetes (3) Type 2 diabetes	MiR-146a	Contribute in remodeling	[25]
	let-7 mimic	Reduced IL-13 levels	[26]
	MiR-145	Pro-inflammatory effect	[27]
	MiR-133a	Modulate RhoA/Rhokinase pathway	[17]
(3) Therapeutic targets	MiR-126	Suppress Th2-driven airway inflammation	[13]
	MiR-106a	Inhibit IL-10	[35]
	MiR-146a	Mediate anti-inflammatory effect of dexamethasone	[36]
	Anti-miR-145	Reduce severity of airway inflammation	[37]
Diabetes mellitus			
(1) Physiological aspects			
	MiR-124a2	Pancreatic β -cell development	[41]
(a) Pancreas development	MiR-375	Formation of pancreatic islets	[42]
(a) Pancreas development	MiR-375	Maintenance pancreatic endocrine mass viability	[43]
(a) Pancreas development	MiR-15a	Targeting UCP-2	[44]
	MiR-30d	Activates MafA expression	[45]
(b) Insulin biosynthesis	MiR-375, miR-122, miR-127-3p, and miR-184	Insulin biosynthesis	[46]
	MiR-133a	Suppress insulin biosynthesis	[47]
	MiR-9	Secretory function of insulin producing cells	[48, 49]
	MiR-375	Interacting with HLA-G rs2910164G/C and rs2292832C/T SNP Decreased expression increase asthma severitydAbnormally expressed in asthma modelsRegulate mast cell functions Polarize Th cells toward Th2 Its blockage diminished Th2 responses Contribute in remodeling Reduced IL-13 levels Pro-inflammatory effectModulate RhoA/Rhokinase pathway Suppress Th2-driven airway inflammation Inhibit IL-10 Mediate anti-inflammatory effect of dexamethasone Reduce severity of airway inflammationPancreatic β -cell development Formation of pancreatic islets Maintenance pancreatic endocrine mass viabilit Targeting UCP-2 Activates MafA expression Insulin biosynthesisSuppress insulin biosynthesis Secretory function of insulin producing cells Regulate insulin secretion Inibit insulin secretion Inibit insulin secretionNotytine-mediated β -cell dysfunction Cytokine-mediated β -cell dysfunction Deregulated in T1D modelInhibit insulin-stimulated AKT activation Mediate insulin resistance Increased expression in T2D Deregulated in plasma of T2D patients	[50]
(c) Insulin secretion	MiR-124a and miR-29	Optimal insulin secretion	[41, 52]
	MiR-33a	Inversely correlates with ABCA1 expression	[89]
	MiR-21, miR-34a, and miR-146	Inhibit insulin secretion	[54]
	MiR-103/107	Insulin sensitivity	[55]
(d) Insulin actions	Lin28/let-7	and Insulin biosynthesis Suppress insulin biosynthesis Secretory function of insulin producing cells Regulate insulin secretion Optimal insulin secretion Inversely correlates with ABCA1 expression Inhibit insulin secretion Insulin sensitivity Regulation of glucose metabolism	[56]
(2) Type 1 diabetes	MiR-29 family	Cytokine-mediated β -cell dysfunction	[59]
	MiRs (124, 128, 192, 194, 204, 375, 672, and 708)	Deregulated in T1D model	[61]
	MiR-143	Inhibit insulin-stimulated AKT activation	[68]
(3) Type 2 diabetes	miR-146a impairment	Mediate insulin resistance	[69]
(3) Type 2 diabetes	MiR-125a	Increased expression in T2D	[70]
	MiR-126	Deregulated in plasma of T2D patients	[77]

TABLE 1: An overview of miRNAs in the major chronic non-neoplastic childhood diseases.

BioMed Research International

Disease	miRNAs	Mechanism	Reference
(4) Complications	MiRs (144, 146a, 150, 182, 192, 30d, and 320)	Biomarkers for diabetes progression	[76]
	MiR-192	Increased in glomeruli of diabetic mice	[78]
	MiR-200b/c, miR-216a, and miR-217	Detected in glomeruli of diabetic mice	[79-81]
	MiR-377	Play a role in DN renal fibrosis	[82]
	MiR-192	Reduced renal fibrosis and improves proteinuria	[89]
	MiR-126, miR-27b, and miR-130a	Proangiogenic miRNAs	[89]
	MiR-98	Modulate TRB2	[90]
	MiR-503	Caused diabetic impaired angiogenesis	[91]
	MiR-126	Related to impaired (EPC)	[92]
	MiR-186, miR-199a, and miR-339	Stem cell therapy of TID	[93]
(5) Therapeutic targets	MiR-21-PDCD4 pathway	Treating autoimmune T1D	[94]
	MiR-375	Facilitate insulin response	[42]
	MiR-181a	Improves hepatic insulin sensitivity	[96]
Epilepsy			
(1) Pathogenesis	MiR-213, miR-132, miR-30c, miR-26a, and miR-375	Prominently upregulated in MTLE acute stage	[102]
	MiR-29a and miR-181c	Prominently downregulated in MTLE acute stage	[102]
	MiR-21	Regulate neurotrophin-3 signaling	[103]
	MiR-let-7e and miR-23 a/b	Deregulated in the MTLE chronic stage	[103]
	MiR-146a	Differently expressed in different stages of MTLE development and may interact with IL-1 β	[107]
	MiR-155	Differently expressed in different stages of MTLE development and may interact with TNF- α	[108]
	MiR-132	Related to synaptic plasticity	[115]
(2) Potential blood biomarker	MiR-34a, miR-22, miR-125a, and miR-21	Showed different expression in the blood	[102]
	Anti-miR-132	Reduced seizure-induced neuronal death	[117]
(3) Therapeutic target	MiR-134 silencing	Neuroprotective effect	[118]
Cystic fibrosis	6	*	
, .	MiR-155	Activation of IL-8-dependent inflammation	[126]
	MiR-138	Regulates CFTR expression	[129]
	MiR-145, -223, and -494	Correlates with decreased CFTR expression	[130]
	MiR-101 and miR-494	Act synergistically on CFTR-reporter inhibition	[131]
	MiR-146	Significantly changed in the sputum of CF patients	[132]

TABLE 1: Continued.

people in the United States alone, of which over 6 million are children. The illness related cost is 6.2 billion US Dollars each year.

In the pediatric population, bronchial asthma is one of the most common chronic lung diseases affecting around 5%–10% of school-age children [6]; it is associated with reduced quality of life and exercise intolerance, accounts for a loss of 10 million school days [7], and is a leading cause of hospitalizations in children [8]. Current available asthma treatment is not effective in preventing airway remodeling processes and fails to prevent asthma exacerbations and hospitalizations even in children on appropriate controller medications. An improved understanding of the molecular mechanisms in asthma through exploring the role of miRNAs is expected to create promising potentials to reveal new approaches for primary prevention and identification of new therapeutic targets in childhood asthma.

miRNAs appear to play an important role in asthma development and pathogenesis. Susceptibility to asthma has been linked to the variation in specific miRNA genes and/or their specific miRNAs. The 3'UTR of HLA-G, a gene, which has been identified as an asthma-susceptibility gene [9], was found to be targeted by miR-148a, miR-148b, and miR-152. The possibility that miRNA variation is a key factor in the risk of developing asthma has been further supported. Significant differences in the genotype and allelic distribution of the pre-miRNAs SNPrs2910164G/C and rs2292832C/T among asthmatics and their controls indicated that this SNP may play a role in asthma development [10]. Another study suggested that decreased expression level of miR-155 plays an important role in the development of asthma and is correlated to asthma disease severity as well [11].

Recent reviews show the involvement of miRNAs in both the immunological and inflammatory components of asthma pathogenesis as well as in the neuronal control of airway smooth muscles. The role of miRNAs in the regulation of immunological pathways in asthma pathogenesis is rather central. The first evidence was obtained through detecting abnormal expression levels of miRNAs in asthma, including miR-146b, miR-223, miR-29b, miR-29c, miR-483, miR-574-5p, miR-672, and miR-690 [12–19].

Extrinsic asthma is an IgE mediated hypersensitivity reaction where the bridging of IgE triggers the release of mast cell mediators. MiR-221 is a likely regulator of mast cell activation [20] and proliferation including mast cells differentiation, migration, adhesion, cytokine production, and survival upon withdrawal of essential cytokines [21].

Asthma is described as Th2 mediated inflammation of the airway [22, 23]. Th2 cells, which play a fundamental role in allergic asthma pathogenesis [12], are polarized by cytokine IL-12p35, the molecular target of miR-21.

