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Advances in managing COPD related to α_1 -antitrypsin deficiency: An under-recognized genetic disorder

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Abstract

α1-Antitrypsin deficiency (AATD) predisposes individuals to chronic obstructive pulmonary disease (COPD) and liver disease. Despite being commonly described as rare, AATD is under-recognized, with less than 10% of cases identified. The following is a comprehensive review of AATD, primarily for physicians who treat COPD or asthma, covering the genetics, epidemiology, clinical presentation, screening and diagnosis, and treatments of AATD. For patients presenting with liver and/or lung disease, screening and diagnostic tests are the only methods to determine whether the disease is related to AATD. Screening guidelines have been established by organizations such as the World Health Organization, European Respiratory Society, and American Thoracic Society. High-risk groups, including individuals with COPD, nonresponsive asthma, bronchiectasis of unknown etiology, or unexplained liver disease, should be tested for AATD. Current treatment options include augmentation therapy with purified AAT for patients with deficient AAT levels and significant lung disease. Recent trial data suggest that lung tissue is preserved by augmentation therapy, and different dosing schedules are currently being investigated. Effective management of AATD and related diseases also includes aggressive avoidance of smoking and biomass burning, vaccinations, antibiotics, exercise, good diet, COPD medications, and serial assessment.

KEYWORDS

 α_1 -antitrypsin, α_1 -antitrypsin deficiency, asthma, augmentation therapy, COPD, emphysema

1 | INTRODUCTION

 α_1 -Antitrypsin (AAT) is a serine protease inhibitor (PI) that protects lung tissue from proteolytic damage by inhibiting neutrophil elastase, a powerful enzyme with broad substrate specificity.¹ Therefore, individuals with AAT deficiency (AATD), an autosomal co-dominant condition, are at increased risk of developing chronic obstructive pulmonary disease (COPD).^{2,3} In addition, these individuals are often at risk of liver disease due to polymerization and retention of AAT in hepatocytes, where the majority of AAT is produced.²

Between 0.02% and 0.04% of individuals suffer from AATD.³ However, despite being commonly described as a rare disease, AATD is highly under-recognized, with less than 10% of individuals with AATD identified ⁴⁻⁶ and with delays of more than 5 years between initial symptoms and diagnosis.⁷ Although younger individuals who are often asymptomatic may understandably escape medical attention, data suggest that, despite a majority of adults with the deficiency allele PI*ZZ manifesting COPD-related symptoms in the fourth and fifth decades, few clinicians identify AATD as the cause.⁵ The lack or delay of a diagnosis of AATD inevitably delays the initiation of augmentation therapy with AAT if indicated, transplantation,

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes. © 2018 The Authors. *Allergy* Published by John Wiley & Sons Ltd. **TABLE 1** Genotypes, serum levels of AAT, and risk of lung and liver disease

Genotype	Serum levels of AAT ^a	Risk of lung disease	Risk of liver disease	Explanatory information
MM	Normal			The PI*M allele encodes normal AAT. ²
Null/null	Absent	+++		Rare null alleles are characterized by absent circulating AAT due to transcriptional or translational errors. ²
ZZ	Very low	+++	+++	The PI*Z allele leads to polymerization in hepatocytes and less frequent binding to neutrophil elastase in the lungs. ²
MZ	Low to normal	+	+	A well-designed study recently observed an increased risk of developing COPD due to exposure to cigarette smoke in individuals with the PI*MZ allele, ⁵² although other studies have found both increased and no association. ¹¹⁹⁻¹²²
SS	Borderline normal to low	+/-		No conclusive evidence links homozygous PI*SS to increased risk for lung or liver disease. However, the PI*S allele is associated with increased degradation of AAT in hepatocytes. ²
SZ	Low	+		The PI*SZ allele has been variably associated with increased risk of disease. A recent study, based on data from 6 previous studies, found an increased odds ratio (3.26, 95% CI: 1.24-8.57) for the development of COPD. ¹²³

^aAAT levels can increase during inflammation. Cigarette smoke and infections lead to an increase in neutrophils and neutrophil elastase in the lungs, thus predisposing exposed individuals with AATD to develop COPD.^{83,84}

+/-, indicates the risk of disease; -, indicates the absence of disease; AAT, α_1 -antitrypsin; AATD, α_1 -antitrypsin deficiency; CI, confidence interval; COPD, chronic obstructive pulmonary disease.

Reproduced with modifications from Henao and Craig.¹²⁴

supportive therapies such as smoking cessation, and genetic and psychological counseling.

Since the discovery of AATD in the 1960s, knowledge and understanding of AAT and its deficiency have progressed significantly, although effective screening methods and non-intravenous therapy have remained elusive. The following is a comprehensive review of AATD primarily for physicians who treat COPD or asthma, highlighting advances into the epidemiology, screening, and treatment of AATD. These data are especially important for allergists because a third of AATD patients enter the healthcare system through allergists, and unfortunately, as noted in reference 9, this often delays the appropriate diagnosis and treatment.^{8,9}

2 | GENETICS, SERUM AAT LEVELS, AND EPIDEMIOLOGY

Individuals with AATD have a homo- or heterozygous mutation of the Serpina1 gene,² of which more than 150 alleles are recognized.⁸ The most common allele (M, for "medium mobility" through an isoelectric gel) encodes normal AAT, with PI*MM the most common homozygous allele. The most common deficiency alleles are Z (slowest) and S (slower), which, in both cases, result from single amino acid substitutions.² The PI*Z allele accounts for 96% of known clinical cases of AATD, although the two most frequent deficiency alleles are thought to be PI*S (50%-60% of carriers) and PI*Z (10%-20% of carriers).^{2,10,11} Rare null and dysfunctional alleles also exist, the former characterized by absent circulating AAT due to transcriptional or translational errors.² Lastly, the epigenetic silencing of Serpina1 has been reported which may explain the AATD effects in genetically unaffected individuals.¹² Different alleles are associated with different risks of developing lung and liver diseases (Table 1). Despite PI*ZZ is the allele most associated with increased likelihood of developing COPD, PI*SZ and PI*MZ also show an increased risk, and data imply that disease progression in PI*SZ patients might be similar to Pi*ZZ.^{2,13} As shown in Table 1, AAT concentrations vary according to genotype, with normal levels in the range of 20-53 μ mol/L.^{2,4,14,15} Individuals with the PI*MM genotype have 105%-164% of normal levels, PI*MS 88%-137%, PI*SS 73%-106%, PI*MZ 66-100%, PI*SZ 49%-66%, while <15%-20% would suggest that an individual has the homozygous PI*ZZ, Z null, or null-null genotype.^{4,15,16}

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While PI*Z and PI*S alleles are particularly prevalent in the north and south of Europe, respectively, deficiency alleles have been detected across diverse populations worldwide.^{10,17-26} By combining data from epidemiological studies, in 2012 de Serres and Blanco ¹⁰ estimated the prevalence of AAT alleles across 10 geographic regions worldwide, encompassing 97 countries and 5.26 billion people. The prevalence of PI*ZZ is 0.1% of deficiency genotypes worldwide (181 894 individuals), with 41% of cases in Northern and Central Europe and 24% in North America.9 PI*SZ accounts for 0.7% (1 269 054 individuals), with 48% of these in Northern and Central Europe, 20% in North and Central America, and 16% in South America.¹⁰ Approximately 75% of deficiency genotypes are PI*MS, and 24% are PI*MZ and PI*SS.¹⁰ In the United States, more than 20 million people (6.6%) are estimated to have at least one deficiency allele; 2.3% and 1.1% of the general population have PI*S or PI*Z, respectively, and 0.01% have PI*ZZ.¹⁰ PI*Z alleles appear to be as prevalent in the United States as the lower ranges in northern European countries, whereas PI*S alleles occur more frequently than in that region.¹⁷

