Research Article



Oxidative stress induces BH₄ deficiency in male, but not female, SHR

Ellen E. Gillis¹, Krystal N. Brinson¹, Olga Rafikova², Wei Chen³, Jacqueline B. Musall¹, David G. Harrison³ and Jennifer C. Sullivan¹

¹Department of Physiology, Augusta University, Augusta, GA, U.S.A.; ²Department of Medicine, University of Arizona, Tucson, AZ, U.S.A.; ³Division of Clinical Pharmacology, Department of Medicine, Vanderbilt University, Nashville, TN, U.S.A.

Correspondence: Jennifer C. Sullivan (jensullivan@augusta.edu)



We previously published that female spontaneously hypertensive rats (SHR) have significantly greater nitric oxide (NO) bioavailability and NO synthase (NOS) enzymatic activity in the renal inner medulla (IM) compared with age-matched males, although the mechanism responsible remains unknown. Tetrahydrobiopterin (BH₄) is a critical cofactor required for NO generation, and decreases in BH₄ as a result of increases in oxidative stress have been implicated in the pathogenesis of hypertension. As male SHR are known to have higher levels of oxidative stress compared with female SHR, we hypothesized that relative BH₄ deficiency induced by oxidative stress in male SHR results in lower levels of NOS activity in renal IM compared with females. Twelve-week-old male and female SHR were randomized to receive tempol (30 mg/kg/day via drinking water) or vehicle for 2 weeks. Tempol treatment did not affect blood pressure (BP) in either sex, but reduced peroxynitrite levels only in males. Females had more total biopterin, dihydrobiopterin (BH₂), and BH₄ levels in renal IMs than males, and tempol treatment eliminated these sex differences. Females had greater total NOS activity in the renal IM than males, and adding exogenous BH₄ to the assay increased NOS activity in both sexes. This sex difference in total NOS and the effect of exogenous BH₄ were abolished with tempol treatment. We conclude that higher oxidative stress in male SHR results in a relative deficiency of BH₄ compared with females, resulting in diminished renal NOS activity in the male.

Introduction

The nitric oxide (NO)/NO synthase (NOS) pathway is critical in blood pressure (BP) regulation [1-3]. Deficiencies in NO are correlated with the incidence and progression of hypertension [1,4-6] and in particular, renal NOS has been shown to be important in modulating BP [7]. The renal inner medulla (IM) has the highest amount of NOS protein expression and enzymatic activity in the kidney [8] and NO regulates inner medullary blood flow and inhibits transport of sodium chloride along the nephron [9,10]. Moreover, both clinical and experimental studies have documented greater NO production and bioavailability in females compared with males. Previously, we published that the renal IM is the only section of the kidney to exhibit sex differences in NOS enzymatic activity in young adult (13 weeks old) spontaneously hypertensive rats (SHR) with greater total NOS enzymatic activity in female SHR compared with males [11-16]. We further showed that female SHR exhibit a sex hormone- and BP-dependent increase in NOS activity with maturation that is not observed in males [15], although why NOS activity does not increase with maturation in males remains unknown. Elucidating the molecular mechanism(s) driving the sexual dimorphism in renal IM NOS activity may provide insight into sex differences in not only the NO action in the kidney, but also BP regulation.

NOS catalyzes the formation of NO from L-arginine and oxygen in a reaction that requires a number of cofactors, including tetrahydrobiopterin (BH₄). In the absence of BH₄, electron flow to molecular

Accepted Manuscript Online: 13 June 2018 Version of Record published: 3 July 2018



oxygen is 'uncoupled' from L-arginine oxidation and NO formation instead resulting in the production of superoxide. In addition to decreasing NO production, superoxide is highly reactive with NO, resulting in the formation of peroxynitrite. Peroxynitrite rapidly oxidizes BH_4 to dihydrobiopterin (BH_2), and as BH_2 is not a NOS cofactor, it can competitively inhibit the binding of BH_4 to NOS [17,18]. As a result, NOS uncoupling serves as both a cause and effect of BH_4 deficiencies. Furthermore, decreases in BH_4 and uncoupled NOS have been implicated in numerous cardiovascular diseases, including hypertension [19,20].

Based on the finding that male SHR have greater levels of oxidative stress compared with female SHR [21,22], coupled with lower levels of NOS activity [15,16], the goal of the current study was to test the hypothesis that relative oxidative stress-induced BH_4 deficiency in male SHR is responsible for lower levels of NOS activity in renal IM compared with females.