Upregulation of miR-21 in the allergic airway indicates its involvement in inflammation. Of similar importance to the pathogenesis of allergic airways disease is miR-126 [13]. The blockade of miR-126 suppressed the asthmatic phenotype in the form of diminished Th2 responses, suppressed inflammation, reduced airway hyperresponsiveness (AHR), inhibited eosinophil recruitment, and lowered mucus secretion [13].

IL-13 induces asthma features, such as epithelial cell hyperplasia, goblet cell metaplasia, deposition of various extracellular matrix proteins in subepithelial regions, and increased airway smooth muscle cell contractility, and seems to be under miRNA control [24].

miR-146a mimics modulate human bronchial epithelial cells (HBEC) survival by upregulating Bcl-XL and STAT3 phosphorylation and appear thereby to contribute to processes of tissue repair and remodeling which are hallmarks in asthma pathogenesis [25].

Intranasal administration of let-7 mimic reduces IL-13 levels in allergic lungs and alleviates these features [26], indicating that let-7 has anti-inflammatory effect through reduction of IL-13.

MiR-145 demonstrated to play an additional central proinflammatory role in the development allergic airways inflammation to house dust mites [27].

In addition to inflammation, dysfunctional neural control of airway smooth muscles (ASMs) is a major component of asthma pathogenesis. A functional cascade that involves Sonic hedgehog (Shh), miR-206 and brain derived neurotrophic factor (BDNF) has been recently uncovered and found to coordinate ASM formation and innervations [28]. Sonic hedgehog signaling blocks miR-206 expression, which results in increased BDNF protein expression. Bronchial epithelium is a major source of many key inflammatory and remodeling molecules [29–32]. These stimulated bronchial epithelial cells with TNF- α and IL-4 revealed that let-7, miR-29a, and miR-155 have been involved in the regulation of allergic inflammation [33].

MiR-133a negatively regulates RhoA in bronchial smooth muscle cells (BSMCs), a new target for asthma therapy [17]. Furthermore, downregulated miR-133a by IL-13 in the BSMCs causes an upregulation of RhoA, presumably resulting in an augmentation of bronchial smooth muscle contraction [34].

miRNAs appear to be attractive new drug targets. Th2driven airway inflammation, mucus hypersecretion, and AHR were shown effectively suppressed by delivery of an antagomir that inhibits miR-126 [13]. Recently, miR-106a was demonstrated to inhibit IL-10 in the posttranscriptional phase, which significantly alleviated most of the features of asthma. This represents the first in vivo proof of a miRNAmediated regulation of IL-10 with a potential to reverse an established asthmatic condition [35].

Glucocorticoids are used as mainstay therapy for asthma. In a murine asthma model, reported downregulation of miR-146a as an effect of dexamethasone, might partially explain its anti-inflammatory mechanism [36]. Antagonizing the function of miR-145 was as effective as glucocorticoid therapy in a trial treating mice treated with anti-miR-145 or dexamethasone and displayed significant reduction in the severity of the inflammatory lesions induced by HDM challenge [27]. The RhoA/Rhokinase pathway has now been proposed as a new target for the treatment of AHR in asthma [37, 38] and modulation of this pathway by miR-133a might provide a new insight into the treatment of AHR [17].

3. miRNAs and Diabetes Mellitus (DM)

Diabetes is one of the most common chronic diseases in the world and is recognized as one of the most important health threats of our time. DM is associated with serious morbidity and chronic disabling complications attributing to its high rate of mortality. Both type 1 (T1D) and type 2 diabetes mellitus (T2D) occur in children. T1D is a chronic autoimmune disease with an increasing incidence in the European pediatric population [39]. T2D, previously considered an adulthood disease, has now an increasing prevalence of early onset T2D secondary to the childhood obesity pandemic [40].

New approaches in investigating diabetes are essential for a deeper understanding of its pathogenesis and for developing novel therapeutic strategies. In recent years, miRNAs have become one of the most encouraging and fruitful fields in biological research and have been implicated as new players in the pathogenesis of diabetes and diabetesassociated complications.

The role of miRNAs in DM starts as early as the development of pancreatic islets. MiR-124a2 and miR-375 are involved in pancreatic beta-cell development [41, 42] and are necessary for proper formation of pancreatic islets in vertebrates. MiR-375 is necessary for the development

of β -cells in mice [42], establishment of normal pancreatic endocrine cell mass in the postnatal period, and maintenance of its viability [43]. Loss of miR-375 results in pancreatic cell defect and chronic hyperglycemia.

miRNAs have been further shown to regulate various physiological events relevant to DM pathophysiology, such as insulin biosynthesis, insulin secretion, insulin action, insulin responsiveness, and energy homeostasis.

miRNAs regulating insulin biosynthesis include miR-15a [44], miR-30d [45], miR-375, miR-122, miR-127-3p, and miR-184 [46]. MiR-15a increases insulin biosynthesis by targeting UCP-2 [44]. MiR-30d increases MafA expression, which promotes the transcription of the insulin gene in pancreatic β -cells [45]. MiR-375, miR-122, miR-127-3p, and miR-184 are suggested to play an important role in β -cell function insulin biosynthesis [46]. Suppression of human islet insulin biosynthesis by high glucose has been demonstrated to be induced by miR-133a decreasing polypyrimidine tract binding protein expression [47].

MiR-9 was found to play a critical role in the control of the secretory function of insulin-producing cells [48, 49].

MiR-375 is the highest expressed miRNA in pancreatic islets of humans and mice and regulates insulin secretion in isolated pancreatic cells [50]. Overexpression of miR-375 reduces insulin secretion through inhibition of exocytosis of insulin granules via translational repression of the cytoplasmic protein myotrophin [50]. Mice lacking miR-375 (375KO) are hyperglycemic and pancreatic β -cell mass is decreased due to impaired proliferation [43]. Li et al. (2010) showed also that miR-375 enhanced palmitate-induced lipo-apoptosis in insulin-secreting NIT-1 cells by repressing myotrophin (V1) protein expression [51]. Optimal insulin secretion in β -cells requires additional appropriate levels of miR-124a, miR-29 [41, 52], and miR-33a. MiR-33a was just recently shown to affect insulin secretion and acts through regulating its expression to correlate inversely with the expression of ABCA1 in pancreatic islets [53]. MiR-21, miR-34a, and miR-146 were shown to function as negative regulators of insulin signaling via inhibition of insulin secretion [54].

Recently, studies have shown the role of miRNAs in insulin sensitivity with emphasis on the importance of miR-103/107 [55]. The Lin28/let-7 pathway is a central regulator of mammalian glucose metabolism through interactions with the insulin-PI3 K-mTOR pathway and T2D-associated genes [56].

T1D, insulin dependent diabetes mellitus (IDDM), is a chronic autoimmune disorder caused by the interaction of environmental factors with an inherited predisposition. Twenty-seven miRNAs were mapped and located in 9 T1D susceptibility regions, rendering these miRNAs candidates for T1D susceptibility genes [57].

Regulatory T cells (Tregs) are known critical regulators of autoimmune diseases, including T1D. miRNA expression profiles in Tregs of T1D patients revealed a significant higher expression of miR-146a and lower expression miR-20b, miR-31, miR-99a, miR-100, miR-125b, miR-151, miR-335, and miR-365 [58]. These results support the hypothesis that changing expression in specific miRNAs can influence the function of Tregs and therefore the pathogenesis of T1D. cytokines. Cytokine-mediated β -cell dysfunction is suggested to be modulated by miR-29, which appeared to be dysregulated in this phase [59]. MiR-326 is expressed at higher levels in T1D subjects with ongoing islet autoimmunity [60]. miRNA array profiling in a T1D model identified eight miRNAs (miR-124, miR-128, miR-192, miR-194, miR-204, miR-375, miR-672, and miR-708) with differential expression that are likely involved in β -cell regulatory networks [61].

Dicer studies provide clear evidences for its role in the T1D pathogenesis. β -cells specific Dicer1 deletion results in aberrant pancreas development and neonatal death [62] and its inactivation leads to development of diabetes due to reduced insulin expression [63]. Targeted disruption of the Dicer1 gene specifically in β -cells leads to progressive reduction in insulin secretion and glucose tolerance and development of diabetes [64].

miRNAs are also emerging as highly tissue and/or cellspecific biomarkers of autoimmunity in T1D. The possibility of measuring miRNA in body fluids such as serum would help to easily recognize these particular markers [65].

T2D is a major health issue that has reached an epidemic status worldwide and is tightly linked to obesity. Obesity is characterized by intracellular accumulation of lipid in the pancreatic islets leading to β -cellular dysfunction and ultimately contributes to the pathogenesis of T2D [66, 67]. T2D is a progressive metabolic disorder characterized by reduced insulin sensitivity, insulin resistance and pancreatic β -cell dysfunction.