3 | CLINICAL PRESENTATION

3.1 | Pulmonary characteristics

Differential diagnosis of AATD-related COPD versus COPD from other etiologies and even asthma with fixed obstruction is complicated by shared signs and symptoms; these similarities partly explain why AATD is highly under-diagnosed. Nevertheless, while not unique to AATD-related COPD, several characteristics indicate that AATD may be the cause of COPD. Individuals with AATD tend to experience earlier onset of symptoms, often in the third or fourth decade, particularly in smokers, whereas COPD of other etiologies often occurs in the fifth decade or later.⁸ There may also be more extensive emphysematous damage to the lung bases, which can be detected by chest radiography or computed tomography (CT) (Figure 1).² In an analysis of 1129 patients enrolled in the National Heart, Lung and Blood Institute (NHLBI) Registry of Individuals with Severe Deficiency of AAT, common symptoms included dyspnea (84%), cough (42%), phlegm (46%), and wheezing and upper respiratory infections (76%).²⁷ Airflow obstruction is often reversible; about 61% of patients in the NHLBI Registry had a 12% and 200 mL increase in forced expiratory volume in one-second (FEV1) or forced vital capacity postbronchodilator.²⁷ Asthma is also common in patients with severe AATD (affecting 35% of individuals in the NHLBI Registry) and may increase the risk of developing AATDrelated COPD.9,27

Another reason why AATD is highly under-diagnosed is that about 40% of individuals with PI deficiency alleles are asymptomatic for COPD.^{1,2,28,29} In CT lung scans, no clinically significant signs of emphysema were found in 14% of 119 patients with severe AATD.³⁰ Similarly, in the Swedish National Registry of AATD, no signs of COPD were evident in 30% of nonsmokers and 16% of smokers.³¹ In a series of postmortems of individuals with AATD, no signs of COPD were found in 20% of cases.³¹

In several studies, the rate of decline of FEV₁ in PI*ZZ patients was anywhere between 23 mL/v and 316 mL/v.³²⁻⁴⁴ Several factors increase this rate of change, including smoking, male sex, age 30-44 years, FEV₁ between 35 and 79% of predicted, serum AAT level, chronic bronchitis symptoms, and bronchodilator responsiveness.^{36,40,43,44} Moreover, lung function declines more rapidly when individuals are exposed to air pollution, ozone, and particulate matter less than 10 μ m in diameter,^{45,46} while respiratory infections may also exacerbate lung disease.⁴⁷ Smoking is a notable risk factor, given the association with earlier onset of respiratory symptoms in patients with AATD.^{47,48} In a recent Swedish study, reporting outcomes 35 years after screening for PI deficiency alleles at birth, PI*ZZ ever-smokers showed early signs of emphysema versus PI*MM ever-smoking controls, whereas PI*ZZ and PI*SZ never-smokers had normal lung function versus never-smoking controls.48 Nevertheless, while the cause of death for nonsmoking PI*ZZ individuals is respiratory failure in an estimated 45%-60% of cases, PI*ZZ nonsmokers are more likely than PI*ZZ ever-smokers to die of liver disease, particularly when elderly, as they do not die earlier of lung disease.⁴⁹⁻⁵¹ With regard to patients with PI*MZ deficiency alleles, a statistically significantly increased risk of developing lung disease was only detected for smokers.⁵² In addition to the emphysematous symptoms of COPD, patients with AATD may also develop bronchiectasis, although it is unclear whether AATD causes bronchiectasis. In a case-control study of patients with a diagnosis of bronchiectasis with and without AATD, no association was found between AAT genes and bronchiectasis.53 In another study of CT scans, clinically significant bronchiectasis was present in 27% of individuals with the PI*Z allele.54 In accordance with these studies, the recommendation is to test for AATD whether the etiology of bronchiectasis remains unknown.^{1,2}

AATD-related COPD has traditionally been associated with more emphysematous damage to the lung bases.² However, emphysema is not always present. In a study of PI*ZZ patients, 20% had emphysema in radiographs ⁵⁵ and, in another study using CT scans,



FIGURE 1 CT scans showing extensive emphysematous damage to the lungs. A, Axial plane of thorax; B, Coronal plane of thorax. CT, computed tomography. CT scans were kindly provided by Prof. Dr. Andreas Rembert Koczulla, Fachzentrum für Pneumologie Schön Klinik Berchtesgadener Land, Germany, and Klinik für Pneumologie, Marburg, Germany

emphysema was present in 86% of PI*Z patients.³⁰ The latter study also demonstrated that a single physiologic parameter should not be used as a surrogate measure of disease severity.³⁰ In particular, basal emphysema was associated with greater impairment of FEV₁, but less impairment of gas exchange and alveolar-arterial oxygen gradient versus apical distribution.³⁰ In the latter study, 36% of PI*Z patients with emphysema had apical damage. Notably, the traditional description of AATD-associated emphysema as predominantly basal and panacinar originates from limited autopsy studies and case series using chest radiographs, which has now been superseded by CT.^{11,55}

3.2 Extrapulmonary characteristics

Individuals with alleles associated with intrahepatocyte polymerization, such as Z, M_{malton}, and S_{iivama}, are predisposed to liver disease, including hepatitis, cirrhosis, and hepatoma.^{2,56} In individuals with AATD, liver disease presents in a bimodal distribution, that is, neonatal hepatitis and cholestasis in infants and chronic liver disease in adults most of who are ZZ and usually over 50 years of age.⁵⁷ It is estimated that 10% of adults may develop symptomatic cirrhosis. In the first 20 years of life, about one-third of PI*ZZ carriers may develop liver disease, although most recover.58,59 In another study, based on 161 infants with liver dysfunction and use of isoelectric focusing, 15% had severe AATD and 12% moderate AATD.⁶⁰ Similarly, in a Swedish study, 18% of 120 newborns with PI*Z alleles during 6 months of follow-up often presented with jaundice, minor laboratory abnormalities, and liver dysfunction. Biopsies revealed typical Periodic acid Schiff diastase-positive intracellular inclusion bodies and AAT-positive staining (Figure 2).^{61,62}

AATD is also associated with various other conditions, including panniculitis (a skin condition that affects an estimated 1 per 1000 individuals with AATD) and granulomatosis with polyangiitis.^{2,27,63-65}

4 SCREENING AND DIAGNOSIS

To address the issue of under-diagnosis, several guidelines for screening and targeted testing of AATD are available. $^{1,66-69}$ In 1997,

the World Health Organization (WHO) recommended screening all patients with COPD and adults and adolescents with asthma by quantitative testing, followed by PI typing for individuals with abnormal results.⁶⁶ In 2015, the Global Initiative for Chronic Obstructive Lung Disease stated that the WHO suggests screening COPD patients from areas with a particularly high prevalence of AATD.⁶⁷ In 2017, the European Respiratory Society (ERS) proposed particularly inclusive guidelines that recommended testing in specific groups of individuals (Table 2).⁶⁹

Screening asymptomatic individuals for AATD is particularly useful because positive correlations exist between smoking cessation and better physical and psychosocial outcomes, due to individuals' awareness of their predisposition to COPD.^{7,70} Similar findings were recently published for adults admitted to a large newborn screening program in Sweden.⁷¹⁻⁷³ However, it is debatable whether this evidence will lead to widespread screening of newborns, as a diagnosis of AATD has been shown to increase stress in families.⁷¹ Additional caveats include the lack of specific treatment for the AATD-associated liver disease, which is the primary cause of childhood morbidity, and the need for additional conclusive evidence that newborn screening ultimately results in better outcomes in longitudinal studies.⁷³ A pilot study of newborn screening for AATD recently commenced in New York State.⁷³

Screening and diagnosis of AATD usually begin with a quantitative measurement of serum AAT concentrations, often using radial immunodiffusion, nephelometry, or latex-enhanced immunoturbidimetry.^{2,14,73-79} Below a protective threshold of 11 μ mol/L (normal range 20-53 μ mol/L), the risk of accelerated airflow obstruction increases. However, a post hoc analysis in the recent RAPID study suggests that patients on augmentation therapy with AAT may experience greater benefit when serum levels increase to well above 11 μ mol/L, relative to patients with lower levels that were also above the protective threshold (Figure 3).^{74,80} Studies are presently being performed to reevaluate the protective threshold.

