

REVIEW ARTICLE

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Advances in toxicology and medical treatment of chemical warfare nerve agents

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Abstract

Organophosphorous (OP) Nerve agents (NAs) are known as the deadliest chemical warfare agents. They are divided into two classes of G and V agents. Most of them are liquid at room temperature. NAs chemical structures and mechanisms of actions are similar to OP pesticides, but their toxicities are higher than these compounds. The main mechanism of action is irreversible inhibition of Acetyl Choline Esterase (AChE) resulting in accumulation of toxic levels of acetylcholine (ACh) at the synaptic junctions and thus induces muscarinic and nicotinic receptors stimulation. However, other mechanisms have recently been described. Central nervous system (CNS) depression particularly on respiratory and vasomotor centers may induce respiratory failure and cardiac arrest. Intermediate syndrome after NAs exposure is less common than OP pesticides poisoning. There are four approaches to detect exposure to NAs in biological samples: (I) AChE activity measurement, (II) Determination of hydrolysis products in plasma and urine, (III) Fluoride reactivation of phosphorylated binding sites and (IV) Mass spectrometric determination of cholinesterase adducts. The clinical manifestations are similar to OP pesticides poisoning, but with more severity and fatalities. The management should be started as soon as possible. The victims should immediately be removed from the field and treatment is commenced with auto-injector antidotes (atropine and oximes) such as MARK I kit. A 0.5% hypochlorite solution as well as novel products like M291 Resin kit, G117H and Phosphotriesterase isolated from soil bacteria, are now available for decontamination of NAs. Atropine and oximes are the well known antidotes that should be infused as clinically indicated. However, some new adjuvant and additional treatment such as magnesium sulfate, sodium bicarbonate, gacyclidine, benactyzine, tezampanel, hemoperfusion, antioxidants and bioscavengers have recently been used for OP NAs poisoning.

Keywords: Nerve agents, Chemical warfare agent, Organophosphorous compounds, Pesticides, Sodium bicarbonate, Magnesium sulfate, Iran

Introduction

Chemical warfare nerve agents (NAs) are one of the important groups of organophosphorous (OP) compounds that have been used as tactical weapons and for terrorism during recent decades. OP compounds have also been used as petroleum additives and pesticides [1]. Although NAs are strongly similar in chemical structure and biological function to many OP pesticides, fatality potency of NAs is generally higher than the OP pesticides [2].

The NAs are traditionally classified into two classes of G and V agents, but also GV compounds (GV:2-dimethylaminoethyl-(dimethylamido)-fluorophosphate) which contained structures of both G and V agent are now exist. The G agents include Tabun (GA; ethyl N, N-dimethylphosphoramidocyanidate), Sarin (GB; 2-fluoro-methylphosphoryloxypropane), Soman (GD; 3-fluoromethyl-phosphoryloxy-2, 2-dimethyl-butane) and Cyclosarin (GF; fluoro-methylphosphoryloxycyclohexane). The important warfare V agents include VE (S-2-diethylaminoethyl O-ethylethylphosphonothioate), VM (2-ethoxy-methylphosphoryl sulfanyl-N,N-diethylethanamine), VG (2 diethoxyphosphorylsulfanyl-N,N-diethylethanamine), VR (Russian VX; N,N-diethyl-2-methyl-2-methylpropoxy phosphorylsulfanylethanamine) and VX (S-2 diisopropylamino O-ethylmethylphosphonothioate) [3-5]. There are no common names for other G

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and V agents. VX is the main and oldest agent of V series which has been produced in large quantities [1,5,6]. Recently a new type of NAs has been claimed to develop named "Novichoks" (means "newcomer" in Russia). This has been attracting increasing attention in recent years, particularly among non-governmental organizations (NGOs). It has been claimed that the toxicity of certain "Novichok" agents may exceed that of VX. The action mechanism is also dissimilar to the other NAs and thus conventional antidotes may be ineffective. Though, to date, there is nothing in details on such chemical has ever been declared in the literature [7,8]. NAs are delivered by missiles, bombs, spray and cluster spray [9].

NAs have fatal effects in acute phase of poisoning and also have considerable long term complications due to irreversible inhibition of Acetyl Choline Esterase (AChE). They have been known as the most lethal agent among chemical warfare agents (CWA) [1,10-13].

It is important to consider the management of civilian casualties due to the possibility of NAs use in terrorist attacks. Despite early treatment and the use of urgent countermeasures (atropine and oxime) in exposure zones, it may take a long time to recover from or even alleviate the complications of NAs exposure. Thus, it was aimed to comprehensively explain clinical manifestations and recent advances in treatment of chemical warfare NAs poisoning in this review article.

History

NAs were first synthesized in 1854 but were not used as a CWA in a large scale until eight decades later [13]. The G agents were first produced in Germany at IG Farben industries by Dr. Gerhard Schrader team in 1930s. They synthesized tabun in 1938 and then sarin. These compounds were named after him and his two co-workers. The letter G for G agents means German [4,14]. The V agents were synthesized after the World War II in the United Kingdom in 1952. The V agents were derived from the word victory; the share of allied forces from World War II [4,6]. NAs had not ever been used on the battlefield until Iran-Iraq war. During the Iran-Iraq conflict in 1983–1988, NAs were infamously used by Iraqi military against Iranian troops and even civilians. Among CWA, Sulfur Mustard and NAs (sarin and tabun, specifically) had been mostly used by Iraq in several chemical massacres [1].

Tabun was the first NAs used in the war at Majnoon Island in February 1984. Several thousands were poisoned by tabun and more than 300 victims died within 30 min. Mortality rate was much more in first years of the war because of the unavailabilities of protective equipment first-aid medications such as atropine and oximes auto-injectors [4,9,15]. Later in 1987 and 1988

another NA named sarin was used against Iranian troops and innocent people in Halabjah massacre [15,16]. It was estimated that over 100,000 individuals were poisoned by chemical attacks during the Iran-Iraq war. Meanwhile, NAs are associated with higher mortality than the other CWAs and had a drastic role in Iraqi missile attacks during the Iran-Iraq war [4].

Other tragedies of NA attacks were Sarin terrorist attacks in 1994 in Matsumoto, Japan and six months later in Tokyo Subways which poisoned 6,100 people including rescue staff with 18 mortalities. These terrorist incidences significantly raised interests in other countries and led to a number of symposia as the seminar on responding to the consequences of chemical and biological terrorism held at Bethesda, Maryland, in July 1995 [17]. United States have had two conflicts with Iraq during 1991 and 2003 which in both wars none of the countries used CWA. Iraq admitted possession of NAs to the USA in 1995 as well as other biologic and chemical weapons. In 1995, the USA also signed the Chemical Weapon Convention. According to this, all the nations that stored CWAs including NAs had to destroy their stockpiles by 2012 [5,18]. The Organization for Prohibition of Chemical Weapons (OPCW) is now responsible to control the CWA threat worldwide. Fortunately, in recent conflict and terrorist events, such as the 11Sept attacks in New York and Washington, Bali island of Indonesia, London and Madrid tube bombings, CWA was not used at all. Nevertheless, the use of CWA, particularly NAs is still a threat.

Chemical structures and properties

NAs are alkylphosphonic acid esters. Tabun has a cyanide group. Sarin and soman are methylphosphonofluoridate. They contain a fluorine substituent group. These NAs have a unique C–P bond that it is not found in OP pesticides and is very hydrolysis resistant. VX contains sulfur and is an alkylphosphonothiolate [19]. The toxicity of these agents are largely more influenced by the chirality around the phosphorus atom than the P(+) isomers. The structural formulae of some main NAs are shown in Figure 1 and some main properties of NAs are presented in Table 1.

