

## Liver Lesions in Hepatitis B Viral Infection

V.J. DESMET, M.D., Ph.D.

*University Hospital St. Rafaël, Department of Medical Research, Catholic  
University Leuven, Leuven, Belgium*

Received August 4, 1987

---

A review is made of the various histological lesions observed in hepatitis B virus-related liver diseases, including different forms of acute and chronic hepatitis, cirrhosis, and hepatocellular carcinoma. The elementary lesions discussed include acidophil necrosis (apoptosis), confluent lytic necrosis in its different patterns, piecemeal necrosis, focal necrosis, and dysplastic hepatocytes. Their pathogenesis is explained in the framework of recent developments in the immunopathology of hepatitis B viral infections.

---

### INTRODUCTION

Infection with the hepatitis B virus (HBV) results in a broad variety of clinical diseases. Evidence supporting this clinical experience came from transmission studies in volunteers nearly half a century ago, in which identical inocula of HBV were injected into healthy recipients and resulted in different clinical responses [1,2].

In 1972, it was first suggested [3] that HBV is not directly cytopathic to the hepatocyte, and that hepatic inflammation and liver cell damage are mediated by various immunological mechanisms. The thesis that host immunity is most important in the causation of liver damage has gained considerable support [4,5], although the factors which determine a particular syndrome in the individual patient remain largely unknown, and direct cytotoxicity of the virus on liver parenchymal cells is not excluded [6,7].

The various hepatic diseases which may result from HBV infection are shown in Fig 1. This paper reviews the main histopathological liver lesions and their presumed pathogenic mechanisms.

### ACUTE VIRAL HEPATITIS B

Acute viral hepatitis B may take the form of a mild disease, either subclinical, anicteric infection or classical icteric hepatitis with spontaneous recovery, or it may present more rarely as a severely necrotizing, life-threatening disease.

All forms of acute viral hepatitis B apparently result from an adequate response of

61

*Abbreviations:* HBcAg: hepatitis B core antigen HBsAg: hepatitis B surface antigen HBV: hepatitis B virus Lex: liver extract LIP: liver immunoregulatory protein or liver-derived inhibitory protein LMA: liver membrane antigen LSP: liver specific protein MHC: major histocompatibility complex NK: natural killer RIF: rosette inhibitory factor

This lecture was presented on April 16, 1987, to inaugurate the Annual Gerald Klatskin Memorial Lectureship in Hepatology. The Klatskin Lectureship has been established by the Department of Internal Medicine through gifts from former fellows, family, and friends, in memory of Dr. Gerald Klatskin (1910–1986), founder of the Liver Study Unit at the Yale University School of Medicine and a member of the Department of Internal Medicine for more than 50 years.

Address reprint requests to: Prof. Dr. V.J. Desmet, Universitair Ziekenhuis St. Rafaël, Laboratorium voor Histo- & Cytochemie, Minderbroedersstraat 12, B-3000 Leuven, Belgium

Copyright © 1988 by The Yale Journal of Biology and Medicine Inc.

All rights of reproduction in any form reserved.

### LIVER LESIONS IN HEPATITIS B VIRAL INFECTION

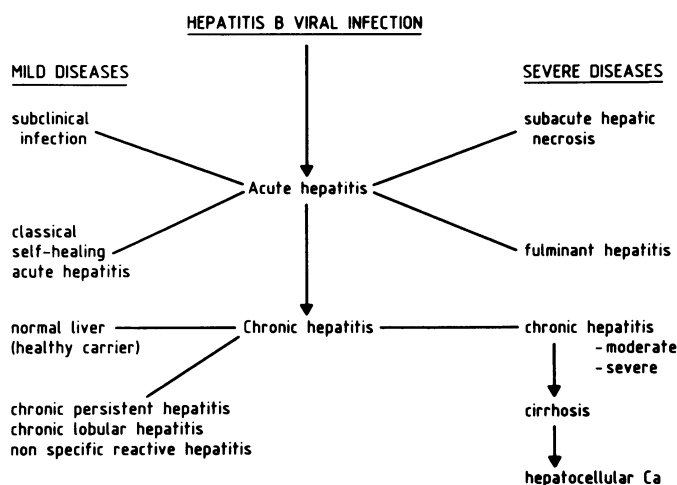


FIG. 1 Liver lesions in hepatitis B viral infection (modified after Thomas et al. [5]).

the host's immune defense, resulting in complete elimination of the virus; acute viral hepatitis B thus represents an "elimination type" of hepatitis [8]. Eradication of the virus implies, however, elimination of virus-infected hepatocytes; this fact is reflected in the complex array of histological lesions observed, including parenchymal damage, hepatocellular necrosis, liver cell regeneration, and inflammatory infiltration.

### CLASSICAL ACUTE HEPATITIS WITH RECOVERY

The histological lesion in uncomplicated acute hepatitis B is a complex one. The sinusoids are crowded with increased numbers of mononuclear cells, mainly macrophages and lymphocytes, with highest density in acinar zone 3. This increased sinusoidal cellularity reflects the intrahepatic immunological reaction against the virus. The sinusoidal lining cells and macrophages have been shown to express strongly HLA-DR or HLA class II antigens [9]. These cells are presumed to function as antigen-presenting cells to T-helper lymphocytes [10].

In order to react to the foreign viral antigen, T-helper lymphocytes need to "see" this antigen in conjunction with self antigens (HLA class II antigens) expressed on the surface of "accessory" or antigen-presenting cells (often indicated by the vague term "macrophages"). The latter secrete the lymphokine interleukin 1, which activates the helper T lymphocytes (Fig. 2). Activated T-helper lymphocytes in turn activate cytotoxic T cells and B lymphocytes through secretion of the lymphokine interleukin 2 (Fig. 2).

Infected hepatocytes supporting viral replication express part of the viral antigens at their plasma membranes: both hepatitis B surface antigen (HBsAg) and hepatitis B core antigen (HBcAg) have been demonstrated on the surface of mechanically isolated hepatocytes in acute viral hepatitis B [11]. In addition to expressing viral antigens, the hepatocyte is also triggered to display at its surface HLA A,B,C (or HLA class I) antigens, which are not expressed on liver parenchymal cells in the normal state [9,12,13].

In this way, the stage is set for a cytotoxic T-cell attack to viral antigens expressed on

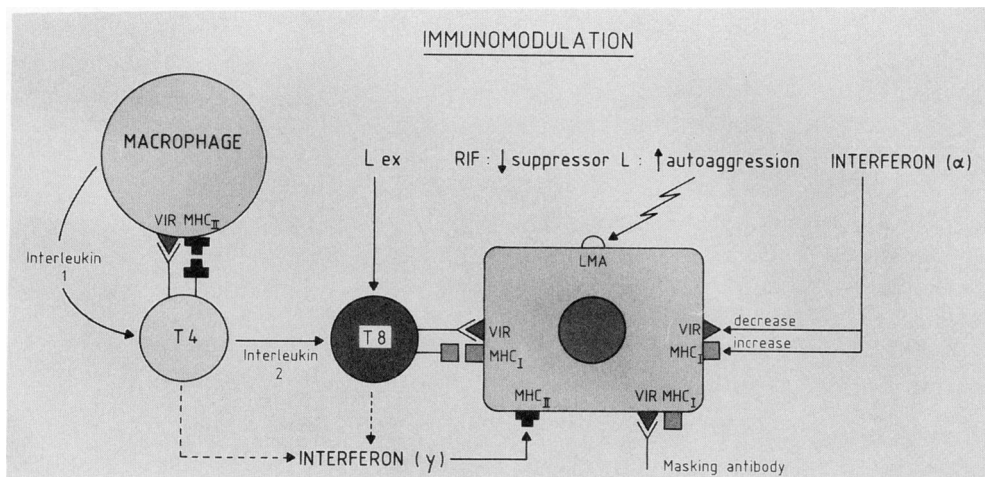


FIG. 2. Abbreviations: **LMA**: liver membrane antigen **Lex**: liver extract **MHC<sub>I</sub>**: major histocompatibility complex antigens, class I **MHC<sub>II</sub>**: major histocompatibility complex antigens, class II **VIR**: viral antigen **RIF**: rosette inhibitory factor **T<sub>4</sub>**: helper-inducer lymphocyte **T<sub>8</sub>**: suppressor-cytotoxic lymphocyte For explanation, see text.

the liver cell's surface, since cytotoxic T cells can now "see" the foreign viral antigen to which they are sensitized in conjunction with the HLA class I antigens. The latter is a necessary constraint of major histocompatibility complex (MHC) compatibility in order to assure their effective reaction [14,15].

On the basis of published histological and immunohistochemical observations, it appears that immunologic elimination of HBV-infected hepatocytes might be mediated by cytotoxic T lymphocytes. Such a mechanism was demonstrated in an elegant animal model of mouse hepatitis [16], and evidence in support of similar mechanisms in human acute viral hepatitis B was recently produced by functional immunologic studies [11].

Immunohistochemical analysis of lymphocyte subsets in acute viral hepatitis B revealed, however, that the infiltrating lymphocytes are not exclusively composed of OKT8+ cytotoxic T lymphocytes; the predominant lymphocyte subset corresponds to natural killer (NK) lymphocytes, which show no genetic restriction and do not require HLA class I antigen display on the target cells [17]. Also, recent functional immunological studies suggest involvement of NK cells [11,18]. These observations fit with what is known in other viral infections, that NK cells are the first line of defense against viral infection.

The result of the lymphocyte attack—either by cytotoxic T cells or natural killer cells or both—is a killing of the target cell.

The mode of cell death in immune-mediated target cell lysis is not classical cell necrosis, but apoptosis [19]. Apoptosis is a special pre-programmed and active mode of cell death. A precise sequence of intracellular changes occurs, including condensation and margination of the nuclear chromatin and formation of numerous blebs at the cell surface, but the intracellular organelles remain well preserved in contrast to what happens in classical cell necrosis. Finally, the cell breaks down into several fragments surrounded by a membrane.