Either when serum AAT levels are found to be low or simultaneously when measuring AAT levels, additional diagnostic assessments can be used to identify AAT alleles and, therefore, strengthen the diagnosis of AATD. These assessments may include phenotyping by



FIGURE 2 Liver damage in patients with α_1 -antitrypsin deficiency. A, Periodic acid Schiff diastase (PAS-d) staining showing intracytoplasmic accumulation of PAS-d–resistant material; B, Immunohistochemical staining of the case patient's hepatocytes; and C, Control immunohistochemical staining of a known α_1 -antitrypsin–deficient patient. From: Rider and Craig⁶¹

TABLE 2 Recommendations for quantitative testing of AAT

No.	Recommendation
1	Patients with a diagnosis of COPD
2	Patients with a diagnosis of adult-onset asthma
3	Individuals with cryptogenic cirrhosis or liver disease
4	Individuals with GPA
5	Adults with bronchiectasis of unknown etiology
6	Adults with necrotizing panniculitis
7	First-degree relatives of patients with AATD

AAT, α_1 -antitrypsin; AATD, α_1 -antitrypsin deficiency; COPD, chronic obstructive pulmonary disease; GPA, granulomatosis with polyangiitis. Reproduced with modifications from Miravitlles et al⁶⁹

isoelectric focusing or genotyping by amplification of PI*S or PI*Z alleles or by deoxyribonucleic acid extraction from circulating mononuclear cells or from mouth swabs for polymerase chain reaction analysis.² Dried blood samples are also widely used for genotyping, often along with testing for AAT levels. Moderate-to-good correlation with serum AAT concentrations can be achieved using dried blood samples.⁸¹ Notably, after enrollment in the Alpha-1 Coded Testing Study, genotyping using dried blood spot kits is available for free (https://alphaoneregistry.org/research_registry). Discrepancies between AAT protein levels and genotype results from dried blood samples need to be investigated further, either by additional assessment of phenotype or using more expensive diagnostic techniques such as gene sequencing.^{79,81,82} Moreover, a recently developed lateral flow assay, which can deliver results within 15 minutes, can detect the most clinically significant PI*Z alleles. However, the test is only designed to detect PI*Z alleles and additional testing is required to determine whether the patient is homoor heterozygous.74

5 | CURRENT TREATMENTS

As shown in Table 3, a series of assessments, medications, and vaccinations should be considered for individuals with AATD, in addition to a healthy lifestyle that includes a good diet, exercise, and avoidance of stimuli of neutrophilic inflammation, including cigarette smoke, air pollution, and infections.^{83,84} Asthma should be treated aggressively.⁸⁵ Similarly, respiratory infections should be treated promptly and, as indicated by a study of completed questionnaires from 267 individuals with AAT, vaccinations against pneumococci (both protein conjugated and polysaccharide pneumococccal vaccines) and influenza should result in fewer exacerbations of lung disease.^{47,85} In addition, maintaining ideal body weight, limiting alcohol consumption and hepatitis A and B vaccine should be administered to prevent exacerbating already compromised liver disease from obesity-, toxicity-, and viral-induced hepatitis. Otherwise, inhaled therapy mimics the treatment of non-AATD COPD. As disease progresses, pulmonary rehabilitation and oxygen therapy may be essential.

5.1 | Transplantation and lung volume reduction surgery

Although the efficacy of lung transplantation for AATD-related COPD is not firmly established, the condition accounted for the fourth-highest percentage (5.8%) of lung transplants in adults between January 1995 and June 2012.⁸⁶ In a retrospective study of 83 lung transplant recipients with PI*ZZ alleles, median survival times were significantly longer (11 years, 95% confidence interval [CI]: 9-14) versus 70 nontransplanted controls (5 years, 95% CI: 4-6) patients.⁸⁷ A recent UK study demonstrated improved quality of life postsurgery, with no difference in mortality when compared with controls.⁸⁸ In most cases, referral for a lung transplant is deferred until FEV₁ decrease to 30% or below.

With regard to liver transplantation, there are few reports of outcomes in patients with AATD.⁸⁹ Reviews of patient databases and case series suggest that AAT levels may normalize following liver transplantation in adults and children, although it is unclear whether this procedure has an impact on pulmonary outcomes.⁹⁰ Further research is required to assess the benefit-risk profile of liver, singlelung, and double-lung transplantation in patients with AATD and the need to augment patients' status postlung transplant.⁶⁹ Evidence for the use of augmentation therapy after lung transplantation is



FIGURE 3 Rates of lung density decrease at total lung capacity versus trough A1PI serum concentrations achieved (RAPID trial). A1PI, α_1 proteinase inhibitor. From: Chapman et al⁸⁰

TABLE 3 Care of the patient with AATD

Vaccination	Medications	Holistic health	Assessments
Polysaccharide pneumococcal vaccine	Short-acting beta-agonists as needed	Good diet to preserve weight. Limit alcohol consumption.	Follow spirometry regularly
Yearly influenza vaccine	Long-acting beta-agonists as per COPD guidelines	Exercise to maintain condition	Check liver function tests regularly
Protein conjugate pneumococcal vaccine	Anticholinergics as per COPD guidelines	Oxygen supplement, if needed	Ultrasound of liver for hepatoma yearly
Tetanus/diphtheria/pertussis vaccine	Inhaled corticosteroids	Respiratory therapy	When FEV ₁ falls below 30% consider lung transplant assessment
	Augmentation with AAT	Lung healthy living	Assess for depression and anxiety at each appointment
		Liver healthy living	Lung cancer screening, if indicated

AAT, α_1 -antitrypsin; AATD, α_1 -antitrypsin deficiency; COPD, chronic obstructive pulmonary disease; FEV₁, forced expiratory volume in 1 s. Constructed using information from Henao and Craig,¹²⁴ Köhnlein et al,⁴⁷ O'Brien et al,⁸³ Alam et al,⁸⁴ and Sutherland and Cherniack.⁸⁵

insufficient, and there is no consensus whether lung transplantation recipients with AATD should receive augmentation therapy. According to the data, only 13%-19% of AATD patients receive augmentation therapy after lung transplantation. Furthermore, the putative influence of previous augmentation therapy on lung recipients who discontinued this therapy following transplantation is entirely unknown.⁹¹

In general, lung volume reduction surgery (LVRS) is not recommended for patients with AATD. In a study of 10 patients with severe AATD, 2-year mortality was higher and exercise tolerance and FEV_1 were worse in patients randomized to LVRS versus medical treatment.⁹² The recent ERS guidelines state that LVRS may be considered in selected patients with AATD, after careful appraisal of risks and benefits, but further studies are needed to confirm the role of such therapy.⁶⁹

5.2 | Augmentation therapy

Intravenous augmentation therapy with infusions of purified AAT from pooled human plasma aims to raise and maintain serum AAT levels above the 11 µmol/L estimated protective threshold value.² Augmentation therapy is recommended only for patients who are below this protective threshold, that is, mainly those with the PI*ZZ genotype.² At present, augmentation therapy is the only approved medication that raises AAT levels both in the plasma and in the epithelial lining fluid (ELF), leading to a reduction in neutrophil elastase activity in the lungs.⁹³⁻⁹⁵ Intravenous augmentation remains the only disease-specific therapy in AATD, and there is evidence that this slows decline in emphysema determined by CT density.⁹⁶ While the commercially available augmentation products (Prolastin, Zemaira/Respreeza, Glassia, and Aralast) have different purification processes and concentrations of AAT, there are only minor differences in storage, need for mixing, infusion rate, and cost, although there is a lack of comparative studies of the effectiveness of these products on lung parameters.97,98

Several recently completed or ongoing randomized clinical trials (including RAPID, EXACTLE, SPARK, and SPARTA) have increased

our understanding of augmentation therapy with purified AAT preparations (Table 4). These new studies benefited from the increased accuracy of CT to detect changes in lung density, relative to the previous gold standard of FEV_1 , which was used in studies in the 1990s.⁸² Thus, the use of CT has overcome the impracticality of performing an adequately powered randomized placebo-controlled trial to assess the development of emphysema using FEV_1 , which changes slowly over time.