The term "nerve gas" is a historical misunderstanding, because all the classic forms of NAs are liquid at room temperature. The first chemical warfare agents (CWA) such as chlorine and phosgene are the reason as they are true gases at standard pressure and temperature [9,11]. All the NAs are liquid in room condition, tasteless and odorless and potentially volatile. However, there are some differences in chemical and density properties (Table 1). G agents' densities are the same as water and they also evaporate at about the same rate. The freezing points are around 0°C (the same as water) and the

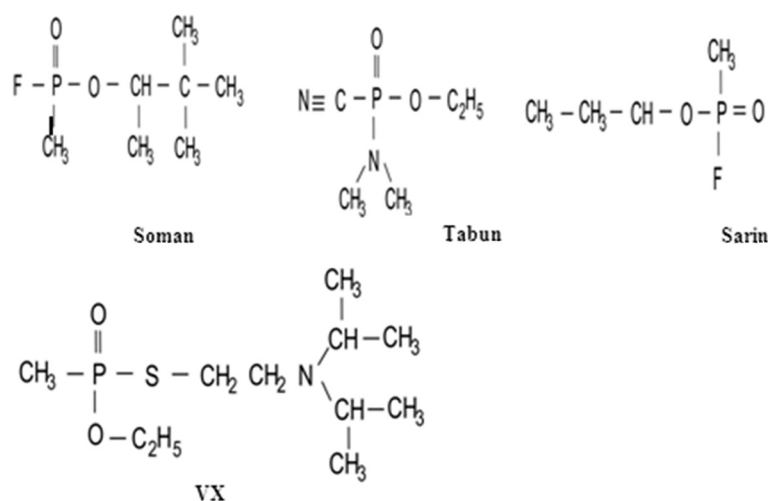


Figure 1 Structural formulae of main nerve agents [1].

boiling point is around 150°C. G agents rapidly spread on skin. They spread rapidly and remain in the environment for several hours and thus are known as “non-persistent agents”. G agents are released from clothing for about 30 minutes after vapor contact [12]. In contrast, V agents specially the VX are more oily, the same as motor vehicle oil and thus evaporate more slowly which is known as “persistent agents” [10-12]. Sarin is

the most volatile agent with a vapor pressure of 4,000 times more than VX as of the least volatile agent [20]. Although VX has less vapor hazard than G agents (due to the least volatility), when it comes to “persistence”, it can contaminate an area for longer time. Due to the oily condition, VX is the most efficiently absorbed NA through the skin [13]. Thickening agents, like acrylates, can be added to some NAs. They alter part of the

Table 1 Physical and chemical properties of main organophosphorous nerve agents

Properties	Tabun (GA)	Sarin (GB)	Soman (GD)	VX
Chemical name	ethyl N, N-dimethylphosphoramidocyanidate	2-fluoro-methylphosphoryloxypropane	3-fluoromethyl-phosphoryloxy-2, 2-dimethyl-butane	S-2 diisopropylamino O-ethylmethylphosphonothioate
CAS No.	77-81-6	107-44-8	96-64-0	50782-69-9
Molecular weight	162.1	140.1	182.2	267.4
State	liquid	liquid	liquid	oily
Odor	Slight fruity odor	None	Slight fruity odor	None
Appearance	Clear colorless; tasteless;	Clear colorless; tasteless;	Clear colorless, ages to brown	Amber color, tasteless
Density (liquid, g/ml)	1.08	1.09	1.02	1.0083
Density (vapor, compared to air)	5.6	4.8	6.3	9.2
Volatility (mg/m ³)	610	22,000	3,900	10.5
Solubility (in water, g/100g)	9.8	Miscible	2.1	Miscible < 24 °C
Solubility (in other solvents)	Soluble in most organic solvents	Soluble in all solvents	Soluble in some solvents	Soluble in all solvents
Boiling point (°C)	220-246	158	167-200	298
Flash point (°C)	77.8	NR	121.1	158.9
Melting point (°C)	-50	-56	NR	-39 (calculated)

CAS, Chemical Abstract Service.

Unless noted, properties are determined at 25°C and pressure of 760 mmHg.

Adapted from references 6, 11, 43 and 88.

physical properties of the new combined component, raising persistency of NAs in the environment [21].

Sarin is water soluble in any ratio and water hydrolyzes it to remove fluorine and produce a nontoxic product compared with the parent compound. Soman and tabun are solved easily in organic solvents, however, moderately mixed with water. VX is slightly water soluble in room temperature. Cold water and organic solvents are strong solvents for VX. G agents are quickly hydrolyzed at alkaline pH solutions. The half-life of sarin in water (pH=7.0) is 5.4 hours while it is 15 min. at a pH of 9.0. Decontaminating with alkaline solution like household bleach solutions (0.5% sodium hypochlorite) is done based on this property [22].

The vapor density of all the NAs is more than one. It means the vapor of NAs are heavier than air and they tend to stay close to the land, thus it would be a risk for people in lower areas and underground shelters [13,21].

Mechanism of action

The main mechanism of action is irreversible inactivation of AChE at the cholinergic synapses leading to accumulation of toxic levels of acetylcholine (ACh) at the synaptic junctions. It over stimulates the cholinergic pathway and consequently desensitizes the cholinergic receptor site. ACh is a neurotransmitter which contributes to nerve conduction in central nervous system (CNS), at autonomic ganglia including parasympathetic postganglionic synapses and sympathetic preganglionic synapses. They also act at the parasympathetic nerve endings like those at neuromuscular junction of skeletal muscles and in the sweat glands [1,4,13].

There are three types of cholinesterase in human body. The main and principal form is AChE which is referred to as "true cholinesterase" and found in neurons, neuromuscular junctions and erythrocyte membrane. AChE is a serine protease that hydrolyzes the neurotransmitter ACh. It is also reported that AChE has some non-hydrolyzing functions. Park S (2004) has stated that AChE has a critical role in the development of apoptosome, a large quaternary protein structure formed in the process of apoptosis, in the body through blocking the interaction between apoptotic protease-activating factor-1 (APAF1) and cytochrome C [23]. Butyrylcholinesterase (BChE) or pseudocholinesterase may have a role in cholinergic neurotransmission, and is occupied in other nervous system functions. It is also important as a biomarker of exposure to OP [24]. BChE inhibition by NAs seems to have no important physiological effect in the absence of other toxicants [25].

Serum cholinesterase (SChE) is the third form. It is a circulating plasma glycoprotein synthesized in the liver including group of enzymes present in cerebrospinal

fluid, liver, glial cells and plasma. SChE does not seem to have any physiological function [4,26].

NAs play their role by binding to serine residue at the active site of AChE molecule and form a phosphate or phosphonate ester [9,11]. Thus, the resulted phosphorylated molecule is incapable of hydrolyzing ACh, and regenerate very slowly. The inhibition will be permanent until the generation of a new enzyme or a reactivator usage such as an oxime [27]. Binding reactions of NAs to esterases such as ChE, AChE, carboxylesterases (CarbE) and other proteins will also occur. Both OP pesticides and NAs lose their acyl radicals in addition to their reaction with the esterases. After binding to AChE and BChE, there is a non-enzymatic time-dependent intra molecular rearrangement which leads to loss of one alkyl group bound to the phosphorus, known as "aging reaction" (The time between NA exposure and irreversible phosphorylation). This leads to a persistent non-reactivable AChE, resistant to the both spontaneous and oxime-induced reactivation [28-30]. The half time of aging varies from a few minutes for soman, five hr. for sarin, 22 hr. for cyclosarin and more than 40 hr. for tabun and VX [9,28,31,32]. Due to reversible binding of soman and sarin to CarbE, there is a hypothesis which supposes a role for CarbE in metabolic detoxification of these agents to their non-toxic metabolites isopropyl methylphosphonic acid (IMPA) and pinacolyl methylphosphonic acid (PMPA) [31-34].

Cholinergic inhibition is not the only mechanism of action of NAs. There are some data showing other probable underlying mechanisms during NAs intoxication. Fonnum and Sterri (1981) reported that toxic effects of soman is due to only 5% of LD50 in rats, about 5µg/kg, which reacts with AChE and the rest lead to various metabolic reaction [35]. It has also been stated that NAs can inhibit enzymes outside of the cholinergic system, mainly serine esterase. It has been reported formerly that NAs alter the persistence and metabolism of some neuropeptides degraded by serine esterase, such as enkephalins, endorphins, and substance P. This may describe some atropine resistant symptoms of NAs [36]. Clement and Copeman (1984) reported longstanding analgesia in mice after exposure to sarin and soman, and nalaxone, an opiate antagonist, alleviates this phenomena. Nevertheless, no exact information is available in opioid receptors following NAs exposure [36]. Duysen and colleagues (2001) studied other probable mechanism of VX on knockout mice. They treated with 0, 50, and 100% AChE activity mice with subcutaneous VX. AChE^{-/-} presented the same cholinergic signs of toxicity as the wild type mice, even though AChE^{-/-} mice have not any AChE whose inhibition could lead to cholinergic signs. It was thus concluded that toxic effects in NAs exposure is due to inhibition or binding to several

proteins, only one of which is AChE [37]. Other involved mechanisms included changes in other enzymes, neurotransmitter, anaphylactoid reactions, immune changes, oxidative stress etc [32,38-41]. Long lasting effects have more reasons beside ChE inhibition. It is formerly reported that NAs also act as secretagogues and can augment bronchial spasm by anaphylactoid reactions. Apart from the cholinergic crisis in NAs poisoning, secondary adverse reactions due to other underlying mechanisms may complicate NAs toxicity. Excitatory amino acids are also involved in both OP pesticides and NAs poisoning. Adenosine receptor agonists have been showed to have good protective activity on this basis [42].