Under the light microscope, apoptosis can be recognized as smaller and larger eosinophilic cell fragments, occasionally containing a pycnotic nucleus (Fig. 3). These

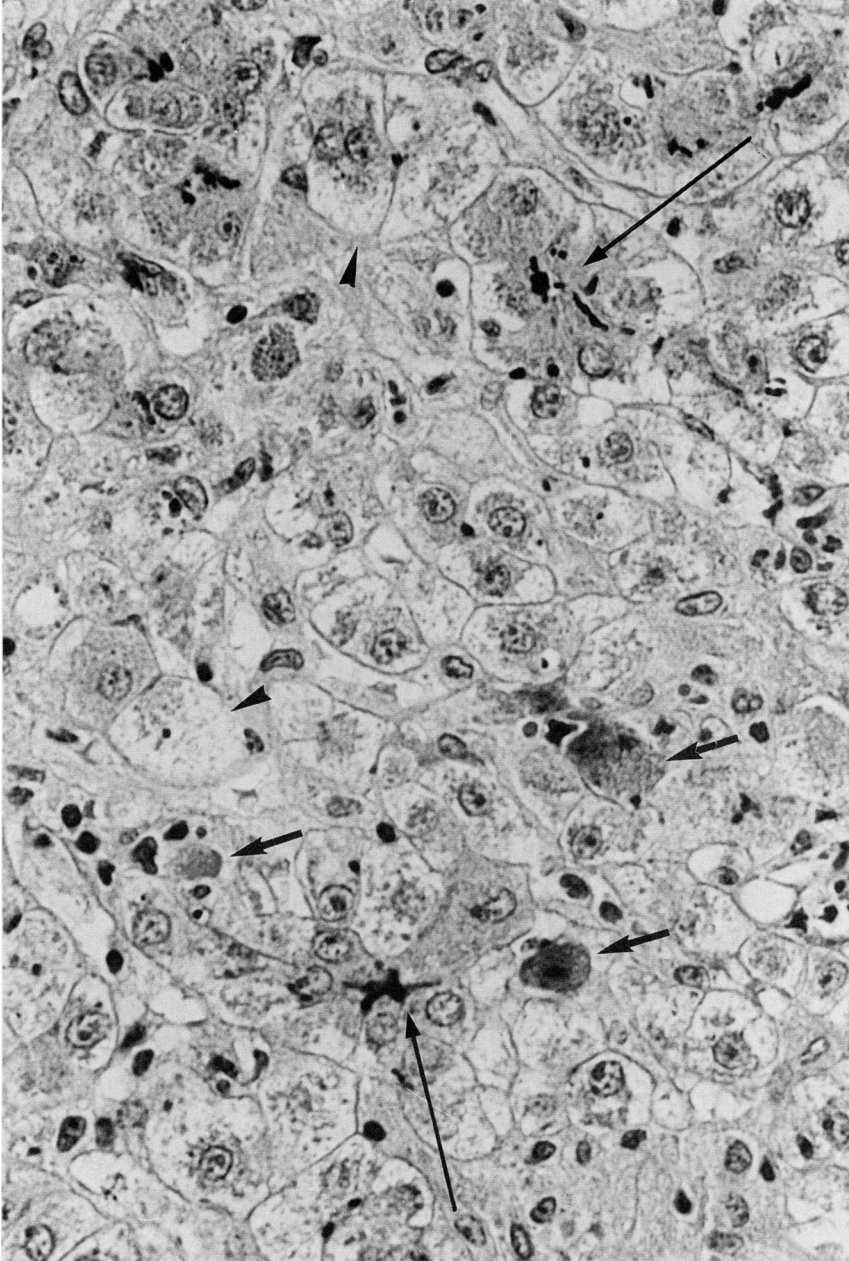


FIG. 3. Centrolobular (zone 3) parenchyma in acute self-limited viral hepatitis B. Several acidophil (apoptotic) bodies are present (arrows), some of them in close contact with lymphocytes. Some hepatocytes appear swollen (liver cell ballooning) (arrowheads); intercellular bile thrombi (long arrows) indicate cholestasis. Hematoxylin and eosin,  $\times 640$

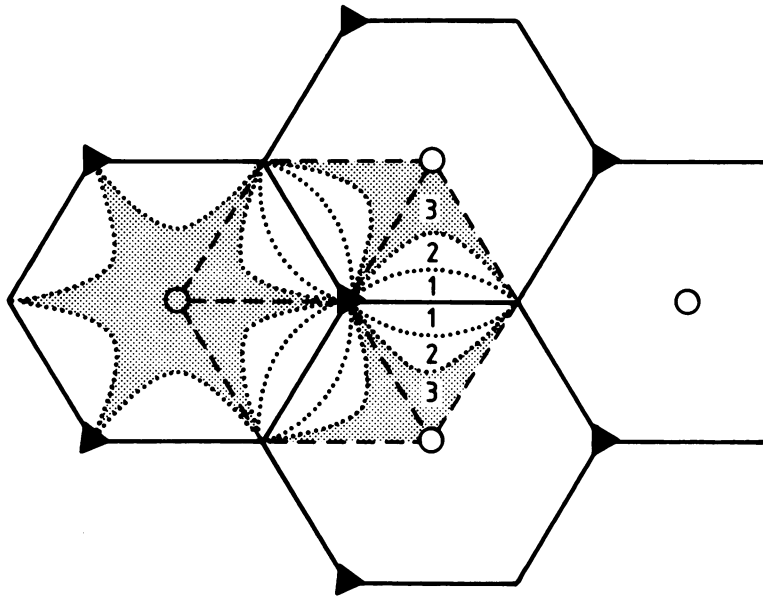


FIG. 4. Schematic drawing of the acinar and lobular concepts of liver architecture. Black triangles represent portal tracts, open circles central veins. The central hexagon, delineated by interrupted lines and centered by a portal tract, represents a complex acinus, composed of three simple acini which are centered by a terminal portal venule (*continuous line*). Each simple acinus can be subdivided in acinar zones 1, 2, and 3, the latter representing the microcirculatory periphery. Necrosis in the periphery of the complex acinus leads to central-central bridging necrosis. Necrosis in the periphery of the simple acinus (zone 3) leads to portal-central bridging necrosis (*shaded area*). This pattern of necrosis creates a star-shaped area of necrosis from the point of view of the classical hexagonal liver lobule (*left hexagon, centered by central vein*).

apoptotic bodies become phagocytosed very soon by neighboring parenchymal cells and by nearby macrophages [20]. Close inspection of liver biopsies taken in patients with acute viral hepatitis B reveals numerous apoptotic bodies in close relationship with lymphocytes. Obviously, it is more difficult to identify the smaller apoptotic bodies, whereas the larger apoptotic fragments are easily recognized. The latter were known for many years as “acidophil necrosis,” “acidophil bodies,” or “Councilman-like bodies” [21] (Fig. 3).

Research on apoptosis has revealed that what was usually considered as single-cell necrosis does not at all correspond to classical cell necrosis but instead represents a lymphocyte-induced triggering of a pre-set program for active self-destruction of the liver cell [22].

Activated lymphocytes release several lymphokines; one of these soluble factors is a cholestasis-inducing lymphokine [23]. This finding may at least in part explain the occurrence of microscopic features of cholestasis in most liver biopsies from patients with acute hepatitis B (Fig. 3).

Not all damaged hepatocytes appear as small, shrunken apoptotic bodies in acute viral hepatitis B. One of the most striking features of the histopathological picture is liver cell pleomorphism, most pronounced in acinar zone 3 [21]. Several hepatocytes

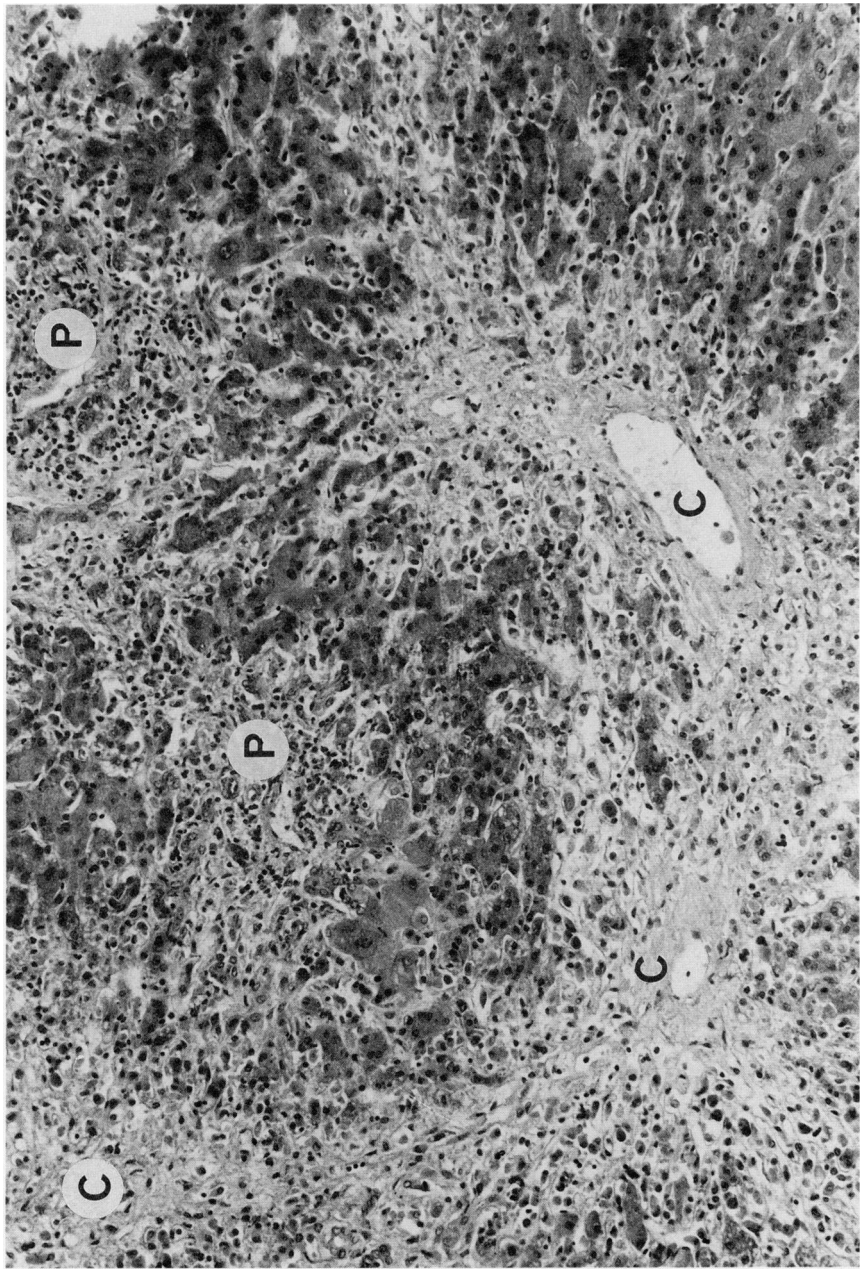


FIG. 5. Central-bridging confluent necrosis. Autopsy specimen from patient with fulminant acute hepatitis B. An area of confluent lytic necrosis links several adjacent central veins (C); the portal tracts (P) show lymphoid infiltration and are surrounded by surviving parenchyma (complex acinus). Hematoxylin and eosin,  $\times 160$

appear enlarged and pale, corresponding to "hydropic swelling" or "ballooning of hepatocytes" (Figs. 3,7). Hydropic swelling is believed to be a precursor stage of lytic necrosis of hepatocytes. The precise mechanism of this type of hepatocellular damage and death is not well known, but there is reason to assume that the primary damage is at the cell membrane [24]. It is this variety of cell damage with leaky cell membranes, which probably is most responsible for the release of intracellular enzymes and which contributes to the rise of serum transaminases.