Experimental

Animals

Twelve-week-old male and female SHR (Envigo, Inc., Indianapolis, Indiana) were studied. All experiments were conducted in accordance with the National Institutes of Health *Guide for the Care and Use of Laboratory Animals* and approved and monitored by the Augusta University Institutional Animal Care and Use Committee. Male and female SHR were randomized to receive tempol, a reactive oxygen species scavenger (30 mg/kg/day; Sigma–Aldrich; n=8-9) or vehicle (tap water; n=11-12) in their drinking water for 2 weeks. Water intake was measured daily and rats were weighed every 3 days to maintain appropriate dosing throughout the study. A separate set of male and female SHR were implanted with telemetry devices for the continuous measurement of BP (n=5). Rats were implanted with telemetry devices for the continuous measurement of 12-week-old male SHR were randomized to receive BH₄ supplementation (20 mg/kg/day; Axxora, LLC, San Diego, CA) or vehicle (saline) via daily ip injection for 1 week (n=8/group). Systolic BP was measured in all rats prior to initiating treatment and following 7 days of treatment via tail-cuff plethsomyography as previously described [24]. For all studies, rats were anesthetized with ketamine/xylazine (50 mg/kg/10 mg/kg, i.p.), and a terminal blood sample was taken and centrifuged for collection of plasma. Kidneys were removed and the renal IM were isolated and snap-frozen in liquid nitrogen.

Dot blot analysis

Plasma samples were diluted and 1.5 μ l diluted plasma was applied to a nitrocellulose membrane forming a dot and allowed to dry overnight. Membranes were then blocked with 1% BSA in TBS, 0.1% Tween 20 (TBST) for 1 h and incubated with anti-3-nitrotyrosine (3-NT) antibody at a final concentration of 1 mg/ml (Calbiochem) at room temperature for 2 h. The membrane was washed with TBST, incubated with secondary antibody, washed again, and imaged using an Odyssey Imaging System (LI-COR Biosciences, Lincoln, NE). To quantitate the total protein in each sample, the membrane was incubated with Ponceau S BioReagent for 15 min. The data were presented as a ratio of 3-NT signal per total protein signal.

NOS enzymatic activity assay

The renal IMs were homogenized as previously described [25] and the whole homogenate was then used in the NOS activity assay in the presence or absence of exogenous 3 μ M BH₄ as previously described [16]. Briefly, total NOS activity was determined based on the rate of L-[³H]citrulline formation from L-[³H]arginine and defined as [³H]arginine to [³H]citrulline conversion inhibited by the nonselective NOS inhibitor *N*-nitro-L-arginine (L-NNA; 1 mmol/l).

Biopterin analysis

Biopterins were measured by HPLC. Briefly, renal IMs were homogenized in 300 μ l ice-cold Lysis buffer (50 mM Tris/HCl, pH 7.4, 1 mM DTT, 1 mM EDTA). Two hundred seventy microliters of extract was added to 30 μ l of a 1:1 mixture of 1.5 M HCLO₄ and 2 M H₃PO₄. Concentrations of BH₄ and BH₂ were determined using HPLC and differential oxidation as previously described [26]. Samples were then centrifuged for 10 min at 14000 *g* at 4°C, and the resulting precipitate was re-suspended in 100 μ l of 1 M NaOH and analyzed for total protein content (Bradford assay) as previously described [26].



Western blot analysis

Renal IMs were homogenized as previously described [25] and the whole homogenate was used in Western blot analysis as previously described [16]. Briefly, protein expression was determined using two-color immunoblots using primary antibodies to GTP cyclohydrolase-1 (GTPCH1; 1:250, Santa Cruz Biotechnology, Santa Cruz, CA, 100 μ g protein/well) and β -actin (A1978, 1:10000; Sigma, St. Louis, MO). Protein concentrations were determined by standard Bradford assay (Bio-Rad, Hercules, CA) using BSA as the standard. β -actin was used to verify equal protein loading, and data were reported normalized to β -actin.

Statistical analysis

All data are expressed as means \pm S.E.M. BP was analyzed using repeated-measures ANOVA to examine within-group effects and Student's *t* test to examine between-group effects. Biopterin levels between control and BH₄ treated male SHR were analyzed using Student's *t* test. All other data were compared using a two-way ANOVA followed by a Newman–Keul's post-hoc. For all comparisons, *P*<0.05 was considered statistically significant. Analyses were performed using GraphPad Prism version 7.0 software (GraphPad Software Inc, La Jolla, CA).

