Hindawi Publishing Corporation Analytical Cellular Pathology Volume 2015, Article ID 150634, 8 pages http://dx.doi.org/10.1155/2015/150634

Review Article

Chemoresistance, Cancer Stem Cells, and miRNA Influences: The Case for Neuroblastoma

Alfred Buhagiar¹ and Duncan Ayers^{2,3}

¹School of Clinical Sciences, Faculty of Medicine and Dentistry, University of Bristol, Bristol BS8 1TH, UK

Correspondence should be addressed to Duncan Ayers; duncan.ayers@googlemail.com

Received 29 April 2015; Revised 26 June 2015; Accepted 1 July 2015

Academic Editor: Nils Ole Schmidt

Copyright © 2015 A. Buhagiar and D. Ayers. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited

Neuroblastoma is a type of cancer that develops most often in infants and children under the age of five years. Neuroblastoma originates within the peripheral sympathetic ganglia, with 30% of the cases developing within the adrenal medulla, although it can also occur within other regions of the body such as nerve tissue in the spinal cord, neck, chest, abdomen, and pelvis. MicroRNAs (miRNAs) regulate cellular pathways, differentiation, apoptosis, and stem cell maintenance. Such miRNAs regulate genes involved in cellular processes. Consequently, they are implicated in the regulation of a spectrum of signaling pathways within the cell. In essence, the role of miRNAs in the development of cancer is of utmost importance for the understanding of dysfunctional cellular pathways that lead to the conversion of normal cells into cancer cells. This review focuses on highlighting the recent, important implications of miRNAs within the context of neuroblastoma basic research efforts, particularly concerning miRNA influences on cancer stem cell pathology and chemoresistance pathology for this condition, together with development of translational medicine approaches for novel diagnostic tools and therapies for this neuroblastoma.

1. Introduction

Neuroblastoma (NB) is a paediatric cancer deriving from the neural-crest cells [1]. The condition is commonly inflicted upon young children prior to the age of five years, and in the USA/UK it accounts for 7-8% of childhood cancers [2–4]. Tumourigenesis typically occurs in the adrenal gland, sympathetic ganglia, and paraganglia or along the spinal cord [5]. It is a highly heterogeneous disease exhibiting variation in clinical appearance from localized to metastatic tissue and can undergo metastasis to the liver, bones, brain, and skin [6].

2. Origin of NB

Two major causes have been identified in the origin of the disease.

The first cause is *Familial origin* which is identified in the *PHOX2B* gene [7]. This loss-of-function mutation is also

present in other congenital diseases such as Hirschsprung's disease and congenital central hypoventilation syndrome [8]. In addition, activating mutations in the anaplastic lymphoma kinase gene have been identified as the leading cause of familial NB [8–10]. ALK forms part of the tyrosine kinases receptors which are associated with cell surface receptors [11]. During early development of nerve cells it is thought the ALK gene helps in the proliferation of the nerve cells and their eventual regulation, with data highlighting that 8% of NB cases are due to mutations in the ALK gene [12]. However, it is not excluded that other genes could be identified as playing major roles in the development of familial NB following future research efforts [8].

The second cause is *Sporadic origin* which results in chromosomal losses [13], such as loss of the lp36.31 heterozygosity, and occurs in 36% of primary tumours. Chromosome gains can also occur, such as *MYCN* amplification, resulting in diploid or tetraploid DNA content within the affected

²Centre for Molecular Medicine and Biobanking, University of Malta, Msida MSD 2080, Malta

³Faculty of Medical and Human Sciences, The University of Manchester, Manchester M1 7DN, UK

cells [13]. Similar to other cancers, NB manifests itself in genetic alteration [14] of genes responsible for cell growth, cycle, and immunity including phenotypic changes that inhibit differentiation. Research conducted over a number of years reveals that the transcription factor MYCN [15] is the most important oncogene [1]. MYCN acts mainly by dysregulating downstream genes involved in key pathways that affect NB formation with putative effects by KIFAP3, OPHN, RGS7, ODC1, TOP2A, TWIST1, and TYMS on the same gene [16]. Moreover, a recent study revealed that although the MYCN gene expression was not amplified, a high level of MYC proteins was present in NB patient group [17]. Additionally, transgenic animal model studies indicated that tumourigenesis does not require gene amplification [18]. Conversely, in the presence of abnormal MYCN expression during the formation of the neural crest, the precursor cells transform into tumourigenic neuroblasts, due to the upregulated MYCN protein level [19]. This pathway was confirmed with the LIN28b gene. A genomic aberration in this gene results in repression of a set of microRNAs of the let-7 family. The let-7 miRNAs function as tumor suppressors through the silencing of oncogenes such as RAS, MYC, and CDK6. The outcome from this suppression was an elevated MYCN protein expression that was elucidated in mouse models. An interesting fact is that LIN28b is a homologue of LIN28, one of the regulators for pluripotency in stem cells, in combination with NANOG, OCT3/4, and SOX2 [20].

The physiological effect of the *MYCN* gene is highly influential. Intriguingly, it has the ability to activate angiogenic factors [21] leading to the formation of novel blood vessels to ensure an adequate supply of nutrients, hence repressing angiogenic inhibitors.

Following the existence of miRNA technology, research was focused on the relationship between *MYCN* [22] and miRNAs. Recent studies [23] highlighted that the gene regulates a number of miRNAs. A selection of these activated miRNAs is associated with clinically aggressive cancer conditions that include the miR-17-92 cluster, miR-18a, miR-7, miR-128, miR-380-5p, and miR-558.