Lytic necrosis of hepatocytes is a rapid event; the cell disappears in a very short time. Hence it follows that lytic necrosis is recognized histologically as an absence of cells. This condition is often described as "drop-out of liver cells," resulting in denudation of the reticulin framework. Lytic necrosis usually affects groups of hepatocytes, leading to the disappearance of confluent areas of parenchyma; this fact explains the term "confluent necrosis" [25].

### SEVERE ACUTE VIRAL HEPATITIS B (SUBACUTE HEPATIC NECROSIS; FULMINANT HEPATITIS)

Confluent necrosis may affect smaller or larger parenchymal territories. When confluent necrosis destroys larger portions of parenchyma, the acute hepatitis is usually of a clinically more severe type, designated as subacute hepatic necrosis and fulminant hepatitis (Fig. 1). One of the remarkable features of confluent lytic necrosis is that it occurs in well-defined areas of the liver parenchyma: it shows a predilection for the microcirculatory periphery [26].

The microcirculatory periphery of the liver parenchyma is better explained by the acinar concept of liver architecture, according to Rappaport, than by the classical lobular concept according to Kiernan [27] (Fig. 4).

Confluent lytic necrosis in the microcirculatory periphery of the complex acinus leads to necrotic bridges between adjacent central veins (central-central bridging necrosis) (Figs. 4,5); whereas confluent necrosis in the microcirculatory periphery of the simple acinus (or acinar zone 3) creates necrotic bridging between portal tracts and central veins (portal-central bridging necrosis) (Figs. 4,6,7). The latter corresponds, at the lobular level, to a star-shaped area of necrosis linking portal tracts and central veins [27] (Figs. 4,6).

These patterns of necrosis correspond to what is often termed "bridging hepatic necrosis" [28]. When still larger masses of parenchymal cells succumb to confluent lytic necrosis, the drop-out of liver cells extends to zone 2 (Fig. 8), and even up to zone 1 of the simple acini, resulting in total destruction of all parenchyma of the liver units: panlobular and multilobular necrosis (Fig. 9). Very often, multilobular necrosis displays an irregular distribution throughout the liver, apparently corresponding to total destruction of some acinar agglomerates, whereas adjacent larger parenchymal units (or acinar agglomerates) are spared.

This predilection of confluent lytic necrosis to occur first in peripheral areas of the microcirculation, the apparent relationship of multilobular necrosis to hepatic angio-architecture, and the relative scarcity of lymphocytes in the denuded areas are features which suggest that in this mode of necrosis other mechanisms are involved than those in focal apoptotic cell death.

The most plausible mechanism is a humoral one, possibly circulating antibodies or immune complexes, associated with activation of complement. Complement components (the complement C5b-9m complex) has been shown to cause a lytic type of



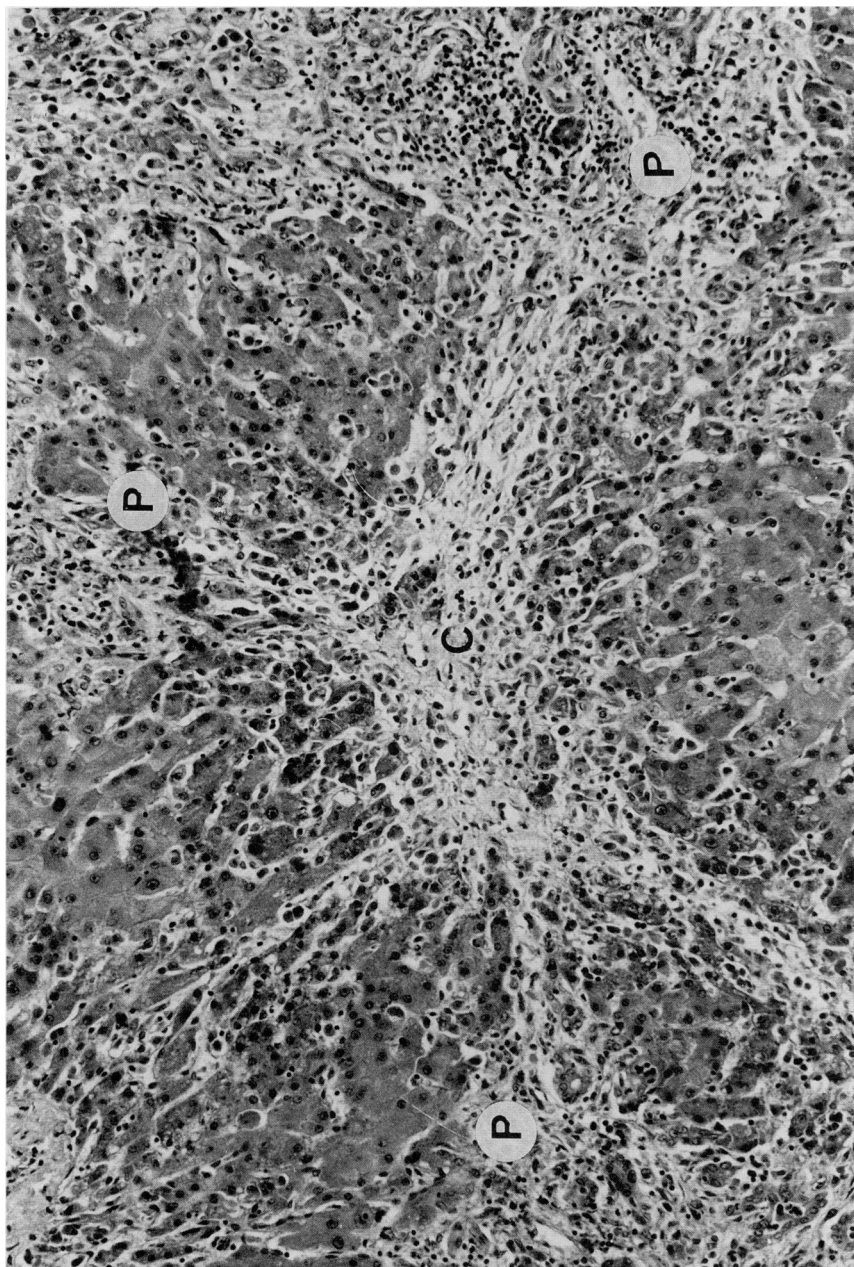


FIG. 6. Portal-central bridging confluent necrosis. Autopsy specimen from patient with fulminant acute hepatitis B (same case as Fig. 5). An area of confluent lytic necrosis links the central vein (C) with surrounding portal tracts (P). Portal-central bridging necrosis creates a star-shaped area of necrosis in the classical hexagonal liver lobule (compare with Fig. 4). Hematoxylin and eosin,  $\times 160$