Results

Tempol decreases measures of oxidative stress independent of BP

BP was measured by telemetry in male and female SHR treated with tempol; Figure 1A. Male SHR had a higher BP compared with female SHR at baseline (140 \pm 2 compared with 132 \pm 2; *P*=0.013). BP remained higher in males throughout the treatment period, although tempol had no effect on BP in either sex (effect of tempol in males, *P*=0.25; effect of tempol in females, *P*=0.096).

Peroxynitrite is formed by the binding of NO and superoxide, and BH₄ is a target for oxidation by peroxynitrite leading to uncoupled NOS [17,18]. Peroxynitrite reacts with tyrosine residues in proteins resulting in the formation of 3-NT [27,28]. Therefore, plasma 3-NT levels were measured in vehicle and tempol-treated male and female SHR using Dot blot analysis; Figure 1B. Male SHR had greater levels of 3-NT than females (effect of sex, P=0.044; Figure 1B). Treatment with tempol reduced 3-NT levels only in males, abolishing the sex difference (effect of tempol, P<0.0001; interaction, P=0.014).

Tempol treatment abolishes the sex difference in IM NOS activity and dependency on exogenous BH_4

The measurement of NOS enzymatic activity via detection of the formation of radiolabeled citrulline (and NO) from arginine is typically performed in the presence of excess amounts of all NOS cofactors, including BH₄. In the current study, we measured NOS enzymatic activity in the renal IM of vehicle control and tempol-treated male and female SHR in the absence and presence of exogenous BH₄; Figure 2. Consistent with our previous publications [15,16], total NOS enzymatic activity was lower in the renal IM of control, vehicle-treated male SHR compared with female SHR (effect of sex, P=0.0004). The inclusion of BH₄ in the assay increased NOS activity in both sexes, and the increase was comparable in males (36 ± 5% increase) and females (30 ± 4% increase; effect of BH₄, P=0.0008; interaction, P=0.55). In contrast, following treatment with tempol there was no sex difference in NOS activity (effect of sex, P=0.36) and inclusion of exogenous BH₄ did not significantly increase NOS activity in either sex (effect of BH₄, P=0.11; interaction, P=0.79).

Male SHR have lower biopterin levels in the renal IM compared with female SHR

Total biopterin, BH₂ and BH₄ levels were quantitated in IM from vehicle control and tempol treated male and female SHR via HPLC analysis; Figure 3. Males have lower levels of total biopterins (effect of sex, P=0.0007), BH₂ (effect of sex, P=0.0021), and BH₄ compared with females (effect of sex, P=0.029). Sex differences in biopterins, BH₂, and BH₄ were abolished by tempol treatment. Treatment with tempol increased total biopterins only in males (effect of tempol, P=0.0013; interaction, P=0.065). BH₂ levels were increased by tempol in both sexes (effect of tempol, P<0.0001; interaction, P=0.643), while BH₄ levels increased only in male SHR with tempol (effect of tempol, P=0.7722; interaction, P=0.0031).





Figure 1. Tempol decreases measures of oxidative stress independent of BP

(A) Mean arterial pressure (MAP) was measured by radiotelemetry in male and female SHR receiving tempol (30 mg/kg/day) in drinking water, n=5. Data are presented as mean \pm S.E.M., *P<0.05 compared with male. (B) Plasma levels of 3-NT were measured in vehicle control (n=5) and tempol-treated (n=8) male and female SHR by Dot blot analysis.

GTP cyclohydrolase expression is comparable in male and female SHR

To determine if sex differences in BH₄ levels were mediated by differences in production, GTPCH1 protein expression was measured; GTPCH1 is the rate-controlling enzyme in BH₄ production. GTPCH1 protein expression is comparable in the renal IM of male and female SHR (Figure 4; P=0.17).

Exogenous BH₄ treatment in male SHR abolishes sex differences in biopterins

To determine if a BH_4 deficiency results in reduced IM NOS activity in male SHR, additional male SHR received BH_4 supplementation for 1 week. BH_4 supplementation increased total biotperin, BH_2 , and BH_4 in the renal IM (Figure 5). BH_4 supplementation of male SHR also abolished sex differences in total biotperin, BH_2 , and BH_4 that





Figure 2. Tempol treatment abolishes the sex difference in IM NOS activity and dependency on exogenous BH₄ Total NOS enzymatic activity in the renal IM of vehicle control ((A); n=11-12) and tempol-treated ((B); n=8-9) male and female SHR (n=8-12) with and without the addition of exogenous BH₄.

were observed between control male and female SHR; biopterin levels in treated males and control females were compared by Student's *t* test (total biopterin, P=0.38; BH₂, P=0.11; BH₄, P=0.86). BH₄ treatment had no effect on body weight, although systolic BP was significantly decreased by BH₄ supplementation (Figure 6A; P=0.0024).