Relative toxicity

Regardless of route of entry, VX is the most potent among NAs. The stability, resistance to detoxification and environmental persistency of VX are higher than the other NAs. It is also less volatile and more impressive at skin penetration. Hence, VX is labeled as a skin penetrant and lethal contact agent rather than inhalation threat [43]. VX at 10 μ M largely reduced cell metabolism within two hours [44]. The G agents are toxic or even fatal by any route of exposure at sufficient concentration.

Based on animal studies, the G agents have lethal inhaled dose of about 1 mg in human. They also represent a skin contact hazard through contaminated cloths, especially when evaporation is minimized. However, the G agents absorption rate is much less rapid in percutaneous than in the inhalation form [1]. VX is easily absorbed through the skin and generally does not have a major inhalation hazard in the zone [1,43]. Overall relative lethality of NAs in animal studies is: VX>Soman>Sarin>Tabun [45].

Metabolism

NAs' metabolism is mostly explained under mechanism of action. The common NAs have an asymmetric center (chiral compounds), which they have two (e.g., sarin) or four enantiomers (e.g., soman) with different toxicity effects on human. Unfortunately, the more toxic enantiomers have longer half life than others in the body. Enzymatic and chemical catalysis of NAs results in the formation of inactive phosphonic acids, which are excreted via renal [46,47]. In in-vitro studies, the elimination half-life of G agents was rather shorter than V agents (less than one hour), whereas VX persists for several hours in intravenous administration and even longer in percutaneous exposure [48].

Oxidation and hydrolysis are principal metabolic reactions which occur mainly by reaction with glutathione and also may happen by glucuronidation and

demethylation. Oxidation gives rise to production more or less toxic products. Tabun causes the largest number of degradation products among G agents. Detoxification of tabun takes place slowly, by the enzyme di-isopropyl-fluorophosphatase; formerly termed tabunase [49]. There are sparse toxicity data available for subset of tabun degradation products. Ethyl-dimethylaminophosphoric acid (EDMPA) is the main product of tabun dimethylamin, which is also produced by hydrolysis of tabun among other reactions. Dimethylamin cause human irritation in the respiratory tracts [50].

Isopropyl-methylphosphonic acid (IMPA) is metabolite of Sarin which subsequently hydrolyses to the high stable methylphosphonic acid (MPA) and resistance to further hydrolysis. MPA mildly irritates rabbit's skin and human skin and eyes. It also produces low oral toxicity in mice and rats [24,51].

In rats, 10 minutes after intravenous sarin, about seventy percent of the plasma level was bound to large protein molecules similar to carboxylesterase [52]. The toxicity of sarin enhanced six to eight time when rats were pretreated with triorthocresyl phosphate (TOCP), a weak anti-ChE OP with irreversible blocking carboxylesterase property [53].

In a study of Little *et al.* (1986) 80 μ g/kg of sarin was injected intravenously to mice. Tissue distribution was recorded for 24 h. Within 1 minute sarin concentration was at the highest in the kidney, liver and plasma. Over the first minute, about half of the labeled sarin was associated with the major sarin metabolite; IMPA and the kidneys contained the highest concentration of sarin and its metabolites. Much lower concentration detected in liver after 24 hr, suggested the main role of the kidneys in detoxification of sarin [54]. In another study of Little *et al.* (1988) with the same method, hypothalamus contained concentration of both sarin and metabolites 2-5 times greater than those in other brain areas. This finding suggests that hypothalamus is important with respect to central effects of NAs [55]. Brain distribution of sarin was detected in 4 of 12 victims who died after the Matsumoto event. In patients of the Matsumoto with sarin exposure the levels of IMPA and MPA correlated with clinical manifestations [56].

Pinacolyl methylphosphonic acid (PMPA) is the predominant hydrolytic product of the soman [57].

The anticholinesterase mechanism of action of V agents is due to the "oxo" group (O) as well as presence of alkyl substituents. VX, as a V agent, is different from G agents in both pharmacodynamics and pharmacokinetics characteristics. It distributes in blood as protonated amine. Its hydrolysis is slower than G compounds and reacts more slowly with A-esterases and CarBE. Oxidation reactions at nitrogen and/or sulfur are another routes for VX metabolism beside hydrolysis [4,24].

Tsuchihashi *et al.* (1998) detected both EMPA and 2-(diisopropylamino-ethyl) methyl sulfide in VX exposed serum samples [58]. These results clarified the first documented detection of the specific VX metabolites in victims' serum and also explained a part of metabolic pathway of VX in human body which has been later used in measuring the VX-inhibited AChE hydrolytic product EMPA [59,60].

Detection and determination methods

Most research on diagnostic methods of NAs exposure has been directed at the most available samples of survivors such as blood (serum, plasma, whole blood, or red cells) and urine. Intact G agents are available in the organism for a few hours; therefore, blood sampling should be obtained in a few hours after OP exposure. Thus intact agents don't seem to be a good target of retrospective detection of exposure [60-62]. There are about four approaches to detect exposure to NAs:

AChE inhibition measurement

Although this method is the most common way to identify NAs exposure, there are some impediments in this procedure. Firstly, it does not identify the exact exposed agent and also its specification is low, because there are some other chemicals contribute to inhibition of AChE. Secondly, inhibition levels less than 20% are not detectable and it cannot be used as a retrospective measurement due to new synthesis of the enzyme. However, it is the most widely used method for evaluation of OP NAs exposure [30,60]. Wang and co-workers (2008) have assessed salivary ChE enzyme activity by using carbon nanotube-based electrochemical sensor. An electrochemical sensor based on a carbon nanotube (CNT)-modified screen-printed carbon electrode and coupled with a microflow injection system was applied for a sensitive, rapid, and simple assessment of salivary ChE enzyme activities of rat. The method provides a noninvasive biomonitoring of contact to OP NAs [61].

Determination of hydrolysis products in plasma and urine

Rapid elimination of intact OP causes that OP-modified enzymes and metabolites are more stable in the organism. Thus, the new methods for identification and quantification of OP biomarkers modifications need to be developed [62]. Analytical methods employed are often based on gas chromatography-mass spectrometry (GC-MS), which derivatized substances before analysis, and liquid chromatography-mass spectrometry (LC-MS) which has advantage of not require derivatization.

Minami *et al.* (1997) detected sarin product MPA in Tokyo subway attack victims' urine, using gas chromatography (GC) with flame photometric detection (GC-FPD) [47]. The GC-FPD can be useful for estimating

the exposure level to sarin and is appropriate for a large number of samples.

Lately, a LC-tandem MS method has been developed for quantitative determination of IMPA in blood and urine. The main disadvantage of using hydrolysis products in NAs exposure detection is rapid elimination rate of these products (a few days) from the organism that restrict their usage in retrospective measurements [60]. John H (2010) presented matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) method for detecting and identifying novel adducts of human serum albumin and suggested the method as a confirmation tool for high-dose exposure to NAs [63,64]. Tabun presents a problem as its initial hydrolysis product, EDMPA and ethyl phosphorocyanidic acid, are not stable and hydrolyze further to ethyl phosphoric acid and then slowly to phosphate. Unfortunately, the general population has a high level of ethyl phosphoric acid, due to plasticizers and pesticides [65]. Several assessment methods of NAs metabolites which were mostly founded on GC-MS and LC-MS released over the past two decades. The trend is toward LC-MS nowadays and also MS-MS, which generally provides lower limits of detection than single-stage MS, and combined with a greater selectivity.

Fluoride reactivation of phosphorylated binding sites

This method is an analysis of phosphorylated binding sites of BuChE in plasma or serum sample. It is based on reactivation of phosphorylated enzyme with fluoride ions. The BuChE has a half-life of 5-16 days and abundant enough for biomonitoring exposure to OPs and NAs (plasma concentration, approximately 80 nMol). In this way, the extent as well as the origin of the toxicity can be determined. The other benefit of this method is ability of BuChE inhibition measurement at levels much less than those which can be measured based on decreasing AChE activity [64,66]. An analogous method was performed for the Tokyo subway sarin exposure based on isolation and trypsinization of inhibited ChE, subsequent treatment with alkaline phosphate, followed by isolation, derivatization, and GC-MS analysis of the released phosphyl moiety [67].

Mass spectrometric determination of cholinesterase adducts

Straightforward isolation of adducted BuChE from plasma is carried out by means of affinity chromatography with a procainamide column. It is followed by pepsin digestion and LC-MS-MS analysis of a specific nonapeptide, containing the phosphonylated active site serine. This method surpasses the priors since it can also deal with aged phosphonylated BuChE [68].