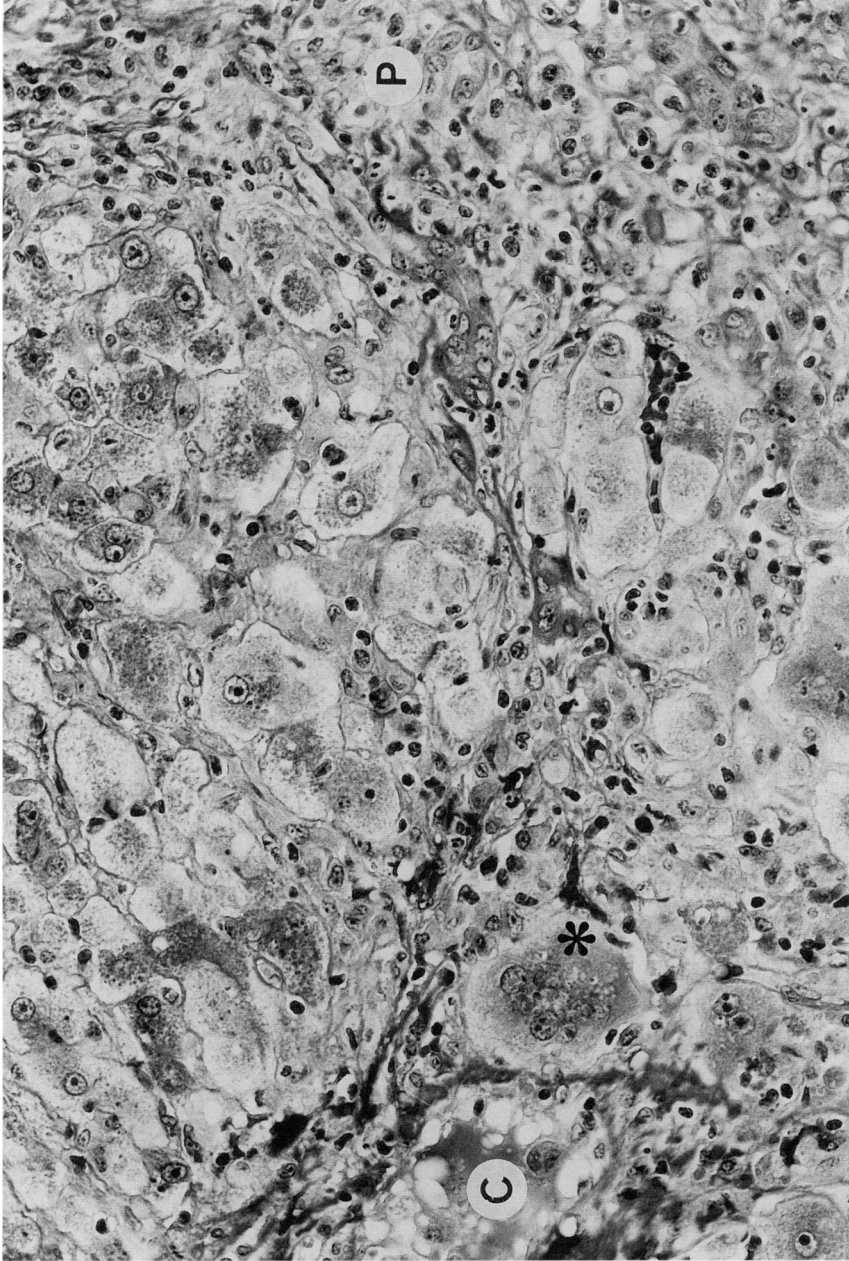


FIG. 7. Portal-central bridging necrosis. Liver biopsy from patient with acute hepatitis B. A streak of necrosis extends from the central vein (C) to the portal tract (P), interrupting parenchymal continuity. Note lymphocytic infiltration and ballooning of surviving hepatocytes. A multinucleated hepatocyte (\*) near the central vein represents an attempt at regeneration. Masson's trichrome stain,  $\times 400$

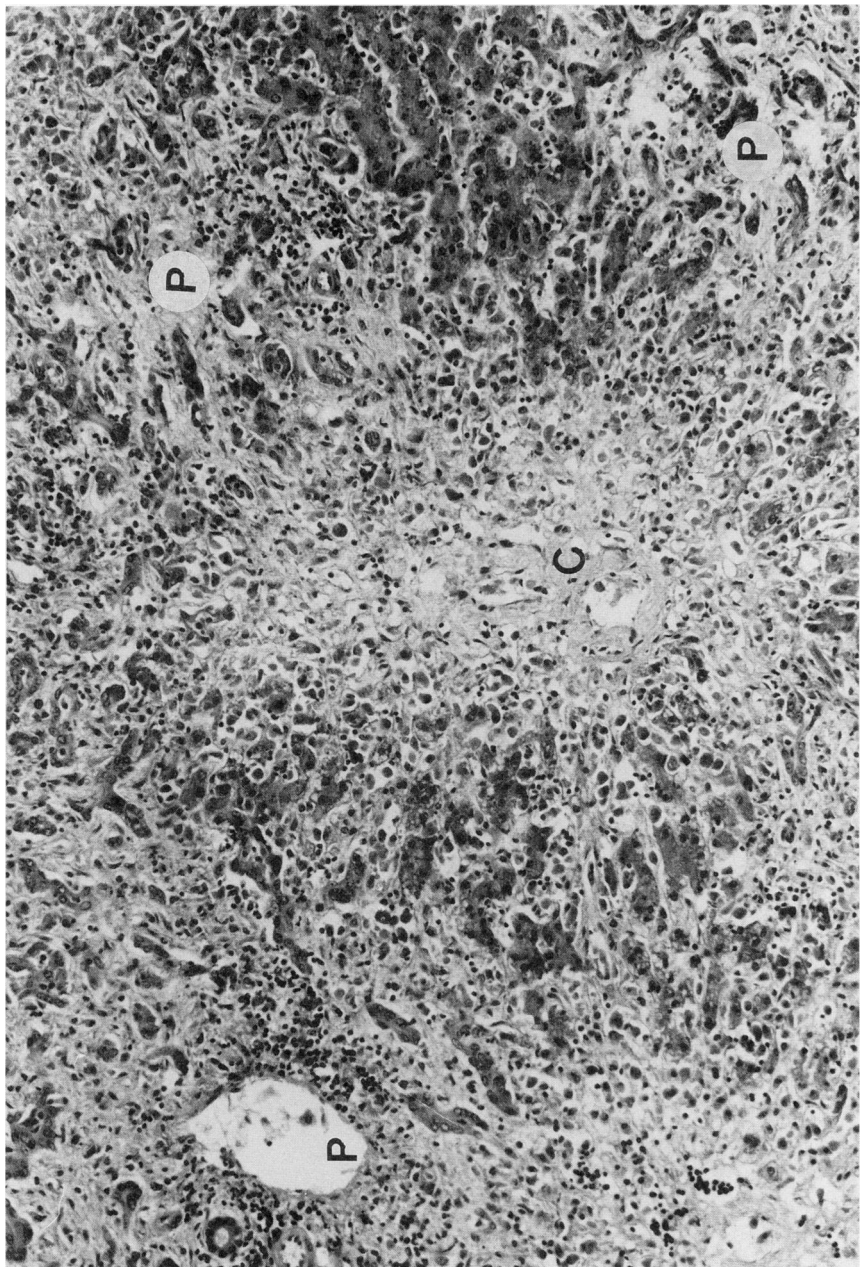


FIG. 8. Submassive confluent lytic necrosis. Autopsy specimen from patient with fulminant hepatitis B (same case as Fig. 5). Most of the liver lobule (acinar zones 2 and 3) is necrotic; the surviving parenchyma (zone 1) lies near the portal tracts (P). Early condensation of mesenchyme occurs around the central vein (C). Hematoxylin and eosin,  $\times 160$

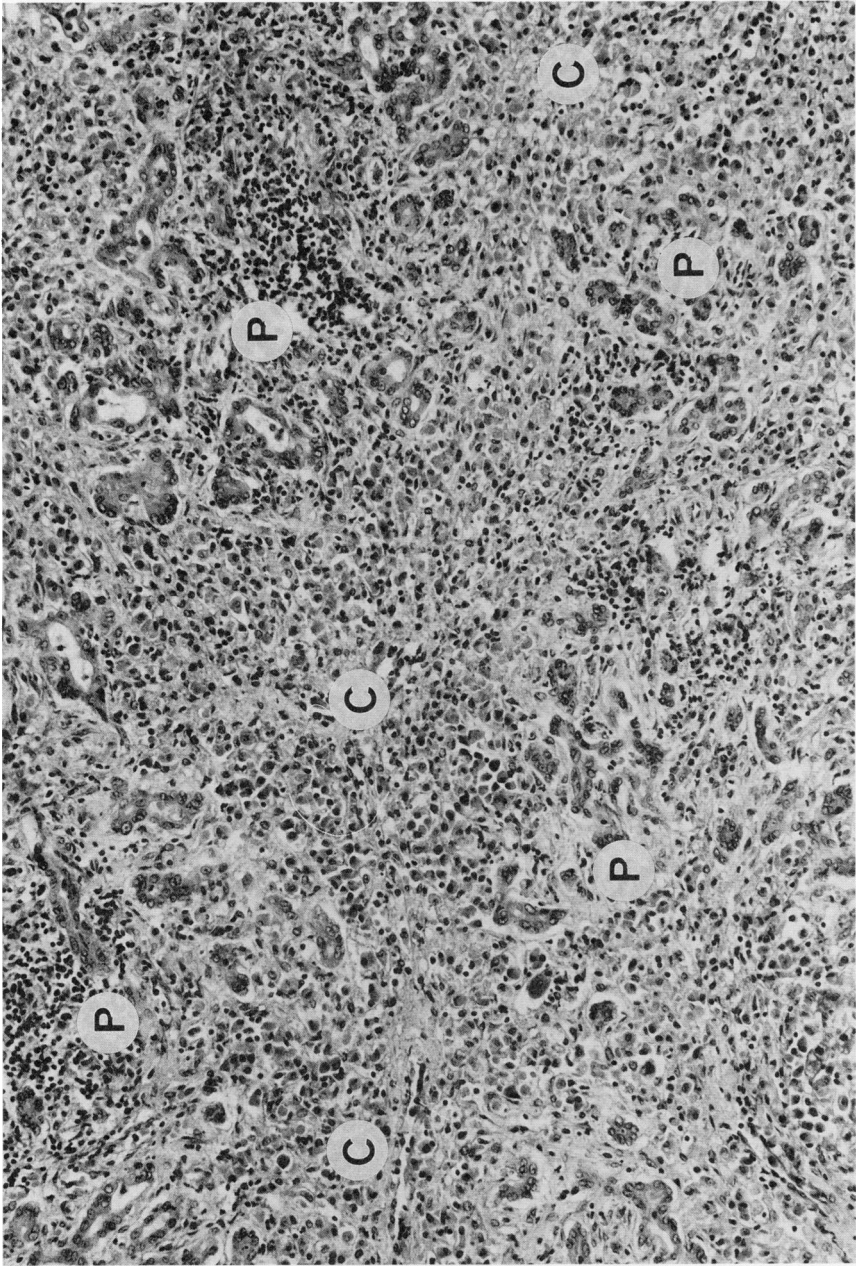


FIG. 9. Multilobular confluent lytic necrosis. Autopsy specimen from patient with fulminant hepatitis B (same case as Fig. 5). Virtually all parenchyma has disappeared in several adjacent lobules; the lobular territories are occupied by some lymphocytes and numerous macrophages. The increased number of ductular profiles around the portal tracts (P) is usually interpreted as an attempt at regeneration. C: central veins. Hematoxylin and eosin,  $\times 160$

necrosis, since it behaves like a membrane-piercing protein [29,30]. Piercing of the cell membrane causes rapid deregulation of ionic pumps, rapid influx of calcium ions, and immediate cell lysis [24].

In summary, the pleomorphic appearance of the liver parenchyma in acute viral hepatitis B seems to reflect a multitude of immunological mechanisms, apparently at work simultaneously or in sequence to eliminate virus-supporting hepatocytes. These mechanisms include: activation of antigen-presenting cells (macrophages), focal apoptosis mediated by infiltrating NK lymphocytes and cytotoxic T lymphocytes, and ballooning of hepatocytes, eventually followed by confluent lytic necrosis, presumably brought about by humoral mechanisms.