A growing body of direct evidence implicates the role miRNAs in T2D and most of its pathophysiological aspects. Recent experiments provide direct evidence that obesity induces overexpression of miR-143, which acts to inhibit insulin-stimulated AKT activation leading to impairment of glucose metabolism [68].

Subclinical inflammation and insulin resistance implicated in T2D patients are a result of impaired function of miR-146a and its downstream signals [69].

MiR-125a was found to be over-expressed in insulin target tissues in a spontaneous rat model of T2D [70]. MiR-125a is suggested to contribute to insulin resistance and play a critical role in insulin signaling [71] through affecting genes involved in the MAPK signaling pathway implicated in T2D [72].

Seven diabetes-related serum miRNAs: miR-9, miR-29a, miR-30d, miR-34a, miR-124a, miR-146a, and miR-375 [73], had been reported previously as key gene regulators involved in the regulation of insulin gene expression, insulin secretion [41, 43, 48], insulin signaling in target tissues [74], and free fatty acid (FFA) mediated β -cell dysfunction [75], all of which are closely related to the pathogenesis of T2D.

Deregulated miRNAs associated with T2D were identified as useful distinguishing serum biomarkers for different stages of diabetes progression and include miR-144, miR-146a, miR-150, miR-182, miR-192, miR-30d and miR-320. The expression profiles of these miRNAs can differentiate between impaired fasting glucose state (IFG) and welldeveloped T2D [76]. The first evidence that plasma miRNAs are deregulated in patients with DM was obtained from the observation that endothelial miR-126 was lost in type 2 diabetic patients [77].

Both T1D and T2D can lead to debilitating microvascular complications such as retinopathy, nephropathy, and neuropathy, as well as macrovascular complications.

A significant association between altered miRNA expression and the development and progression of the various diabetes complications has been recently reported. Several studies have demonstrated a role for miRNAs in diabetic nephropathy (DN) and was first demonstrated by Kato et al. in 2007. The authors found increased expression of miR-192 in glomeruli from mice with both type 1 and type 2 diabetes as well as in TGF- β treated cultured mesangial cells (MCs) [78]. TGF- β signaling events are crucial in regulating fibrotic effects in MCs and other renal cells through subtle molecular mechanisms that are yet not fully clear.

Of particular interest is a group of miRNAs including miR-200b/c, miR-216a, and miR-217, which were found to be upregulated in mouse renal mesangial cells (MMC) treated with TGF- β and in glomeruli of mouse models for diabetes [79–81]. These key miRNAs are highly expressed in the kidney and can act as effectors of TGF- β actions and high glucose in diabetic kidney disease.

Renal fibrosis is a component of DN and it was found that miR-377 induces fibronectin (ECM protein) expression in MCs via downregulation of manganese superoxide dismutase and p21 activated kinase indicating its role in pathogenesis of microvascular complications [82]. Specific reduction of renal miR-192 on the other hand decreases renal fibrosis and improves proteinuria, lending support for the possibility of an anti-miRNA-based translational approach to the treatment of DN [83].

Diabetic retinopathy (DR) is one of the leading causes of blindness. miRNAs are involved in the pathogenesis of DR through the modulation of multiple pathogenetic pathways and may be novel therapeutic targets for the treatment of DR [84–86].

Diabetic individuals are two to four times more likely to have vascular and heart disease compared to the normal population, and 75% of diabetes related deaths are due to heart diseases. Cardiac involvement in diabetes includes coronary atherosclerosis, diabetic cardiomyopathy, and autonomic neuropathy.

Accumulating evidence suggests that miRNAs are involved in the process of angiogenesis by modulating new vessel formation through their upregulation or downregulation [87, 88]. Among downregulated miRNAs in DM patients, miR-126, miR-27b, and miR-130a have been identified as proangiogenic miRNAs [89].

Tribble 2 (TRB2) plays important roles in the pathogenesis of T2D large artery complications at early stage and seems to be modulated by miR-98. Thus, targeting TRB2 and miR-98 could be considered as novel therapeutic strategies for T2D early large artery complication [90].

Caporali et al. have augmented our understanding of miRNA biology in vascular pathophysiology in diabetic patients through detecting the causal role of miR-503 in diabetes-induced impairment of endothelial function and reparative angiogenesis [91]. MiR-126 downregulation in endothelial progenitor cells (EPC) from diabetes patients leads to impairment in their functions via targeting gene Spred-1 [92].

Many miRNAs are promising to have a future role in the development of treatments of DM. Human embryonic stem (hES) cells have proven to possess unlimited selfrenewal and pluripotency and thus have the potential to provide an unlimited supply of different cell types for tissue replacement. Hence, hES cells are considered in the effort to find replacement for damaged islet β -cells especially T3 cells (T3pi).

Pancreatic islet-like cell clusters derived from T3 cells showed very high expression of miRNAs including miR-186, miR-199a, and miR-339, which downregulate the expression of LIN28, PRDM1, CALB1, GCNT2, RBM47, PLEKHH1, RBPMS2 and PAK6. Therefore, manipulation of these miR-NAs may be useful to increase the proportion of beta cells and insulin synthesis in the differentiated T3pi for cell therapy of TID [93].

A unique regulatory pathway of β -cell death involves miR-21. MiR-21 targets the tumor suppressor gene PDCD4 and its upstream transcriptional activator nuclear factor- κ B (NF- κ B); thus, targeting the miR-21–PDCD4 pathway may represent a unique strategy for treating autoimmune T1D [94].

As reported previously, miR-375 negatively regulates insulin secretion, and attenuation of miR-375 through the cAMP-PKA pathway may facilitate the insulin response in pancreatic β -cells [53].

Sirtuin-1 (SIRT1) is a potential therapeutic target to combat insulin resistance and T2D [95]. SIRT1 is regulated by miR-181a and improves hepatic insulin sensitivity. Inhibiting miR-181a might be a potential new strategy for treating insulin resistance and T2D [96].

Islet transplantation represents a potentially interesting strategy for T1D therapy. However, allogeneic islet grafts require immunosuppressive therapy to avoid rejection. Therefore, immune system modulation is necessary for functional stabilization of the transplantation. Adequate knowledge of the role of miRNAs in the regulation of immune function could result also in the possibility to design a novel immunosuppressive therapy for pancreatic islet transplantation.

4. miRNAs and Epilepsy

Epileptic disorders are serious chronic brain disorders among the most frequent neurologic problems that occur in childhood. Approximately 2% of the population is affected by epilepsy (lifetime prevalence), and in the majority (threefourths), the onset of epilepsy occurs in the pediatric age group. At least 12% of patients with childhood-onset epilepsy will have a period of intractability during long-term followup [97], for which epilepsy surgery has become an increasing treatment option [98]. Children with seizures are at increased risk for mental health impairments, developmental and physical comorbidities, increasing needs for care coordination, and specialized services [99]. Attention has been recently drawn to the role of miRNAs in pediatric CNS diseases [2], including epilepsy, shedding new light on the molecular mechanism promising novel therapeutic targets and effective antiepileptogenic medications.

Loss of Dicer in neurons or astrocytes results in miRNA downregulation, neuronal dysfunction, apoptosis, seizures, and cognitive deficits [100]. This observation was confirmed by a study showing reduced mature miRNAs levels in the human temporal lobe epilepsy (TLE) as a result of Dicer loss [101]. These findings suggest that loss of Dicer and failure of mature miRNA expression may be a feature of the pathophysiology of hippocampal sclerosis (HS) in patients with TLE and future efforts might be directed to determining whether restitution of Dicer to such tissue restores mature miRNA production and influences the epileptic phenotype.

Status epilepticus (SE) induces a cascade of molecular changes that contribute to the development of epilepsy. In the acute stage of mesial temporal lobe epilepsy (MTLE) development in rats, 19 miRNAs were up-regulated, amongst which miR-213, miR-132, miR-30c, miR-26a, and miR-375 were the most prominent upregulated miRNAs. Seven miR-NAs were downregulated including miR-29a and miR-181c [102]. Neurotrophin-3 (NT-3) is a neurotrophic factor that has been implicated in the development of epilepsy in several rodent models. MiR-21 was identified as a candidate for regulating neurotrophin-3 signaling in the hippocampus following SE suggesting that miR-21 downregulates NT-3 which is responsible for increased neuronal cell loss following SE [103]. MiR-21 is also upregulated in children with MTLE [104].

Deregulated miRNAs may be involved directly or indirectly in the pathogenesis in both the acute and chronic stages of MTLE. One hundred and twenty-five miRNAs have been identified in the hippocampus of lithium-pilocarpineinduced TLE and normal rats, including 23 miRNAs that were expressed differentially in the chronic stage of MTLE development. Five miRNAs were found downregulated and include miR-let-7e. Eighteen miRNAs were found upregulated and include miR-23 a/b [105].