A review article by Robin Black published in 2010 provides details of bio-analytical methods for NAs detection [65].

Clinical presentations

The complexity and persistent nature of NAs induce several organic complications among poisoned patients. Compared with other OP compounds and CWA such as sulfur mustard, NAs have relatively more acute lethal toxicity and are known as lethal agents and the deadliest CWAs [5,69]. Severity of clinical manifestations are affected by many environmental factors such as temperature, humidity, wind direction, personal protective equipment, activity level of the soldier and the time during soldier remains in the zone [69,70]. Despite primary treatment and use of urgent countermeasures (atropine and oxime), it may take long to recover or even alleviate the complications. Clinical manifestations can be divided into acute and late complications.

Acute effects

The NAs are fatal in acute phase of heavy exposure. Thus, life threatening complications should be considered by clinicians. Depression of respiratory and vasomotor centers in the brain can induce life threatening manifestations and may lead to respiratory failure [71-73]. Hypoxia is also a life threatening effect which may lead to cerebral edema, convulsions, and histopathological brain damage [4].

ACh accumulation at the muscarinic and nicotinic receptor sites is the reason of most systemic complications. Initial symptoms and signs are mostly related to local effect rather than systemic toxicity.

Ocular system

The most common sign in the eye is miosis which is more observed in vapor exposure. The miosis duration is varied from several days to as long as 9 weeks [74,75]. Sharp or aching ocular pain is due to ciliary spasm and can be associated with headache [76]. Impaired visual acuity, tearing and bloodshot appearance, due to subconjunctival vascular dilation, are other common features [13,73,76].

Respiratory system

Rhinorrhea is generally considered as a local irritation effect but can also occur due to the systemic toxicity. Rhinorrhea is always heavier than those caused by hay fever or cold and the severity is dose-dependent [77]. Bronchorrhea, wheezing, bronchiolar smooth muscle constriction, and ventilator failure may be seen due to large exposure to vapor of OP NAs [73,77].

Cardiovascular system

The expected effect is increased vagal tone which leads to bradycardia and atrioventricular block. In fact, the heart rate can actually increase. It may be due to accumulation of ACh in sympathetic ganglia and at the adrenal medulla, or because of fear and anxiety of the patient. Ventricular arrhythmias are rare [78]. Ludomirsky et al. (1982) reported Q-T prolongation in 14 and malignant tachy-arrhythmias in 6 patients out of 15 accidental OP poisoned patients [79]. Tabun, sarin and VX at 5 to 10 times LD50 caused circulatory arrest a few minutes after apnea in non-treated guinea pigs. Histopathological studies suggesting myocarditis have been reported in animal experiments, though, not conclusively proved in human studies [80].

Nervous system

High doses of NAs can cause fatigue, muscle weakness, and even flaccid paralysis. Generalized fasciculation can continue more than other acute complications [77]. Jalali et al. (2011) studied patients with moderate and severe OP pesticide poisoning 10–210 days post exposure by means of electromyography (EMG) and nerve conduction velocity (NCV). On EMG, sensory-motor peripheral polyneuropathy was observed which a distal sensory deficit was predominantly. The Dysfunction of Sensory nerve (84.4%) was significantly higher than motor nerve (18.7%). Sensory nerve dysfunction in the lower extremities was more common than motor nerves, which was mainly a distal sensory deficit [81]. Seizure is also recorded as an acute effect which can be prolonged with status epilepticus. Apnea may happen abruptly and does not resolve without antidotal therapy [6]. Victims of low dose NAs exposure may experience headache, dizziness, restlessness, anxiety, mental confusion, ataxia, irritability, insomnia, bad dreams, depression, forgetfulness, impaired judgment, and lack of concentration even in absence of any physical signs [1,4,6].

Skin and mucosal membrane

Systemic signs and symptoms can occur about two to three hours after exposure via skin. However, Skin penetrating powers of NAs are different. VX is absorbed through skin nearly eight times more rapidly than other NAs. NA skin absorption increases markedly as surrounding temperature rises from 18 to 46°C [1,82]. Generalized sweating is a common complication following prolonged dermal or inhaled exposure [77].

Gastrointestinal system

Mobility and secretion of gastrointestinal system increase according to excess accumulation of ACh. Nausea and vomiting occur among the first signs followed by dermal exposure and can be due to nervous system

complications. Diarrhea is an infrequent symptom. Among 111 patients examined after Tokyo sarin attack, 60.4% complained from nausea, 36.9% reported vomiting, and diarrhea was observed in just 5.4% of the patients [82]. Hyperglycemia may occur due to adrenal medulla stimulation that raises the blood concentrations of circulatory norepinephrine and epinephrine [6].

Genitourinary system

Urinary system has a critical role in excretion of the NAs. During 24 hours, 76-100% of the radioactive substance presented in the urine of volunteers who received 32P-dimethoate orally [83]. Micturition can occur after large dermal contact and after the inhalation of considerable amounts of vapors [6].

Intermediate syndrome (IMS)

IMS occurs 24–96 hours after exposure to organophosphate pesticides or theoretically after NAs exposure [84]. Recovery begins 4 to 18 days later. IMS is characterized by reversible weakness in proximal muscles, especially chest muscles, and cranial nerve palsies [85,86]. Although the etiology of IMS is not well defined, delayed AChE inhibition, down regulation or desensitization of postsynaptic ACh receptors, muscle necrosis, oxidative stress-related myopathy and failure of postsynaptic ACh release are some proposed involved mechanisms [87]. Plasma AChE of less than 200 units is a predictor and the 30 Hz repetitive nerve stimulation depreciatory response could be a useful marker for the IMS [88]. Because of potential dangers of IMS, clinicians must be aware of the syndrome and should perform neuromuscular studies and use mechanical ventilation if necessary. However, there is limited data regarding the occurrence of IMS after NAs exposure [89]. IMS has not been observed obviously after NAs intoxication in animal nor has it been noted in the handful of persons with high contact to NAs [90].

Organophosphate-induced delay neuropathy (OPIDN)

It is defined by sensory and motor disorder of the peripheral nervous system 2–4 weeks after exposure and characterized by progressive weakness, impaired reflexes and distal paraesthesia [91]. Inhibition of an enzyme called neuropathy target enzyme (NTE) in CNS is responsible in OPIDN [92]. Degeneration of myelinated axons and inhibition of NTE is the probable etiology of OPIDN. After 1–4 weeks post exposure, approximately 30% of the patients represent cholinergic irritation (nose secretion, increased salivation, pharyngitis, and laryngitis) followed by paralysis of the leg muscles which persists for 1–2 month but does not leave any changes in sensitive innervation. Then, denervation and atrophy of the leg muscles is observed [86,91,92].

Late complications

The NAs are less likely to cause chronic diseases in comparison with other CWAs. However, NAs poisoning was reported to have association with late complications in both experimental animals and human beings. Hypoxic encephalopathy is one of the most remarkable long-term neurologic effects of NAs reported by Newmark [11]. Cardiomyopathy has been reported in soman and sarin intoxicated rats, which may be contributory cause of death, however it is not reported in human cases yet [80]. Neurological assessment of 43 Iranian veterans 22–27 years post exposure revealed fatigue, paraesthesia and headache as the most common symptoms and sensory nerve impairments as the most common observed clinical complication. The authors concluded that late neurological complications of CWAs poisoning are notable [93]. Sensory nerve dysfunction is more prevalent than motor nerves, which predominantly was a distal sensory deficit [94].

Engel *et al.* (2004) described fatigue as one of the presentations of “Gulf war syndrome” as well as depression and chronic pain [95]. Electroencephalogram (EEG) studies on sarin patients showed considerable slowing with bursts of high voltage waves at a rate of five per second epileptic type changes of EEG, 11 months after the exposure [96,97]. Asthenia, insomnia, fatigue, blurred vision, narrowing of the visual field, shoulder stiffness, slight fever, and asthenia was associated with grades of sarin contact 1 and 3 years after Tokyo subway explosion [98]. Long-term psychological effects are also recorded. Fullerton and co-authors (1990) on a review of article mentioned temporary psychological effects such as depression, insomnia, fatigue, nervousness, irritability, and memory impairment as long-term complication of acute and chronic exposure to NAs [99]. Page (2003) on a telephone survey of 4,022 sarin exposed patients 28 years post exposure reported significantly more concentration lack and sleep disturbances in the patients in comparison with the controls [100]. Grauer *et al.* (2008) has studied late neuronal and behavioral deficit after sarin exposure to rats. The glial activation following neural damage was also established. The data showed long lasting impairment of brain function after single sarin exposure in rats that developing with time [101]. There is not acceptable evidences on carcinogenicity, mutagenicity and teratogenicity of NAs [32,41,76].