3. Clinical Outcomes

Current treatments in NB are varied due to the heterogeneous nature of the disease. The outcome of the treatment depends on a multitude of factors, including age of patient, histology, genetic abnormalities [24], and stage of disease. NB is a high risk disease, with almost 20% of children experiencing relapse [25]. Poor prognosis of the disease leads to inadequate therapy, undetected residual tumour, and inefficient modalities of treatment; this leads to the need to standardize criteria for evaluation of the disease. Consequently a stage and risk classification was developed for clinical presentations of NB [26]. A panel of experts, the International NB Staging System (INSS), bases its categorization on the stage of the disease and subsequent removal by surgery [27]. If the disease does not need surgery, another panel (the International NB Risk Group Staging System, INRGSS) bases its categorization on clinical pretreatment [28]. Data collected from imaging, including CT scans and MRI, give an evaluation of the

disease. In addition to the acquired imaging data, bone marrow biopsy and status of the *MYCN* gene are also utilized for NB risk classification. Through the application of these criteria, new improved therapies with high survivability percentage could be achieved.

4. MiRNAs and Cancer

Since their discovery in 1993 in Caenorhabditis elegans [29], novel miRNA sequences have been identified in both plants and animals, with the database cataloging all miRNA sequences still increasing in number [30]. Such miRNAs are defined as single strands of not more than 25 nucleotides that are generated from genes located on the chromosome [30]. The generation of these miRNAs follows a four-step process involving the following [31]: (i) cropping: this involves the heterodimer Drosha cleaving the mature pri-miRNA into a smaller size called the pre-miRNA; (ii) exportation: this premiRNA is exported from the nucleus by combining with Exportin-5 and Ran-GTP; (iii) dicing: once in the cytoplasm they are cleaved into a smaller strand by RNase III enzyme known as the dicer, releasing the two small complementary short strands; (iv) formation of the RNA Interference Silencing Complex (RISC) complex, a ribonucleoprotein complex forming part of the argonaute family: one strand (the guide) forms a complex with ribonucleoprotein while the other strand (the passenger) is destroyed. The complex then binds to the mRNA leading to translational repression.

Since their discovery, the premise was established that there is a connection between miRNA activity and cancer [32]. The tumorigenic effect of miRNAs is typically induced by the dysregulation in miRNA expression through multiple mechanisms that include translocation, deletion, mutations, and rearrangements [32]. High-throughput screening studies and functional genomics research revealed the significant variations in miRNA expression profiles of normal cells in comparison with the cancer inflicted counterpart cells [30]. This striking evidence led to the concept of the miRnome, which is the characteristic malignant form. It was also elucidated that most cancer conditions demonstrate an overall downregulation in over 50% of the miRnome's individual miRNAs, though a few key tumourigenic miRNAs were found to be upregulated [33]. A feature of miRNAs is their tissue specificity. Apparently, miRNA expression is modulated according to the cancer cell population which is different from that of a normal cell population [30] (see Table 1). To be tumorigenic, miRNAs must induce production of growth factors (e.g., let-7) [34], be unresponsive to external antigrowth factors (e.g., miR-17/19) [35], and avoid cell apoptosis (e.g., miR-34a) [30], indefinite proliferation (e.g., miR-372/373) [36], blood vessels formation also known as angiogenesis (e.g., miR-210) [37], metastasis, and migration, with an increase in tumor macrophages and inflammation (e.g., miR-10b) [38, 39].

There are already 4469 miRNAs discovered, of which 1881 are precursors and 2588 are mature (http://www.mirbase.org/) and the list is increasing. To a large extent, some of the miRNAs involved in specific types of cancer progression have been identified [32, 33]:

Table 1: List of miRNAs identified to influence clinical progression in specific cancer conditions.

Cancer type	miRNA	
Lung cancer	miR-21, miR-125 miR-574-5p miR-155, miR-197, and miR-182	
Breast cancer		
Prostate cancer	miR-375, miR-9, miR-141, and miR-200b	
Colon cancer miR-221, miR-17-5, miR-29b-2, miR-22 miR-128b, miR-24-1, and miR-155		
Stomach cancer	miR-21, miR-191, miR-223, miR-107, miR-214, miR-221, and miR-25	

5. Chemoresistance, Cancer Stem Cells, and miRNA Influences

One of the modalities in treating cancer is chemotherapy and a recurring problem encountered during cancer treatment is chemoresistance. Resistance to chemotherapy is believed to cause over 90% failure [40] in metastatic cancer. This therapeutic failure is attributed to two mechanisms, intrinsic and extrinsic resistance [41]. Intrinsic resistance originates from cancer cells that have already the capability of withstanding chemotherapy [42]. Extrinsic resistance is the acquired capability of counteracting chemotherapy via genetic and epigenetic mutations in genes during continual chemotherapeutic treatments [43]. Seven major mechanisms have been presently elucidated [44] to explain multi drug resistance (MDR) in cancer:

- (A) Genetic alterations that empower the cells with an added function in their enzymatic pathways, such as the well-studied PI3K/Akt/mTOR [45]: The phosphorylation by these enzymes produces substrates that regulate apoptosis. These substrate proteins regulate cell cycle activity by being inactivated due to phosphorylation and consequently suspending apoptosis [46, 47]. Another example is the p53 mutation which is a well-known tumour suppressor [48]. Mutation within the *Tp53* gene results exacerbates metastasis and invasiveness [49]. Compared with other cancers which exhibit tumour expression via loss of function mutation, p53 suffers from point mutation [50] resulting in a single amino acid substitution. The mutation alters the resistance to drug toxicity by downregulating genes responsible for apoptosis [51].
- (B) Drug export modulations that are mediated by the ATP-binding cassette (ABC) genes [43, 52]: chemoresistance occurs since MDR proteins actively pump drugs outside the tumour cell membrane, thus reducing the intracellular effects of the drug on the tumour cell.
- (C) DNA mutations that allow cancer cells to accumulate genetic changes that favour selection towards chemoresistance [53]: this predisposition towards transformation makes cancer cells more resistant to chemotherapy.
- (D) Inhibition of apoptosis by inactivation of genes that encode for caspases: The latter are cysteine proteases that play an essential role in programmed cell death [54]. The failure of therapies to kill cancer cells is mainly due to the drugs that

are oriented towards inducing apoptosis. Tumour cells have managed to [55] avoid cell death by overexpressing proteins which have antiapoptotic properties.