The RAPID trials are the largest clinical trials of augmentation therapy completed to date, with a treatment period of 4 years, and are the only studies designed to investigate the disease-modifying effects of treatment. RAPID was a 2-year, multicenter, randomized placebo-controlled trial of 60 mg/kg weekly AAT (Zemaira, CSL Behring, KOP, PA, USA) in 180 patients with AATD. Most patients continued into the 2-year RAPID open-label extension trial, in which all patients were treated with active therapy; thus, the patients formed two groups: early-start (4 years of active treatment) and delayedstart (2 years of placebo followed by 2 years of active treatment). In both RAPID trials, the loss of lung parenchyma was statistically significantly slowed by approximately 34% in individuals treated with AAT, as ascertained by CT-measured lung density at total lung capacity.^{80,99,100} The RAPID trials support the efficacy of augmentation therapy in slowing disease progression during 4 years of treatment and, as lost lung density during placebo treatment never recovered following augmentation therapy in the delayed-start group, the trials highlight the importance of early initiation of augmentation therapy (Figure 4). In a post hoc analysis of the RAPID trials, treatment with AAT was also associated with reduced elastin degradation as evident by biomarkers isodesmosine and desmosine (Figure 5).¹⁰¹

In the EXACTLE randomized placebo-controlled trial in 77 patients with AATD, CT scans suggested that patients could benefit from treatment with AAT (Prolastin C; Grifols, Barcelona, Spain) 60 mg/kg weekly.⁴¹ In the SPARK randomized crossover trial in 30 patients with AATD, more physiologic levels of serum AAT were gained following treatment with AAT (Prolastin C; Grifols) at a dose of 120 mg/kg weekly vs. 60 mg/kg.¹⁰² Importantly, while

Study; reference	Year	Study design	Duration	Experimental drug; dosage and regimen	Comparator	No. of patients	Primary efficacy parameter
EXACTLE ⁴¹	2009	Prospective, multicenter, randomized, double-blind, placebo-controlled, parallel- group trial	2-2.5 y	Prolastin C 60 mg/kg weekly	Placebo	77	Change in the 15th percentile lung density by CT
SPARK ¹⁰²	2013	Prospective, multicenter, randomized, double-blind crossover trial	16 wk	Prolastin C 120 mg/kg weekly	Prolastin C 60 mg/kg weekly	30	AUC _{0-7 days} , C _{max} , elimination rate, t _{1/2} , t _{max} , C _{trough}
SPARTA ¹⁰³	2013	Prospective, multicenter, randomized, double-blind, placebo-controlled trial	156 wk	Prolastin C 120 mg/kg weekly	Prolastin C 60 mg/kg weekly or placebo	339	Change in the 15th percentile lung density by CT
RAPID ^{80,100}	2014	Prospective, multicenter, randomized, double-blind, placebo-controlled, parallel- group trial followed by an open-label extension trial	4 y early-start (4 y of active treatment): delayed-start (2 y of placebo followed by 2 y of active treatment)	Zemaira 60 mg/kg weekly	Placebo	180	Change in the 15th percentile lung density by CT
AAT, α_1 -antitrypsin;	AUC, are	a under the curve; C _{max} , maximum pl	lasma concentration; C _{trough} , lowest	concentration prior to ac	dministration of next d	ose; CT, computed	tomography; $t_{1/2}$, half-life; t_{max} , time

Randomized clinical trials in AAT deficiency

TABLE 4





FIGURE 4 Estimated lung density decline over 48 mo (RAPID trial). n, number of patients. From: McElvaney et al¹⁰⁰

improvements in lung function were not detected in the RAPID, EXACTLE, or SPARK trials (as the trials were not sufficiently powered to show this effect), augmentation therapy appears to be an effective treatment, which slows lung deterioration in patients with severe AATD and COPD, as noted by lung density measured by CT. As a follow-up to SPARK, the ongoing SPARTA randomized placebocontrolled trial in 339 patients with AATD is comparing AAT (Prolastin C; Grifols) 120 mg/kg weekly and 60 mg/kg weekly, to assess both the change in 15th percentile lung density and the number of severe COPD exacerbations.¹⁰³

These recent randomized trials have added to less conclusive research from the 1990s and 1980s. In 1997, in a nonrandomized study of AAT 60 mg/kg weekly involving 295 patients, decline in FEV₁ was slower in treated versus untreated patients.³⁵ In 1999, the first randomized controlled trial of augmentation therapy performed with 56 patients; while no difference in FEV₁ was detected between the 250 mg/kg at 4-week intervals versus albumin, the decline in lung tissue was reduced in CT scans.³⁸ Since the late 1980s, several studies have demonstrated that 60 mg/kg AAT infused at weekly intervals maintained AAT levels above the protective threshold throughout the week in both plasma and ELF and, in the SPARK trial, 120 mg/kg weekly AAT was associated with increased serum AAT levels and was well tolerated.^{102,104-106} However, when administered at bi-weekly intervals in another study, the 120 mg/kg dose did not maintain AAT levels above the protective threshold.¹⁰⁷ At present, while augmentation therapy is only approved at doses of 60 mg/kg weekly in the United States, some clinicians prescribe higher doses when patients are not gaining adequate benefit (Figure 3).82

Currently, the commercially available augmentation products have comparable safety profiles. No deaths or viral transmissions have been reported.² Adverse events, which are usually rare and transient, include headache, nausea, and dizziness (<0.03 events per patient month).³⁶ Anaphylactic reactions have been reported for four patients, all of whom recovered.^{1,106,108} As pooled human plasma can contain small amounts of immunoglobulin A (IgA), anaphylaxis may be triggered in IgA-deficient individuals with anti-IgA antibodies.

to maximum plasma concentration.



FIGURE 5 Changes in DES/IDES plasma levels from baseline in the RAPID trials. A1Pl, α_1 proteinase inhibitor; DES/ IDES, desmosine/isodesmosine; n, number of patients. Figure used with permission from Chronic Obstructive Pulmonary Diseases: Journal of the COPD Foundation¹⁰¹

Therefore, it is suggested that patients should be checked for IgA deficiency before initiating augmentation therapy.¹

At present, augmentation therapy is recommended only for adult patients with high-risk deficiency alleles, serum AAT levels below normal, nonreversible airflow obstruction by spirometry, and who avoid respiratory irritants, such as cigarette smoke.¹⁰³ Augmentation therapy is not recommended for individuals with heterozygous PI alleles when serum AAT levels are above the protective threshold.¹⁰⁹ For patients with severe COPD related to AATD, the efficacy of augmentation therapy is lower when FEV₁ is <30% predicted, although some clinicians argue that these patients may benefit even when changes in airflow obstruction and FEV₁ decline are small.^{36,110,111}

6 | EXPERT ANALYSIS AND FUTURE RESEARCH

Since the discovery of AATD in the 1960s, knowledge and understanding of AAT and its deficiency have progressed significantly, although effective screening methods and optimal therapy have remained elusive. Further advances are required to improve the detection of AATD and to improve treatment outcomes. It appears likely that the greatest barrier to diagnosis is lack of awareness of the condition and of screening guidelines among clinicians,^{1,66-68,74} while other diagnostic hurdles include similarities between COPD related to AATD and other etiologies and the asymptomatic nature of AATD in many individuals. Some physicians may also avoid testing because they are unaware of effective treatments for AATD-related COPD although, in one survey, only 8% of physicians believed no therapy existed.^{74,112}