Management of NAs poisoning

First aid advices (hot zone)

Treatment for a severe NA exposure must be started immediately, and even seconds are important for making the difference between life and death. The first aid for victims of NAs is their immediate removal from the field or contaminated area. The rescuers should worn

protective devices to prevent exposure. They must pay attention to Airway- Breathing-Circulation (ABC), and put the unconscious casualty in recovery position to prevent aspiration as the consequence of possible vomiting. When the victim is apneic and medical aid station is not near, anybody who wants to assist might consider mouth-to-mouth ventilation [102]. The rescuer should be sure about presence of vapor hazard before initiation, though, is not always possible. Only less than 10% of inspired sarin is expired [103]. It is estimated that hazard of expired breathing of casualty is minor [102].

It must be remembered that usually the casualties of NA attack are not pure chemical victims and they might simultaneously have other blunt or penetrating injuries that need evaluation and treatment. Thus they must be completely assessed.

As quickly as possible, decontamination and antidote therapy, based on severity, should be initiated [1]. Antidotes could be injected with several type of auto injectors by own victim or everybody finds him.

There are some types of auto-injectors that have different amount of antidotes, such as MARK I kit and antidote treatment nerve agent auto-injector (ATNAA) [102,104]. MARK I kit, which is the most popular one, is composed of 2 mg atropine (0.7 ml) and 600 mg 2-pyridine aldoxime methyl chloride (2-PAMCl) [105]. The ATNAA, designated by the Department of Defense of U.S., contains 2.1 mg/0.7 mL atropine and 600 mg/2 mL 2-PAM, and has ability of simultaneously injection both of them through single needle [102,106].

Every soldier carries 3 kits and one auto-injector containing 10 mg diazepam when there is a suspicious of NA attack [105].

One MARK I should be given to a casualty with only miosis and severe rhinorrhea. The second one should be added depends on the severity of respiratory distress. Applying three MARK I kits and diazepam is necessary when severe breathing difficulty or apnea, cyanosis, muscle fasciculation or twitching, seizure or loss of consciousness are present [105,107-110].

The dose of atropine of MARK I kit is between therapeutically desirable dose and safely administrable dose to a non-intoxicated person [102]. The major disadvantage of 2 mg of atropine is decreasing in sweating. The walking tolerance of 35 soldiers who were treated with 2 mg of atropine significantly decreased because of raising their body temperature resulted in limitation of sweating [111].

Absorption of antidotes when administered with autopens is more rapid than by intramuscular needle-and-syringe, because injection by autopens sprays the liquid throughout the muscle as the needle goes in, while the classical types of needle-and-syringe make a "globe" or puddle of liquid in muscle [102].

Decontamination

NAs vapor readily absorb through inhalation and eye contact and they rapidly produce local and systemic effects. However, the absorption of the liquid types is readily through the skin, their effects might be postponed for several minutes and even up to 18 hours [110]. Contaminated skin or clothing of victims can contaminate others by close contact or through off-gassing vapor [110]. Thus, decontamination should be performed as soon as possible to reduce skin absorption of NAs and prevention of the rescuers contamination, members of medical team and other patients [1,105]. This is the best that all casualties before transport are decontaminated [110].

Medical and paramedical personnel who manage and assist to casualties either in the field, during transportation or in the hospital, must protect themselves from NAs contamination [105]. Eight staff who had taken apart in Matsumoto incident had mild symptoms of sarin poisoning [112]. Two important things for this purpose are applying personal protective equipment and decontamination of patients before entering a clinic or a ward [1]. Surgical or similar mask and gloves are not sufficient, and personnel should apply mask containing a charcoal filter, heavy rubber gloves and proper cloths. They should avoid skin contact with victims before decontamination [1,105,109,113].

Furthermore, patient handling equipment, such as gurney and back boards, should be decontaminated to prevent cross contamination. Because of easier cleaning of fiberglass back boards, it would rather to use this type of the equipment [110].

The decontamination has two important parts: physical and chemical. Early physical removal of the agent is more preferred than chemically. It means that decontamination should not be delayed because of unavailability of suitable solution and it must be started by the best means available such as water, soap plus water or other common household products to prevent NA absorption. In sarin contaminated animals which were flushed with water, dose requirement of Sarin for inducing the same mortality rate as non decontaminated animals have been raised up to 10.6 times. In This method, physical removal predominates over hydrolysis [114].

Desirable secondary objective is detoxification (destruction chemically) of the NA. Chemical decontamination may be performed by methods: water/soap wash, oxidation, and acid/base hydrolysis [114]. G agents and VX have phosphorus groups that can be hydrolyzed [114-116]. Furthermore, VX contains sulfur molecules which are readily subject to oxidation reactions [114,115]. Oxidation/Hydrolysis is one of the main routes of CWAs decontamination [114].

Oxidative chlorination with "active chlorine" is the most important reactions of chemical decontamination. A 0.5% sodium or calcium hypochlorite solution (household bleach) for decontamination of skin followed by copious water rinsing is recommended. A 5% solution of the household bleach should be used for contaminated tools [1,114,117].

The hydrolysis rate of NAs increases at pH values higher than 8 and it also four times increases for every 10°C rise in temperature of water [118]. However, there are some potent detoxificant solutions, such as NaOH. They can damage the skin and tissues [114]. VX and the G agents are quite well hydrolyzed by alkaline pH, hypochlorite, as well as sulfur mustard [15,70,114,117,118].

Washing with water/soap, fresh water or sea water can also remove warfare agents through hydrolysis, slower than the other type of physical decontamination [15,18,119]. High lipid solubility of NAs significantly limits the hydrolysis rate [114]. Applying alkaline soap may increase the detoxification through rising water solubility and alkaline hydrolysis [4,114].

All contaminated clothes, shoes and jewelry of victim, should be taken off and flooded in a 5% solution of hypochlorite or put inside a plastic bag and sealed. Decontamination of intertriginous areas, axilla, groin, under the nails and hair, is also essential [1,120]. If the casualty only exposed to NA vapor, skin decontamination is not necessary, whereas his/her clothes should be all removed [105].

M291 resin kit, that contains carbonaceous adsorbent, a polystyrene polymeric, and ion exchange resins, is well suited for field use due to small and dry thus soldier are able to carry it easily [114,121,122]. It can be used on the skin, the face, and around wounds. As powder is scrubbed over the contaminated skin, its carbonaceous material rapidly adsorbs the agents and physically removes them from skin. Then the trapped agents in the interior of the resin particles will be neutralized through chemical detoxification due to the presence of basic and acidic groups in the resin. These groups destruct the agents by way of acid and base hydrolysis [114,123].

It seems that dry powders like flour, earth, and soap detergents are useful. In emergency condition, pushed flour over contaminated area, followed by wiping with wet tissue paper, has been efficient against soman, VX, and sulfur mustard [124].

The ability of mass production of G agent degrading enzyme is possible with over-producing recombinant cell line that has encoded genes of OP acid anhydrolases [125]. In addition, for detoxification of NAs, *Escherichia coli*, which OP hydrolase was expressed on surface, is immobilized by utilization of cell immobilization technology [126]. Phosphotriesterase extracted from the soil bacteria *Pseudomonas diminuta* is also applied for

recognition and decontamination of insecticides and CWAs [127]. One BChE mutant G117H, was prepared through protein engineering techniques, can hydrolyze V and G agents, however, it does not react so fast [1]. There is some sponge made by a polyurethane matrix that is covalently coupled cholinesterases. They can trap and detoxificate NAs from contaminated surface [1,4,128].

As mentioned previously, NAs casualty are not pure chemical victims and they may suffer some types of other injuries and wounds which have or not bandage dressing and need to decontaminate. The toxicity of NAs could reach to wound tissues and increases injuries and toxicity. In this state, due to rapid absorption, a small drop could be lethal [128]. VX absorption is less quickly than other NAs and may persist longer in the wound [110]. All bandages should be removed and wounds be decontaminated by flushed water and remove all foreign materials from wounds. Wounds will be bandaged again only if bleeding recurs. Tourniquets and Splints are also replaced with clean ones, under physician supervision, and original sites should be decontaminated [13,114].

Cross contamination of surgeon works with contaminated wounds result by NAs on foreign bodies in the wound and by the thickened agents. It does not arise from off gassing. Thus, surgical personnel do not need to chemical-protective mask when there is not foreign body or the thickened agents. The surgeons and their assistants should wear a pair of thin, butyl rubber gloves or double latex surgical gloves and change them when they sure that the wound is free of foreign bodies or the thickened agents [110,129,130].