- (E) The cellular environment also plays an important role in MDR: It was elucidated that the hypoxic conditions typically prevailing in tumour cells induce such cells to produce hypoxia inducible factors. The chaotic conditions and aggressive proliferation of cancer cause oxygen insufficiency [56, 57]. These hypoxic factors elicit a multitude of factors such as gene expression, protein overexpression, and antiapoptotic inhibitors which collectively compromise the effectiveness of chemotherapies [58].
- (F) Stromal cells, which are a form of connective tissue and are also developed in cancer: In this microenvironment it was established that these stromal cells, together with macrophages, synonymous with inflammation and endothelial cells, offer a large influence to the cancer cells in their progression and metastasis [59]. Cancer-associated fibroblasts secrete protein chemokines which promote metastasis in cancer stem cells [60].
- (G) The role of cancer stem cells is also a major contributing factor in the resurgence of cancer: Stem cells have the capacity to renew and differentiate into diverse specific cell types [61]. Stem cells have been identified from three sources: the inner cell mass of embryos, induced pluripotent from normal somatic cells [62], and somatic adult stem cells [63]. In 1994 it was discovered that cancer stem cells [64] exist in leukemia but since then, they have been discovered in other solid tumors. Similarly to normal stem cells, these cancer stem cells (CSCs) have the ability to regenerate and produce cells that differentiate into a tumour condition which exhibits MDR properties [65].

Asymmetric division in CSCs enables self-renewal and reconstitutes the tumour cells, synonymous of remission [66]. By definition, asymmetric division is the homeostatic control of cells that is maintained by producing copies identical to itself plus differentiated cells. These differentiated cells will be carrying the different characteristics attributed to exposure of external signals and or genetic mutations that maintain tumourogenic capabilities [67]. Tumor-initiating genetic aberrations are usually affected [68] via signaling pathways such as Wnt, Notch, Oct-4, and Hedgehog. The similarity of normal stem cells and CSCs is mediated through these pathways [69]. In Wnt/ β controls differentiation, Notch proteins located on cell membranes are implicated in a number of stem cells, including neuronal and haematopoietic tissue [70]. Oct-4, which is a transcription factor, regulates embryonal carcinomas [71] and the maintaining of undifferentiated carcinoma cells and also in pancreatic cancer [72]. Furthermore, it was also recognized that adult stem cells that were converted into CSCs engage in oncogenic pathways involving these genetic aberrations [73, 74]. It was also elucidated that miRNAs are able to promote reprogramming of cells even in induced pluripotent stem cell lines, such that cells become independent from their origin [75]. It was later recognized that transcription factor STAT3 is required for the maintenance of multipotency [76] in glioblastoma stem cells. It was also established that NB cell lines with CD114+ CSC expression markers demonstrated that such cells have a cell cycle typical for induced pluripotent and embryonic stem cells [77]. Thus in the face of these facts cancer stem cells show the same chemoresistance properties exhibited by normal stem cells and it is very hard to design new drugs because of the toxicity to normal cells.

Clearly, the clinical obstacle associated with drug resistance is the reduced effect of chemotherapeutic drugs, mainly due to efflux from the targeted cells via the ABC transporter cassette and P-glycoprotein (P-gp), which is a subfamily B member (ABCB1) [78]. P-gp in humans is encoded by the ABCB1 gene [79]. Studies have elucidated that miRNAs play a major role in the metabolism and subsequent uptake of the drug therapy [80]. Treatment of MCF-7 breast carcinoma cell line with antagomiR-451 via transfection and doxorubicin revealed an augmented sensitivity to the drug uptake [81]. The accumulation of the drug was enhanced through miR-451 activity, which influenced the P-gp pathway [82]. Furthermore, research elucidated that miRNAs play a role in the metabolism of chemotherapeutic drugs and their associated membrane transporters and receptors [79]. An interesting property of stem cells which differentiates them from other cells is their ability to express the ABC transporter mechanism through the encoding genes ABCB1, ABCG2, and ABCC1 at high levels [83]. This high level of expression makes them efficient in achieving multidrug resistance. It was elucidated that miR-328 regulates the expression of ABCG2 in MCF-7 cells [80] which were transfected by an antagomiR for miR-328, further enhancing the similarity between normal stem cells and cancer stem cells. It was also reported that, in pancreatic cancer, miRNAs play a crucial role in the induction of MDR. Using miRdeep2, an algorithmic software, four distinct miRNAs (miR-181a-5p, miR-218-5p, miR-130a-3p, and miR-424-3p) were identified to be downregulated with a subsequent decrease in MDR [84].

6. miRNAs and NB

MicroRNAs which are also involved [85] in the transcriptional process can regulate the expression of these genes (see Table 2). The development of NB depends on these expressed genes and the key role that miRNAs play in their expression. Cancer epidemiology has elucidated that frequently miRNAs are located on genomic regions and fragile sites that are implicated in the disease [86]. The best studied oncomiR cases that exhibited a specific miRNA signature for NB are miR-17-92 and miR-21. The polycistronic miR-17-92 consists of seven miRNAs that regulate the PTEN, E2Fs, and TGF- β receptor II sites [35] and has shown that the transcription of the cluster is regulated by MYCN, an oncogene responsible for tumourigenesis. The cluster inhibits *p21* gene responsible [87] for cell cycle progression and apoptosis, ensuing in MYCN amplified NB cells making them more resistant to chemotherapy. Other NB miRNAs are [88] dysregulated as shown in Table 2

The role of miRNAs in cancer treatment is very promising but still highly complex. An all-inclusive approach is a possible route to overcome MDR combined with a personalized therapy. MiRNA profiling can predict the outcome of response to treatment. Through modulation of the expression

TABLE 2: Previous research identifying miRNAs involved in NB pathogenesis and clinical progression.

