

# Synthesis and In Silico Study of 4-Substituted 1-Aminoanthraquinones

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**Abstract**—Eight new 4-substituted 1-amino-9,10-anthraquinones containing a primary amino group were synthesized by nucleophilic substitution of bromine in 1-amino-4-bromo-9,10-anthraquinones. 1-Amino-4-[2-(hydroxyethyl)amino]-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid containing a biogenic amine fragment (2-aminoethanol) was converted into the corresponding 1-triazenyl derivatives. The structure of the synthesized compounds was determined on the basis of the LC/MS and <sup>13</sup>C and <sup>1</sup>H NMR data, and their drug likeness was estimated in silico. Compounds with a good drug likeness score were analyzed by DIGEP-Pred, their possible interactions with proteins were simulated using STRING, and their biological activity was interpreted using the Kyoto Encyclopedia of Genes and Genomes.

**Keywords:** bromaminic acid, Ullmann reaction, LC/MS, 4-substituted 9,10-anthraquinones, triazenes, genes

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## INTRODUCTION

Formerly, anthraquinone derivatives were historically important natural dyes. Later, it was found that the planar tricyclic anthraquinone system gives rise to a broad spectrum of biologically important properties (Fig. 1) [1–2]. At present, anthraquinone derivatives are extensively studied as therapeutic agents against COVID-19, specifically acting against 3CLpro and PLpro proteases [3].

A large number of anthraquinone derivatives contain a sulfonate group in the 2-position. These compounds can be synthesized from bromaminic acid sodium salt **1** (Fig. 2) which is the basic starting material for the preparation of biologically active anthraquinone derivatives and numerous dyes [4–6]. In fact, bromaminic acid and its salts are widely used as intermediate products for the synthesis of anthraquinone derivatives, including acid dyes, via substitution of the 4-bromine atom by (aryl)alkylamino group [7–9].

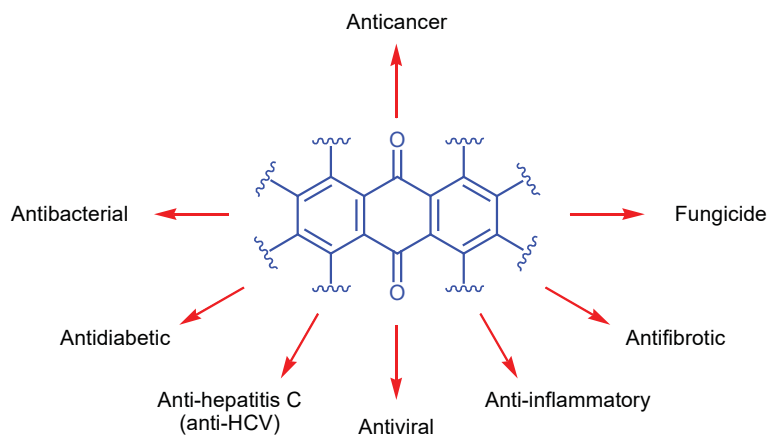
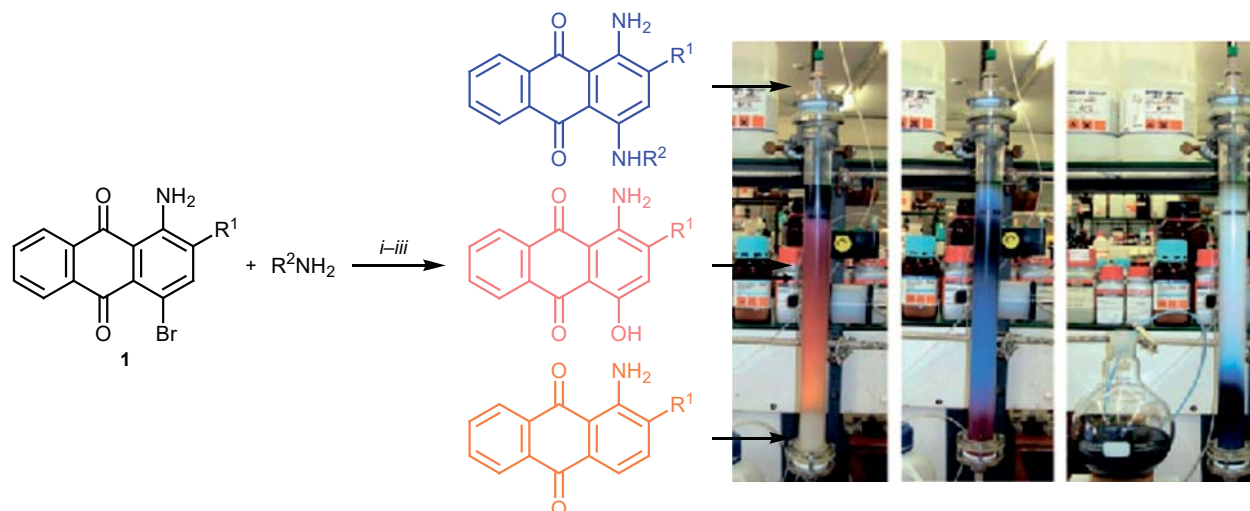