The role of neuroinflammation is emerging as a key element in the pathogenesis of MTLE, the most common form of partial-onset epilepsies that usually begins in childhood. Aronica et al. were the first to report an altered expression pattern of miR-146a associated with inflammation in epileptic rats and TLE patients, adding a new insight to molecular mechanisms in proepileptogenic inflammatory signaling processes [106]. MiR-146a and interleukin-1 β (IL- 1β) are differently expressed in the various stages of MTLE development in an immature rat model and in children. The different expression pattern of both IL-1 β and miR-146a at various stages suggests an interactive relationship. Consequently, modulation of the IL-1 β -miR-146a axis may be a new target for antiepileptic therapy [107]. Furthermore, we just very recently found that miR-155 and tumor necrosis factor alpha (TNF- α) showed the same pattern of expressions in the three stages of MTLE development in immature rat model and are upregulated in children with MTLE. We found also a direct relationship between them on the astrocyte level [108].

A genome-wide miRNA profiling study revealed segregated miRNA signatures and deregulation of 165 miRNAs in MTLE patients. The immune response was most prominently targeted by the deregulated miR-221 and miR-222. These miRNAs regulate endogenous ICAM1 expression and were selectively coexpressed with ICAM1 in astrocytes in MTLE patients, which suggest that miRNA changes in MTLE patients affect the expression of immunomodulatory proteins facilitating the immune response [109].

Increasing evidences highlight the role of synaptic plasticity in the development of MTLE [110, 111]. Recently miRNAs have been proposed to target neuronal mRNAs localized near the synapse, exerting a pivotal role in modulating local protein synthesis, and presumably affecting adaptive mechanisms such as synaptic plasticity [112, 113]. Using an in vivo model for increasing excitatory activity in the cortex and the hippocampus indicates that the distribution of some miRNAs can be modulated by enhanced neuronal (epileptogenic) activity.

The dynamic modulation in the local distribution of miRNAs seems to play key roles in controlling localized protein synthesis at the synapse [114]. Pilocarpine-induced seizures led to a robust, rapid, and transient increase in the primary transcript of miR-132 (pri-miR-132) followed by a subsequent rise in mature miR-132 indicating that miR-132 is an activity-dependent in vivo, and may contribute to the long-lasting proteomic changes required for neuronal plasticity [115].

Taking a step in using miRNAa as blood biomarkers for epilepsy, Liu et al. described a unique expression of blood miRNAs 24 hours after induction of kainate seizures [116]. Also Hu et al. demonstrated a possible correlation between hippocampal and peripheral blood miRNAs in post-SE rats, through detecting similar expression patterns in miR-34a, miR-22, and miR-125a (upregulated) while miR-21 had decreased [102].

Very recently, in vivo microinjection of locked nucleic acid-modified oligonucleotides depleted hippocampal miR-132 levels and reduced seizure-induced neuronal death, thus strongly suggesting that miRNAs are important regulators of seizure-induced neuronal death [117]. We found in our study that brain-specific miR-124 and miR-134 were upregulated in the seizure related stages of MTLE in immature rat model and children with MTLE, suggesting that downregulation of these miRNAs may have anti-convulsive effects [104]. It was demonstrated additionally that silencing miR-134 exerts prolonged seizure-suppressant and neuroprotective actions giving promising hope for miRNAs to be useful as potential therapeutic target for epilepsy treatment [118]. Whether antimiRNAs could function as anticonvulsants or would be true antiepileptogenic requires more experimental work.

5. miRNAs and Cystic Fibrosis

Cystic fibrosis (CF) is the most common lethal genetic disease in the Caucasian populations and occurs in approximately 1 in 2500 births [119]. It is caused by mutations in cystic fibrosis transmembrane conductance regulator (CFTR) gene. The most frequent mutation is deletion of a phenylalanine residue at position 508 (Δ F508).

The life expectancy of patients with CF has dramatically increased over the past decades [120], and the median survival of patients born in 2000 is expected to be above 50 years [121]. Despite significant advances in treatment regimes, CF remains a condition for which no effective cure exists and still has a mortality rate of >90% as a result of respiratory failure [122].

Investigating the expression and function of miRNAs in CF will shed light on previously unidentified regulatory mechanisms and further direct the development of future therapeutic strategies.

Emerging evidence suggests that changes in miRNAs expression are associated with CF [123–126]. It is hypothesized that unique miRNA expression profiles exist in CF versus non-CF bronchial epithelial cells and that these differential molecular miRNA signatures can regulate proinflammatory gene expression [124].

To date, several groups have examined the potential role of miRNAs in molecular pathways involved in the pathogenesis of CF, especially lung inflammation [127, 128]. MiR-155 is suggested playing an important role in the activation of IL-8dependent inflammation in CF [126].

Several studies demonstrate that miRNAs regulate expression of the CFTR gene post transcriptionally. MiR-138 was discovered to regulate CFTR expression through its interaction with the transcriptional regulatory protein SIN3A. Treating airway epithelia with an miR-138 mimic indeed increased CFTR mRNA and enhanced CFTR abundance and transepithelial Cl (–) permeability independent of elevated mRNA levels. Anti-miR-138 had the opposite effects [129].

A role of miRNAs in targeting CFTR has been supported. hsa-miR-384, hsa-miR-494 and hsa-miR-1246 are involved in the post-transcriptional regulation of the CFTR channel synthesis. In individuals carrying the DF508 CFTR mutation, increased expression of miR-145, miR-223, and miR-494 in bronchial epithelium showed correlation with decreased CFTR expression [130].

Furthermore, miR-101 and miR-494 seem to act synergistically on CFTR-reporter inhibition with a more than additive effect on the post-translational control, which could have a physiological relevance in the complex disease phenotypes observed in CF [131].

The hallmark of CF lung disease is chronic infection by *Pseudomonas aeruginosa* that gradually increases from childhood through early adolescence. Rao et al. detected miRNAs in *P. aeruginosa* infected sputum of CF patients. A significant change in miR-146 expression in these patients was associated with the Toll-like receptor family, a family which includes the primary evolutionarily conserved sensors of pathogen-associated molecular patterns and is known to trigger host inflammatory and immune responses [132].

CF affects epithelial organs including the intestine, where both meconium ileus and distal intestinal obstruction syndrome can occur as complications. Bazett et al. [125] investigated whether miRNAs contribute to the different phenotypic changes observed in the CF intestine by initially measuring the miRNA signature of this tissue with an array. They concluded that altered miRNA expression is a feature that putatively influences both metabolic abnormalities and the altered tissue homeostasis component of CF intestinal disease [122].

The fact that a miRNA-regulated network directs gene expression from chromosome to cell membrane indicates that one individual miRNA can control a cellular process more broadly than recognized previously. This discovery will provide therapeutic avenues for restoring CFTR function to cells affected by the most common cystic fibrosis mutation and mandates miRNA-based research in this field [129].

6. Conclusion

Despite the inherent limitations, much progress has been made towards developing effective treatments for pediatric chronic diseases, offering hope for millions of children with these disorders. The role of miRNAs in the pathogenesis of these diseases makes them promising targets worth studying if our goal is to secure normal growth and development. Research efforts directed towards a greater understanding of the mechanisms and functional significance of the aberrant expression of miRNAs in these major chronic non-neoplastic diseases will assist in the development of less toxic therapies and provide better markers for disease classification. We believe that the discovery of miRNAs will open new research avenues for pediatric chronic diseases, which are expected to advance this area of research from its infancy to the mature stages.

Conflict of Interests

The authors declare that they have no conflict of interests.

References

- A. J. Janse, C. S. Uiterwaal, R. J. Gemke, J. L. Kimpen, and G. Sinnema, "A difference in perception of quality of life in chronically ill children was found between parents and pediatricians," *Journal of Clinical Epidemiology*, vol. 58, no. 5, pp. 495–502, 2005.
- [2] A. Omran, D. Elimam, S. Shalaby, J. Peng, and F. Yin, "MicroR-NAs: a light into the "Black Box" of neuropediatric diseases," *Neuromolecular Medicine*, vol. 14, no. 4, pp. 244–261, 2012.
- [3] A. Omran, D. Elimam, K. Webster, L. Shehadeh, and F. Yin, "MicroRNAs: a new piece in the paediatric cardiovascular disease puzzle," *Cardiology in the Young*, pp. 1–14, 2013.
- [4] Y. Bossé, P. D. Paré, and C. Y. Seow, "Airway wall remodeling in asthma: from the epithelial layer to the adventitia," *Current Allergy and Asthma Reports*, vol. 8, no. 4, pp. 357–366, 2008.
- [5] A. M. Vignola, F. Mirabella, G. Costanzo et al., "Airway remodeling in asthma," *Chest*, vol. 123, supplement 3, pp. 417S– 422S, 2003.
- [6] G. P. Anderson, "Endotyping asthma: new insights into key pathogenic mechanisms in a complex, heterogeneous disease," *The Lancet*, vol. 372, no. 9643, pp. 1107–1119, 2008.
- [7] L. J. Akinbami, J. E. Moorman, P. L. Garbe, and E. J. Sondik, "Status of childhood asthma in the United States, 1980–2007," *Pediatrics*, vol. 123, no. 3, pp. S131–S145, 2009.