miRNA	Dysregulated expression	Target gene	References
miR-17-92*	Upregulated	DKK3, CDKN1A, BIM, and MEF2D ESR1	[22, 89–91]
miR-21	Upregulated	PTEN	[92]
miR-128	Upregulated	NTRK	[93]
miR-380-5p	Upregulated	p53	[94]
miR-558	Upregulated	HPSE	[95]
miR-375	Upregulated	ELAVL4	[96]
let-7	Downregulated	MYCN	[19, 97]
miR-9	Downregulated	MMP-14	[98]
miR-15	Downregulated	BcL2	[99, 100]
miR-103	Downregulated	CDK5R1	[101, 102]
miR-107	Downregulated	CDK5R1	[101, 102]
miR-124	Downregulated	SOX9and PTBP1	[96, 103, 104]
miR-29a	Downregulated	Cdk6 and CDC6	[105]
miR152	Downregulated	DNMT1	[106]
miR-125b	Downregulated	TrkC	[107]
miR-204	Downregulated	BCL2 and NTRK2	[108]
miR-363	Downregulated	ADAM15 and MYO1B	[109]
miR-335	Downregulated	SOX4 and TNC	[109]

^{*}Including miR-18a, miR-19a, miR-20a, and miR-92.

levels of these miRNAs, cancer cell lines demonstrated a marked response to therapeutic agents [110]. Through cross-validation analysis, miRNAs circulating in the blood can be screened and used as a diagnostic tool in early detection of cancer [111]. Therefore, their detection is also useful as biomarkers in predicting the response to treatment in a particular tumour, although more research is needed to validate precisely the response. It has been observed that 30% of mammalian physiological processes are regulated by miRNAs and [112] therefore one can implicate that tumour regression can be regulated via these physiological pathways by modulating tumour suppressor miRNAs. Safe protocols for clinical trials in targeting cancer are still the biggest hurdle; the mode of delivery of these modulators is still in its infancy.

7. Conclusion

A pressing challenge in cancer treatment is to overcome MDR. The clinical value of miRNAs in pediatric tumours is very hopeful considering the growing number of scientific literature. Therefore it is imperative that further research is increased further, especially directed towards the use of miRNA-targeted therapy. A recent literature study revealed that out of 11,000 articles on miRNA [112] and cancer only about 500 articles were written directly or indirectly related to cancer therapy via miRNA modulation. It is an attractive field

in the research of oncology especially when one considers the failure in current use of xenobiotics and radiation. Combined with cancer stem cell understanding, miRNA offers a new way of how to control remission and metastasis in tumours with a better prognosis.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- [1] S. Gherardi, E. Valli, D. Erriquez, and G. Perini, "MYCN-mediated transcriptional repression in neuroblastoma: the other side of the coin," *Frontiers in Oncology*, vol. 3, article 42, 2013.
- [2] MD Anderson Cancer Center, Neuroblastoma Cancer Facts & Information, 2015, http://www.mdanderson.org/patient-and-cancer-information/cancer-information/cancer-types/neuroblastoma/index.html.
- [3] Neuroblastoma in children, 2013, http://www.macmillan.org .uk/Cancerinformation/Cancertypes/Childrenscancers/Typesofchildrenscancers/Neuroblastoma.aspx.
- [4] J. Buechner and C. Einvik, "N-myc and noncoding RNAs in neuroblastoma," *Molecular Cancer Research*, vol. 10, no. 10, pp. 1243–1253, 2012.
- [5] R. P. Castleberry, "Neuroblastoma," European Journal of Cancer, vol. 33, no. 9, pp. 1430–1438, 1990.
- [6] L. Moreno, L. V. Marshall, and A. D. J. Pearson, "At the frontier of progress for paediatric oncology: the neuroblastoma paradigm," *British Medical Bulletin*, vol. 108, no. 1, pp. 173–188, 2013.
- [7] F. Bourdeaut, D. Trochet, I. Janoueix-Lerosey et al., "Germline mutations of the paired-like homeobox 2B (PHOX2B) gene in neuroblastoma," *Cancer Letters*, vol. 228, no. 1-2, pp. 51–58, 2005.
- [8] J. M. Maris, "Recent advances in neuroblastoma," The New England Journal of Medicine, vol. 362, no. 23, pp. 2202–2211, 2010
- [9] Y. P. Mossé, M. Laudenslager, L. Longo et al., "Identification of ALK as a major familial neuroblastoma predisposition gene," *Nature*, vol. 455, no. 7215, pp. 930–935, 2008.
- [10] T. R. Webb, J. Slavish, R. E. George et al., "Anaplastic lymphoma kinase: role in cancer pathogenesis and small-molecule inhibitor development for therapy," *Expert Review of Anticancer Therapy*, vol. 9, no. 3, pp. 331–356, 2009.
- [11] Y. Chen, J. Takita, Y. L. Choi et al., "Oncogenic mutations of ALK kinase in neuroblastoma," *Nature*, vol. 455, no. 7215, pp. 971–974, 2008.
- [12] S. C. Bresler, D. A. Weiser, P. J. Huwe et al., "ALK mutations confer differential oncogenic activation and sensitivity to ALK inhibition therapy in neuroblastoma," *Cancer Cell*, vol. 26, no. 5, pp. 682–694, 2014.
- [13] J. M. Maris and K. K. Matthay, "Molecular biology of neuroblastoma," *Journal of Clinical Oncology*, vol. 17, no. 7, pp. 2264–2279, 1999.
- [14] M. A. Hayat, Ed., *Neuroblastoma*, vol. 1 of *Pediatric Cancer*, Springer, Dordrecht, The Netherlands, 2012.
- [15] M. Yoshimoto, S. R. C. de Toledo, E. M. Monteiro Caran et al., "MYCN gene amplification," *The American Journal of Pathology*, vol. 155, no. 5, pp. 1439–1443, 1999.