NOS activity was then measured in the absence and presence of BH₄ in control and BH₄-treated male SHR. Supplementation of male SHR with BH₄ resulted in a significant decrease in total NOS activity compared with vehicle-treated SHR (P<0.0001), however, the increase in NOS activity observed with the inclusion of BH₄ in the assay was blunted (Figure 6B; P=0.0075).

Discussion

Although there are numerous reports of sex differences in NO production and NO bioavailability, the molecular mechanisms responsible remain unknown. The goal of the present study was to further explore the mechanisms that result in sex differences in the NO system. Our novel hypothesis in the current study is that increased oxidative stress in male SHR induces a relative BH₄ deficiency compared with female SHR, resulting in lower renal NOS activity.







SHR (*n*=4–5).

The main findings of the current study are: (i) there is an oxidative stress-induced sex difference in biopterins, where female SHR have greater biopterin levels than males and (ii) the relative deficiency in BH_4 contributes to lower renal NOS activity in male SHR compared with female SHR. These data suggest that mechanisms to maximize BH_4 levels will offer cardiovascular protection to hypertensive males in particular. Indeed, treating male SHR with BH_4 significantly decreases BP, although this is not associated with an increase in total NOS activity. Additional studies are needed to fully understand the mechanisms by which BH_4 lowers BP.

We have previously reported that there is a sex difference in NOS activity in the renal IM of SHR that cannot be explained by sex difference in NOS protein expression alone [15,16]. The relative abundance of NOS in female SHR may contribute to the lower BP observed in female SHR compared with males [6]. However, the mechanisms







contributing to sex difference in NOS activity have not yet been clearly elucidated. The goal of the current study was to test the hypothesis that relative BH₄ deficiency induced by oxidative stress in male SHR results in lower levels of NOS activity in renal IM compared with females. Male SHR have been reported numerous times to have greater levels of oxidative stress in the kidney compared with female SHR [21,25,29], although peroxynitrite in particular is known to rapidly oxidize BH₄ to BH₂ [17,18]. In the current study we show greater renal levels of 3-NT in male SHR compared with females, and confirmed that treatment with tempol abolishes the sex difference in 3-NT independent of an effect on BP. These findings lend support to the hypothesis that male SHR will have greater BH₄ oxidation which may result in decrease in NOS activity. Based on the central role of the NO/NOS pathway in modulating BP and vascular tone, sex differences in biopterin levels may significantly contribute to sex differences in overall cardiovascular health.

 BH_4 is an essential cofactor for NO production. BH_4 levels are decreased in many models of cardiovascular disease [30] including hypertension [31]. A role for BH_4 in BP control is further supported by the finding that BH_4 supplementation lowers BP in a male rat model of pulmonary arterial hypertension [32] and male mice with disrupted BH_4 synthesis are hypertensive [33]. Despite growing interest and evidence for the role of BH_4 in the pathogenesis of hypertension, there is a scarcity of data in the literature examining the impact of sex on the BH_4 system. We report that total biopterin, BH_2 , and BH_4 levels are all greater in females compared with males. These data support the hypothesis that there is an impaired biopterin system in the renal IM of male SHR. This hypothesis is supported by findings of lower biopterin and BH_4 content in cardiac and vascular tissue in male SHR compared with normotensive male WKY [34,35].

NOS enzymatic activity is typically measured in the presence of excess amounts of BH_4 . However, to determine if NOS enzymatic activity is dependent on exogenous BH_4 or if sufficient amounts are present in the tissue homogenate, we performed the NOS activity assay in the presence and absence of excess BH_4 . Despite sex differences in BP, NOS activity, and oxidative stress, vehicle control male and female SHR exhibited a comparable increase in NOS activity with the inclusion of excess BH_4 . However, treatment with tempol abolished the dependency of the NOS activity assay on excess BH_4 in both sexes, suggesting that oxidative stress is a greater determinant of dependency on BH_4 than sex of the animal. It should be noted that we confirmed that tempol treatment did not alter NOS protein expression (data not shown).



Figure 5. Exogenous BH₄ treatment in male SHR abolishes sex differences in biopterins HPLC analysis of total biopterins (A), BH₂ (B), and BH₄ levels (C) in renal IM of vehicle control and BH₄-treated male SHR (*n*=4).