As Hypochlorite solution has potential for corneal injuries, it is contraindicated for the eyes [105]. If the eyes have been exposed to liquid NAs, they should be irrigated within minutes of exposure with running water or saline by leaning the head to the side, pulling eyelids apart with fingers, and pouring solution gentle [1,110]. However, flushing is not necessary for an eye exposure to NA vapor [110]. Hypochlorite solution is also contraindicated for irrigation of the abdominal cavity and not recommended for brain and spinal cord injuries [114]. Do not induce emesis in cases of NAs ingestion, administer activated charcoal without delay, if the victim is conscious and able to swallow [110].

Treatment

General

The patients should be evaluated completely because the NA poisoning may be complicated with multiple traumas or other CWAs. Conscious patients with full muscular power will require minimal care. Victims with

possibility of liquid exposure need to be observed at least 18 hours [110].

Airway, breathing, and circulation should be evaluated. Administer antidotes without delay. Intubate the trachea in case of respiratory compromise and suck excessive bronchial secretions [110]. Severely hypoxic patients need supplemental oxygen with positive end-expiratory pressure [1,2]. It is important that improved tissue oxygenation before administration of atropine to reduce ventricular fibrillation risk [4].

Anticholinergics and Atropine sulfate

Atropine is a parasympatholytic and competitive antagonism of ACh on muscarinic receptors [15,131]. It is an antidote for the muscarinic signs but not nicotinic and CNS symptoms of NAs poisonings [6]. Therefore, atropine has not been able to neutralize fasciculation, weakness, flaccid paralysis, or respiratory arrest which is resulted by blocking neuromuscular nicotinic receptors [4,132]. In addition, atropine does not restore blocked AChE and thus it is not curative [6,102]. But it is very effective in reversing bradycardia, drying the secretions of exocrine glands and reducing smooth muscle constriction result in decreasing bronchoconstriction and hypermotility of gastrointestinal [15,102,105]. The goal of treatment with atropine sulfate is, alleviation of bronchoconstriction and bradycardia and drying the secretions. Thus, its dosage should be titrated based on these aims and there is no clarified exact dose for atropine [1]. Balali-Mood has recommended the following protocol based on his experience on Iranian OP pesticides intoxicated patients and the Iranian soldiers who exposed to NAs,: Atropine is started at 2 mg, as available in auto-injector, and will be added based on patient response up to sings of mild to moderate atropinization (tongue dryness, reduced secretion of oropharyngeal and bronchial tree, tachycardia, and flushing) be appeared [1,4,15]. The same dose that induces initial atropinization should be constantly infused in 500 mL dextrose 5% to sustain mild atropinization and repeated based on need until the patient becomes asymptomatic. The main objective is the dryness of the mucosal membrane [1,4]. However, according to clinical experience of Balali-Mood, dose requirement of atropine for NAs is much lower than for the severe OP poisoning [1,4]. It may be due to greater fat solubility and slower metabolic rate of OP pesticides in compare with NAs [102].

Foroutan, who had treated Iranian casualties in the field and another hospital of Iran, reported another similar atropine administration protocol. He has recommended 4 mg atropine for initial dose. Then after 1 to 2 minutes, he administered another 5 mg intravenously over 5 minutes unless atropinization signs had presented. He checked pulse rate through infusion. He

titrated his dose according to pulse rate and tried to set at 60–110 beat per minute in adults [102,133].

The endpoints of atropinization has been recommended by different authors, Balali-Mood [1,4], Foroutan [133], Eddleston [134], Sidell [102], are very similar; ease of respiration, lack of bronchoconstriction, drying of respiratory secretions, and a heart rate > 80 beats per minute.

Atropine absorbs via bronchial tree, thus it could be administrated in hypotensive patients through endotracheal tube or intratracheally, and it will be shown local and systemic effect [1,4,15]. “Medical aerosolized nerve agent antidote” (MANAA) is an inhaled form of atropine which is used by United States military physicians and has been approved by FDA [102].

Atropine could not reverse NAs induced miosis, except in high doses [102,105] and the pupils’ size is not a response indicator [102]. If 15 to 30 minutes after the vapor exposure has terminated, a victim has only presented miosis, atropine administration may not be indicated [102]. However, if victim, with only miosis, is visited immediately after nerve agent vapor exposure, he/she should receive one Mark I kit or ATNAA [102]. Ophthalmic application of atropine like hemotropine could reduce severe eyes or head pain associated with nausea and the miosis. As these topical application are able to cause prolong blurred vision, they should not be used without appropriated reason such as severe pain.

Atropine Side effects include delirium, inhibition of sweating that induces heat related illness [105].

It is predictable that any compound that could block cholinergic might have antidotal activity [102]. More lipid soluble anticholinergic substances could penetrate the CNS more readily than atropine and display greater antidotal activity [96,135].

Benactyzine is a lipid soluble anticholinergic drug which has been used as antidepressant [136,137]. Although, its administration for these indications is limited due to the side effects, it is shown that the CNS effects of NAs intoxication are reversed more rapidly by benactyzine than atropine [102,135]. Furthermore, benactyzine inhibits sweating or impairs accommodation much less than atropine. Therefore, it seems that it is more suitable than atropine for soldiers particularly in warm environments [102]. Benactyzine could also terminate NAs induced seizure more effectively than diazepam, in guinea pig model [138]. Some countries use “TAB” for immediately nerve agent treatment. It contains TM B-4 (an oxime), atropine, and benactyzine [102].

Oximes

Oximes are mainly pyridinium compounds which are divided into mono and bi-s pyridium. Their general formula is $R_1R_2C=NOH$, where R_1 and R_2 represent any

carbon group or hydrogen [1,139]. They are nucleophilic substances and reactivate the phosphorylated cholinesterase enzyme by breaking the nerve agent-enzyme bond [13,105]. Therefore, it is believed that Oximes are more physiologic antidotes than atropine for NAs poisoning [102,140].

According to mechanism of oximes action, it seems that they might completely reverse the NAs effects. However, there are five reasons that they are practically less effective than atropine as follows:

- (I) It is possible that NAs act through mechanisms other than ChE inhibition [102,107,141].
- (II) The oximes are unable to reverse apparent clinically muscarinic signs and they act on the nicotinic sites [4,15,105].
- (III) Most of oximes are quaternary drugs with limited CNS penetration, thus they could not improve central effects of NAs intoxication [142,143]. Although a prodrug of 2-PAM, that is tertiary amine, has penetrated into the blood brain barrier, it quickly undergoes oxidation in the brain to produce an active 2-PAM and reactivate OP-inhibited AChE in the CNS. However synthesis of this prodrug is complicated now due to rapid autoxidation [144].
- (IV) A meta-analysis results of six clinical trials on OP poisoning demonstrate a high relative risk for death among oxime-exposed [2,17]. It also showed the necessary to have ventilation of patients who received oxime was 1.53 more than others. And the incidence of IMS for patients who received oxime was 1.57 higher than patients treated without oxime. The authors of this paper concluded that oximes are not only effective in the management of OP poisoning, but also they can be dangerous and worsen the patient's clinical situation [145].
- (V) When cholinesterase attached to NAs gets to be aged, it will become resistant to reactivation by oxime or water [107,139]. This reaction limits the efficacy of oxime in fast aging NAs as soman [105,107].

Efficacy of different types of oximes against NAs are not equal (Table 2) [4].

Boskovic had evaluated efficacy of HI-6, HGG-12, and paralidoxime (2-PAM) in conjunction with atropine and diazepam on soman and tabun intoxicated dogs. He reported that HI-6, HGG-12, and 2-PAM had showed the best protective effect in soman-poisoned dogs. However none of them had shown significant protection against tabun [146]. In another in vitro study, both H oximes (HLO-7, HI-6) and BI-6 were found to be more effective in reactivation of sarin and VX-inhibited AChE

Table 2 Relative effects of oximes in organophosphorous nerve agent poisoning

Oximes	Soman	Tabun	Sarin	Cycoserin	VX
HI6	++++	+ / ++	+++	++ / +++	+++
HLO7	+++	-	++++	++++	+++
HGG12	+++	-	NA	NA	NA
2-PM	++	-	++ / +++	+ / -	++ / +++
TMB4	NA	++	NA	NA	NA
BI6	NA	-	++	NA	++
obidoxime	+ / +++	++	++	++	+++
pyrimidoxime	++	++	++	+	++
K oximes	-	++ / +	NA	NA	NA

NA=No data available - =no effective += less effective ++= mild effective. +++= moderately effective ++++= most effective.

than 2-PAM and obidoxime. However, HLO-7 was less effective than HI-6. The HLO-7, HI-6 and BI-6 could not reactive tabun-inhibited AChE efficiently [147,148]. Reactivating potency of AChE inhibited by soman, sarin, cyclosarin, and VX is decreased in the order of HLO7>HI-6>obidoxime>2-PAM [1]. Therapy of intoxicated rat with GV demonstrated best antidotal effect of combination of benactyzine, atropine and HI-6 [149].