- [16] K. De Preter, J. Vandesompele, P. Heimann et al., "Human fetal neuroblast and neuroblastoma transcriptome analysis confirms neuroblast origin and highlights neuroblastoma candidate genes," *Genome Biology*, vol. 7, no. 9, article R84, 2006.
- [17] L. J. Valentijn, J. Koster, F. Haneveld et al., "Functional MYCN signature predicts outcome of neuroblastoma irrespective of MYCN amplification," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 47, pp. 19190–19195, 2012.
- [18] L. M. Hansford, W. D. Thomas, J. M. Keating et al., "Mechanisms of embryonal tumor initiation: distinct roles for MycN expression and MYCN amplification," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 34, pp. 12664–12669, 2004.
- [19] J. J. Molenaar, R. Domingo-Fernández, M. E. Ebus et al., "LIN28B induces neuroblastoma and enhances MYCN levels via let-7 suppression," *Nature Genetics*, vol. 44, no. 11, pp. 1199– 1206, 2012.
- [20] J. Yu, M. A. Vodyanik, K. Smuga-Otto et al., "Induced pluripotent stem cell lines derived from human somatic cells," *Science*, vol. 318, no. 5858, pp. 1917–1920, 2007.
- [21] J. M. Shohet, "Redefining functional MYCN gene signatures in neuroblastoma," *Proceedings of the National Academy of Sciences* of the United States of America, vol. 109, no. 47, pp. 19041–19042, 2012.
- [22] P. Mestdagh, E. Fredlund, F. Pattyn et al., "MYCN/c-MYC-induced microRNAs repress coding gene networks associated with poor outcome in MYCN/c-MYC-activated tumors," Oncogene, vol. 29, no. 9, pp. 1394–1404, 2010.
- [23] F. Zhi, R. Wang, Q. Wang et al., "MicroRNAs in neuroblastoma: small-sized players with a large impact," *Neurochemical Research*, vol. 39, no. 4, pp. 613–623, 2014.
- [24] C. Bottino, A. Dondero, F. Bellora et al., "Natural killer cells and neuroblastoma: tumor recognition, escape mechanisms, and possible novel immunotherapeutic approaches," *Frontiers in Immunology*, vol. 5, article 56, 2014.
- [25] National Cancer Institute, Neuroblastoma Treatment, 2015, http://www.cancer.gov/cancertopics/pdq/treatment/neuroblastoma/HealthProfessional/page9.
- [26] S. Mueller and K. K. Matthay, "Neuroblastoma: biology and staging," Current Oncology Reports, vol. 11, no. 6, pp. 431–438, 2009
- [27] G. M. Brodeur, J. Pritchard, F. Berthold et al., "Revisions of the international criteria for neuroblastoma diagnosis, staging and response to treatment," *Progress in Clinical and Biological Research*, vol. 385, pp. 363–369, 1994.
- [28] T. Monclair, G. M. Brodeur, P. F. Ambros et al., "The International Neuroblastoma Risk Group (INRG) staging system: an INRG Task Force report," *Journal of Clinical Oncology*, vol. 27, no. 2, pp. 298–303, 2009.
- [29] R. C. Lee, R. L. Feinbaum, and V. Ambros, "The *C. elegans* heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14," *Cell*, vol. 75, no. 5, pp. 843–854, 1993.
- [30] G. Di Leva, M. Garofalo, and C. M. Croce, "MicroRNAs in cancer," *Annual Review of Pathology*, vol. 9, pp. 287–314, 2014.
- [31] R. W. Carthew and E. J. Sontheimer, "Origins and mechanisms of miRNAs and siRNAs," *Cell*, vol. 136, no. 4, pp. 642–655, 2009.
- [32] F. L. Kisseljov, "MicroRNAs and cancer," *Molecular Biology*, vol. 48, no. 2, pp. 197–206, 2014.
- [33] S. Volinia, G. A. Calin, C.-G. Liu et al., "A microRNA expression signature of human solid tumors defines cancer gene targets,"