 BH_4 bioavailability is determined by: (i) the activity of GTPCH1, the rate-controlling enzyme in BH_4 production and (ii) oxidative stress, which can oxidize BH_4 to BH_2 , resulting in NOS uncoupling and the production of reactive oxygen species [36]. We found no differences in GTPCH1 protein expression in the renal IM of male and female SHR. Therefore it is unlikely that the relative deficiency in BH_4 in male SHR is due to alterations in BH_4 production compared with the female SHR. Activity of GTPCH1 was not assessed in the current study, therefore it is difficult to completely rule out a sex difference in production. Instead, our data support the hypothesis that oxidative stress is an important determinant of BH_4 availability in male SHR, as treatment with tempol significantly increases BH_4 levels in males, while BH_4 levels in the female SHR treated with tempol do not change. However, without direct measurement of the rate of synthesis of BH_4 in male compared with female IM lysates it is not possible to definitively







demonstrate that oxidative destruction of BH₄ is the major mechanism responsible for sex-dependent differences in BH₄ concentrations, NOS activity, and BP in SHR.

Interestingly, BH₂ levels are also greater in females compared with males, however the ratio of BH₂ to BH₄ is comparable between the sexes (males: 0.41 ± 0.14 ; females: 0.43 ± 0.05). Therefore, it is unlikely that there are sex differences in BH₂ competition for NOS binding. Consistent with our hypothesis, tempol increased BH₄ levels, although BH₂ levels were also increased in both male and female SHR following tempol treatment and total biopterin increased in male SHR. While we cannot account for the mechanism mediating the increase in BH₂, it is likely due to the fact that systemic tempol treatment does not abolish all oxidative stress in the renal IM. As a result, the increase in BH₂ levels in female SHR increase in the absence of a change in total biopterins, suggesting a sex-specific effect of oxidative stress on biopterins. Future studies will be designed to examine this sex difference.

To further assess the role of endogenous BH_4 in modulating NOS activity in male SHR and explore the potential therapeutic role of BH_4 treatment, male SHR were treated with exogenous BH_4 for 1 week. Supplementation with BH_4 decreased dependency of NOS enzymatic activity on excess BH_4 , further supporting the hypothesis that a BH_4 deficiency in male SHR contributes to lower levels of NOS activity compared with females. Interestingly, total NOS activity was decreased by 7 days of BH_4 treatment. Future studies will investigate the mechanism driving the decrease in NOS activity; however, the corresponding increase in BH_2 with BH_4 supplementation may compete with BH_4 for binding to NOS thereby resulting in a decrease in total NOS activity. BH_4 supplementation also decreased BP in male SHR, although it is unlikely that a decrease in BP alone was responsible for the decreased NOS activity as we



have previously published that decreasing BP in male SHR with hydrochlorothiazide and reserpine has no effect on renal NOS activity [15]. Indeed, the finding that BH_4 supplementation reduces BP despite reducing NOS activity in the IM suggests that inner medullary NOS is not a significant determinant of BP in male SHR. Instead, our result is consistent with previous reports in male SHR indicating that BH_4 reduces testosterone synthesis thereby reducing BP [37]. In addition to decreasing BP, BH_4 has also been shown to ameliorate cardiac hypertrophy and diastolic dysfunction in male SHR [35]. Genomic analysis of *GCH1*, which encodes GTPCH1, in humans found a sex-specific risk of hypertension in patients with a specific polymorphism of the *GCH1* gene, with females having significantly higher BP than males and lower NO production [38]. Given this finding in human patients, along with the currently reported sex difference in BH_4 levels, it is important to consider sex as a biological variable in future studies assessing the therapeutic potential of BH_4 in hypertension.

In conclusion, although it is well established that there is a sexual dimorphism in NO bioavailability [11-14], the molecular mechanisms responsible are still being investigated. NOS enzymatic activity is tightly regulated by numerous biochemical pathways [39], including the availability of the cofactor BH_4 . We found sex differences in BH_4 in the renal IM of SHR which corresponds to sex differences in NOS activity and NO production. Further studies are needed to better elucidate the potential role of BH_4 as a therapeutic agent in both sexes.

Clinical perspectives

- Premenopausal females have a lower incidence of hypertension than age-matched males, however the mechanisms contributing to this are not well understood.
- This is the first study to report a sex difference in BH₄ levels in a rodent model of hypertension. BH₄ is an essential cofactor for the production of NO, a potent vasodilator that plays an essential role in BP regulation.
- Targetting BH₄ may serve as a novel therapeutic pathway for the treatment of hypertension in both sexes.