To increase awareness, one investigated intervention was to issue an alert in electronic health records to test for AATD. The alert consisted of a pop-up reminder on the computer of all main-campus Cleveland Clinic physicians using the hospital's electronic health record system, suggesting to order an AAT serum level or phenotype test when the results of the patient's pulmonary function test show airflow obstruction consistent with \geq Global Initiative for Chronic Obstructive Lung Disease stage 1 (ie, FEV₁/forced vital capacity <0.70, with FEV₁<80% predicted). However, this alert did not result in a higher diagnostic rate for severe AATD, possibly due to a high baseline detection rate before the electronic alert or due to the small percentage of physicians (19%) who requested testing after the alert.¹¹³ It remains to be determined whether the use of electronic medical records, combined with increased awareness of AATD and testing guidelines, will increase the detection rate for AATD. With regard to diagnostic techniques, advances in "next-generation sequencing" may provide a highly sensitive, fast, and economical screening test for AATD.⁷⁴

Several support groups for patients with AATD have now been set up around the world and have been praised as a paradigm for confronting rare diseases.¹¹⁴ In addition to providing psychological support and helping to increase awareness of AATD, these groups have been integral to the development and maintenance of extensive registries for use in retrospective and prospective trials, and have raised large sums of money for research.^{7,115} As of April 2017, AlphaNet has contributed over \$50 million to research funding via the Alpha-1 Foundation. AlphaNet operates as a self-sustaining model where fees for services provided at no cost to individuals with AATD are covered by biologic and the pharmaceutical companies developing or manufacturing therapies for AATD.¹¹⁴

Ongoing and recent research has explored novel therapies for the treatment of AATD. Gene therapy is presently being studied, but the limitation is inflammation of the liver, which can possibly be overcome by immunosuppressive therapy. In the mouse model, there is selective advantage of the hepatocyte with wild-type AAT over the hepatocyte with the Z mutant, which may allow the repopulation of the liver with the edited hepatocytes and increase production of normal AAT.¹¹⁶

When developing potential treatments for AATD, in addition to assessing their effectiveness on AATD-associated pulmonary and liver parameters, product availability and financial viability also need

to be considered. At present, augmentation therapy is the only approved medication that effectively raises AAT levels both in serum and ELF. However, the costs of acquiring human plasma-derived AAT are high, and treatment costs of \$60 000 to \$150 000 per year (depending on body weight, pricing, and the costs of nursing care) exceed the standard criterion for cost-effectiveness of \$50 000 per quality-adjusted life-year.^{2,117} Thus, more cost-effective treatment would be welcomed. With regard to efficacy, some studies indicate that the protective threshold of serum AAT (11 μ mol/L, relative to 20-53 μ mol/L in healthy individuals) may need to be revised.^{27,74,80} The results of dose-ranging studies of augmentation therapy are eagerly awaited.¹⁰² Similarly, while previous trials were not successful, further research into increasing the interval between infusions may result in appropriate efficacy, potentially with lower treatment costs.

A recent attempt to develop a guideline on management and treatment of AATD was published in 2016.¹¹⁸ This publication was more of a consensus than a guideline, but is important reading for those that are managing patients with AATD, and agrees with previous recommendations with only a few outstanding exceptions. One exception is that while augmentation should be given to those AATD patients with FEV₁ \leq 65%, treating at FEV₁ >65% should be decided upon on a case-by-case basis. In addition, augmentation therapy is recommended for individuals with necrotizing panniculitis. Importantly, the guideline also clarifies augmentation should not be offered to patients with the PI*MZ genotype.

In summary, several key issues should be borne in mind. AATD is highly under-recognized; less than 10% of cases are diagnosed. For patients presenting with liver and/or lung disease, screening and diagnostic tests are the only methods to determine whether they are related to AATD. In accordance with guidelines issued by bodies such as the WHO, ERS, and American Thoracic Society, individuals at high risk of having AATD should be tested, including individuals with COPD, nonresponsive asthma, bronchiectasis of unknown etiology, or unexplained liver disease.^{1,66} PI*ZZ is the allele most associated with increased likelihood of developing COPD, although individuals with PI*SZ may be at risk, and PI*MZ alleles show increased risk, but less than ZZ, null Z, or null-null. Current therapy is comprised of aggressive treatment of asthma, vaccinations, and antibiotics against respiratory infections, maintaining ideal body weight, exercise, good diet, limiting alcohol, COPD medications and, if necessary, augmentation with AAT.

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CONFLICT OF INTERESTS

Timothy J. Craig does research for Biocryst, CSL Behring, Grifols, and Shire. He speaks for CSL Behring, Pharming and Grifols. He

consults with Biocryst and CSL Behring. He is also on the Advisory Board of the HAE-A. Maria Paula Henao has no conflict of interests or financial disclosures.

REFERENCES

- American Thoracic Society, European Respiratory Society. American Thoracic Society/European Respiratory Society statement: standards for the diagnosis and management of individuals with alpha-1 antitrypsin deficiency. Am J Respir Crit Care Med. 2003;168:818-900.
- Stoller JK, Aboussouan LS. A review of α1-antitrypsin deficiency. Am J Respir Crit Care Med. 2012;185:246-259.
- 3. Wanner A, Sandhaus RA, editors. *Alpha-1 Antitrypsin: Role in Health and Disease*. New York, NY: Humana Press; 2016.
- McElvaney NG. Diagnosing α1-antitrypsin deficiency: how to improve the current algorithm. *Eur Respir Rev.* 2015;24:52-57.
- Stoller JK, Brantly M. The challenge of detecting alpha-1 antitrypsin deficiency. COPD. 2013;10(Suppl 1):26-34.
- Campos M, Shmuels D, Walsh J. Detection of alpha-1 antitrypsin deficiency in the US. Am J Med. 2012;125:623-624.
- Stoller JK, Smith P, Yang P, Spray J. Physical and social impact of alpha 1-antitrypsin deficiency: results of a survey. *Cleve Clin J Med.* 1994;61:461-467.
- Craig TJ. Suspecting and testing for alpha-1 antitrypsin deficiencyan allergist's and/or immunologist's perspective. J Allergy Clin Immunol Pract. 2015;3:506-511.
- Kelbel T, Morros D, Walker D, Henao MP, Craig T. The allergist's role in detection of severe alpha-1 antitrypsin deficiency. J Allergy Clin Immunol Pract. 2017;5:3102-3106.
- De Serres FJ, Blanco I. Prevalence of α1-antitrypsin deficiency alleles PI*S and PI*Z worldwide and effective screening for each of the five phenotypic classes PI*MS, PI*MZ, PI*SS, PI*SZ, and PI*ZZ: a comprehensive review. *Ther Adv Respir Dis.* 2012;6:277-295.
- Brantly ML, Paul LD, Miller BH, Falk RT, Wu M, Crystal RG. Clinical features and history of the destructive lung disease associated with alpha-1-antitrypsin deficiency of adults with pulmonary symptoms. *Am Rev Respir Dis.* 1988;138:327-336.
- Qiu W, Baccarelli A, Carey VJ, et al. Variable DNA methylation is associated with chronic obstructive pulmonary disease and lung function. Am J Respir Crit Care Med. 2012;185:373-381.
- Green CE, Vayalapra S, Hampson JA, Mukherjee D, Stockley RA, Turner AM. PiSZ alpha-1 antitrypsin deficiency (AATD): pulmonary phenotype and prognosis relative to PiZZ AATD and PiMM COPD. *Thorax.* 2015;70:939-945.
- Bornhorst JA, Greene DN, Ashwood ER, Grenache DG. α1-Antitrypsin phenotypes and associated serum protein concentrations in a large clinical population. *Chest.* 2013;143:1000-1008.
- Ferrarotti I, Thun GA, Zorzetto M, et al. Serum levels and genotype distribution of α1-antitrypsin in the general population. *Thorax*. 2012;67:669-674.
- Brode SK, Ling SC, Chapman KR. Alpha-1 antitrypsin deficiency: a commonly overlooked cause of lung disease. CMAJ. 2012;184:1365-1371.
- Luisetti M, Seersholm N. Alpha1-antitrypsin deficiency. 1: epidemiology of alpha1-antitrypsin deficiency. *Thorax*. 2004;59:164-169.
- Vandeville D, Martin JP, Ropartz C. Alpha 1-antitrypsin polymorphism of a Bantu population: description of a new allele PiL. *Humangenetik*. 1974;21:33-38.
- Harada S, Miyake K, Suzuki H, Oda T. New phenotypes of serum alpha1-antitrypsin in Japanese detected by gel slab isoelectric focusing. *Hum Genet*. 1977;38:333-336.
- Massi G, Vecchio FM. Alpha-1-antitrypsin phenotypes in a group of newborn infants in Somalia. *Hum Genet*. 1977;38:265-269.