Fig. 1. Biological activity of anthraquinone derivatives.



$R^1 = \text{SO}_3\text{Na}$ ,  $R^2 = \text{Alk}$ , Ar; *i*:  $\text{CuCl}$ ,  $\text{Na}_2\text{CO}_3$ ,  $\text{Na}_2\text{SO}_3$ ,  $\text{H}_2\text{O}$ , r.t., 8–24 h or  $120^\circ\text{C}$ , 8–10 h; *ii*:  $\text{CuSO}_4$ ,  $\text{Na}_2\text{CO}_3$ ,  $\text{H}_2\text{O}$ ,  $120^\circ\text{C}$ , 12–48 h; *iii*: phosphate buffer, pH 6–7,  $\text{Cu}(0)$ , microwave irradiation,  $100\text{--}120^\circ\text{C}$ , 5–24 min.

**Fig. 2.** Synthesis of 4-substituted 1-aminoanthraquinones.

There are almost no published data on chemical properties of triazenyl-substituted anthraquinones [10, 11]. Triazene moiety is a known alkylating carcinolytic group. It was introduced into anthraquinone molecule by azo coupling of anthraquinone-1-diazonium salt with various aliphatic and aromatic amines.

## RESULTS AND DISCUSSION

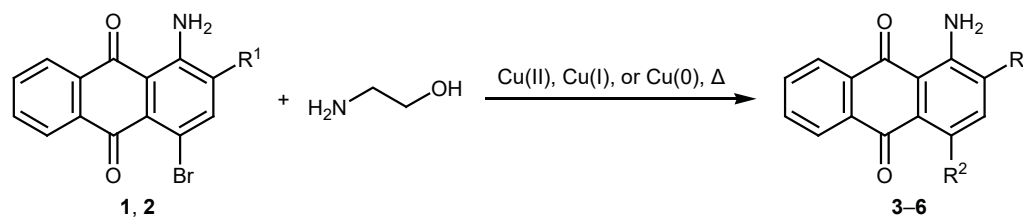
While developing an optimal procedure for the substitution of bromine in bromaminic acid **1**, we tried different known nucleophilic substitution methods [12, 13] and performed a series of reactions of bromaminic acid **1** and its 2-methyl analog **2** with 2-aminoethanol under different conditions; however, the yields were not always satisfactory (Scheme 1). We found that the most efficient procedure was to react compound **1** or **2** with 2-aminoethanol in aqueous medium in the presence of a mixture of copper(II) and iron(II) salts [14]. In this case, the yield of target 1-amino-4-[(2-hydroxyethyl)amino]-9,10-dioxo-9,10-dihydroan-

thracene-2-sulfonic acid (**5**) was 96% ( $m/z$  362.0 [ $M + H$ ]<sup>+</sup>). The structure of **5** was confirmed by <sup>1</sup>H and <sup>13</sup>C NMR spectra and elemental analysis. Compound **6** was obtained in 65% yield, and its <sup>1</sup>H NMR spectrum showed signals of the ethylene moiety at  $\delta$  3.50 and 3.67 ppm. Under these conditions, bromaminic acid **1** was reacted with other primary aliphatic amines to obtain 1,4-diaminoanthraquinone derivatives **7–12** (Scheme 2).