Acknowledgements

We thank the technical expertise of Tantiana Burns.

Competing interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

This work was supported by the NIH [grant number HL127091]; and the AHA [grant numbers JCS: 17EIA33410565].

Author contribution

K.N.B and J.C.S. contributed to the experimental design of the present study. E.E.G., K.N.B., O.R., W.C., J.B.M., and D.G.H. performed experiments and collected data. E.E.G., K.N.B., W.C., and J.C.S analyzed the data and prepared the figures. E.E.G., K.N.B., and J.C.S wrote the manuscript. All authors reviewed the final manuscript.

Abbreviations

3-NT, 3-nitrotyrosine; BH₂, dihydrobiopterin; BH₄, tetrahydrobiopterin; BP, blood pressure; GTPCH1, GTP cyclohydrolase-1; IM, inner medulla; NOS, nitric oxide synthase; SHR, spontaneously hypertensive rat; TBS-T, TBS, 0.1% Tween 20.

References

- 1 Baylis, C., Mitruka, B. and Deng, A. (1992) Chronic blockade of nitric oxide synthesis in the rat produces systemic hypertension and glomerular damage. *J. Clin. Invest.* **90**, 278–281, https://doi.org/10.1172/JCl115849
- 2 Ribeiro, M.O., Antunes, E., de Nucci, G., Lovisolo, S.M. and Zatz, R. (1992) Chronic inhibition of nitric oxide synthesis. A new model of arterial hypertension. *Hypertension* **20**, 298–303, https://doi.org/10.1161/01.HYP.20.3.298
- 3 Gamboa, A., Shibao, C., Diedrich, A., Choi, L., Pohar, B., Jordan, J. et al. (2007) Contribution of endothelial nitric oxide to blood pressure in humans. *Hypertension* **49**, 170–177, https://doi.org/10.1161/01.HYP.0000252425.06216.26
- © 2018 The Author(s). This is an open access article published by Portland Press Limited on behalf of the Biochemical Society and distributed under the Creative Commons Attribution License 4.0 (CC BY).



