

First Complete Genome Sequence of a Genogroup II Genotype 18 Porcine Norovirus, Strain QW125

Tomoichiro Oka,^{a,b} Linda J. Saif,^a Qihong Wang (王秋红)^a

Food Animal Health Research Program, Ohio Agricultural Research and Development Center, Department of Veterinary Preventive Medicine, College of Veterinary Medicine, The Ohio State University, Wooster, Ohio, USA^a; Department of Virology II, National Institute of Infectious Diseases, Tokyo, Japan^b

Noroviruses, members of the family *Caliciviridae*, are genetically diverse. We report the first complete genome sequence of a genogroup II genotype 18 porcine norovirus, strain QW125. A protein BLAST search revealed that identity scores of this strain compared to other norovirus strains were highest in the predicted protease region.

Received 26 April 2013 Accepted 1 May 2013 Published 13 June 2013

Citation Oka T, Saif LJ, Wang Q. 2013. First complete genome sequence of a genogroup II genotype 18 porcine norovirus, strain QW125. *Genome Announc.* 1(3):e00344-13. doi:10.1128/genomeA.00344-13.

Copyright © 2013 Oka et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Qihong Wang, wang.655@osu.edu.

Noroviruses (NoVs) have been detected from humans, swine, cattle, sheep, rodents, cats, dogs, and lions (1–4). The NoV genome is a positive-sense, single-stranded RNA molecule with three open reading frames (ORFs). ORF1 encodes 6 to 7 nonstructural proteins (NS1–2, NS3 [NTPase], NS4, NS5 [VPg], NS6 [protease], and NS7 [polymerase]). ORF2 and ORF3 encode major (VP1) and minor (VP2) structural proteins, respectively. Based on the complete VP1 sequences, NoVs are classified into at least five genogroups (GI to GV), which are further divided into multiple genotypes (5). The GII.11, -18, and -19 NoVs were detected from swine (6); however, only two full-length genome sequences of GII.11 (AB126320 and HQ392821), and none for GII.18 and -19, are available in GenBank. We determined the first full-length genome sequence of a porcine GII.18 NoV.

The Po/NoV/GII.18/OH-QW125/2003/US strain (QW125) was detected from the feces of a finisher pig. The 3′-end 3.3-kb cDNA fragment corresponding to the partial polymerase, VP1 and VP2 genes, and 3′-untranslated region (UTR) of QW125 has been determined previously (6). In this study, the 5′-end 4.5-kb fragment corresponding to the 5′-UTR and most of ORF1 was amplified using seminested PCR with forward primer Noro-F1-F (7) and QW125-specific reverse primers designed in the polymerase gene. The 5′-end sequence of the genome was confirmed by 5′ rapid amplification of cDNA ends (5′-RACE) method. The amplified cDNA fragment was cloned into pCR4 Blunt-Topo vector (Invitrogen) and sequenced by primer walking using a set of QW125-specific primers and an automated sequencer, ABI3730 (Applied Biosystems). The full-length genome sequence was assembled using the Sequencher 4.10.1 program (GeneCodes).

The genome of QW125 consists of 7,612 nucleotides (nt), excluding the poly(A) tail. Similar to other NoVs, the genome was predicted to encode three ORFs, nucleotide positions 8 to 5083 (ORF1), 5064 to 6737 (ORF2), and 6737 to 7564 (ORF3). The 5′- and 3′-UTRs were 7 nt and 48 nt long, respectively. The ORF1-encoded polyprotein was predicted to be protease processed to NS1–2 (amino acid [aa] 1 to 327), NS3 (aa 328 to 693), NS4 (aa 694 to 867), NS5 (aa 868 to 1000), NS6 (aa 1001 to 1181), and NS7 (aa

1182 to 1691) based on the cleavage sites, characteristic motifs, and protein sizes (8). The first three aa residues of polyprotein and VP1 were MMA and MMM, respectively, and differed from those (MKM) of other complete genomes available for NoV GII strains. A protein BLAST search revealed that the highest identity scores of each region of the QW125 strain to those of other human and animal NoV strains were 63% (NS1–2), 84% (NS3), 48% (NS4), 84% (NS5), 91% (NS6), 82% (NS7), 71% (VP1), and 57% (VP2), respectively. The putative NS6 (protease) region had the highest aa identity, suggesting a strict evolutionary constraint of this region.

The accumulation of complete genome sequences of multiple genogroups and genotypes of NoVs is useful to establish new classification schemes, assess the potential interspecies transmission of NoVs, and investigate the viral protein functions, virus-host interactions, and pathogenesis of NoVs.

Nucleotide sequence accession number. The genome sequence of the QW125 strain has been deposited in GenBank under the accession no. [AY823305](https://www.ncbi.nlm.nih.gov/nuclot/AY823305).

ACKNOWLEDGMENTS

This work was supported by grants from 2011-68003-30395 (L. A. Jaykus, PD; L.J.S. and Q.W., subaward PIs) from the U.S. Department of Agriculture. Salaries and research support were provided by state and federal funds provided to the Ohio Agricultural Research and Development Center (OARDC), the Ohio State University.

Sequencing was performed at the Molecular and Cellular Imaging Center, OARDC, the Ohio State University.

REFERENCES

- Pinto P, Wang Q, Chen N, Dubovi EJ, Daniels JB, Millward LM, Buonavoglia C, Martella V, Saif LJ. 2012. Discovery and genomic characterization of noroviruses from a gastroenteritis outbreak in domestic cats in the US. *PLoS One* 7:e32739.
- Martella V, Campolo M, Lorusso E, Cavicchio P, Camero M, Bellacicco AL, Decaro N, Elia G, Greco G, Corrente M, Desario C, Arista S, Banyai K, Koopmans M, Buonavoglia C. 2007. Norovirus in captive lion cub (*Panthera leo*). *Emerg. Infect. Dis.* 13:1071–1073.
- Martella V, Lorusso E, Decaro N, Elia G, Radogna A, D'Abramo M,

- Desario C, Cavalli A, Corrente M, Camero M, Germinario CA, Bányai K, Di Martino B, Marsilio F, Carmichael LE, Buonavoglia C. 2008. Detection and molecular characterization of a canine norovirus. *Emerg. Infect. Dis.* 14:1306–1308.
4. Tse H, Chan WM, Lam CS, Lau SK, Woo PC, Yuen KY. 2012. Complete genome sequences of novel rat noroviruses in Hong Kong. *J. Virol.* 86: 12435–12436.
5. Green KY. 2007. Caliciviridae: the noroviruses, p 949–979. *In* Knipe DM, Howley PM (ed), *Fields virology*, 5th ed. Lippincott Williams & Wilkins, Philadelphia, PA.
6. Wang QH, Han MG, Cheetham S, Souza M, Funk JA, Saif LJ. 2005. Porcine noroviruses related to human noroviruses. *Emerg. Infect. Dis.* 11: 1874–1881.
7. Takanashi S, Wang Q, Chen N, Shen Q, Jung K, Zhang Z, Yokoyama M, Lindesmith LC, Baric RS, Saif LJ. 2011. Characterization of emerging GII.g/GII.12 noroviruses from a gastroenteritis outbreak in the United States in 2010. *J. Clin. Microbiol.* 49:3234–3244.
8. Oka T, Yokoyama M, Katayama K, Tsunemitsu H, Yamamoto M, Miyashita K, Ogawa S, Motomura K, Mori H, Nakamura H, Wakita T, Takeda N, Sato H. 2009. Structural and biological constraints on diversity of regions immediately upstream of cleavage sites in calicivirus precursor proteins. *Virology* 394:119–129.