- Proceedings of the National Academy of Sciences of the United States of America, vol. 103, no. 7, pp. 2257–2261, 2006.
- [34] X. Wang, L. Cao, Y. Wang, X. Wang, N. Liu, and Y. You, "Regulation of let-7 and its target oncogenes (review)," *Oncology Letters*, vol. 3, no. 5, pp. 955–960, 2012.
- [35] E. Mogilyansky and I. Rigoutsos, "The miR-17/92 cluster: a comprehensive update on its genomics, genetics, functions and increasingly important and numerous roles in health and disease," Cell Death and Differentiation, vol. 20, no. 12, pp. 1603– 1614, 2013.
- [36] X. Chen, B. Hao, G. Han et al., "miR-372 regulates glioma cell proliferation and invasion by directly targeting PHLPP2," *Journal of Cellular Biochemistry*, vol. 116, no. 2, pp. 225–232, 2015.
- [37] Y. C. Chan, J. Banerjee, S. Y. Choi, and C. K. Sen, "miR-210: the master hypoxamir," *Microcirculation*, vol. 19, no. 3, pp. 215–223, 2012.
- [38] P. Tsukerman, R. Yamin, E. Seidel et al., "MiR-520d-5p directly targets TWIST1 and downregulates the metastamiR miR-10b," *Oncotarget*, vol. 5, no. 23, pp. 12141–12150, 2014.
- [39] L. Ma, J. Teruya-Feldstein, and R. A. Weinberg, "Tumour invasion and metastasis initiated by microRNA-10b in breast cancer," *Nature*, vol. 449, no. 7163, pp. 682–688, 2007.
- [40] D. B. Longley and P. G. Johnston, "Molecular mechanisms of drug resistance," *Journal of Pathology*, vol. 205, no. 2, pp. 275– 292, 2005.
- [41] D. Ayers and A. Nasti, "Utilisation of nanoparticle technology in cancer chemoresistance," *Journal of Drug Delivery*, vol. 2012, Article ID 265691, 12 pages, 2012.
- [42] T. R. Wilson, D. B. Longley, and P. G. Johnston, "Chemoresistance in solid tumours," *Annals of Oncology*, vol. 17, supplement 10, pp. x315–x324, 2006.
- [43] J.-P. Gillet and M. M. Gottesman, "Mechanisms of multidrug resistance in cancer," in *Multi-Drug Resistance in Cancer*, vol. 596 of *Methods in Molecular Biology*, pp. 47–76, Humana Press, Clifton, NJ, USA, 2010.
- [44] M. Rebucci and C. Michiels, "Molecular aspects of cancer cell resistance to chemotherapy," *Biochemical Pharmacology*, vol. 85, no. 9, pp. 1219–1226, 2013.
- [45] A. K. Nagaraja, C. J. Creighton, Z. Yu et al., "A link between mir-100 and FRAP1/mTOR in clear cell ovarian cancer," *Molecular Endocrinology*, vol. 24, no. 2, pp. 447–463, 2010.
- [46] M. R. Abedini, E. J. Muller, R. Bergeron, D. A. Gray, and B. K. Tsang, "Akt promotes chemoresistance in human ovarian cancer cells by modulating cisplatin-induced, p53-dependent ubiquitination of FLICE-like inhibitory protein," *Oncogene*, vol. 29, no. 1, pp. 11–25, 2010.
- [47] H. Wu, Y. Cao, D. Weng et al., "Effect of tumor suppressor gene PTEN on the resistance to cisplatin in human ovarian cancer cell lines and related mechanisms," *Cancer Letters*, vol. 271, no. 2, pp. 260–271, 2008.
- [48] G. Gadea, M. de Toledo, C. Anguille, and P. Roux, "Loss of p53 promotes RhoA-ROCK-dependent cell migration and invasion in 3D matrices," *Journal of Cell Biology*, vol. 178, no. 1, pp. 23–30, 2007.
- [49] N. Arsic, G. Gadea, E. Lagerqvist et al., "The p53 isoform $\Delta 133p53\beta$ promotes cancer stem cell potential," *Stem Cell Reports*, vol. 4, no. 4, pp. 531–540, 2015.
- [50] P. A. J. Muller, P. T. Caswell, B. Doyle et al., "Mutant p53 drives invasion by promoting integrin recycling," *Cell*, vol. 139, no. 7, pp. 1327–1341, 2009.

- [51] D. Lai, S. Visser-Grieve, and X. Yang, "Tumour suppressor genes in chemotherapeutic drug response," *Bioscience Reports*, vol. 32, no. 4, pp. 361–374, 2012.
- [52] W. B. Nagengast, T. H. O. Munnink, E. C. F. Dijkers et al., "Multidrug resistance in oncology and beyond: from imaging of drug efflux pumps to cellular drug targets," *Methods in Molecular Biology*, vol. 596, pp. 15–31, 2010.
- [53] D. Hanahan and R. A. Weinberg, "Hallmarks of cancer: the next generation," *Cell*, vol. 144, no. 5, pp. 646–674, 2011.
- [54] S. Ghavami, M. Hashemi, S. R. Ande et al., "Apoptosis and cancer: mutations within caspase genes," *Journal of Medical Genetics*, vol. 46, no. 8, pp. 497–510, 2009.
- [55] F. H. Igney and P. H. Krammer, "Death and anti-death: tumour resistance to apoptosis," *Nature Reviews Cancer*, vol. 2, no. 4, pp. 277–288, 2002.
- [56] P. Vaupel and A. Mayer, "Hypoxia and anemia: effects on tumor biology and treatment resistance," *Transfusion Clinique et Biologique*, vol. 12, no. 1, pp. 5–10, 2005.
- [57] P. Vaupel and A. Mayer, "Hypoxia in cancer: significance and impact on clinical outcome," *Cancer and Metastasis Reviews*, vol. 26, no. 2, pp. 225–239, 2007.
- [58] J.-P. Cosse and C. Michiels, "Tumour hypoxia affects the responsiveness of cancer cells to chemotherapy and promotes cancer progression," *Anti-Cancer Agents in Medicinal Chemistry*, vol. 8, no. 7, pp. 790–797, 2008.
- [59] R. Straussman, T. Morikawa, K. Shee et al., "Tumour microenvironment elicits innate resistance to RAF inhibitors through HGF secretion," *Nature*, vol. 487, no. 7408, pp. 500–504, 2012.
- [60] Y. Mao, E. T. Keller, D. H. Garfield, K. Shen, and J. Wang, "Stromal cells in tumor microenvironment and breast cancer," *Cancer and Metastasis Reviews*, vol. 32, no. 1-2, pp. 303–315, 2013.
- [61] R. Lanza and A. Atala, Eds., Essentials of Stem Cell Biology, Academic Press, Amsterdam, The Netherlands, 3rd edition, 2013
- [62] K. Takahashi, K. Tanabe, M. Ohnuki et al., "Induction of pluripotent stem cells from adult human fibroblasts by defined factors," *Cell*, vol. 131, no. 5, pp. 861–872, 2007.
- [63] A. Bongso, Stem Cells: From Bench to Bedside, E. H. Lee, Ed., World Scientific Publishing, Hackensack, NJ, USA, 2nd edition, 2010.
- [64] T. Lapidot, C. Sirard, J. Vormoor et al., "A cell initiating human acute myeloid leukaemia after transplantation into SCID mice," *Nature*, vol. 367, no. 6464, pp. 645–648, 1994.
- [65] P. Borst, "Cancer drug pan-resistance: pumps, cancer stem cells, quiescence, epithelial to mesenchymal transition, blocked cell death pathways, persisters or what?" *Open Biology*, vol. 2, no. 5, Article ID 120066, 2012.
- [66] S. J. Morrison and J. Kimble, "Asymmetric and symmetric stemcell divisions in development and cancer," *Nature*, vol. 441, no. 7097, pp. 1068–1074, 2006.
- [67] L. Shahriyari and N. L. Komarova, "Symmetric vs. asymmetric stem cell divisions: an adaptation against cancer?" *PLoS ONE*, vol. 8, no. 10, Article ID e76195, 2013.
- [68] Z. Yu, T. G. Pestell, M. P. Lisanti, and R. G. Pestell, "Cancer stem cells," *International Journal of Biochemistry and Cell Biology*, vol. 44, no. 12, pp. 2144–2151, 2012.
- [69] I. Ischenko, H. Seeliger, M. Schaffer, K.-W. Jauch, and C. J. Bruns, "Cancer stem cells: how can we target them?" *Current Medicinal Chemistry*, vol. 15, no. 30, pp. 3171–3184, 2008.
- [70] S. Chiba, "Notch signaling in stem cell systems," *Stem Cells*, vol. 24, no. 11, pp. 2437–2447, 2006.