- [8] C. F. Kelley, D. M. Mannino, D. M. Homa, A. Savage-Brown, and F. Holguin, "Asthma phenotypes, risk factors, and measures of severity in a national sample of US children," *Pediatrics*, vol. 115, no. 3, pp. 726–731, 2005.
- [9] Z. Tan, G. Randall, J. Fan et al., "Allele-specific targeting of microRNAs to HLA-G and risk of asthma," *American Journal* of Human Genetics, vol. 81, no. 4, pp. 829–834, 2007.
- [10] X.-W. Su, Y. Yang, M.-L. Lv et al., "Association between singlenucleotide polymorphisms in pre-mirnas and the risk of asthma in a Chinese population," *DNA and Cell Biology*, vol. 30, no. 11, pp. 919–923, 2011.
- [11] Y. Y. Zhang, M. Zhong, M. Y. Zhang, and K. Lv, "Expression and clinical significance of miR-155 in peripheral blood CD4⁺; T cells of patients with allergic asthma," *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*, vol. 28, no. 5, pp. 540–543, 2012.
- [12] T. X. Lu, A. Munitz, and M. E. Rothenberg, "MicroRNA-21 is up-regulated in allergic airway inflammation and regulates IL-12p35 expression," *Journal of Immunology*, vol. 182, no. 8, pp. 4994–5002, 2009.
- [13] J. Mattes, A. Collison, M. Plank, S. Phipps, and P. S. Foster, "Antagonism of microRNA-126 suppresses the effector function of T H2 cells and the development of allergic airways disease," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 44, pp. 18704–18709, 2009.
- [14] A. E. Williams, H. Larner-Svensson, M. M. Perry et al., "MicroRNA expression profiling in mild asthmatic human airways and effect of corticosteroid therapy," *PLoS ONE*, vol. 4, no. 6, article e5889, 2009.
- [15] S. Polikepahad, J. M. Knight, A. O. Naghavi et al., "Proinflammatory role for let-7 microRNAS in experimental asthma," *Journal of Biological Chemistry*, vol. 285, no. 39, pp. 30139–30149, 2010.
- [16] A. Rodriguez, E. Vigorito, S. Clare et al., "Requirement of bic/microRNA-155 for normal immune function," *Science*, vol. 316, no. 5824, pp. 608–611, 2007.
- [17] Y. Chiba, M. Tanabe, K. Goto, H. Sakai, and M. Misawa, "Downregulation of miR-133a contributes to up-regulation of RhoA in bronchial smooth muscle cells," *American Journal of Respiratory* and Critical Care Medicine, vol. 180, no. 8, pp. 713–719, 2009.
- [18] M. Kumar, U. Mabalirajan, A. Agrawal, and B. Ghosh, "Proinflammatory role of let-7 miRNAs in experimental asthma?" *Journal of Biological Chemistry*, vol. 285, no. 48, p. le20, 2010.
- [19] N. Garbacki, E. di Valentin, V. A. Huynh-Thu et al., "MicroRNAs profiling in murine models of acute and chronic asthma: a relationship with mRNAs targets," *PLoS ONE*, vol. 6, no. 1, article e16509, 2011.
- [20] R. J. Mayoral, M. E. Pipkin, M. Pachkov, E. van Nimwegen, A. Rao, and S. Monticelli, "MicroRNA-221-222 regulate the cell cycle in mast cells," *Journal of Immunology*, vol. 182, no. 1, pp. 433–445, 2009.
- [21] R. J. Mayoral, L. Deho, N. Rusca et al., "MiR-221 influences effector functions and actin cytoskeleton in mast cells," *PLoS ONE*, vol. 6, no. 10, article e26133, 2011.
- [22] G. M. Walsh, "Targeting eosinophils in asthma: current and future state of cytokine-and chemokine-directed monoclonal therapy," *Expert Review of Clinical Immunology*, vol. 6, no. 5, pp. 701–704, 2010.
- [23] H. Y. Kim, R. H. Dekruyff, and D. T. Umetsu, "The many paths to asthma:phenotype shaped by innate and adaptive immunity," *Nature Immunology*, vol. 11, no. 7, pp. 577–584, 2010.

- [24] J. T. Schroeder, A. P. Bieneman, K. L. Chichester, L. Breslin, H. Xiao, and M. C. Liu, "Pulmonary allergic responses augment interleukin-13 secretion by circulating basophils yet suppress interferon-α from plasmacytoid dendritic cells," *Clinical and Experimental Allergy*, vol. 40, no. 5, pp. 745–754, 2010.
- [25] X. Liu, A. Nelson, X. Wang et al., "MicroRNA-146a modulates human bronchial epithelial cell survival in response to the cytokine-induced apoptosis," *Biochemical and Biophysical Research Communications*, vol. 380, no. 1, pp. 177–182, 2009.
- [26] M. Kumar, T. Ahmad, A. Sharma et al., "Let-7 microRNAmediated regulation of IL-13 and allergic airway inflammation," *Journal of Allergy and Clinical Immunology*, vol. 128, no. 5, pp. 1077.e10–1085.e10, 2011.
- [27] A. Collison, J. Mattes, M. Plank, and P. S. Foster, "Inhibition of house dust mite-induced allergic airways disease by antagonism of microRNA-145 is comparable to glucocorticoid treatment," *Journal of Allergy and Clinical Immunology*, vol. 128, no. 1, pp. 160–167, 2011.
- [28] K. Radzikinas, L. Aven, Z. Jiang et al., "A Shh/miR-206/BDNF cascade coordinates innervation and formation of airway smooth muscle," *Journal of Neuroscience*, vol. 31, no. 43, pp. 15407–15415, 2011.
- [29] P. J. Barnes, "Immunology of asthma and chronic obstructive pulmonary disease," *Nature Reviews Immunology*, vol. 8, no. 3, pp. 183–192, 2008.
- [30] H. Hammad and B. N. Lambrecht, "Dendritic cells and epithelial cells: linking innate and adaptive immunity in asthma," *Nature Reviews Immunology*, vol. 8, no. 3, pp. 193–204, 2008.
- [31] S. T. Holgate, "The epithelium takes centre stage in asthma and atopic dermatitis," *Trends in Immunology*, vol. 28, no. 6, pp. 248– 251, 2007.
- [32] R. P. Schleimer, A. Kato, R. Kern, D. Kuperman, and P. C. Avila, "Epithelium: at the interface of innate and adaptive immune responses," *Journal of Allergy and Clinical Immunology*, vol. 120, no. 6, pp. 1279–1284, 2007.
- [33] Y. Zhai, Z. Zhong, C.-Y. A. Chen et al., "Coordinated changes in mRNA turnover, translation, and RNA processing bodies in bronchial epithelial cells following inflammatory stimulation," *Molecular and Cellular Biology*, vol. 28, no. 24, pp. 7414–7426, 2008.
- [34] Y. Chiba and M. Misawa, "MicroRNAs and their therapeutic potential for human diseases: MiR-133a and bronchial smooth muscle hyperresponsiveness in asthma," *Journal of Pharmacological Sciences*, vol. 114, no. 3, pp. 264–268, 2010.
- [35] A. Sharma, M. Kumar, T. Ahmad et al., "Antagonism of mmumir-106a attenuates asthma features in allergic murine model," *Journal of Applied Physiology*, vol. 113, no. 3, pp. 459–464, 2012.
- [36] M. J. Feng, F. Shi, C. Qiu, and W. K. Peng, "MicroRNA-181a, -146a and -146b in spleen CD4⁺ T lymphocytes play proinflammatory roles in a murine model of asthma," *International Immunopharmacology*, vol. 13, no. 3, pp. 347–353, 2012.
- [37] D. Schaafsma, R. Gosens, J. Zaagsma, A. J. Halayko, and H. Meurs, "Rho kinase inhibitors: a novel therapeutical intervention in asthma?" *European Journal of Pharmacology*, vol. 585, no. 2-3, pp. 398–406, 2008.
- [38] H. Kume, "RhoA/Rho-kinase as a therapeutic target in asthma," *Current Medicinal Chemistry*, vol. 15, no. 27, pp. 2876–2885, 2008.
- [39] C. C. Patterson, G. G. Dahlquist, E. Gyürüs, A. Green, G. Soltész, and EURODIAB Study Group, "Incidence trends for

childhood type 1 diabetes in Europe during 1989–2003 and predicted new cases 2005–20: a multicentre prospective registration study," *The Lancet*, vol. 373, no. 9680, pp. 2027–2033, 2009.