Almost all these reactions were accompanied by side formation of 1-amino-4-hydroxy-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid (**4**) due to concurrent attack of hydroxy nucleophile (Table 1). Furthermore, there was a clear relation between the  $pK_a$  value of the amine and the purity of the product (Scheme 2).

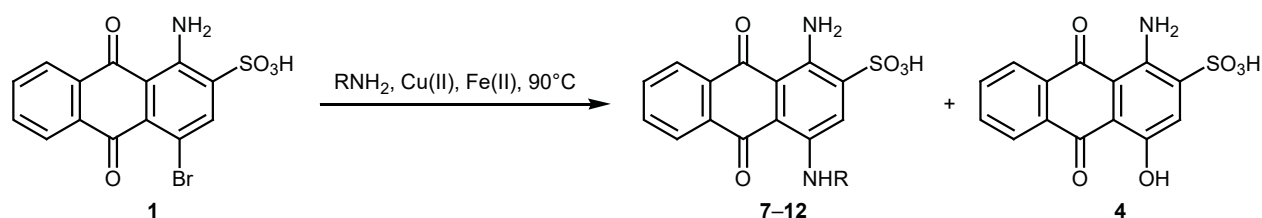
**Synthesis of triazenes.** In the next step of our study, aminoanthraquinone **5** was subjected to diazotization with sodium nitrite in aqueous medium in the presence of HCl at  $0\text{--}5^\circ\text{C}$  [15–16] (Scheme 3). The subsequent azo coupling of diazonium salts **13** with secondary and primary amines afforded triazenes **14–**

**Scheme 1.**



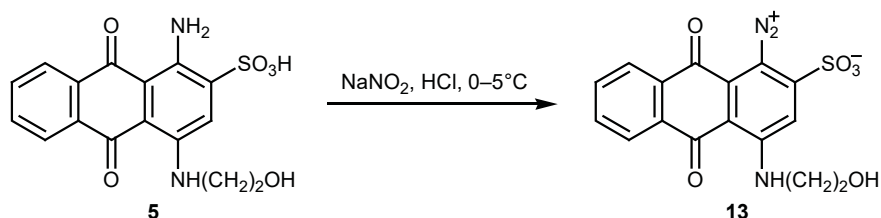
**1**,  $R^1 = \text{SO}_3\text{H}$ ; **2**,  $R^1 = \text{Me}$ ; **3**,  $R^1 = \text{SO}_3\text{H}$ ,  $R^2 = \text{H}$ ; **4**,  $R^1 = \text{SO}_3\text{H}$ ,  $R^2 = \text{OH}$ ;  
**5**,  $R^1 = \text{SO}_3\text{H}$ ,  $R^2 = \text{NHCH}_2\text{CH}_2\text{OH}$ ; **6**,  $R^1 = \text{Me}$ ,  $R^2 = \text{NHCH}_2\text{CH}_2\text{OH}$ .

Scheme 2.

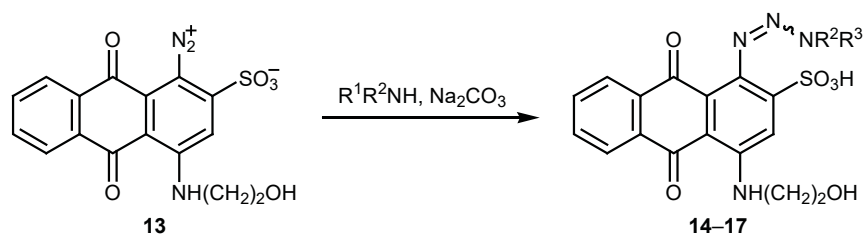


7, R = *cyclo*-C<sub>5</sub>H<sub>9</sub> (65%, p*K*<sub>a</sub> = 10.5); 8, R = *i*-Pr (68%, p*K*<sub>a</sub> = 10.6); 9, R = furan-2-ylmethyl (84%, p*K*<sub>a</sub> = 10.62); 10, R = morpholin-4-ylamino (85%, p*K*<sub>a</sub> = 10.67); 11, R = Pr (98%, p*K*<sub>a</sub> = 10.71); 12, R = Bu (100%, p*K*<sub>a</sub> = 10.77).