It is estimated that both newly developed K oximes (K074, K075) have higher efficacy in antidotal effect on acute tabun poisoning [150,151]. Some studies on tabun intoxicated mice and rats have been shown that K074 is more potent reverser tabun-inhibited brain AChE in rat than the other commonly used oximes [152,153]. Also K074 and K075 were effective in reversing tabun-inhibited blood AChE of rat almost as much as obidoxime [152,153]. Both of them have presented much more therapeutic efficacy in tabun intoxicated mice than obidoxime and HI-6 [150]. In rats intoxicated with tabun, reactivation of inhibited AChE in brain tissue was increased in the order HI-6 < K048 < obidoxime. This reactivation was prominent in frontal part and HI-6 was not a good reactivator against of tabun intoxication [154]. In another report, oxime effectiveness against tabun poisoning decreased in order Trimedoxime (TM B4)> 2-PAM > K127> K117 [102,155].

HI-6 is more effective in treatment cyclosarin toxicity of mice and reversing rat cyclosarin -inhibited AChE of blood and brain than other oximes such as obidoxime and K oximes [150,153]. Based on other studies, HI-6 and HLo7 have been extremely effective against cyclosarin, although obidoxime was fairly effective and the least effective agents were pyrimidoxime and PAM-2Cl [4].

In soman-intoxicated guinea pigs, HI-6 is slightly more effective than HLO-7 [156]. If efficient doses of HI-6 is administrated it can achieve efficient concentration in bran to reactivate inhibited AChE [1]. The signs of

soman poisoning have positive correlation to AChE inhibition and negative correlation to the level of unbound HI-6 in the brain [157]. Also, brain uptake of HI-6 is significantly reduced by soman intoxication [157]. The K oximes (K074, K075) and obidoxime had no effect on reversing rat soman-inhibited AChE of blood and brain, however, HI-6 was very effective [153].

HLO-7 may reactivate phosphorylated muscular AChE by sarin, cyclosarin, soman, and tabun in decreasing order [158]. HLO-7 is extremely effective in tabun intoxicated guinea pigs in compare of HI-6 and pyrimidoxime [156].

Pro-2-PAM is a pro-drug dihydropyridine derivative of the 2-PAM. The pro-2-PAM has showed reactivation of sarin or VX-inhibited AChE of brain tissues and peripheral, in a dose-dependent manner, although it has been greater efficient in peripheral tissues compared to brain [159]. Pro-2-PAM has blocked sarin- or VX-induced seizures as well [159]. Though, Pro-2-PAM had no reactivation of cyclosarin-inhibited AChE in brain or muscle tissues [159]. This oxime also had no effect against cyclosarin-induced seizures [159].

The results reinforce the theory that therapeutic response of oximes depends on NA type [150]. Although, other factors such as cost and availability of the oxime and its side effects influence the selection of the oxime [1]. For example, toxicity of obidoxime (especially with high doses) is higher than 2-PAM and HI-6. However HI-6 is not as commercially available as obidoxime or 2-PAM in several countries [4]. While, the majority of our knowledge about side effects of oximes are limited to animal studies, the human experiences are limited to apply 2-PAM and obidoxime either in pesticides or war/terrorism [1]. In the United Kingdom the methanesulfonate salt of 2-PAM is the standard oxime, whereas, in other European countries TMB4 and obidoxim are used. Pralidoxime iodide is used in Japan. 2-PAM was chosen for use in the United States. HI-6 is used in Canada [102].

Administration of 2-PAM should be started at a dose of 30 mg/kg (up to 2 grams) intravenously over 30 minutes and followed by continues infusion of 8–10 mg/kg/hr (up to 650 mg/h) in dextrose 5% solution. It could be continued till the full healing or atropine is required [1,4,160]. Animal studies showed that a plasma level about 4 µg/mL could reverse sarin-induced neuromuscular block. Administration of 2-PAM with the Combo-Pen or MARK 1 auto-injector (600 mg) intramuscularly could produce a plasma concentration about 6.5 µg/mL in an average soldier (8.9 mg/kg in a 70-kg male) [102]. The oximes should be initially administered after or at the same time of atropine [102,108,129].

In humans, 2-PAM adverse effects are minimal at therapeutic doses [102,160]. Transient dizziness, blurred

vision, diplopia and elevations in diastolic blood pressure may be depended to the administration rate. Some of other reported adverse effects include: headache, drowsiness, tachycardia, increased systolic blood pressure, hyperventilation, decreased renal function, muscular weakness, nausea, vomiting and pain at the injection site [102,129,160]. Administration of 45 mg/kg 2-PAM can elevate systolic and diastolic blood pressure up to 90 mm Hg and 30 mm Hg, respectively [102]. The elevations may persist for several hours. Hypertensive effect could be minimized by giving the oxime more slowly (over 30–40 min) and reversed by phentolamine 5 mg, intravenously [102,160]. Rapid intravenous administration of 2-PAM has produced sudden cardiac and respiratory arrest due to laryngospasm and muscle rigidity [161-163]. Due to side effects administration of more than 2.5 g of oxime through 1 to 1.5 hours is forbidden [102].

More than 80% of 2-PAM is excreted unchanged through the kidneys within 3–12 hours [164,165]. The main suggested mechanism of 2-PAM kidney excretion is active tubular excretory mechanism [102,164,165]. Heat, exercise, renal failure and thiamine could decrease clearance and excretion of 2-PAM [102,164,165].

Initially and daily doses of obidoxime is not recommended more than 500 mg and 750 mg/day, respectively, due to its hepatotoxicity. During obidoxime therapy regular control of Liver function tests should be done [1]. Also liver enzymes concentrations must be observed in patients that receive doses of 1200 to 1800 mg through auto-injector contain 2-PAM. Enzymes concentrations return to normal within 2 weeks [160].

Convulsion and Diazepam

The results of a study about efficacy of diazepam in treatment of NAs have shown that it would be an excellent adjunct therapy [1]. Convulsions should be controlled by utilizing diazepam (0.2 to 0.5 mg/kg in children and 5 to 10 mg in adults) [110,130].

It has not only symptomatic anticonvulsant effect but also has more specific effect on cholinergic and GABAergic systems [1]. In severe cases of NA exposure, convulsion (or what are described as “convulsive jerks” or “spasms”) starts within seconds after losing consciousness and collapsing the casualty. It will persist for several minutes until the victim becomes flaccid and apneic [102]. It is not reported that the convulsion has recurred after atropine and oxime therapy and ventilation support. In these cases, specific anticonvulsive therapy is not required [102].

In animal models, diazepam has been revealed to control NA-induced seizures/convulsions [138,166,167]. It also reduces brain lesion induced by NAs [138,168]. Also, food and drug administration (FDA) have

approved diazepam for treatment of status epilepticus seizures via intramuscular route [102]. Thus, auto-injector of diazepam (contain 10 mg) as Convulsive Antidote Nerve Agent (CANA), is given to US military personal for immediate anticonvulsant treatment of NAs casualties in the field [102]. CANA is not considered for self-use, it rather uses in a soldier exhibits severe effects from a NA by others [102,169]. However, it is recommended to self-injection following the third Mark I or ATNAA, if soldier needs to receive all three kits [102].

Other anticonvulsant benzodiazepines e.g. lorazepam and midazolam, are effective in stopping NA-induced seizure [102,138,167,168,170]. Midazolam, however, is more potent and more rapid than diazepam in stopping NA-induced seizure [171]. It is recommended that midazolam replace diazepam as the urgent anticonvulsant treatment for NA-induced seizures. Barbiturates, phenytoin, and other anticonvulsants are not effective against NA-induced seizure [110].

Some anticholinergic drugs like atropine, benactyzine, aprocphen, azaprocphen, trihexyphenidyl, procyclidine, biperiden and scopolamine, had been tested for their ability to terminate soman induced seizure, in compare of diazepam, in guinea pigs. When drugs had been given 5 min after seizure onset, all these anticholinergic compounds except atropine, were able to terminate seizures at lower doses than diazepam. Seizures were rapidly terminated by procyclidine, Benactyzine, and aprocphen. At 40 min after seizure onset, the most potent compound was diazepam that was followed by scopolamine, benactyzine and biperiden [138].