The viral antigen(s) which function(s) as target antigen(s) for the immune elimination is (are) not so extensively studied in acute as in chronic hepatitis B; the obvious reason is that diagnostic liver biopsies are usually not taken during the incubation period, but rather at the height of transaminase elevation.

In this fully developed stage of acute hepatitis, only little or no HBsAg or HBcAg can be demonstrated in the liver. This state is consistent with the concept that acute hepatitis B is an "elimination type" of disease, resulting in eradication of the virus and assuring a self-limited course of the disease [8]. The necrotic hepatocytes are phagocytosed by Kupffer cells, which acquire a bulky cytoplasm, loaded with coarse, golden-yellow PAS-positive and diastase-resistant pigment granules termed ceroid pigment. These ceroid-laden macrophages tend to cluster in the parenchyma and finally migrate toward the portal tracts. The loss of parenchymal cells is compensated for by hepatocellular regeneration, which usually leads to complete "restitutio ad integrum" in uncomplicated acute viral hepatitis B.

## CHRONIC VIRAL HEPATITIS B

In about 10 percent of patients infected with HBV, the disease which develops corresponds to chronic hepatitis (Fig. 1).

## CLASSIFICATION OF CHRONIC HEPATITIS

Histologically, the term chronic hepatitis refers to a broad spectrum of hepatic inflammatory conditions, ranging from near-normal liver histology to complete cirrhotic transformation of the liver architecture. Classification remains a problem, because of the multiplicity of simultaneous lesions, the imprecise borderline between subgroups, the subjective evaluation of individual tissue alterations, the problem of sampling error, and terminologic confusion.

In 1968, an international Liver Study Group proposed a very simple classification of chronic hepatitis [31]. Distinction was made between chronic persistent hepatitis, a milder form of disease with better prognosis, and chronic aggressive hepatitis, a more severe category of disease with frequent evolution into cirrhosis. This classification emphasized piecemeal necrosis (see below) as the important histological hallmark of progressive forms of chronic hepatitis.

In contrast, Dr. Gerald Klatzkin and his colleagues from Yale maintained that the real pacemaker for cirrhotic evolution is bridging hepatic necrosis [32–35]. This controversy became the subject of a long debate. Patterns and extent of piecemeal necrosis and bridging hepatic necrosis were examined in several studies which also intended to evaluate the use of steroid treatment.

These studies were evaluated in a recent editorial in *Hepatology* [36], ending with the main conclusion that in fact the problem is not finally settled: larger and better

stratified series of patients need to be studied. Nevertheless, a unifying picture seems to emerge in that both piecemeal necrosis and bridging hepatic necrosis are prognostically important lesions. The concept that lobular necro-inflammatory lesions, especially more extensive bridging necrosis, characterize more severe variants of the disease, was incorporated in further classifications of chronic hepatitis [37–39].

As a result of ongoing studies, recent schemes of chronic hepatitis have become more complicated but still distinguish between milder forms and more severe variants of the disease (Table 1) [40].

The prototype form of mild chronic viral hepatitis is chronic persistent hepatitis. Histologically, this condition corresponds to “portal” hepatitis [41]. The portal tracts are infiltrated by lymphoid cells, which remain confined within the limits of the portal connective tissue. The lobular parenchyma shows minimal lesions: occasional focal necrosis and rare apoptotic bodies.

The more severe category of chronic hepatitis (chronic active or aggressive hepatitis) comprises several variants with different degrees of liver damage, all characterized by piecemeal necrosis (Table 1).

In mild to moderate chronic aggressive hepatitis, the predominant lesion is periportal piecemeal necrosis; the lobular parenchyma shows only minor changes, including some scattered foci of focal necrosis. This type of chronic hepatitis, although mild, may slowly progress into cirrhosis [42].

More severe variants of chronic aggressive hepatitis show a more complex histologic picture; not only is piecemeal necrosis more extensive, but in addition the lobular lesions are impressive and may include bridging hepatic necrosis (Table 1).

Bridging hepatic necrosis obviously entails more extensive and more rapid dissection of the lobular architecture. The most severe form of confluent lytic necrosis, i.e., multilobular necrosis, may be an especially ominous feature [43]. Dr. G. Klatzkin and his colleagues deserve the credit for having emphasized the fundamental importance of bridging hepatic necrosis in accelerating the evolution of chronic hepatitis into cirrhosis [32–35].

Collapse of the reticulin framework in areas of extensive confluent necrosis leads to the formation of fibrous scars; these can be considered as “passive” septa, since they derive primarily from passive collapse of a denudated frame of fibers with subsequent new collagen formation [25]. Passive septa show an excess of fibers over cells; they are paucicellular septa, carrying little inflammatory infiltration, and remain sharply delineated from the surrounding parenchyma. This appearance is in contrast to extending wedges of piecemeal necrosis, rich in cells and poorly delineated, which have been termed active septa.

## NATURAL HISTORY OF CHRONIC VIRAL HEPATITIS B

For several years, classification of chronic hepatitis has been the decision base for starting treatment with steroids and/or immunosuppressives. This procedure is no longer the case for chronic viral hepatitis type B [44].

Histological definition of the degree of disease activity and the stage of progression of the disease remains, however, important in the light of newer insights into the natural history of the disease.

From several studies [5,45,46] it appeared that three phases can be recognized over the years during the long course of chronic hepatitis B.

In the first phase, which can be considered as a period of “immune tolerance” to the HBV, extensive viral replication takes place; this process is reflected in positive

TABLE 1  
Classification of Chronic Hepatitis

Category	Interstitium		Parenchyma	Architecture
CLH		Mild portal hepatitis	Marked spotty necrosis	Normal
CPH		Portal hepatitis	Spotty necrosis	Normal
Chronic septal hepatitis	Minimal			
	Marked	Minimal		Minimal
NSRH		Mild portal hepatitis	Minimal spotty necrosis	Passive septa—no nodules
		Mild (variable) portal hepatitis	Occasional spotty necrosis	Normal
CAH		PMN	Kupffer cell activation	
	Minimal			
Moderate		Minimal PMN	Mild spotty necrosis	Normal
	Severe	Moderate PMN	Marked spotty necrosis	± Disturbed
Very severe		Severe PMN	Severe spotty necrosis	Disturbed
		active septa	Focal confluent necrosis	
		Severe PMN	Zonal confluent necrosis	
		active septa	BIIN	
Cirrhosis		Bile duct lesion		—PC CN
				—Panlobular CN
				—MLN
				Severely disturbed
	Active	PMN		
	Inactive	Portal hepatitis	CN	Nodules + active septa
			Spotty necrosis	Nodules + passive septa

*Abbreviations:* CLH: chronic lobular hepatitis CPH: chronic persistent hepatitis BHN: bridging hepatic necrosis CAH: chronic active (aggressive) hepatitis PMN: piecemeal necrosis CN: confluent necrosis NSRH: nonspecific reactive hepatitis PC: portal-central MLN: multilobular necrosis

serological reactions for HBeAg, hepatitis B viral DNA, and DNA polymerase activity. Biochemically, the disease is mild, without marked elevation of transaminase levels. Histologically, the liver biopsy reveals only low inflammatory activity, corresponding to chronic persistent hepatitis or non-specific reactive hepatitis.

This form of chronic persistent hepatitis is of the so-called HBcAg-predominance type [47]: the majority of the hepatocytes are positively stained for HBcAg, mainly in their nuclei, and HBsAg is often found in a membranous localization in the liver cell membranes. Nuclei which are overloaded with HBcAg show on hematoxylin-eosin stained sections a peculiar appearance, described as "sanded nuclei" [48]. This stage represents the high-replicative phase of chronic hepatitis.

After some time, and for reasons unknown, the immune system succeeds in eliminating substantial numbers of virus-infected cells. This phase can be considered the "immune-clearance" phase of the disease, characterized by low levels of viral replication and in many patients by seroconversion from HBeAg to anti-HBe antibody. This phase is preceded by or associated with a flare-up of transaminases and is characterized histologically by more impressive parenchymal lesions, recognizable as chronic lobular or chronic active hepatitis. The episode of parenchymal necrosis may be quite severe, with multiple areas of focal necrosis and even bridging hepatic necrosis [49]. Such a necrotizing episode may turn the liver into cirrhosis.

The ensuing third phase is the "non-replicative" phase: one finds no more signs of viral replication and the patient is anti-HBe antibody-positive. Transaminases are only slightly elevated, and, accordingly, liver histology reveals only mild inflammatory lesions of chronic persistent hepatitis. This form of chronic persistent hepatitis is, however, of the so-called HBsAg-predominance type [47]: large numbers of parenchymal cells display a massive accumulation of HBsAg in the cytoplasm, and only few or no nuclei are positive for HBcAg. Such parenchymal cells, containing large amounts of HBsAg in their endoplasmic reticulum, represent the so-called ground-glass hepatocytes [50]. It should be emphasized, however, that the ground-glass appearance of hepatocytes may also be caused by other mechanisms [51].

Apparently, several virus-infected cells have escaped immune elimination. This condition might be due to defective display of viral antigens at the liver cell surface, inadequate display of HLA class I antigens, or masking of membrane-expressed viral antigen(s) by antibody (Fig. 2).

Several studies [52,53] indicate that the latter mechanism is operating in chronic hepatitis B: HBcAg-positive liver cells are covered with immunoglobulin IgG which could be shown to have anti-HBc antibody specificity. This condition adds to the complexity of phenomena occurring in the liver in HBV infection and is part of a complex network of immunomodulation (Fig. 2).

## NECRO-INFLAMMATORY LESIONS IN CHRONIC ACTIVE VIRAL HEPATITIS B

### *Piecemeal Necrosis*

Piecemeal necrosis is not unlike interphase dermatitis, observed, for example, in lichen ruber planus or phototoxic dermatitis [54]. In these conditions, mononuclear inflammatory cells concentrate at the interphase between epidermal keratinocytes and dermal connective tissue. The infiltrate is predominantly composed of helper T lymphocytes in the derma, whereas cytotoxic T cells infiltrate between the epidermal keratinocytes, which undergo individual acidophil necrosis (so-called Civatte bodies).