- [71] H. J. Kraft, S. Mosselman, H. A. Smits et al., "Oct-4 regulates alternative platelet-derived growth factor α receptor gene promoter in human embryonal carcinoma cells," *Journal of Biological Chemistry*, vol. 271, no. 22, pp. 12873–12878, 1996.
- [72] L. Gao, Y. Yang, H. Xu et al., "MiR-335 functions as a tumor suppressor in pancreatic cancer by targeting OCT4," *Tumour Biology*, vol. 35, no. 8, pp. 8309–8318, 2014.
- [73] P. A. Beachy, S. S. Karhadkar, and D. M. Berman, "Tissue repair and stem cell renewal in carcinogenesis," *Nature*, vol. 432, no. 7015, pp. 324–331, 2004.
- [74] L. Li and W. B. Neaves, "Normal stem cells and cancer stem cells: the niche matters," *Cancer Research*, vol. 66, no. 9, pp. 4553–4557, 2006.
- [75] P. Neveu, M. J. Kye, S. Qi et al., "MicroRNA profiling reveals two distinct p53-related human pluripotent stem cell states," *Cell Stem Cell*, vol. 7, no. 6, pp. 671–681, 2010.
- [76] M. M. Sherry, A. Reeves, J. K. Wu, and B. H. Cochran, "STAT3 is required for proliferation and maintenance of multipotency in glioblastoma stem cells," *Stem Cells*, vol. 27, no. 10, pp. 2383– 2392, 2009.
- [77] D. M. Hsu, S. Agarwal, A. Benham et al., "G-CSF receptor positive neuroblastoma subpopulations are enriched in chemotherapy-resistant or relapsed tumors and are highly tumorigenic," *Cancer Research*, vol. 73, no. 13, pp. 4134–4146, 2013.
- [78] M. M. Gottesman, T. Fojo, and S. E. Bates, "Multidrug resistance in cancer: role of ATP-dependent transporters," *Nature Reviews Cancer*, vol. 2, no. 1, pp. 48–58, 2002.
- [79] T. Zheng, J. Wang, X. Chen, and L. Liu, "Role of microRNA in anticancer drug resistance," *International Journal of Cancer*, vol. 126, no. 1, pp. 2–10, 2010.
- [80] Y.-Z. Pan, M. E. Morris, and A.-M. Yu, "MicroRNA-328 negatively regulates the expression of breast cancer resistance protein (BCRP/ABCG2) in human cancer cells," *Molecular Pharmacology*, vol. 75, no. 6, pp. 1374–1379, 2009.
- [81] H. Zhu, H. Wu, X. Liu et al., "Role of MicroRNA miR-27a and miR-451 in the regulation of MDR1/P-glycoprotein expression in human cancer cells," *Biochemical Pharmacology*, vol. 76, no. 5, pp. 582–588, 2008.
- [82] O. Kovalchuk, J. Filkowski, J. Meservy et al., "Involvement of microRNA-451 in resistance of the MCF-7 breast cancer cells to chemotherapeutic drug doxorubicin," *Molecular Cancer Therapeutics*, vol. 7, no. 7, pp. 2152–2159, 2008.
- [83] M. Dean, T. Fojo, and S. Bates, "Tumour stem cells and drug resistance," *Nature Reviews Cancer*, vol. 5, no. 4, pp. 275–284, 2005.
- [84] A. Gisel, M. Valvano, I. G. El Idrissi et al., "MiRNAs for the detection of multidrug resistance: overview and perspectives," *Molecules*, vol. 19, no. 5, pp. 5611–5623, 2014.
- [85] Y. Shi, J. Wang, Z. Xin et al., "Transcription factors and microRNA-co-regulated genes in gastric cancer invasion in ex vivo," PLoS ONE, vol. 10, no. 4, Article ID e0122882, 2015.
- [86] G. A. Calin, C. Sevignani, C. D. Dumitru et al., "Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 9, pp. 2999–3004, 2004.
- [87] L. Fontana, M. E. Fiori, S. Albini et al., "Antagomir-17-5p abolishes the growth of therapy-resistant neuroblastoma through p21 and BIM," *PLoS ONE*, vol. 3, no. 5, Article ID e2236, 2008.