- [40] G. Danaei, M. M. Finucane, Y. Lu et al., "National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2.7 million participants," *The Lancet*, vol. 378, no. 9785, pp. 31–40, 2011.
- [41] N. Baroukh, M. A. Ravier, M. K. Loder et al., "MicroRNA-124a regulates foxa2 expression and intracellular signaling in pancreatic β-cell lines," *Journal of Biological Chemistry*, vol. 282, no. 27, pp. 19575–19588, 2007.
- [42] D. M. Keller, E. A. Clark, and R. H. Goodman, "Regulation of microRNA-375 by cAMP in pancreatic β-cells," *Molecular Endocrinology*, vol. 26, no. 6, pp. 989–999, 2012.
- [43] M. N. Poy, J. Hausser, M. Trajkovski et al., "miR-375 maintains normal pancreatic α- and β-cell mass," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 14, pp. 5813–5818, 2009.
- [44] L.-L. Sun, B.-G. Jiang, W.-T. Li, J.-J. Zou, Y.-Q. Shi, and Z.-M. Liu, "MicroRNA-15a positively regulates insulin synthesis by inhibiting uncoupling protein-2 expression," *Diabetes Research* and Clinical Practice, vol. 91, no. 1, pp. 94–100, 2011.
- [45] X. Zhao, R. Mohan, and X. Tang, "MicroRNA-30d induces insulin transcription factor MafA and insulin production by targeting mitogen-activated protein 4 kinase 4 (Map4k4) in pancreatic β cells," *Journal of Biological Chemistry*, vol. 287, no. 37, pp. 31155–31164, 2012.
- [46] C. Bolmeson, J. L. S. Esguerra, A. Salehi, D. Speidel, L. Eliasson, and C. M. Cilio, "Differences in islet-enriched miRNAs in healthy and glucose intolerant human subjects," *Biochemical* and Biophysical Research Communications, vol. 404, no. 1, pp. 16–22, 2011.
- [47] R. G. Fred, C. H. Bang-Berthelsen, T. Mandrup-Poulsen, L. G. Grunnet, and N. Welsh, "High glucose suppresses human islet insulin biosynthesis by inducing mir-133a leading to decreased polypyrimidine tract binding protein-expression," *PLoS ONE*, vol. 5, no. 5, article e10843, 2010.
- [48] V. Plaisance, A. Abderrahmani, V. Perret-Menoud, P. Jacquemin, F. Lemaigre, and R. Regazzi, "MicroRNA-9 controls the expression of Granuphilin/Slp4 and the secretory response of insulin-producing cells," *Journal of Biological Chemistry*, vol. 281, no. 37, pp. 26932–26942, 2006.
- [49] D. Ramachandran, U. Roy, S. Garg, S. Ghosh, S. Pathak, and U. Kolthur-Seetharam, "Sirt1 and mir-9 expression is regulated during glucose-stimulated insulin secretion in pancreatic βislets," *FEBS Journal*, vol. 278, no. 7, pp. 1167–1174, 2011.
- [50] M. N. Poy, L. Eliasson, J. Krutzfeldt et al., "A pancreatic isletspecific microRNA regulates insulin secretion," *Nature*, vol. 432, no. 7014, pp. 226–230, 2004.
- [51] Y. Li, X. Xu, Y. Liang et al., "miR-375 enhances palmitateinduced lipoapoptosis in insulin-secreting NIT-1 cells by repressing myotrophin (V1) protein expression," *International Journal of Clinical and Experimental Pathology*, vol. 3, no. 3, pp. 254–264, 2010.
- [52] T. J. Pullen, G. da Silva Xavier, G. Kelsey, and G. A. Rutter, "miR-29a and miR-29b contribute to pancreatic β-cell-specific silencing of monocarboxylate transporter 1 (MCT1)," *Molecular and Cellular Biology*, vol. 31, no. 15, pp. 3182–3194, 2011.

- [53] N. Wijesekara, L.-H. Zhang, M. H. Kang et al., "miR-33a modulates ABCA1 expression, cholesterol accumulation, and insulin secretion in pancreatic islets," *Diabetes*, vol. 61, no. 3, pp. 653–658, 2012.
- [54] E. Roggli, A. Britan, S. Gattesco et al., "Involvement of microRNAs in the cytotoxic effects exerted by proinflammatory cytokines on pancreatic β-cells," *Diabetes*, vol. 59, no. 4, pp. 978– 986, 2010.
- [55] M. Trajkovski, J. Hausser, J. Soutschek et al., "MicroRNAs 103 and 107 regulate insulin sensitivity," *Nature*, vol. 474, no. 7353, pp. 649–653, 2011.
- [56] H. Zhu, N. Shyh-Chang, A. V. Segr et al., "The Lin28/let-7 axis regulates glucose metabolism," *Cell*, vol. 147, no. 1, pp. 81–94, 2011.
- [57] L. Zhou, H. He, J. X. Mi, C. Li, B. Lee, and Q.-S. Mi, "MicroRNA genes: are they susceptibility candidates for human type 1 diabetes?" *Annals of the New York Academy of Sciences*, vol. 1150, pp. 72–75, 2008.
- [58] R. Hezova, O. Slaby, P. Faltejskova et al., "microRNA-342, microRNA-191 and microRNA-510 are differentially expressed in T regulatory cells of type 1 diabetic patients," *Cellular Immunology*, vol. 260, no. 2, pp. 70–74, 2010.
- [59] E. Roggli, S. Gattesco, D. Caille et al., "Changes in microrna expression contribute to pancreatic β-cell dysfunction in prediabetic nod mice," *Diabetes*, vol. 61, no. 7, pp. 1742–1751, 2012.
- [60] G. Sebastiani, F. A. Grieco, I. Spagnuolo, L. Galleri, D. Cataldo, and F. Dotta, "Increased expression of microRNA miR-326 in type 1 diabetic patients with ongoing islet autoimmunity," *Diabetes/Metabolism Research and Reviews*, vol. 27, no. 8, pp. 862–866, 2011.
- [61] C. H. Bang-Berthelsen, L. Pedersen, T. Fløyel, P. H. Hagedorn, T. Gylvin, and F. Pociot, "Independent component and pathwaybased analysis of miRNA-regulated gene expression in a model of type 1 diabetes," *BMC Genomics*, vol. 12, article 97, 2011.
- [62] F. C. Lynn, P. Skewes-Cox, Y. Kosaka, M. T. McManus, B. D. Harfe, and M. S. German, "MicroRNA expression is required for pancreatic islet cell genesis in the mouse," *Diabetes*, vol. 56, no. 12, pp. 2938–2945, 2007.
- [63] T. Melkman-Zehavi, R. Oren, S. Kredo-Russo et al., "miRNAs control insulin content in pancreatic β-cells via downregulation of transcriptional repressors," *EMBO Journal*, vol. 30, no. 5, pp. 835–845, 2011.
- [64] M. Kalis, C. Bolmeson, J. L. S. Esguerra et al., "Beta-cell specific deletion of dicer1 leads to defective insulin secretion and diabetes mellitus," *PLoS ONE*, vol. 6, no. 12, article e29166, 2011.
- [65] S. Gilad, E. Meiri, Y. Yogev et al., "Serum microRNAs are promising novel biomarkers," *PLoS ONE*, vol. 3, no. 9, article e3148, 2008.
- [66] J. D. Johnson, "Proteomic identification of carboxypeptidase E connects lipid-induced β-cell apoptosis and dysfunction in type 2 diabetes," *Cell Cycle*, vol. 8, no. 1, pp. 38–42, 2009.
- [67] K. S. Gwiazda, T.-L. B. Yang, Y. Lin, and J. D. Johnson, "Effects of palmitate on ER and cytosolic Ca²⁺ homeostasis in β-cells," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 296, no. 4, pp. E690–E701, 2009.
- [68] S. D. Jordan, M. Krüger, D. M. Willmes et al., "Obesity-induced overexpression of miRNA-143 inhibits insulin-stimulated AKT activation and impairs glucose metabolism," *Nature Cell Biol*ogy, vol. 13, no. 4, pp. 434–448, 2011.