The number of epidermal, HLA class II antigen-positive Langerhans cells is increased, whereas the keratinocytes themselves show increased expression of HLA class I antigens [55] and de novo display of HLA class II antigens [56]. The whole process causes an indistinct borderline between epidermis and derma.

Interphase dermatitis represents a cell-mediated immunological reaction, associated with destruction of epidermal keratinocytes. The antigen against which the reaction is mounted remains unknown.

Piecemeal necrosis in the liver can also be described as an interphase hepatitis [57]. Here also the epithelial-mesenchymal borderline is blurred, lymphocytes invade the epithelial territories, and some hepatocytes appear as acidophil bodies (Fig. 10).

The predominant type of lymphocyte in the invading fronts of piecemeal necrosis is the OKT8+ suppressor/cytotoxic T lymphocyte, whereas OKT4+ helper/inducer T lymphocytes are more numerous in the central area of the portal connective tissue [58].

Some of the portal and periportal mononuclear cells do not correspond to lymphocytes but represent accessory cells, reticulum cells, or antigen-presenting cells.

One type of reticulum cell shows the ultrastructural features of follicular dendritic cells which are well known in the lymph follicles or so-called B areas of lymphoid tissues like lymph nodes and spleen. They are mainly found in the central parts of portal tracts [58,59].

In the peripheral areas of piecemeal necrosis, another type of reticulum cell can be observed. It has the electron microscopic features of interdigitating reticulum cells, which were often described in the paracortical areas or so-called T zones of lymph nodes [59]. These cells are the homologues of the Langerhans cells in the skin. Dendritic and interdigitating reticulum cells appear to function as antigen-presenting cells, respectively, for B and T lymphocytes [60].

The morphological similarity of these reticulum cells to those observed in lymph nodes and spleen, and their preferential localization in central or peripheral areas of piecemeal necrosis, allow us to consider the central part of the portal tract as the B zone, and the invading periphery as the T zone of piecemeal necrosis [58].

Endothelial cells also play an important role in immunological reactions. One of the crucial features is to influence the exit of lymphocytes from blood to interstitium. In areas of piecemeal necrosis, numerous lymphocytes are crowded together (Fig. 10), suggesting that the capillaries in those areas must be a preferential zone of exit from blood to tissue, not unlike the high-endothelial venules in the paracortical areas of lymph nodes.

Ultrastructural investigation of piecemeal necrosis reveals that in these areas the sinusoidal lining endothelial cells lose their fenestrae, show swelling of their cytoplasm, and accumulate electron-dense granules, assuming morphological features resembling those of the lining cells in high-endothelial venules [61,62]. Such transformed endothelial cells presumably may further change into so-called fibroblastic reticulum cells, which are thought to be involved in collagen production. As such, these phenotypic shifts of endothelial cells may represent a link between inflammation and fibroplasia in piecemeal necrosis [58,61].

Liver parenchymal cells show marked changes in expression of HLA antigens in areas of piecemeal necrosis. Normal hepatocytes do not express HLA-ABC antigens; in areas of piecemeal necrosis, however, liver parenchymal cells do display these antigens at their surface [9]. In the same areas, hyperplastic sinusoidal lining cells,

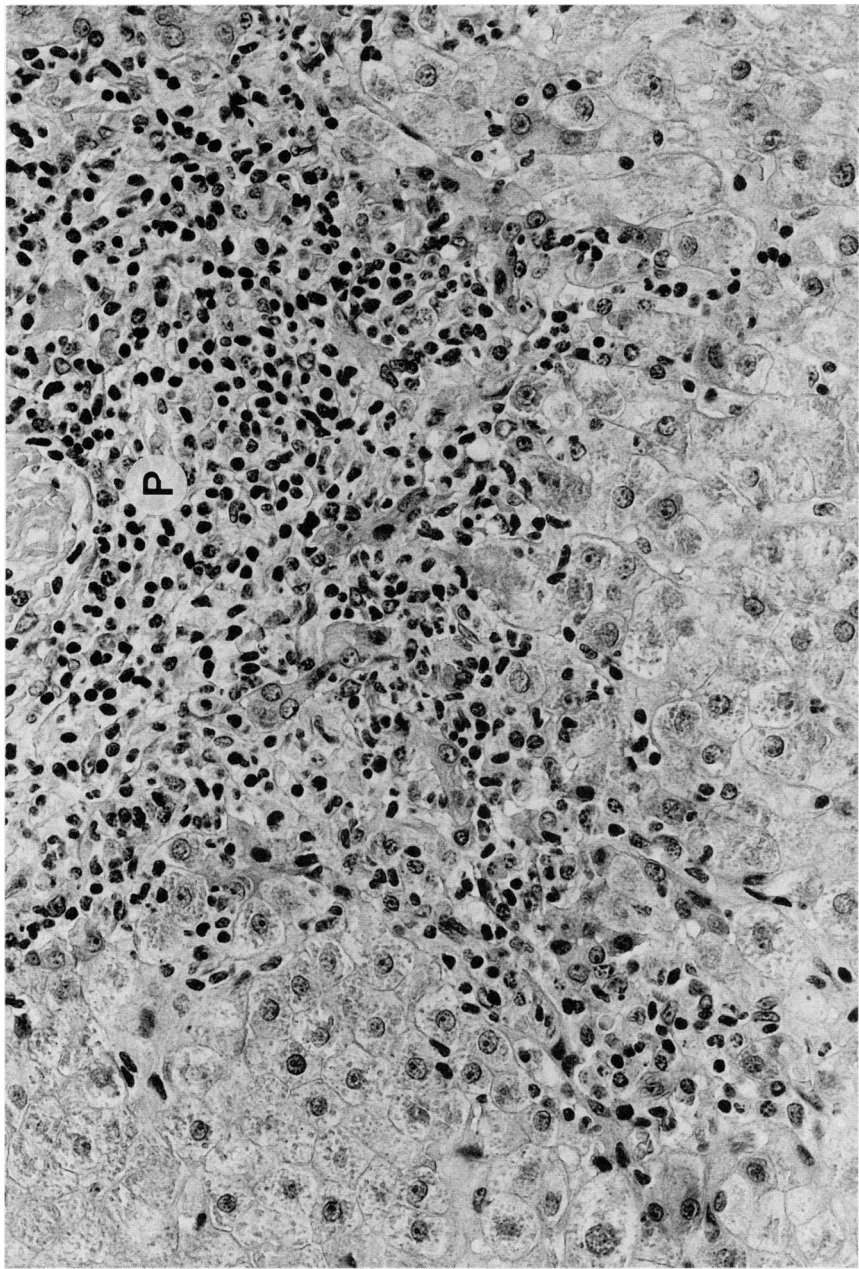


FIG. 10. Piecemeal necrosis in chronic active hepatitis B. The interphase between portal tract (P) and the lobular parenchyma is blurred by infiltration of numerous mononuclear cells, mainly lymphocytes, accompanied by loss of parenchymal cells in the same area. Hematoxylin and eosin,  $\times 400$

strongly positive for HLA class II antigens, are found to encircle hepatocytes [9]. The meaning of increased hepatocellular HLA class I antigen expression may be that nearby cytotoxic T lymphocytes have to "see" their target antigen in conjunction with HLA class I antigens; the activation and proliferation of these cytotoxic T cells may be enhanced by increased HLA-DR (class II) antigen expression on the surrounding sinusoidal lining cells [9].

The mode of cell death of the target hepatocytes in piecemeal necrosis appears to correspond to apoptosis [63].

The nature of the target antigen is not known. Apparently, it is not a viral antigen, since neither HBcAg nor HBsAg are preferentially localized in zones of piecemeal necrosis. Some studies suggested that the target antigen may be a self-antigen, termed liver membrane antigen [64]. The latter studies, however, indicate that the effector cells are not cytotoxic T cells, but rather killer cells involved in antibody-dependent cellular cytotoxicity.

It is not impossible that both T-cell cytotoxicity and antibody-dependent killer cell activity occur together in piecemeal necrosis.

### *Focal Necrosis*

Much attention has been focused on the mechanism of elimination of virus-infected cells in chronic hepatitis, causing lobular parenchymal damage and necrosis appearing as focal or spotty necrosis.

Functional immunologic studies [52,65–66] have indicated that in chronic viral hepatitis B, focal necrosis is due to cell-mediated immunity, with cytotoxic T lymphocytes functioning as effector cells against viral nucleocapsid antigen (HBcAg and/or HBeAg) as the target antigen.

Immunohistochemical studies on tissue sections [67] reveal that, in areas of focal necrosis, the infiltrating lymphocytes are OKT8+ suppressor/cytotoxic cells; hepatocytes in these foci display not only HLA class I, but also HLA class II antigens on their membrane, and often contain HBcAg in their nuclei, cytoplasm, and plasma membrane [67]. Immuno-electron microscopy allows us to confirm that HLA class II antigen [67] and HBcAg [68,69] are indeed expressed at the level of the liver cell plasma membrane.

These results are consistent with the conclusions obtained from functional immunological studies that HBcAg is a target antigen, against which effector cytotoxic T cells react when it is expressed together with HLA class I antigens at the liver cell membrane (Fig. 2). The meaning of hepatocellular HLA class II antigen expression is not readily explained, although this expression may allow the hepatocyte to present the viral antigen(s) to T lymphocytes, thus enhancing the immunological response. The "aberrant" expression of HLA class II antigens on epithelial cells has been observed in other tissues as well and indicates that antigen presentation may not be exclusively a function of antigen-presenting reticulum cells [67].

## IMMUNOMODULATION IN CHRONIC VIRAL HEPATITIS B

In addition to immunomodulation by anti-HBc antibody, further immunomodulation is brought about by interferons. Alpha interferon from different cellular sources may influence expression of viral antigens and enhance the display of HLA class I antigens [70]. Gamma interferon, produced by sensitized T lymphocytes, may induce enhanced expression of HLA class I and class II antigens on hepatocytes [12,67].