- 21. De Serres FJ, Blanco I, Fernández-Bustillo E. PI, S and PI Z alpha-1 antitrypsin deficiency worldwide. A review of existing genetic epidemiological data. *Monaldi Arch Chest Dis.* 2007;67:184-208.
- 22. De Serres FJ, Blanco I, Fernández-Bustillo E. Estimates of PI* S and PI* Z Alpha-1 antitrypsin deficiency alleles prevalence in the Caribbean and North, Central and South America. *Monaldi Arch Chest Dis.* 2009;71:96-105.
- 23. Spínola C, Brehm A, Spínola H. Alpha-1-antitrypsin deficiency in the Cape Verde islands (Northwest Africa): high prevalence in a sub-Saharan population. *Respir Med.* 2010;104:1069-1072.
- Aljarallah B, Ali A, Dowaidar M, Settin A. Prevalence of α-1-antitrypsin gene mutations in Saudi Arabia. Saudi J Gastroenterol. 2011;17:256-260.
- 25. De Serres FJ, Blanco I, Fernández-Bustillo E. Estimated numbers and prevalence of PI*S and PI*Z deficiency alleles of alpha1-antitrypsin deficiency in Asia. *Eur Respir J.* 2006;28:1091-1099.
- Blanco I, de Serres FJ, Carcaba V, Lara B, Fernández-Bustillo E. Alpha-1 antitrypsin deficiency PI*Z and PI*S gene frequency distribution using on maps of the world by an inverse distance weighting (IDW) multivariate interpolation method. *Hepat Mon.* 2012;12(10 HCC):e7434.
- McElvaney NG, Stoller JK, Buist AS, et al. Baseline characteristics of enrollees in the National Heart, Lung and Blood Institute Registry of alpha 1-antitrypsin deficiency. Alpha 1-Antitrypsin Deficiency Registry Study Group. *Chest.* 1997;111:394-403.
- Silverman EK, Miletich JP, Pierce JA, et al. Alpha-1-antitrypsin deficiency. High prevalence in the St. Louis area determined by direct population screening. *Am Rev Respir Dis.* 1989;140:961-966.
- 29. Petrache I, Hajjar J, Campos M. Safety and efficacy of alpha-1-antitrypsin augmentation therapy in the treatment of patients with alpha-1-antitrypsin deficiency. *Biol Targets Ther.* 2009;3:193-204.
- Parr DG, Stoel BC, Stolk J, Stockley RA. Pattern of emphysema distribution in alpha1-antitrypsin deficiency influences lung function impairment. *Am J Respir Crit Care Med.* 2004;170:1172-1178.
- Eriksson S. A 30-year perspective on alpha 1-antitrypsin deficiency. Chest. 1996;110(Suppl 6):2375-2425.
- Buist AS, Burrows B, Eriksson S, Mittman C, Wu M. The natural history of air-flow obstruction in PiZ emphysema. Report of an NHLBI workshop. Am Rev Respir Dis. 1983;127:S43-S45.
- Wu MC, Eriksson S. Lung function, smoking and survival in severe alpha 1-antitrypsin deficiency, PiZZ. J Clin Epidemiol. 1988;41:1157-1165.
- Seersholm N, Kok-Jensen A, Dirksen A. Decline in FEV1 among patients with severe hereditary alpha 1-antitrypsin deficiency type PiZ. Am J Respir Crit Care Med. 1995;152:1922-1925.
- 35. Seersholm N, Wencker M, Banik N, et al. Does alpha1-antitrypsin augmentation therapy slow the annual decline in FEV1 in patients with severe hereditary alpha1-antitrypsin deficiency? Wissenschaftliche Arbeitsgemeinschaft zur Therapie von Lungenerkrankungen (WATL) alpha1-AT study group. *Eur Respir J.* 1997;10:2260-2263.
- Survival and FEV1 decline in individuals with severe deficiency of alpha1-antitrypsin. The Alpha-1-Antitrypsin Deficiency Registry Study Group. Am J Respir Crit Care Med. 1998;158:49-59.
- Piitulainen E, Eriksson S. Decline in FEV1 related to smoking status in individuals with severe alpha1-antitrypsin deficiency (PiZZ). Eur Respir J. 1999;13:247-251.
- Dirksen A, Dijkman JH, Madsen F, et al. A randomized clinical trial of alpha(1)-antitrypsin augmentation therapy. *Am J Respir Crit Care Med.* 1999;160:1468-1472.
- Wencker M, Fuhrmann B, Banik N, Konietzko N; Wissenschaftliche Arbeitsgemeinschaft zur Therapie von Lungenerkrankungen. Longitudinal follow-up of patients with alpha(1)-protease inhibitor deficiency before and during therapy with IV alpha(1)-protease inhibitor. *Chest.* 2001;119:737-744.

- Dawkins PA, Dawkins CL, Wood AM, Nightingale PG, Stockley JA, Stockley RA. Rate of progression of lung function impairment in alpha1-antitrypsin deficiency. *Eur Respir J.* 2009;33:1338-1344.
- 41. Dirksen A, Piitulainen E, Parr DG, et al. Exploring the role of CT densitometry: a randomised study of augmentation therapy in alpha1-antitrypsin deficiency. *Eur Respir J*. 2009;33:1345-1353.
- Tonelli AR, Rouhani F, Li N, Schreck P, Brantly ML. Alpha-1-antitrypsin augmentation therapy in deficient individuals enrolled in the Alpha-1 Foundation DNA and Tissue Bank. Int J Chron Obstruct Pulmon Dis. 2009;4:443-452.
- Demeo DL, Sandhaus RA, Barker AF, et al. Determinants of airflow obstruction in severe alpha-1-antitrypsin deficiency. *Thorax.* 2007;62:806-813.
- Castaldi PJ, DeMeo DL, Kent DM, et al. Development of predictive models for airflow obstruction in alpha-1-antitrypsin deficiency. Am J Epidemiol. 2009;170:1005-1013.
- Banauch GI, Brantly M, Izbicki G, et al. Accelerated spirometric decline in New York City firefighters with α₁-antitrypsin deficiency. *Chest.* 2010;138:1116-1124.
- Wood AM, Harrison RM, Semple S, Ayres JG, Stockley RA. Outdoor air pollution is associated with rapid decline of lung function in alpha-1-antitrypsin deficiency. *Occup Environ Med.* 2010;67:556-561.
- Köhnlein T, Janciauskiene S, Welte T. Diagnostic delay and clinical modifiers in alpha-1 antitrypsin deficiency. *Ther Adv Respir Dis*. 2010;4:279-287.
- Piitulainen E, Montero LC, Nystedt-Düzakin M, et al. Lung function and CT densitometry in subjects with alpha-1-antitrypsin deficiency and healthy controls at 35 years of age. COPD. 2015;12:162-167.
- Stoller JK, Tomashefski J, Crystal RG, et al. Mortality in individuals with severe deficiency of alpha1-antitrypsin: findings from the National Heart, Lung, and Blood Institute Registry. *Chest*. 2005;127:1196-1204.
- Tanash HA, Nilsson PM, Nilsson J-A, Piitulainen E. Clinical course and prognosis of never-smokers with severe alpha-1-antitrypsin deficiency (PiZZ). *Thorax*. 2008;63:1091-1095.
- Seersholm N, Kok-Jensen A. Clinical features and prognosis of life time non-smokers with severe alpha 1-antitrypsin deficiency. *Thorax.* 1998;53:265-268.
- Molloy K, Hersh CP, Morris VB, et al. Clarification of the risk of chronic obstructive pulmonary disease in α1-antitrypsin deficiency PiMZ heterozygotes. Am J Respir Crit Care Med. 2014;189:419-427.
- Cuvelier A, Muir JF, Hellot MF, et al. Distribution of alpha(1)-antitrypsin alleles in patients with bronchiectasis. *Chest.* 2000;117:415-419.
- Parr DG, Guest PG, Reynolds JH, Dowson LJ, Stockley RA. Prevalence and impact of bronchiectasis in alpha1-antitrypsin deficiency. *Am J Respir Crit Care Med.* 2007;176:1215-1221.
- Lomas DA, Evans DL, Finch JT, Carrell RW. The mechanism of Z alpha 1-antitrypsin accumulation in the liver. *Nature*. 1992;357:605-607.
- 56. Gishen P, Saunders AJ, Tobin MJ, Hutchison DC. Alpha 1-antitrypsin deficiency: the radiological features of pulmonary emphysema in subjects of Pi type Z and Pi type SZ: a survey by the British Thoracic Association. *Clin Radiol.* 1982;33:371-377.
- 57. Fairbanks KD, Tavill AS. Liver disease in alpha 1-antitrypsin deficiency: a review. Am J Gastroenterol. 2008;103:2136-2141.
- Sveger T. The natural history of liver disease in alpha 1-antitrypsin deficient children. Acta Paediatr Scand. 1988;77:847-851.
- Psacharopoulos HT, Mowat AP, Cook PJ, Carlile PA, Portmann B, Rodeck CH. Outcome of liver disease associated with alpha 1 antitrypsin deficiency (PiZ). Implications for genetic counselling and antenatal diagnosis. Arch Dis Child. 1983;58:882-887.
- Topic A, Prokic D, Stankovic I. Alpha-1-antitrypsin deficiency in early childhood. *Fetal Pediatr Pathol*. 2011;30:312-319.