Scheme 3.



Scheme 4.



14, R<sup>1</sup> = R<sup>2</sup> = CH<sub>2</sub>CH<sub>2</sub>OH; 15, R<sup>1</sup>R<sup>2</sup>N = morpholin-4-yl; 16, R<sup>1</sup> = R<sup>2</sup> = Et; 17, R<sup>1</sup> = H, R<sup>2</sup> = 4-HOC(O)C<sub>6</sub>H<sub>4</sub>.

17 (Scheme 4). This reaction was not always smooth, depending on the properties of the initial amine and stability of the diazo coupling products. The structure of 14–17 was confirmed by spectral data. The <sup>1</sup>H NMR spectra of 14–17 showed aromatic protons signals in the region δ 7.50–8.70 ppm.

**Biological activity and drug likeness.** Among the 12 synthesized compounds, we identified triazene derivatives with good drug likeness scores. In particular, 4-aminobenzoic acid derivative 17 (*M* 510.08) showed the highest drug likeness score (+0.06). Table 2

contains the detailed drug likeness parameters of the synthesized compounds.

Triazene 14 modulated the largest number of genes (10). The effect on the CHEK1 gene responsible for the p53 signaling pathway directly involved in immune strengthening was revealed. Furthermore, the gene set enrichment analysis showed modulation of 15 different biological pathways, among which cancer pathways are modulated mainly by regulation of three genes (SP1, IL23A, NFE2L2). The results of gene set enrichment analysis of protein modulation by anthraquinone

Table 1. Synthesis of compounds 7–12

Compound no.	Yield, %	Impurity of 4, %
7	65	25
8	68	20
9	84	15
10	85	5
11	98	1
12	100	–

**Table 2.** Drug likeness parameters of compounds 5–17<sup>a</sup>

Compd. no.	Formula	Molecular weight	NHBA	NHBD	Log <i>P</i>	TPSA, Å <sup>2</sup>	<i>V</i> , Å <sup>3</sup>	DLS
5	C <sub>16</sub> H <sub>14</sub> N <sub>2</sub> O <sub>6</sub> S	362.06	6	5	-0.57	114.29	307.29	-0.73
6	C <sub>17</sub> H <sub>16</sub> N <sub>2</sub> O <sub>3</sub>	296.12	3	4	2.76	71.87	288.83	-0.16
7	C <sub>19</sub> H <sub>18</sub> N <sub>2</sub> O <sub>5</sub> S	386.09	5	5	1.40	97.61	340.44	-0.19
8	C <sub>17</sub> H <sub>16</sub> N <sub>2</sub> O <sub>5</sub> S	360.08	5	4	0.72	96.76	317.25	-0.66
9	C <sub>19</sub> H <sub>18</sub> N <sub>2</sub> O <sub>6</sub> S	402.09	6	4	0.24	106.48	355.56	-0.17
10	C <sub>18</sub> H <sub>17</sub> N <sub>3</sub> O <sub>6</sub> S	403.08	7	4	-0.53	110.07	353.50	-0.27
11	C <sub>17</sub> H <sub>16</sub> N <sub>2</sub> O <sub>5</sub> S	360.08	5	4	0.82	97.63	317.22	-0.57
12	C <sub>18</sub> H <sub>18</sub> N <sub>2</sub> O <sub>5</sub> S	374.09	5	4	1.34	97.63	335.12	-0.69
14	C <sub>18</sub> H <sub>18</sub> N <sub>4</sub> O <sub>8</sub> S	450.08	10	5	-0.74	150.24	380.77	-0.79
15	C <sub>20</sub> H <sub>22</sub> N <sub>4</sub> O <sub>6</sub> S	446.13	8	3	1.18	117.64	399.55	-0.60
16	C <sub>20</sub> H <sub>20</sub> N <sub>4</sub> O <sub>7</sub> S	460.11	9	3	0.04	126.10	402.62	-0.23
17	C <sub>23</sub> H <sub>18</sub> N <sub>4</sub> O <sub>8</sub> S	510.08	10	5	1.01	155.40	434.50	0.06

<sup>a</sup> NHBA is the number of hydrogen bond acceptor centers, NHBD is the number of hydrogen bond donor centers, Log*P* is the lipophilicity coefficient, TPSA is the topological polar surface area, *V* is the molecular volume, and DLS is the drug likeness score.