Further components in immunomodulation are immunoregulatory molecules. One such factor is a lipoprotein termed rosette inhibitory factor (RIF) [71]. Incorporation of the HBV genome into infected hepatocytes induces a state of disordered liver cell metabolism, resulting in the induction of abnormal immunoregulatory molecules (including RIF), which are responsible for inhibition of normal suppressor cell function, resulting, in turn, in antibody responses to autologous molecules such as actin, liver specific protein (LSP), and liver membrane antigen (LMA) [72] (Fig. 2). Factors such as Lex (liver extract) [71,73], LIP (liver immunoregulatory protein) [74], and another LIP (liver-derived inhibitory protein) [75] may be responsible for inhibition of cytotoxic effector lymphocytes near necrotizing hepatocytes. Such immune inhibitory effect of factors released from damaged hepatocytes may confine the inflammatory reaction within restricted tissue areas and help to explain the topographical restriction of necro-inflammatory lesions like piecemeal necrosis and focal necrosis in chronic hepatitis B.

### CIRRHOSIS

Continuous or relapsing parenchymal damage in chronic hepatitis B is followed by parenchymal regeneration. The development of active and passive septa causes disturbances in the normal spatial relationship between hepatic parenchyma and interstitial connective tissue. Parenchymal regeneration in a nodular fashion, in between the restructured or restructuring fibrous scaffold, leads to progressive development of cirrhosis (Fig. 1).

Post-hepatic cirrhosis is often of the macronodular variety, characterized by irregular size of the parenchymal nodules and a variable thickness of the fibrous septa.

Even in the cirrhotic stage, the necrotizing and inflammatory process may continue to operate in the already altered and architecturally ruined liver; this situation corresponds to "active" cirrhosis. In liver biopsies from these patients, signs of activity are reflected in the same histological features which also characterize chronic active hepatitis in the non-cirrhotic stage: that is, piecemeal necrosis, spotty necrosis, and confluent necrosis [40].

### HEPATOCELLULAR CARCINOMA

Evidence has become compelling that HBV is an important etiological factor in the causation of primary hepatocellular carcinoma [76] (Fig. 1).

Integration of viral DNA into the host DNA of some immune-protected hepatocytes is thought to represent the initiation factor in the sequence of events that finally result in the development of hepatocellular carcinoma [77]. Initially, integration is a random event, with random insertion of random viral DNA segments into cellular DNA. It is followed by gene rearrangements (duplication, deletion, and so on) of viral and host DNA, leading to altered gene expression and cellular transformation [77]. Cycles of cellular necrosis due to the hepatic process and ensuing regeneration are believed to fulfill the role of promotion [78].

After a long period of selection, a clonal proliferation of cells ensues, endowed with properties of unlimited proliferation and immortality. This stage represents the early stage of malignant growth.

It may occur in non-cirrhotic liver, as is often the case in patients from Africa and the Far East, representing areas with high endemicity of HBV, where infection often

occurs in the perinatal period. In the Western world, hepatocellular carcinoma develops more often on a background of cirrhosis. The tumor cells contain integrated viral DNA but express HBsAg only to a limited extent, and even less HBcAg.

Dysplastic liver cells represent a cellular lesion which has been considered a histological precursor lesion of malignant hepatocytes [79]. Such liver parenchymal cells are mainly characterized by enlargement, irregular size, and hyperchromatic staining of their nuclei. They are frequently observed in HBsAg-positive liver cirrhosis and may occasionally themselves express HBsAg. Whether dysplastic liver cells truly represent a precursor stage of malignant transformation of hepatocytes remains a matter of debate [80].

Early foci of hepatocellular carcinoma may be difficult to diagnose in biopsy specimens from a cirrhotic liver. It would be useful to dispose of reliable markers of malignant hepatocytes. Immunohistochemical staining for hepatic transferrin receptor has been suggested as a useful diagnostic tool [81] which merits further investigation.

### CONCLUSION

In conclusion, infection with HBV may lead to a wide spectrum of liver lesions, including diverse cytological alterations, various types and patterns of necrosis, different modes of inflammatory infiltration, normal and aberrant patterns of regeneration and hyperplasia, variable degrees and patterns of fibrosis, including the irreversible stage of liver cirrhosis, and even malignant transformation of hepatocytes.

I had the good fortune to discuss several of these lesions with Dr. Gerald Klatskin on multiple occasions, usually during coffee breaks and social parties at liver meetings. I learned a lot from his distinctive stories and detailed descriptions. Dr. G. Klatskin not only was a leader in clinical hepatology but also a most competent student with encyclopedic knowledge of microscopic liver changes.

With great affection and deep gratitude, I dedicate this review of "liver lesions in hepatitis B virus infection" to the memory of the legendary Dr. Gerald Klatskin.

### REFERENCES

1. Mac Callum FO, Bauer DJ: Homologous serum jaundice, transmission experiments with human volunteers. *Lancet* i:622-627, 1944
2. Neeffe JR, Gellis SS, Stokes J: Homologous serum hepatitis and infectious "epidemic" hepatitis; studies in volunteers bearing on immunological and other characteristics of etiological agents. *Am J Med* 1:3-22, 1946
3. Dudley FJ, Fox RA, Sherlock S: Cellular immunity and hepatitis-associated, australia antigen liver disease. *Lancet* ii:723-726, 1972
4. Mondelli M, Eddleston ALWF: Mechanisms of liver cell injury in acute and chronic hepatitis B. *Semin Liver Dis* 4:47-58, 1984
5. Thomas HC, Pignatelli M, Scully LJ: Viruses and immune reactions in the liver. *Scand J Gastroenterol* 20 (Supplement):105-117, 1985
6. Burrell CG, Gowans EJ, Rowland R, et al: Correlation between liver histology and markers of hepatitis B virus replication in infected patients: a study by in situ hybridisation. *Hepatology* 4:20-24, 1984
7. Dienstag JL: Studies of cell-mediated immunity in chronic hepatitis B virus infection: the elusive goal of virus and host antigen specificity. In *Advances in Hepatitis Research*. Edited by FV Chisari. New York, Masson Publishing, 1984, pp 163-167
8. Bianchi L: The immunopathology of acute type B hepatitis. *Springer Semin Immunopathol* 3:421-438, 1981
9. Van den Oord JJ, Desmet VJ: Verteilungsmuster der Histokompatibilitäts-Hauptantigene in normalen und pathologischen Lebergewebe. *Leber Magen Darm* 14:244-254, 1984

10. Benacerraf B: Role of MHC gene products in immune regulation. *Science* 212:1229–1238, 1981
11. Mondelli MU, Bortolotti F, Pontisso P, et al: Definition of hepatitis B virus (HBV)-specific target antigens recognized by cytotoxic T cells in acute HBV infection. *Clin Exp Immunol* 68:242–250, 1987
12. Ikeda T, Pignatelli M, Lever AML, et al: Relationship of HLA protein display to activation of 2-5 A synthetase in HBe antigen or anti-HBe positive chronic HBV infection. *Gut* 27:1498–1501, 1986
13. De Vos R, De Wolf-Peeters C, Van den Oord J, et al: Ultrastructural immunocytochemical demonstration of MHC class I antigens in human pathological liver tissue. *Hepatology* 5:1071–1075, 1985
14. Zinkernagel RM, Doherty PC: Restriction of in vitro T cell mediated cytotoxicity in lymphocyte choriomeningitis within a syngeneic or semiallogeneic system. *Nature* 248:702–705, 1974
15. Doherty PC, Allen JE: Role of the major histocompatibility complex in targeting effector T cells into a site of virus infection. *Eur J Immunol* 16:1237–1242, 1986
16. Zinkernagel RM, Haenseler E, Leist T, et al: T cell-mediated hepatitis in mice infected with lymphocytic choriomeningitis virus. Liver cell destruction by H-2 Class I-restricted virus-specific cytotoxic T cells as a physiological correlate of the <sup>51</sup>Cr-release assay? *J Exp Med* 164:1075–1093, 1986
17. Eggink HF, Houthoff HJ, Huitema S, et al: Cellular and humoral immune reactions in chronic active liver disease. II. Lymphocyte subsets and viral antigen in liver biopsies of patients with acute and chronic hepatitis B. *Clin Exp Immunol* 56:121–128, 1984
18. Chemello L, Mondelli M, Bortolotti F, et al: Natural killer activity in patients with acute viral hepatitis. *Clin Exp Immunol* 64:59–64, 1986
19. Stacey NH, Bishop CJ, Halliday JW, et al: Apoptosis as the mode of cell death in antibody-dependent lymphocytotoxicity. *J Cell Sci* 74:169–179, 1985
20. Wyllie AH: Cell death: a new classification separating apoptosis from necrosis. In *Cell Death in Biology and Pathology*. Edited by ID Bowen, RA Lockshin. London, Chapman and Hall, 1981, pp 9–34
21. MacSween NMR: Pathology of viral hepatitis and its sequelae. *Clin Gastroenterol* 9:23–46, 1980
22. Searle J, Harmon BV, Bishop CJ, et al: The significance of cell death by apoptosis in hepatobiliary disease. *J Gastroenterol Hepatol* 2:77–96, 1987
23. Mizoguchi Y, Miyajima K, Sakagami Y, et al: Detection of the cholestatic factor in the liver tissue of patients with acute intrahepatic cholestasis. *Ann Allergy* 56:304–307, 1986
24. Desmet VJ, De Vos R: Structural analysis of acute liver injury. In *Mechanism of Hepatocyte Injury and Death*. Edited by D Keppler, L Bianchi, W. Reutter. Lancaster, UK, MTP Press, 1984, pp 11–30
25. Review by an International Group: Morphological criteria in viral hepatitis. *Lancet* i:333–337, 1971
26. Review by an International Group: Acute and chronic hepatitis revisited. *Lancet* ii:914–919, 1977
27. Rappaport AM: The microcirculatory acinar concept of normal and pathological hepatic structure. *Beitr Pathol* 157:215–243, 1976
28. Conn HO: Chronic hepatitis: reducing an iatrogenic enigma to a workable puzzle. *Gastroenterology* 70:1182–1184, 1976
29. Bhakdi S, Tranum-Jensen J: Membrane damage by channel-forming proteins. *Trends Biochem Sci* 8:134–136, 1983
30. Tranum-Jensen J, Bhakdi S: Freeze-fracture analysis of the membrane lesion of human complement. *J Cell Biol* 87:618–626, 1983
31. De Groote J, Desmet VJ, Gedigk P, et al: A classification of chronic hepatitis. *Lancet* ii:626–628, 1968
32. Klatskin G: Subacute hepatic necrosis and postnecrotic cirrhosis due to anicteric infections with the hepatitis virus. *Am J Med* 25:333–358, 1958
33. Boyer JL, Klatskin G: Pattern of necrosis in acute viral hepatitis. Prognostic value of bridging (subacute hepatitis necrosis). *N Engl J Med* 283:1063–1071, 1970
34. Boyer JL: The diagnosis and pathogenesis of clinical variants in viral hepatitis. *Am J Clin Pathol* 65:898–908, 1976
35. Boyer JL: Chronic hepatitis. A perspective on classification and determinants of prognosis. *Gastroenterology* 70:1161–1171, 1976
36. Combes B: The initial morphologic lesion in chronic hepatitis, important or unimportant? *Hepatology* 6:518–522, 1986
37. Desmet V: Chronic hepatitis (including primary biliary cirrhosis). In *The Liver (IAP Monograph Number 13)*. Edited by EA Gall, FK Mostofi. Baltimore, Williams and Wilkins Co, 1972, pp 286–341
38. Scheuer PJ: Changing views on chronic hepatitis. *Histopathology* 10:1–4, 1986
39. Popper H: Changing concepts of the evolution of chronic hepatitis and the role of piecemeal necrosis. *Hepatology* 3:758–762, 1983