- 4 Baylis, C. (2005) Changes in renal hemodynamics and structure in the aging kidney; sexual dimorphism and the nitric oxide system. *Exp. Gerontol.* **40**, 271–278, https://doi.org/10.1016/j.exger.2005.01.008
- 5 Lee, J., Bae, E.H., Ma, S.K. and Kim, S.W. (2016) Altered nitric oxide system in cardiovascular and renal diseases. *Chonnam Med. J.* 52, 81–90, https://doi.org/10.4068/cmj.2016.52.2.81
- 6 Brinson, K.N., Elmarakby, A.A., Tipton, A.J., Crislip, G.R., Yamamoto, T., Baban, B. et al. (2013) Female SHR have greater blood pressure sensitivity and renal T cell infiltration following chronic NOS inhibition than males. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **305**, R701–10, https://doi.org/10.1152/ajpregu.00226.2013
- 7 Lee, J. (2008) Nitric oxide in the kidney: its physiological role and pathophysiological implications. *Electrolyte Blood Press.* 6, 27–34, https://doi.org/10.5049/EBP.2008.6.1.27
- 8 Wu, F., Park, F., Cowley, Jr, A.W. and Mattson, D.L. (1999) Quantification of nitric oxide synthase activity in microdissected segments of the rat kidney. *Am. J. Physiol.* **276**, F874–81
- 9 Garvin, J.L., Herrera, M. and Ortiz, P.A. (2011) Regulation of renal NaCl transport by nitric oxide, endothelin, and ATP: clinical implications. *Annu. Rev. Physiol.* **73**, 359–376, https://doi.org/10.1146/annurev-physiol-012110-142247
- 10 Hyndman, K.A. and Pollock, J.S. (2013) Nitric oxide and the A and B of endothelin of sodium homeostasis. *Curr. Opin. Nephrol. Hypertens.* 22, 26–31, https://doi.org/10.1097/MNH.0b013e32835b4edc
- 11 Neugarten, J., Ding, Q., Friedman, A., Lei, J. and Silbiger, S. (1997) Sex hormones and renal nitric oxide synthases. J. Am. Soc. Nephrol. 8, 1240–1246
- 12 Forte, P., Kneale, B.J., Milne, E., Chowienczyk, P.J., Johnston, A., Benjamin, N. et al. (1998) Evidence for a difference in nitric oxide biosynthesis between healthy women and men. *Hypertension* **32**, 730–734, https://doi.org/10.1161/01.HYP.32.4.730
- 13 Glushkovskaya-Semyachkina, O.V., Anishchenko, T.G., Sindyakova, T.A., Leksina, O.V. and Berdnikova, V.A. (2006) Sex-related differences in nitric oxide content in healthy and hypertensive rats at rest and under stress conditions. *Bull. Exp. Biol. Med.* **142**, 9–11, https://doi.org/10.1007/s10517-006-0277-y
- 14 Majmudar, N.G., Robson, S.C. and Ford, G.A. (2000) Effects of the menopause, gender, and estrogen replacement therapy on vascular nitric oxide activity. J. Clin. Endocrinol. Metab. 85, 1577–1583, https://doi.org/10.1210/jcem.85.4.6530
- 15 Sasser, J.M., Brinson, K.N., Tipton, A.J., Crislip, G.R. and Sullivan, J.C. (2015) Blood pressure, sex, and female sex hormones influence renal inner medullary nitric oxide synthase activity and expression in spontaneously hypertensive rats. J. Am. Heart Assoc. 4, https://doi.org/10.1161/JAHA.114.001738
- 16 Sullivan, J.C., Pardieck, J.L., Hyndman, K.A. and Pollock, J.S. (2010) Renal NOS activity, expression, and localization in male and female spontaneously hypertensive rats. *Am. J. Physiol.* **298**, R61–R69
- 17 Kuzkaya, N., Weissmann, N., Harrison, D.G. and Dikalov, S. (2003) Interactions of peroxynitrite, tetrahydrobiopterin, ascorbic acid, and thiols: implications for uncoupling endothelial nitric-oxide synthase. J. Biol. Chem. 278, 22546–22554, https://doi.org/10.1074/jbc.M302227200
- 18 Milstien, S. and Katusic, Z. (1999) Oxidation of tetrahydrobiopterin by peroxynitrite: implications for vascular endothelial function. *Biochem. Biophys. Res. Commun.* **263**, 681–684, https://doi.org/10.1006/bbrc.1999.1422
- 19 Harrison, D.G., Cai, H., Landmesser, U. and Griendling, K.K. (2003) Interactions of angiotensin II with NAD(P)H oxidase, oxidant stress and cardiovascular disease. J. Renin Angiotensin Aldosterone Syst. 4, 51–61, https://doi.org/10.3317/jraas.2003.014
- 20 Cosentino, F. and Luscher, T.F. (1999) Tetrahydrobiopterin and endothelial nitric oxide synthase activity. *Cardiovasc. Res.* **43**, 274–278, https://doi.org/10.1016/S0008-6363(99)00134-0
- 21 Bhatia, K., Elmarakby, A.A., El-Remessy, A.B. and Sullivan, J.C. (2012) Oxidative stress contributes to sex differences in angiotensin II-mediated hypertension in spontaneously hypertensive rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **302**, R274–R282, https://doi.org/10.1152/ajpregu.00546.2011
- 22 Fortepiani, L.A. and Reckelhoff, J.F. (2005) Role of oxidative stress in the sex differences in blood pressure in spontaneously hypertensive rats. *J. Hypertens.* 23, 801–805, https://doi.org/10.1097/01.hjh.0000163149.05083.13
- 23 Sullivan, J.C., Bhatia, K., Yamamoto, T. and Elmarakby, A.A. (2010) Angiotensin (1-7) receptor antagonism equalizes angiotensin II-induced hypertension in male and female spontaneously hypertensive rats. *Hypertension* 56, 658–666, https://doi.org/10.1161/HYPERTENSIONAHA.110.153668
- 24 Sullivan, J.C., Pollock, J.S. and Pollock, D.M. (2006) Superoxide-dependent hypertension in male and female endothelin B receptor-deficient rats. *Exp. Biol. Med.* **231**, 818–823
- 25 Sullivan, J.C., Sasser, J.M. and Pollock, J.S. (2007) Sexual dimorphism in oxidant status in spontaneously hypertensive rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **292**, R764–R768, https://doi.org/10.1152/ajpregu.00322.2006
- 26 Widder, J.D., Chen, W., Li, L., Dikalov, S., Thony, B., Hatakeyama, K. et al. (2007) Regulation of tetrahydrobiopterin biosynthesis by shear stress. *Circ. Res.* **101**, 830–838, https://doi.org/10.1161/CIRCRESAHA.107.153809
- 27 Tsikas, D. (2017) What we-authors, reviewers and editors of scientific work-can learn from the analytical history of biological 3-nitrotyrosine. J. Chromatogr. B Analyt. Technol. Biomed. Life Sci. 1058, 68–72, https://doi.org/10.1016/j.jchromb.2017.05.012
- 28 Ahsan, H. (2013) 3-Nitrotyrosine: a biomarker of nitrogen free radical species modified proteins in systemic autoimmunogenic conditions. *Hum. Immunol.* 74, 1392–1399, https://doi.org/10.1016/j.humimm.2013.06.009
- 29 Sullivan, J.C., Semprun-Prieto, L., Boesen, E.I., Pollock, D.M. and Pollock, J.S. (2007) Sex and sex hormones influence the development of albuminuria and renal macrophage infiltration in spontaneously hypertensive rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 293, R1573–R1579, https://doi.org/10.1152/ajpregu.00429.2007
- 30 Forstermann, U. and Sessa, W.C. (2012) Nitric oxide synthases: regulation and function. *Eur. Heart J.* **33**, 829–837, 37a-37d, https://doi.org/10.1093/eurheartj/ehr304