**Table 3.** Analysis of proteins modulated by anthraquinone derivatives

Pathway ID	Description	Number of genes	Matching genes
hsa05200	Pathways in cancer	3	SP1, IL23A, NFE2L2
hsa05164	Influenza A	2	NXT2, SP1
hsa05133	Pertussis	2	SP1, IL23A
ko04625	C-Type lectin receptor signaling pathway	2	SP1, IL23A
hsa05166	HTLV-I infection	1	CHEK1
hsa05014	Amyotrophic lateral sclerosis (ALS)	1	SP1
hsa05152	Tuberculosis	1	IL23A
hsa05224	Breast cancer	1	SP1
hsa05204	Chemical carcinogenesis	1	CBR1
hsa05225	Hepatocellular carcinoma	1	NFE2L2
hsa05203	Viral carcinogenesis	1	CHEK1
hsa04216	Ferroptosis	1	GSS
hsa05323	Rheumatoid arthritis	1	IL23A
hsa01524	Platinum drug resistance	1	TOP2A
hsa04115	p53 signaling pathway	1	CHEK1



**1-Amino-4-(butylamino)-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid (12).** Yield 100%, blue solid, mp 290–292°C. <sup>1</sup>H NMR spectrum,  $\delta$ , ppm: 1.00 s (3H, CH<sub>3</sub>), 1.60 s (2H, CH<sub>2</sub>), 7.80 s (3H, H<sub>arom</sub>), 8.20 s (2H, H<sub>arom</sub>), 10.70 s (1H, NH). Mass spectrum:  $m/z$ : 375.5 [ $M + H$ ]<sup>+</sup>. C<sub>18</sub>H<sub>18</sub>N<sub>2</sub>O<sub>5</sub>S. *M* 375.

**1-(Diazyn-1-ium-1-yl)-4-[(2-hydroxyethyl)amino]-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid (13).** A 50-mL round-bottom flask equipped with a magnetic stirrer was charged with a solution of anthraquinone **5** (0.1 mmol) in 1 M aqueous HCl (5.0 mL). The solution was cooled to 0–5°C in an ice bath, a solution of sodium nitrite (0.2 mmol) in 0.5 mL of distilled water was added dropwise, maintaining the temperature at 0–5°C, and the mixture was stirred for 5 min at that temperature.

**Triazene derivatives 14–17 (general procedure).** The mixture containing diazonium salt **13** was allowed to warm up to room temperature, a solution of the corresponding amine (0.15 mmol) in 5 mL of ethanol was added, and the mixture was stirred for ~30 s at room temperature. The progress of the reaction was monitored by change of the color of the reaction mixture (the color changed from blue to red after diazotization, and the final product was purple), as well as by RP-TLC using acetone–water (2:3) as eluent. The product was purified by reversed phase column chromatography (RP-18) using water as eluent.

**1-[3,3-Bis(2-hydroxyethyl)triaz-1-en-1-yl]-4-[(2-hydroxyethyl)amino]-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid (14).** Yield 80%. <sup>1</sup>H NMR spectrum,  $\delta$ , ppm: 2.99 s (4H, CH<sub>2</sub>), 3.40 d (4H, CH<sub>2</sub>,  $J = 11.2$  Hz), 3.65 s (4H, CH<sub>2</sub>), 5.08 s (OH), 5.25 s (2H, OH), 7.85 s (2H, H<sub>arom</sub>), 8.15–8.20 m (2H, H<sub>arom</sub>), 8.69 s (2H, H<sub>arom</sub>), 9.87 s (1H, NH). Mass spectrum:  $m/z$  360.9 [ $M + H$ ]<sup>+</sup>. C<sub>16</sub>H<sub>12</sub>N<sub>2</sub>O<sub>6</sub>S. *M* 361.