40. Desmet VJ: Histopathology of chronic viral hepatitis. In *Viral Hepatitis*. Edited by F Callea, M Zorzi, VJ Desmet. Berlin, Springer Verlag, 1986, pp 32–40
41. Popper H, Schaffner F: The vocabulary of chronic hepatitis. *N Engl J Med* 284:1154–1156, 1971
42. De Groote J, Fevery J, Lepoutre L: Long-term follow-up of chronic active hepatitis of moderate severity. *Gut* 19:510–513, 1978
43. Baggenstoss AH, Soloway RD, Summerskill WHJ: Chronic active liver disease. The range of histologic lesions, their response to treatment and evolution. *Hum Pathol* 3:183–198, 1972
44. Smith CI, Gregory PB: Management of patients with HBsAg positive chronic active liver disease. In *Chronic Active Liver Disease. Contemporary issues in Gastroenterology*, volume 2. Edited by S Cohen, RD Soloway. New York, Churchill Livingstone, 1983, pp 73–80.
45. Hoofnagle J, Dusheiko GM, Seel LB: Seroconversion from hepatitis B antigen to antibody in chronic type B hepatitis. *Ann Intern Med* 94:744–748, 1981
46. Liaw Y-F, Chu C-M, Su I-J, et al: Clinical and histological events preceding hepatitis B e antigen seroconversion in chronic type B hepatitis. *Gastroenterology* 84:216–219, 1983
47. Bianchi L, Gudan F: Immunopathology of hepatitis B. In *Progress in Liver Diseases*, Volume VI. Edited by H Popper, F Schaffner. New York, Grune and Stratton, 1979, pp 371–392
48. Bianchi L, Gudan F: Sanded nuclei in hepatitis B. Eosinophilic inclusions in liver cell nuclei due to excess in hepatitis B core antigen formation. *Lab Invest* 35:1–5, 1976
49. Liaw Y-F, Yang SS, Chen TJ, et al: Acute exacerbation in hepatitis B e antigen—positive chronic type B hepatitis—a clinico-pathological study. *J Hepatol* 1:227–233, 1985
50. Hadziyannis S, Gerber M, Vissoulis C, et al: Cytoplasmic hepatitis B antigen in “ground-glass” hepatocytes of carriers. *Arch Pathol* 96:327–330, 1973
51. Callea F, De Vos R, Togni R, et al: Fibrinogen inclusions in liver cells: a new type of ground-glass hepatocyte. Immune light and electron microscopic characterization. *Histopathology* 10:65–73, 1986
52. Alberti A, Trevisan A, Fattovich G, et al: The role of hepatitis B virus replication and hepatocyte membrane expression in the pathogenesis of HBV-related hepatic damage. In *Advances in Hepatitis Research*. Edited by FV Chisari. New York, Masson Publishing, 1984, pp 134–143
53. Pignatelli M, Waters J, Lever A, et al: Cytotoxic T-cell responses to the nucleocapsid proteins of HBV in chronic hepatitis. Evidence that antibody modulation may cause protracted infection. *J Hepatol* 4:15–21, 1987
54. Ackerman AB: Superficial perivascular dermatitis. In *Histologic Diagnosis of Inflammatory Skin Diseases. A Method by Pattern Analysis*. Philadelphia, Lea and Febiger, 1978, pp 203–219
55. Havrilt TJ, Ruiter DJ, Mihm MC Jr, et al: Distribution of major histocompatibility antigens in normal skin. *Br J Dermatol* 109:623–633, 1983
56. Lampert IA: Expression of HLA-DR (Ia-like) antigen on epidermal keratinocytes in human dermatoses. *Clin Exp Immunol* 57:93–100, 1984
57. Desmet VJ: Histopathology of chronic hepatitis. In *Chronic Hepatitis and Primary Biliary Cirrhosis*, AASLD Postgraduate Course. 1982, pp 155–177
58. Desmet VJ: New aspects of piecemeal necrosis. In *Trends in Hepatology*. Edited by L Bianchi, W Gerok, H Popper. Lancaster, MTP Press, 1985, pp 183–200
59. Bardadin KA, Desmet VJ: Interdigitating and dendritic reticulum cells in chronic active hepatitis. *Histopathology* 8:657–667, 1984
60. Unanue ER, Beller DI, Lu CY, et al: Opinion: Antigen presentation: comments on its regulation and mechanism. *J Immunol* 132:1–5, 1984
61. Bardadin KA, Desmet VJ: Ultrastructural observations on sinusoidal endothelial cells in chronic active hepatitis. *Histopathology* 9:171–181, 1985
62. Freemont AJ: A possible route for lymphocyte migration into diseased tissues. *J Clin Pathol* 36:161–166, 1983
63. Kerr JFR, Cooksley WGE, Searle J, et al: Hypothesis: The nature of piecemeal necrosis in chronic active hepatitis. *Lancet* ii:827–828, 1979
64. Montano L, Aranguibel F, Boffhill M, et al: An analysis of the composition of the inflammatory infiltrate in autoimmune and hepatitis B virus-induced chronic liver disease. *Hepatology* 3:292–296, 1983
65. Mondelli M, Naumov N, Eddleston ALWF: The immunopathogenesis of liver cell damage in chronic hepatitis B virus infection. In *Advances in Hepatitis Research*. Edited by FV Chisari. New York, Masson Publishing, 1984, pp 144–151
66. Thomas HC, Pignatelli M, Goodall A, et al: Immunologic mechanisms of cell lysis in hepatitis B virus infection. In *Viral Hepatitis and Liver Disease*. Edited by GN Vyas, JL Dienstag, JH Hoofnagle. Orlando, Grune and Stratton, 1984, pp 167–177



67. Van den Oord JJ, De Vos R, Desmet V: In situ distribution of major histocompatibility complex products and viral antigens in chronic hepatitis B virus infection: evidence that HBc-containing hepatocytes may express HLA-DR antigens. *Hepatology* 6:981–989, 1986
68. Kojima T, Desmet VJ: Hepatitis B core antigen (HBcAg) in liver cell plasma membrane: immunoelectron microscopic study. *Hepatology* 4:780, 1984
69. Kojima J, Bloemen J, Desmet VJ: Immune electron microscopic demonstration of hepatitis B core antigen (HBcAg) in liver cell plasma membranes. *Liver* 7:191–200, 1987
70. Montano L, Miescher GC, Goodall AH, et al: Hepatitis B virus and HLA antigen display in the liver during chronic hepatitis B virus infection. *Hepatology* 2:557–561, 1982
71. Chisari FV: Hepatic immunoregulatory molecules and the pathogenesis of hepatocellular injury in viral hepatitis. In *Advances in Hepatitis Research*. Edited by FV Chisari. New York, Masson Publishing, 1984, pp 168–178
72. Chisari FV, Edgington TS: An integrating immunoregulatory hypothesis for the immunopathogenesis of liver disease associated with hepatitis B virus infection. In *Persistent Viruses*. Edited by JC Stevens, GJ Todaro, CF Fox. New York, Academic Press, 1978, pp 499–520
73. Chisari FV, Nakamura M, Milich DR, et al: Production of two distinct and independent hepatic immunoregulatory molecules by the perfused rat liver. *Hepatology* 5:735–743, 1985
74. Brattig NW, Schrempf-Decker GE, Brockl CW, et al: Immunosuppressive serum factors in viral hepatitis. II. Further characterization of serum inhibition factor as an albumin-associated molecule. *Hepatology* 3:647–655, 1983
75. Grol M, Schumacher K: Purification and biochemical characterization of human liver-derived inhibitory protein LIP. *J Immunol* 130:323–326, 1983
76. Popper H, Gerber MA, Thung SN: The relation of hepatocellular carcinoma to infections with hepatitis B and related viruses in man and animals. *Hepatology* 2:1S–9S, 1982
77. Shafritz DA, Rogler CE: Molecular characterization of viral forms observed in persistent hepatitis infections, chronic liver disease and hepatocellular carcinoma in woodchucks and humans. In *Viral Hepatitis and Liver Disease*. Edited by GN Vyas, JL Dienstag, JH Hoofnagle. Orlando, Grune and Stratton, 1984, pp 225–243
78. Popper H: The relation between Hepatitis B virus infection and hepatocellular carcinoma. *Hepatogastroenterology* 33:2–5, 1986
79. Anthony PP, Vogel CL, Barker LF: Liver cell dysplasia: a premalignant condition. *J Clin Pathol* 26:217–233, 1973
80. Anthony PP: Liver cell dysplasia: what is its significance? *Hepatology* 7:394–395, 1987
81. Sciò R, Paterson AC, Van Eyken P, et al: Transferrin receptor expression in human hepatocellular carcinoma. An immunohistochemical study of 34 cases. *Histopathology*, in press