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- Rider NL, Craig TJ. Liver enzyme elevation and normal pulmonary function in an adult with a declining forced expiratory volume in 1 second. Allergy Asthma Proc. 2008;29:345-348.
- 62. Sveger T. Liver disease in alpha1-antitrypsin deficiency detected by screening of 200,000 infants. N Engl J Med. 1976;294:1316-1321.
- Esnault VL, Testa A, Audrain M, et al. Alpha 1-antitrypsin genetic polymorphism in ANCA-positive systemic vasculitis. *Kidney Int*. 1993;43:1329-1332.
- O'Donoghue DJ, Guickian M, Blundell G, Winney RJ. Alpha-1-proteinase inhibitor and pulmonary haemorrhage in systemic vasculitis. *Adv Exp Med Biol.* 1993;336:331-335.
- 65. Gross B, Grebe M, Wencker M, Stoller JK, Bjursten LM, Janciauskiene S. New findings in PiZZ alpha1-antitrypsin deficiencyrelated panniculitis. Demonstration of skin polymers and high dosing requirements of intravenous augmentation therapy. *Dermatol Basel Switz*. 2009;218:370-375.
- Alpha 1-antitrypsin deficiency: memorandum from a WHO meeting. Bull World Health Organ. 1997;75:397-415.
- Vestbo J, Hurd SS, Agustí AG, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2013;187:347-365.
- Marciniuk DD, Hernandez P, Balter M, et al. Alpha-1 antitrypsin deficiency targeted testing and augmentation therapy: a Canadian Thoracic Society clinical practice guideline. *Can Respir J.* 2012;19: 109-116.
- Miravitlles M, Dirksen A, Ferrarotti I, et al. European Respiratory Society statement: diagnosis and treatment of pulmonary disease in α1-antitrypsin deficiency. *Eur Respir J.* 2017;50:1-24.
- Thelin T, Sveger T, McNeil TF. Primary prevention in a high-risk group: smoking habits in adolescents with homozygous alpha-1antitrypsin deficiency (ATD). Acta Paediatr. 1996;85:1207-1212.
- Teckman J, Pardee E, Howell RR, et al. Appropriateness of newborn screening for α1-antitrypsin deficiency. J Pediatr Gastroenterol Nutr. 2014;58:199-203.
- 72. Tretter JT. Adding α-1 antitrypsin deficiency to the newborn screen. J Pediatr Gastroenterol Nutr. 2015;60:e37.
- 73. Teckman J. Author's response. J Pediatr Gastroenterol Nutr. 2015;60:e38.
- Greulich T, Vogelmeier CF. Alpha-1-antitrypsin deficiency: increasing awareness and improving diagnosis. *Ther Adv Respir Dis.* 2016;10:72-84.
- Steiner SJ, Gupta SK, Croffie JM, Fitzgerald JF. Serum levels of alpha1-antitrypsin predict phenotypic expression of the alpha1-antitrypsin gene. *Dig Dis Sci.* 2003;48:1793-1796.
- Bals R, Koczulla R, Kotke V, Andress J, Blackert K, Vogelmeier C. Identification of individuals with alpha-1-antitrypsin deficiency by a targeted screening program. *Respir Med.* 2007;101:1708-1714.
- Ferrarotti I, Scabini R, Campo I, et al. Laboratory diagnosis of alpha1-antitrypsin deficiency. *Transl Res.* 2007;150:267-274.
- Snyder MR, Katzmann JA, Butz ML, et al. Diagnosis of alpha-1-antitrypsin deficiency: an algorithm of quantification, genotyping, and phenotyping. *Clin Chem.* 2006;52:2236-2242.
- Miravitlles M, Herr C, Ferrarotti I, et al. Laboratory testing of individuals with severe alpha1-antitrypsin deficiency in three European centres. *Eur Respir J.* 2010;35:960-968.
- Chapman KR, Burdon JGW, Piitulainen E, et al. Intravenous augmentation treatment and lung density in severe α1 antitrypsin deficiency (RAPID): a randomised, double-blind, placebo-controlled trial. *Lancet*. 2015;386:360-368.
- Costa X, Jardi R, Rodriguez F, et al. Simple method for alpha1-antitrypsin deficiency screening by use of dried blood spot specimens. *Eur Respir J*. 2000;15:1111-1115.
- Campos MA, Lascano J. α1 Antitrypsin deficiency: current best practice in testing and augmentation therapy. *Ther Adv Respir Dis.* 2014;8:150-161.