**4-[(2-Hydroxyethyl)amino]-1-[(E)-(morpholin-4-yl)diazonyl]-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid (15).** Yield 90%. mp >300°C. <sup>1</sup>H NMR spectrum,  $\delta$ , ppm: in DMSO-*d*<sub>6</sub>: 3.37 t (1H, CH<sub>2</sub>, morpholine), 3.43 t (2H, CH<sub>2</sub>, morpholine), 3.70–3.75 m (3H, CH<sub>2</sub>, morpholine), 4.95 s (5H, CH<sub>2</sub>), 7.62 s (1H, 3-H), 7.86 s (1H, H<sub>arom</sub>), 7.92 t (1H, H<sub>arom</sub>,  $J = 8.0$  Hz), 8.17 d (1H, H<sub>arom</sub>,  $J = 6.4$  Hz), 8.23 d (1H, H<sub>arom</sub>,  $J = 7.6$  Hz), 9.89 s (1H, OH); in DMSO-*d*<sub>6</sub>–CCl<sub>4</sub>: 3.46 s (2H, CH<sub>2</sub>), 3.73 s (1H, CH<sub>2</sub>), 7.67 d (1H, 3-H,  $J = 6.8$  Hz), 7.87–7.90 m (2H, H<sub>arom</sub>), 8.18 t (1H, H<sub>arom</sub>,  $J = 6.0$  Hz), 8.28 d (1H, H<sub>arom</sub>,  $J = 4.8$  Hz), 9.90 s (1H, OH). Mass spectrum:  $m/z$  460 [ $M$ ]<sup>+</sup>.

**1-(3,3-Diethyltriaz-1-en-1-yl)-4-[(2-hydroxyethyl)amino]-9,10-dioxo-9,10-dihydroanthracene-2-sulfonic acid (16).** Yield 52%. <sup>1</sup>H NMR spectrum,  $\delta$ , ppm: 1.18 s (3H, CH<sub>3</sub>), 2.86 d (3H, CH<sub>3</sub>,  $J = 4.8$  Hz), 3.40 s (2H, CH<sub>2</sub>), 7.66–7.70 m (1H, 3-H), 8.04–8.30 m (8H, H<sub>arom</sub>), 9.01 s (1H, OH). Mass spectrum:  $m/z$  425.9 [ $M$ ]<sup>+</sup>.

**4-(3-{4-[(2-Hydroxyethyl)amino]-2-sulfo-9,10-dioxo-9,10-dihydroanthracen-1-yl}triaz-2-en-1-yl)-benzoic acid (17).** Yield 95%, mp >300°C. <sup>1</sup>H NMR spectrum,  $\delta$ , ppm: 1.10 s (1H, CH<sub>2</sub>), 3.46–3.50 m (2H, CH<sub>2</sub>,  $J = 16.0$  Hz), 7.20 d (8H, H<sub>arom</sub>,  $J = 7.6$  Hz), 7.80 d (3H, H<sub>arom</sub>,  $J = 7.6$  Hz), 7.90 s (4H, NH, OH). Mass spectrum:  $m/z$  466 [ $M - C_{22}H_{17}O_6CH_4$ ]<sup>+</sup>.

## CONCLUSIONS

The results of computer simulation suggest the possibility of therapeutic effect of the synthesized substituted anthraquinone derivatives, which needs to be verified using experimental protocols.

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## CONFLICT OF INTEREST

The authors declare the absence of conflict of interest.

## SUPPLEMENTARY INFORMATION

The online version contains supplementary material available at <https://doi.org/10.1134/S1070428021040126>.