- [69] M. Balasubramanyam, S. Aravind, K. Gokulakrishnan et al., "Impaired miR-146a expression links subclinical inflammation and insulin resistance in Type 2 diabetes," *Molecular and Cellular Biochemistry*, vol. 351, no. 1-2, pp. 197–205, 2011.
- [70] B. M. Herrera, H. E. Lockstone, J. M. Taylor et al., "MicroRNA-125a is over-expressed in insulin target tissues in a spontaneous rat model of Type 2 Diabetes," *BMC Medical Genomics*, vol. 2, article no. 54, 2009.
- [71] M. Fujishiro, Y. Gotoh, H. Katagiri et al., "Three mitogenactivated protein kinases inhibit insulin signaling by different mechanisms in 3T3-L1 adipocytes," *Molecular Endocrinology*, vol. 17, no. 3, pp. 487–497, 2003.
- [72] J. A. Engelman, A. H. Berg, R. Y. Lewis, M. P. Lisanti, and P. E. Scherer, "Tumor necrosis factor α-mediated insulin resistance, but not dedifferentiation, is abrogated by MEK1/2 inhibitors in 3T3-L1 adipocytes," *Molecular Endocrinology*, vol. 14, no. 10, pp. 1557–1569, 2000.
- [73] L. Kong, J. Zhu, W. Han et al., "Significance of serum microR-NAs in pre-diabetes and newly diagnosed type 2 diabetes: A Clinical Study," *Acta Diabetologica*, vol. 48, no. 1, pp. 61–69, 2011.
- [74] A. He, L. Zhu, N. Gupta, Y. Chang, and F. Fang, "Overexpression of micro ribonucleic acid 29, highly up-regulated in diabetic rats, leads to insulin resistance in 3T3-L1 adipocytes," *Molecular Endocrinology*, vol. 21, no. 11, pp. 2785–2794, 2007.
- [75] P. Lovis, E. Roggli, D. R. Laybutt et al., "Alterations in MicroRNA expression contribute to fatty Acid-Induced pancreatic β-Cell dysfunction," *Diabetes*, vol. 57, no. 10, pp. 2728–2736, 2008.
- [76] D. S. Karolina, A. Armugam, S. Tavintharan et al., "MicroRNA 144 impairs insulin signaling by inhibiting the expression of insulin receptor substrate 1 in type 2 diabetes mellitus," *PLoS ONE*, vol. 6, no. 8, article e22839, 2011.
- [77] A. Zampetaki, S. Kiechl, I. Drozdov et al., "Plasma microRNA profiling reveals loss of endothelial miR-126 and other MicroR-NAs in type 2 diabetes," *Circulation Research*, vol. 107, no. 6, pp. 810–817, 2010.
- [78] M. Kato, J. Zhang, M. Wang et al., "MicroRNA-192 in diabetic kidney glomeruli and its function in TGF-β-induced collagen expression via inhibition of E-box repressors," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 9, pp. 3432–3437, 2007.
- [79] M. Kato, L. Wang, S. Putta et al., "Post-transcriptional upregulation of Tsc-22 by Ybx1, a target of miR-216a, mediates TGF-β-induced collagen expression in kidney cells," *Journal of Biological Chemistry*, vol. 285, no. 44, pp. 34004–34015, 2010.
- [80] M. Kato, L. Arce, M. Wang, S. Putta, L. Lanting, and R. Natarajan, "A microRNA circuit mediates transforming growth factor-β1 autoregulation in renal glomerular mesangial cells," *Kidney International*, vol. 80, no. 4, pp. 358–368, 2011.
- [81] M. Kato, S. Putta, M. Wang et al., "TGF-β activates Akt kinase through a microRNA-dependent amplifying circuit targeting PTEN," *Nature Cell Biology*, vol. 11, no. 7, pp. 881–889, 2009.
- [82] Q. Wang, Y. Wang, A. W. Minto et al., "MicroRNA-377 is upregulated and can lead to increased fibronectin production in diabetic nephropathy," *FASEB Journal*, vol. 22, no. 12, pp. 4126– 4135, 2008.
- [83] S. Putta, L. Lanting, G. Sun, G. Lawson, M. Kato, and R. Natarajan, "Inhibiting microRNA-192 ameliorates renal fibrosis in diabetic nephropathy," *Journal of the American Society of Nephrology*, vol. 23, no. 3, pp. 458–469, 2012.
- [84] B. Kovacs, S. Lumayag, C. Cowan, and S. Xu, "MicroRNAs in early diabetic retinopathy in streptozotocin-induced diabetic

rats," *Investigative Ophthalmology & Visual Science*, vol. 52, no. 7, pp. 4402–4409, 2011.

- [85] V. A. O. Silva, A. Polesskaya, T. A. Sousa et al., "Expression and cellular localization of microRNA-29b and RAX, an activator of the RNA-dependent protein kinase (PKR), in the retina of streptozotocin-induced diabetic rats," *Molecular Vision*, vol. 17, pp. 2228–2240, 2011.
- [86] J.-H. Wu, Y. Gao, A.-J. Ren et al., "Altered microRNA expression profiles in retinas with diabetic retinopathy," *Ophthalmic Research*, vol. 47, no. 4, pp. 195–201, 2012.
- [87] H. Hermeking, "The miR-34 family in cancer and apoptosis," *Cell Death and Differentiation*, vol. 17, no. 2, pp. 193–199, 2010.
- [88] Y. Suárez and W. C. Sessa, "MicroRNAs as novel regulators of angiogenesis," *Circulation Research*, vol. 104, no. 4, pp. 442–454, 2009.
- [89] C. Urbich, A. Kuehbacher, and S. Dimmeler, "Role of microR-NAs in vascular diseases, inflammation, and angiogenesis," *Cardiovascular Research*, vol. 79, no. 4, pp. 581–588, 2008.
- [90] S. Xie, N. Xie, Y. Li et al., "Upregulation of TRB2 induced by miR-98 in the early lesions of large artery of type-2 diabetic rat," *Molecular and Cellular Biochemistry*, vol. 361, no. 1-2, pp. 305– 314, 2012.
- [91] A. Caporali, M. Meloni, C. Völlenkle et al., "Deregulation of microRNA-503 contributes to diabetes mellitus-induced impairment of endothelial function and reparative angiogenesis after Limb Ischemia," *Circulation*, vol. 123, no. 3, pp. 282–291, 2011.
- [92] S. Meng, J. T. Cao, B. Zhang, Q. Zhou, C. X. Shen, and C. Q. Wang, "Downregulation of microRNA-126 in endothelial progenitor cells from diabetes patients, impairs their functional properties, via target gene Spred-1," *Journal of Molecular and Cellular Cardiology*, vol. 53, no. 1, pp. 64–72, 2012.
- [93] B.-Z. Chen, S.-L. Yu, S. Singh et al., "Identification of microR-NAs expressed highly in pancreatic islet-like cell clusters differentiated from human embryonic stem cells," *Cell Biology International*, vol. 35, no. 1, pp. 29–37, 2011.
- [94] Q. Ruan, T. Wang, V. Kameswaran et al., "The microRNA-21-PDCD4 axis prevents type 1 diabetes by blocking pancreatic β cell death," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 29, pp. 12030–12035, 2011.
- [95] F. Liang, S. Kume, and D. Koya, "SIRT1 and insulin resistance," *Nature Reviews Endocrinology*, vol. 5, no. 7, pp. 367–373, 2009.
- [96] B. Zhou, C. Li, W. Qi et al., "Downregulation of miR-181a upregulates sirtuin-1 (SIRT1) and improves hepatic insulin sensitivity," *Diabetologia*, vol. 55, no. 7, pp. 2032–2043, 2012.
- [97] A. Geerts, O. Brouwer, H. Stroink et al., "Onset of intractability and its course over time: The Dutch Study of Epilepsy in Childhood," *Epilepsia*, vol. 53, no. 4, pp. 741–751, 2012.
- [98] M. S. Perry and M. Duchowny, "Surgical management of intractable childhood epilepsy: curative and palliative procedures," *Seminars in Pediatric Neurology*, vol. 18, no. 3, pp. 195– 202, 2011.
- [99] S. A. Russ, K. Larson, and N. Halfon, "A national profile of childhood epilepsy and seizure disorder," *Pediatrics*, vol. 129, no. 2, pp. 256–264, 2012.
- [100] J. Tao, H. Wu, Q. Lin et al., "Deletion of astroglial dicer causes non-cell autonomous neuronal dysfunction and degeneration," *Journal of Neuroscience*, vol. 31, no. 22, pp. 8306–8319, 2011.