12

- 31 Hong, H.J., Hsiao, G., Cheng, T.H. and Yen, M.H. (2001) Supplemention with tetrahydrobiopterin suppresses the development of hypertension in spontaneously hypertensive rats. *Hypertension* **38**, 1044–1048, https://doi.org/10.1161/hy1101.095331
- 32 Schreiber, C., Eilenberg, M.S., Panzenboeck, A., Winter, M.P., Bergmeister, H., Herzog, R. et al. (2017) Combined oral administration of L-arginine and tetrahydrobiopterin in a rat model of pulmonary arterial hypertension. *Pulm. Circ.* **7**, 89–97, https://doi.org/10.1086/689289
- 33 Sumi-Ichinose, C., Suganuma, Y., Kano, T., Ihira, N., Nomura, H., Ikemoto, K. et al. (2017) Sepiapterin reductase gene-disrupted mice suffer from hypertension with fluctuation and bradycardia. *Physiol. Rep.* 5, https://doi.org/10.14814/phy2.13196
- 34 Li, H., Witte, K., August, M., Brausch, I., Godtel-Armbrust, U., Habermeier, A. et al. (2006) Reversal of endothelial nitric oxide synthase uncoupling and up-regulation of endothelial nitric oxide synthase expression lowers blood pressure in hypertensive rats. *J. Am. Coll. Cardiol.* **47**, 2536–2544, https://doi.org/10.1016/j.jacc.2006.01.071
- 35 Chang, P., Wang, Q., Xu, H., Yang, M., Lin, X., Li, X. et al. (2015) Tetrahydrobiopterin reverse left ventricular hypertrophy and diastolic dysfunction through the PI3K/p-Akt pathway in spontaneously hypertensive rats. *Biochem. Biophys. Res. Commun.* 463, 1012–1020, https://doi.org/10.1016/j.bbrc.2015.06.051
- 36 Crabtree, M.J., Smith, C.L., Lam, G., Goligorsky, M.S. and Gross, S.S. (2008) Ratio of 5,6,7,8-tetrahydrobiopterin to 7,8-dihydrobiopterin in endothelial cells determines glucose-elicited changes in NO vs. superoxide production by eNOS. Am. J. Physiol. Heart Circ. Physiol. 294, H1530–H1540, https://doi.org/10.1152/ajpheart.00823.2007
- 37 Fortepiani, L.A. and Reckelhoff, J.F. (2005) Treatment with tetrahydrobiopterin reduces blood pressure in male SHR by reducing testosterone synthesis. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **288**, R733–6, https://doi.org/10.1152/ajpregu.00500.2004
- 38 Zhang, L., Rao, F., Zhang, K., Khandrika, S., Das, M., Vaingankar, S.M. et al. (2007) Discovery of common human genetic variants of GTP cyclohydrolase 1 (GCH1) governing nitric oxide, autonomic activity, and cardiovascular risk. *J. Clin. Invest.* **117**, 2658–2671, https://doi.org/10.1172/JCl31093
- 39 Proskuryakov, S.Y., Konoplyannikov, A.G., Skvortsov, V.G., Mandrugin, A.A. and Fedoseev, V.M. (2005) Structure and activity of NO synthase inhibitors specific to the L-arginine binding site. *Biochemistry* **70**, 8–23