## REFERENCES

- Hussain, H., Al-Harrasi, A., Al-Rawahi, A., Green, I., Csuk, R., Ahmed, I., Shan, A., Abbas, G., Rehman, N., and Ullah, R., *Expert Opin. Ther. Pat.*, 2015, vol. 25, p. 1053. <https://doi.org/10.1517/13543776.2015.1050793>
- Malik, E. and Muller, C., *Med. Res. Rev.*, 2016, vol. 36, p. 705. <https://doi.org/10.1002/med.21391>
- Khanal, P., Patil, B.M., Chand, J., and Naaz, Y., *Nat. Prod. Bioprospect.*, 2020, vol. 10, p. 325. <https://doi.org/10.1007/s13659-020-00260-2>
- Baqi, Y. and Muller, C.E., *Org. Lett.*, 2007, vol. 9, p. 1271. <https://doi.org/10.1021/ol070102v>

5. Baqi, Y. and Muller, C.E., *Nat. Protoc.*, 2010, vol. 5, p. 945.  
<https://doi.org/10.1038/nprot.2010.63>
6. Baqi, Y., Alzeler, K., Koze, M., and Muller, C.E., *J. Med. Chem.*, 2009, vol. 52, p. 3784.  
<https://doi.org/10.1021/jm9003297>
7. Malik, E.M., Baqi, Y., and Müller, C.E., *Beilstein J. Org. Chem.*, 2015, vol. 11, p. 2326.  
<https://doi.org/10.3762/bioci.11.253>
8. Roy, S., Large, J.R., Akande, A.M., Kshatri, A., Webb, T.I., Domene, C., Sergeant, G.P., Mchale, N.G., Thornbury, K.D., and Hollywood, M.A., *Eur. J. Med. Chem.*, 2004, vol. 75, p. 426.  
<https://doi.org/10.1016/j.ejmech.2014.01.035>
9. Fiene, A., Baqi, Y., Malik, E.M., Newton, P., Li, W., Lee, S.-Y., Hartland, L.E., and Muller, C.E., *Bioorg. Med. Chem.*, 2016, vol. 24, p. 4363.  
<https://doi.org/10.1016/j.bmc.2016.07.027>
10. Sabadakh, O.P., Taras, T.N., Luchkevich, E.R., and Novikov, V.P., *Russ. J. Org. Chem.*, 2015, vol. 51, p. 277.  
<https://doi.org/10.1134/S1070428015020244>
11. Bulgakova, N.A. and Gornostaev, L.M., *J. Org. Chem.*, 2001, vol. 37, p. 1351.  
<https://doi.org/10.1023/A:1013164528653>
12. Topanov, A.P., Mashevskaya, I.V., Dmitriev, M.V., and Maslivets, A.N., *Russ. J. Org. Chem.*, 2020, vol. 56, p. 719.  
<https://doi.org/10.1134/S1070428020040247>
13. Kaur, G., Utreja, D., Jain, N., and Dhillon, N.K., *Russ. J. Org. Chem.*, 2020, vol. 56, p. 113.  
<https://doi.org/10.1134/S1070428020010182>
14. Shupenyuk, V.I., Taras, T.N., Sabadakh, O.P., Luchkevich, E.R., and Kornii, Y., *Fr.-Ukr. J. Chem.*, 2020, vol. 8, p. 58.  
<https://doi.org/10.17721/fujcV8I1P58-65>
15. Baqi, Y. and Muller, C.E., *Tetrahedron Lett.*, 2012, vol. 53, p. 6739.  
<https://doi.org/10.1016/j.tetlet.2012.09.011>
16. Glushkova, M.A., Popkov, S.V., and Burdeinyi, M.L., *Russ. J. Org. Chem.*, 2020, vol. 56, p. 390.  
<https://doi.org/10.1134/S1070428020030045>
17. Lagunin, A., Ivanov, S., Rudik, A., Filimonov, D., and Poroikov, V., *Bioinformatics*, 2013, vol. 29, p. 2062.  
<https://doi.org/10.1093/bioinformatics/btt322>
18. Szklarczyk, D., Morris, J.H., Cook, H., Kuhn, M., Wyder, S., Simonovic, M., Santos, A., Doncheva, N.T., Roth, A., Bork, P., Jensen, L.J., and von Mering, C., *Nucleic Acids Res.*, 2017, vol. 45, p. D362.  
<https://doi.org/10.1093/nar/gkw937>