- [101] R. C. McKiernan, E. M. Jimenez-Mateos, I. Bray et al., "Reduced mature microRNA levels in association with dicer loss in human temporal lobe epilepsy with hippocampal sclerosis," *PLoS ONE*, vol. 7, no. 5, article e35921, 2012.
- [102] K. Hu, C. Zhang, L. Long et al., "Expression profile of microRNAs in rat hippocampus following lithium-pilocarpineinduced status epilepticus," *Neuroscience Letters*, vol. 488, no. 3, pp. 252–257, 2011.
- [103] R. M. Risbud, C. Lee, and B. E. Porter, "Neurotrophin-3 mRNA a putative target of miR21 following status epilepticus," *Brain Research*, vol. 1424, pp. 53–59, 2011.
- [104] J. Peng, A. Omran, M. U. Ashhab et al., "Expression patterns of miR-124, miR-134, miR-132, and miR-21 in an immature rat model and children with mesial temporal lobe epilepsy," *Journal* of *Molecular Neuroscience*, vol. 50, no. 2, pp. 291–297, 2013.
- [105] Y.-J. Song, X.-B. Tian, S. Zhang et al., "Temporal lobe epilepsy induces differential expression of hippocampal miRNAs including let-7e and miR-23a/b," *Brain Research*, vol. 1387, pp. 134–140, 2011.
- [106] E. Aronica, K. Fluiter, A. Iyer et al., "Expression pattern of miR-146a, an inflammation-associated microRNA, in experimental and human temporal lobe epilepsy," *European Journal of Neuroscience*, vol. 31, no. 6, pp. 1100–1107, 2010.
- [107] A. Omran, J. Peng, C. Zhang et al., "Interleukin-1β and microRNA-146a in an immature rat model and children with mesial temporal lobe epilepsy," *Epilepsia*, vol. 53, no. 7, pp. 1215– 1224, 2012.
- [108] M. U. Ashhab, A. Omran, H. Kong et al., "Expressions of tumor necrosis factor-alpha and microrna-155 in immature rat model of status epilepticus and children with mesial temporal lobe epilepsy," *Journal of Molecular Neuroscience*, 2013.
- [109] A. A. Kan, S. van Erp, A. A. H. A. Derijck et al., "Genome-wide microRNA profiling of human temporal lobe epilepsy identifies modulators of the immune response," *Cellular and Molecular Life Sciences*, vol. 69, no. 18, pp. 3127–3145, 2012.
- [110] A. Brooks-Kayal, "Molecular mechanisms of cognitive and behavioral comorbidities of epilepsy in children," *Epilepsia*, vol. 52, no. 1, pp. 13–20, 2011.
- [111] L. Wu, J. Peng, C. Wei et al., "Characterization, using comparative proteomics, of differentially expressed proteins in the hippocampus of the mesial temporal lobe of epileptic rats following treatment with valproate," *Amino Acids*, vol. 40, no. 1, pp. 221–238, 2011.
- [112] S. I. Ashraf, A. L. McLoon, S. M. Sclarsic, and S. Kunes, "Synaptic protein synthesis associated with memory is regulated by the RISC pathway in Drosophila," *Cell*, vol. 124, no. 1, pp. 191– 205, 2006.
- [113] P. Rajasethupathy, F. Fiumara, R. Sheridan et al., "Characterization of small RNAs in aplysia reveals a role for miR-124 in constraining synaptic plasticity through CREB," *Neuron*, vol. 63, no. 6, pp. 803–817, 2009.
- [114] I. Pichardo-Casas, L. A. Goff, M. R. Swerdel et al., "Expression profiling of synaptic microRNAs from the adult rat brain identifies regional differences and seizure-induced dynamic modulation," *Brain Research*, vol. 1436, pp. 20–33, 2012.
- [115] A. S. Nudelman, D. P. Dirocco, T. J. Lambert et al., "Neuronal activity rapidly induces transcription of the CREB-regulated microRNA-132, in vivo," *Hippocampus*, vol. 20, no. 4, pp. 492– 498, 2010.

- [116] D.-Z. Liu, Y. Tian, B. P. Ander et al., "Brain and blood microRNA expression profiling of ischemic stroke, intracerebral hemorrhage, and kainate seizures," *Journal of Cerebral Blood Flow and Metabolism*, vol. 30, no. 1, pp. 92–101, 2010.
- [117] E. M. Jimenez-Mateos, I. Bray, A. Sanz-Rodriguez et al., "miRNA expression profile after status epilepticus and hippocampal neuroprotection by targeting miR-132," *American Journal of Pathology*, vol. 179, no. 5, pp. 2519–2532, 2011.
- [118] E. M. Jimenez-Mateos, T. Engel, P. Merino-Serrais et al., "Silencing microRNA-134 produces neuroprotective and prolonged seizure-suppressive effects," *Nature Medicine*, vol. 18, no. 7, pp. 1087–1094, 2012.
- [119] F. Ratjen and G. Döring, "Cystic fibrosis," *The Lancet*, vol. 361, no. 9358, pp. 681–689, 2003.
- [120] J. A. Dodge, P. A. Lewis, M. Stanton, and J. Wilsher, "Cystic fibrosis mortality and survival in the UK: 1947–2003," *European Respiratory Journal*, vol. 29, no. 3, pp. 522–526, 2007.
- [121] M. E. Hodson, N. J. Simmonds, W. J. Warwick et al., "An international/multicentre report on patients with cystic fibrosis (CF) over the age of 40 years," *Journal of Cystic Fibrosis*, vol. 7, no. 6, pp. 537–542, 2008.
- [122] R. L. Gibson, J. L. Burns, and B. W. Ramsey, "Pathophysiology and management of pulmonary infections in cystic fibrosis," *American Journal of Respiratory and Critical Care Medicine*, vol. 168, no. 8, pp. 918–951, 2003.
- [123] W. Xu, C. Hui, S. S. B. Yu, C. Jing, and H. C. Chan, "MicroRNAs and cystic fibrosis—an epigenetic perspective," *Cell Biology International*, vol. 35, no. 5, pp. 463–466, 2011.
- [124] I. K. Oglesby, I. M. Bray, S. H. Chotirmall et al., "miR-126 is downregulated in cystic fibrosis airway epithelial cells and regulates TOM1 expression," *Journal of Immunology*, vol. 184, no. 4, pp. 1702–1709, 2010.
- [125] M. Bazett, A. Paun, and C. K. Haston, "MicroRNA profiling of cystic fibrosis intestinal disease in mice," *Molecular Genetics and Metabolism*, vol. 103, no. 1, pp. 38–43, 2011.
- [126] S. Bhattacharyya, N. S. Balakathiresan, C. Dalgard et al., "Elevated miR-155 promotes inflammation in cystic fibrosis by driving hyperexpression of interleukin-8," *Journal of Biological Chemistry*, vol. 286, no. 13, pp. 11604–11615, 2011.
- [127] A. R. Kuhn, K. Schlauch, R. Lao, A. J. Halayko, W. T. Gerthoffer, and C. A. Singer, "MicroRNA expression in human airway smooth muscle cells: Role of miR-25 in regulation of airway smooth muscle phenotype," *American Journal of Respiratory Cell and Molecular Biology*, vol. 42, no. 4, pp. 506–513, 2010.
- [128] S. A. Moschos, A. E. Williams, M. M. Perry, M. A. Birrell, M. G. Belvisi, and M. A. Lindsay, "Expression profiling in vivo demonstrates rapid changes in lung microRNA levels following lipopolysaccharide-induced inflammation but not in the antiinflammatory action of glucocorticoids," *BMC Genomics*, vol. 8, article 240, 2007.
- [129] S. Ramachandran, P. H. Karp, P. Jiang et al., "A microRNA network regulates expression and biosynthesis of wild-type and ΔF508 mutantcystic fibrosis transmembrane conductance regulator," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 33, pp. 13362–13367, 2012.
- [130] A. E. Gillen, N. Gosalia, S.-H. Leir, and A. Harris, "MicroRNA regulation of expression of the cystic fibrosis transmembrane conductance regulator gene," *Biochemical Journal*, vol. 438, no. 1, pp. 25–32, 2011.
- [131] F. Megiorni, S. Cialfi, C. Dominici, S. Quattrucci, and A. Pizzuti, "Synergistic post-transcriptional regulation of the cystic fibrosis

transmembrane conductance regulator (CFTR) by miR-101 and miR-494 specific binding," *PLoS ONE*, vol. 6, no. 10, article e26601, 2011.

[132] J. R. Rao, D. Nelson, J. E. Moore et al., "Non-coding small (micro) RNAs of Pseudomonas aeruginosa isolated from clinical isolates from adult patients with cystic fibrosis," *British Journal of Biomedical Science*, vol. 67, no. 3, pp. 126–132, 2010.