- [88] H. Mei, Z.-Y. Lin, and Q.-S. Tong, "The roles of microRNAs in neuroblastoma," *World Journal of Pediatrics*, vol. 10, no. 1, pp. 10–16, 2014.
- [89] J. Lovén, N. Zinin, T. Wahlström et al., "MYCN-regulated microRNAs repress estrogen receptor-alpha (ESR1) expression and neuronal differentiation in human neuroblastoma," Proceedings of the National Academy of Sciences of the United States of America, vol. 107, no. 4, pp. 1553–1558, 2010.
- [90] R. L. Stallings, "MicroRNA involvement in the pathogenesis of neuroblastoma: potential for microRNA mediated therapeutics," *Current Pharmaceutical Design*, vol. 15, no. 4, pp. 456–462, 2009.
- [91] P. Mestdagh, A.-K. Boström, F. Impens et al., "The miR-17-92 microRNA cluster regulates multiple components of the TGF- β pathway in neuroblastoma," *Molecular Cell*, vol. 40, no. 5, pp. 762–773, 2010.
- [92] Y. Chen, Y.-H. Tsai, Y. Fang, and S.-H. Tseng, "Micro-RNA-21 regulates the sensitivity to cisplatin in human neuroblastoma cells," *Journal of Pediatric Surgery*, vol. 47, no. 10, pp. 1797–1805, 2012.
- [93] M. Guidi, M. Muiños-Gimeno, B. Kagerbauer, E. Martí, X. Estivill, and Y. Espinosa-Parrilla, "Overexpression of miR-128 specifically inhibits the truncated isoform of NTRK3 and upregulates BCL2 in SH-SY5Y neuroblastoma cells," *BMC Molecular Biology*, vol. 11, article 95, 2010.
- [94] A. Swarbrick, S. L. Woods, A. Shaw et al., "MiR-380-5p represses p53 to control cellular survival and is associated with poor outcome in MYCN-amplified neuroblastoma," *Nature Medicine*, vol. 16, no. 10, pp. 1134–1140, 2010.
- [95] H. Qu, L. Zheng, J. Pu et al., "miRNA-558 promotes tumorigenesis and aggressiveness of neuroblastoma cells through activating the transcription of heparanase," *Human Molecular Genetics*, vol. 24, no. 9, pp. 2539–2551, 2015.
- [96] L. Samaraweera, K. B. Grandinetti, R. Huang, B. A. Spengler, and R. A. Ross, "MicroRNAs define distinct human neuroblastoma cell phenotypes and regulate their differentiation and tumorigenicity," *BMC Cancer*, vol. 14, article 309, 2014.
- [97] A. M. Lozier, M. E. Rich, A. P. Grawe et al., "Targeting ornithine decarboxylase reverses the LIN28/Let-7 axis and inhibits glycolytic metabolism in neuroblastoma," *Oncotarget*, vol. 6, no. 1, pp. 196–206, 2015.
- [98] H. Zhang, M. Qi, S. Li et al., "MicroRNA-9 targets matrix metalloproteinase 14 to inhibit invasion, metastasis, and angiogenesis of neuroblastoma cells," *Molecular Cancer Therapeutics*, vol. 11, no. 7, pp. 1454–1466, 2012.
- [99] W. C. S. Cho, "OncomiRs: the discovery and progress of microRNAs in cancers," *Molecular Cancer*, vol. 6, article 60, 2007
- [100] A. Bottoni, D. Piccin, F. Tagliati, A. Luchin, M. C. Zatelli, and E. C. D. Uberti, "miR-15a and miR-16-1 down-regulation in pituitary adenomas," *Journal of Cellular Physiology*, vol. 204, no. 1, pp. 280–285, 2005.
- [101] S. Moncini, A. Salvi, P. Zuccotti et al., "The role of miR-103 and miR-107 in regulation of CDK5R1 expression and in cellular migration," *PLoS ONE*, vol. 6, no. 5, Article ID e20038, 2011.
- [102] P. Zuccotti, C. Colombrita, S. Moncini et al., "HnRNPA2/B1 and nELAV proteins bind to a specific U-rich element in CDK5R1 3'-UTR and oppositely regulate its expression," *Biochimica et Biophysica Acta—Gene Regulatory Mechanisms*, vol. 1839, no. 6, pp. 506–516, 2014.
- [103] L.-C. Cheng, E. Pastrana, M. Tavazoie, and F. Doetsch, "MiR-124 regulates adult neurogenesis in the subventricular zone stem

- cell niche," Nature Neuroscience, vol. 12, no. 4, pp. 399-408, 2009
- [104] X. Cao, S. L. Pfaff, and F. H. Gage, "A functional study of miR-124 in the developing neural tube," *Genes & Development*, vol. 21, no. 5, pp. 531–536, 2007.
- [105] I. Y. Cheung, T. A. Farazi, I. Ostrovnaya et al., "Deep MicroRNA sequencing reveals downregulation of miR-29a in neuroblastoma central nervous system metastasis," *Genes Chromosomes* and Cancer, vol. 53, no. 10, pp. 803–814, 2014.
- [106] S. Das, N. Foley, K. Bryan et al., "MicroRNA mediates DNA demethylation events triggered by retinoic acid during neuroblastoma cell differentiation," *Cancer Research*, vol. 70, no. 20, pp. 7874–7881, 2010.
- [107] P. Laneve, L. Di Marcotullio, U. Gioia et al., "The interplay between microRNAs and the neurotrophin receptor tropomyosin-related kinase C controls proliferation of human neuroblastoma cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 19, pp. 7957–7962, 2007.
- [108] J. Ryan, A. Tivnan, J. Fay et al., "MicroRNA-204 increases sensitivity of neuroblastoma cells to cisplatin and is associated with a favourable clinical outcome," *British Journal of Cancer*, vol. 107, no. 6, pp. 967–976, 2012.
- [109] J. Qiao, S. Lee, P. Paul et al., "MiR-335 and miR-363 regulation of neuroblastoma tumorigenesis and metastasis," *Surgery*, vol. 154, no. 2, pp. 226–233, 2013.
- [110] P. E. Blower, J.-H. Chung, J. S. Verducci et al., "MicroRNAs modulate the chemosensitivity of tumor cells," *Molecular Can*cer Therapeutics, vol. 7, no. 1, pp. 1–9, 2008.
- [111] J. Wang, J. Chen, P. Chang et al., "MicroRNAs in plasma of pancreatic ductal adenocarcinoma patients as novel bloodbased biomarkers of disease," *Cancer Prevention Research*, vol. 2, no. 9, pp. 807–813, 2009.
- [112] F. H. Sarkar, Ed., MicroRNA Targeted Cancer Therapy, Springer, New York, NY, USA, 2014.