- 83. O'Brien ME, Pennycooke K, Carroll TP, et al. The impact of smoke exposure on the clinical phenotype of alpha-1 antitrypsin deficiency in Ireland: exploiting a national registry to understand a rare disease. COPD. 2015;12(Suppl 1):2-9.
- Alam S, Li Z, Janciauskiene S, Mahadeva R. Oxidation of Z α1-antitrypsin by cigarette smoke induces polymerization: a novel mechanism of early-onset emphysema. Am J Respir Cell Mol Biol. 2011;45:261-269.
- Sutherland ER, Cherniack RM. Management of chronic obstructive pulmonary disease. N Engl J Med. 2004;350:2689-2697.
- Yusen RD, Christie JD, Edwards LB, et al. The Registry of the International Society for Heart and Lung Transplantation: Thirtieth Adult Lung and Heart-Lung Transplant Report-2013; focus theme: age. J Heart Lung Transplant. 2013;32:965-978.
- Tanash HA, Riise GC, Hansson L, Nilsson PM, Piitulainen E. Survival benefit of lung transplantation in individuals with severe α1-antitrypsin deficiency (PiZZ) and emphysema. J Heart Lung Transplant. 2011;30:1342-1347.
- Stone HM, Edgar RG, Thompson RD, Stockley RA. Lung transplantation in alpha-1-antitrypsin deficiency. COPD. 2016;13:146-252.
- Townsend SA, Edgar RG, Ellis PR, Kantas D, Newsome PN, Turner AM. Systematic review: the natural history of alpha-1 antitrypsin deficiency, and associated liver disease. *Aliment Pharmacol Ther*. 2018;47:877-885.
- Hood JM, Koep LJ, Peters RL, et al. Liver transplantation for advanced liver disease with alpha-1-antitrypsin deficiency. N Engl J Med. 1980;302:272-275.
- Conrad A, Janciauskiene S, Köhnlein T, et al. Impact of alpha 1-antitrypsin deficiency and prior augmentation therapy on patients' survival after lung transplantation. *Eur Respir J.* 2017;50:1-4.
- Stoller JK, Gildea TR, Ries AL, Meli YM, Karafa MT; National Emphysema Treatment Trial Research Group. Lung volume reduction surgery in patients with emphysema and alpha-1 antitrypsin deficiency. Ann Thorac Surg. 2007;83:241-251.
- Stoller JK, Aboussouan LS. Alpha1-antitrypsin deficiency. 5: intravenous augmentation therapy: current understanding. *Thorax*. 2004;59:708-712.
- Sandhaus RA. Alpha1-antitrypsin deficiency. 6: new and emerging treatments for alpha1-antitrypsin deficiency. *Thorax*. 2004;59:904-909.
- Gadek JE, Hosea SW, Gelfand JA, et al. Replacement therapy in hereditary angioedema: successful treatment of acute episodes of angioedema with partly purified C1 inhibitor. N Engl J Med. 1980;302:542-546.
- Edgar RG, Patel M, Bayliss S, Crossley D, Sapey E, Turner AM. Treatment of lung disease in alpha-1 antitrypsin deficiency: a systematic review. Int J Chron Obstruct Pulmon Dis. 2017;12:1295-1308.
- Stoller JK, Aboussouan LS. Alpha1-antitrypsin deficiency. Lancet. 2005;365:2225-2236.
- Lomas DA, Dickens JA. Why has it been so difficult to prove the efficacy of alpha-1-antitrypsin replacement therapy? Insights from the study of disease pathogenesis. *Drug Des Devel Ther*. 2011;5:391-405.
- 99. Chorostowska-Wynimko J. Disease modification in emphysema related to alpha-1 antitrypsin deficiency. *COPD*. 2016;13:807-815.
- 100. McElvaney NG, Burdon J, Holmes M, et al. Long-term efficacy and safety of α 1 proteinase inhibitor treatment for emphysema caused by severe α 1 antitrypsin deficiency: an open-label extension trial (RAPID-OLE). *Lancet Respir Med.* 2017;5:51-60.
- 101. Ma S, Lin YY, Cantor JO, et al. The effect of alpha-1 proteinase inhibitor on biomarkers of elastin degradation in alpha-1 antitrypsin deficiency: an analysis of the RAPID/RAPID Extension trials. Chronic Obstr Pulm Dis (Miami). 2017;4:34-44.
- Campos MA, Kueppers F, Stocks JM, et al. Safety and pharmacokinetics of 120 mg/kg versus 60 mg/kg weekly intravenous infusions

of alpha-1 proteinase inhibitor in alpha-1 antitrypsin deficiency: a multicenter, randomized, double-blind, crossover study (SPARK). *COPD*. 2013;10:687-695.

- 103. Sorrells S, Camprubi S, Griffin R, Chen J, Ayguasanosa J. SPARTA clinical trial design: exploring the efficacy and safety of two dose regimens of alpha1-proteinase inhibitor augmentation therapy in alpha1-antitrypsin deficiency. *Respir Med.* 2015;109:490-499.
- Hubbard RC, Crystal RG. Alpha-1-antitrypsin augmentation therapy for alpha-1-antitrypsin deficiency. *Am J Med.* 1988;84:52-62.
- 105. Hubbard RC, Crystal RG. Augmentation therapy of alpha 1-antitrypsin deficiency. *Eur Respir J Suppl.* 1990;9:44s-52s.
- 106. Wewers MD, Crystal RG. Alpha-1 antitrypsin augmentation therapy. COPD. 2013;10:64-67.
- Barker AF, Iwata-Morgan I, Oveson L, Roussel R. Pharmacokinetic study of alpha1-antitrypsin infusion in alpha1-antitrypsin deficiency. *Chest.* 1997;112:607-613.
- Meyer FJ, Wencker M, Teschler H, et al. Acute allergic reaction and demonstration of specific IgE antibodies against alpha-1-protease inhibitor. *Eur Respir J.* 1998;12:996-997.
- Sandhaus RA, Turino G, Stocks J, et al. Alpha1-Antitrypsin augmentation therapy for PI*MZ heterozygotes: a cautionary note. *Chest.* 2008;134:831-834.
- 110. Stockley RA, Turner AM. α-1-Antitrypsin deficiency: clinical variability, assessment, and treatment. *Trends Mol Med.* 2014;20:105-115.
- 111. Chapman KR, Stockley RA, Dawkins C, Wilkes MM, Navickis RJ. Augmentation therapy for alpha1 antitrypsin deficiency: a metaanalysis. *COPD*. 2009;6:177-184.
- 112. Greulich T, Ottaviani S, Bals R, et al. Alpha1-antitrypsin deficiency diagnostic testing and disease awareness in Germany and Italy. *Respir Med.* 2013;107:1400-1408.
- 113. Jain A, McCarthy K, Xu M, Stoller JK. Impact of a clinical decision support system in an electronic health record to enhance detection of α_1 -antitrypsin deficiency. *Chest.* 2011;140:198-204.
- 114. Walsh JW. The alpha-1 constellation of voluntary health organizations as a paradigm for confronting rare diseases. In: Wanner A, Sandhaus RA, eds. *Alpha-1 Antitrypsin: Role in Health and Disease*. New York, NY: Humana Press; 2016:157-170.

115. Stoller JK. Augmentation therapy for severe α1-antitrypsin deficiency: is the jury still out on a trial? *Thorax*. 1998;53:1007-1009.

- 116. Borel F, Tang Q, Gemoux G, et al. Survival advantage of both human hepatocytes xenografts and genome edited hepatocytes for treatment of α -1 antitrypsin deficiency. *Mol Ther.* 2017;25:2477-2489.
- 117. Silverman EK, Sandhaus RA. Clinical practice. Alpha1-antitrypsin deficiency. N Engl J Med. 2009;360:2749-2757.
- 118. Sandhaus RA, Turino G, Brantly ML, et al. The diagnosis and management of alpha-1 antitrypsin deficiency in the adult. *Chronic Obstr Pulm Dis* (*Miami*). 2016;3:668-682.
- 119. Lieberman J, Winter B, Sastre A. Alpha 1-antitrypsin Pi-types in 965 COPD patients. *Chest.* 1986;89:370-373.
- 120. Shigeoka JW, Hall WJ, Hyde RW, et al. The prevalence of alphaantitrypsin heterozygotes (Pi MZ) in patients with obstructive pulmonary disease. *Am Rev Respir Dis.* 1976;114:1077-1084.
- 121. Cox DW, Hoeppner VH, Levison H. Protease inhibitors in patients with chronic obstructive pulmonary disease: the alpha-antitrypsin heterozygote controversy. *Am Rev Respir Dis.* 1976;113:601-606.
- Morse JO, Lebowitz MD, Knudson RJ, Burrows B. Relation of protease inhibitor phenotypes to obstructive lung diseases in a community. N Engl J Med. 1977:296:1190-1194.
- 123. Dahl M, Hersh CP, Ly NP, Berkey CS, Silverman EK, Nordestgaard BG. The protease inhibitor PI*S allele and COPD: a meta-analysis. *Eur Respir J.* 2005;26:67-76.
- 124. Henao MP, Craig TJ. Understanding alpha-1 antitrypsin deficiency: a review with an allergist's outlook. *Allergy Asthma Proc.* 2017;38:98-107.

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