Anti-NMDA and anti-glutamate drugs

Glutamate, as an excitatory amino acid plays a role in the continuance of OP-induced seizures through

overactivation of the N-methyl-D-aspartate (NMDA) [172,173]. Some of them are listed in Table 3.

Gacyclidine is a novel anti-NMDA compound which was approved for human use in neurotraumatology [174,175]. In an animal study, soman intoxicated primates were pretreated with pyridostigmine and treated by atropine, 2-PAM, and diazepam. Another group received additional gacyclidine. Only gacyclidine was able to ensure complete recovery of NA poisoning for rapid normalization of EEG activity, clinical recovery and neuroprotection [175]. Early administration of gacyclidine added classical emergency medication could prevent the mortality [1]. In an animal study, it could prevent the neuropathology three weeks after soman exposure, however, it has not high CNS penetration [4]. Administration of gacyclidine at zero to thirty minutes after intoxication obtained optimal neuropathological protection [175].

Tezampanel, a glutamate receptor antagonist, which is specific for kainate sub-type receptors, had been useful against soman-induced seizures when administered one hr after exposure. It reduced the length of status epilepticus recorded by electroencephalographic in a 24 hours period after soman exposure. It also showed useful effects in protection of neuropathy induced by soman, as well [176].

Ketamine is a noncompetitive NMDA receptor antagonist. In one study, the effects of ketamine/atropine sulfate combinations were evaluated as delayed therapy in soman-poisoned guinea pigs. Ketamine could stop seizure effectively and highly reduced seizure-related brain damage when treated one hour post-challenge. Co-administration of ketamine and benzodiazepines increased its efficacy [177]. These effects of ketamine have been proved by another animal study on mice [178]. Ketamine plus atropine combinations have been

Table 3 New recommended treatments for organophosphorous nerve agents

Category	Drug	Benefit
<i>Anti-NMDA and anti-glutamate drugs</i>	Gacyclidine	Early administration could prevent the mortality
	Tezampanel	It reduced the length of status epilepticus induced by soman exposure. Useful in protection of neuropathy induced by soman
	Ketamine	Could stop seizure and reduced seizure-related brain damage, protection against OP nerve agent poisoning of peripheral and CNS AChE
	Huperzine A	Useful effects on seizures and status epilepticus prevention in post-exposure,
<i>Magnesium Sulphate:</i>		Administration in the first day decreases hospitalization period and improve outcomes in patients
<i>Antioxidants:</i>	Vitamin E	Therapeutic effects in OPs induced oxidative stress
<i>Bioscavengers:</i>	BChE purified from human plasma (HuBChE)	Therapeutic blood concentration of BChE can be kept for at least 4 days after a single dose administration
	Fetal bovine serum AChE (FBSAChE)	Protected against multiple LD50s of organophosphate NAs
	Fresh frozen plasma (FFP)	No significant effect

revealed suppression of neutrophil granulocyte infiltration and partially suppression of glial activity as important neuroprotective effects. It could also reduce related pro-inflammatory proteins and mRNA excess and aroused by the soman poisoning [178].

As CNS toxic effects result from increased excretory release of glutamate, neuroprotection can be implemented via anticholinergic effects [4]. Huperzine A is a naturally alkaloid found in the Firmoss *Huperzia serrata* [179,180]. It is a reversible AChE inhibitor, like donepezil, rivastigmine or galantamine [179-181] and NMDA receptor antagonist [182]. Huperzine A is able to cross the blood-brain barrier [183]. Huperzine A has revealed useful effects on seizures and status epilepticus prevention in post-exposure by blocking NMDA-induced excitation [4]. These properties make it useful in protection against OP nerve agent poisoning of peripheral and CNS AChE [184].

Blood alkalization by sodium bicarbonate:

Effects of sodium bicarbonate in OPs poisoning were assessed in moderate to severe intoxicated patients [1,4,118]. The aim was to achieve and maintain the arterial blood pH between 7.45 and 7.55 [1,4,118]. After correction arterial acidosis with intravenous sodium bicarbonate solution, it is administered 3–5 mg/kg/24hr as continuous infusion until recovery or until atropine discontinued [1,4,118]. Dose adjustment is done based on regular arterial blood gas analysis.

Esteratic portion of OP molecules are hydrolysed in alkaline pH. Increasing one unit of pH is accompanied with 10 fold increasing in OP hydrolysis [185]. The arterial pH of higher than 7.50 makes the OP hydrolysis faster through hydrolysis acceleration [186].

Alkalinization of blood in pH more than 7.50 results in increasing urinary pH. It stimulates extraction of weak acids. The most of NAs and their metabolites are weak acids [187]. Administration of sodium bicarbonate helps to control the cardio toxicity through augmentation sodium pump channel [188]. It is also estimated that alkalization facilitates recovery from OP poisoning thorough preventing the cardio-respiratory arrest, increasing the bio-availability of oximes, augmentation therapeutic activity of atropine and direct effect of sodium bicarbonate on neuromuscular functions [185].

Ventilation

Ventilatory support is a main part of treatment of a casualty with severe respiratory compromise [102,105]. In animal studies, giving antidote intramuscularly at the onset of signs had been sufficient to reverse the effects of NAs, however, additional ventilation promotes the effectiveness of antidotes [102,105]. Although, some authors believe antidote therapy and intensive care

management can reduce morbidity and mortality rate, the risk of respiratory failure or incapacitation do not avoid with the available antidotes (atropine, oximes) [1].

When NA vapor exposure is minimum and victim has mild to moderate dyspnea, it may be reversed by the administration of atropine [102]. Because of reversing bronchoconstriction caused by atropine, intubation of conscious patient with respiratory distress could be delayed [1]. Atropine could not reverse central respiratory arrest. If casualty suffers from respiratory distress and is elderly or has cardiac or pulmonary underlying disease, he/she needs supplementary inhalation oxygen in addition to the antidotes. Patient with losing consciousness, generalized muscular twitching or convulsive jerks has apnotic or impaired respiration. Thus, they require appropriate respiratory support and should be intubated with assisted ventilation as soon as possible [1,102].

Increasing bronchial secretion is one reason of respiratory problem in NAs exposed victims. These secretions incline toward thickening, mucoid, and “ropy,” and could plug up the airways. Frequent suctioning and postural drainage of the airways can be helpful [102]. A large amount of secretions and broncho-constriction usually cause high airway resistance of 50–70 cm water [102,105]. Very high airway resistance results in causing some mechanical ventilators to malfunction [1]. After atropine administration, resistance decreases to 40 cm H₂O or less, and the secretions reduce. Thus, ventilation is set up and respiratory support should be adjusted after starting atropine [102]. The NAs need to respiratory support much short than that applied for severe OP insecticide poisoning, because of higher fat solubility of OP pesticides that they tend to store more than NAs [15,102,168]. Unlike other CWAs such as sulfur mustard, chlorine and phosgene which induce pulmonary edema, intoxication with NAs may only require ventilator support for 20 min – 3 hours [102].

Therapy for Cardiac Arrhythmias

NA intoxication could promote transient arrhythmias, however, it may happen after atropine administration in a normal subject [102]. High doses (5–20 LD₅₀) of NAs (sarin, soman, tabun and VX) intoxication in guinea pigs had caused an obvious sinus bradycardia and a consequent complete atrioventricular block within 1–2 minutes, followed by idioventricular rhythm, while, no ventricular tachyarrhythmias had been observed in these animals just before death. In this animal model, atropine and atropine plus oxime reversed right away sinus rhythm in animals which had sufficient respiration [72]. In contrast, treatment in animals without sufficient respiratory support, especially in tabun and soman (10LD₅₀) poisoning, converted sinus rhythm to deleterious ventricular tachycardia through one minute after

treatment [72]. However, this kind of arrhythmia has not been reported in humans [102]. Balali-Mood, based on his experience on Iranian soldiers intoxicated in Iran-Iraq war and patients with OP pesticides poisoning, prohibits physicians from administering atropine to patients with tissue hypoxia e.g. cyanosis of lip and fingers. He advised to correct hypoxia, by clearing the airways and giving oxygen, before inducing atropinisation [1,4]. Valero has reported that propranolol could control cardiac tachycardia and ST depression secondary to large dose of atropine in a young woman who ingested organophosphate pesticide accidentally [189].

Hemoperfusion

Evaluation of Effects of hemoperfusion (HP) via coated resin adsorbent synchrome E-5 in intoxicated dogs had been shown that HP in VX and sarin intoxication is only partially effective [190].

Yokoyama reported a 45 year old woman who intoxicated by sarin during Tokyo subway attack. She suffered serious NA poisoning with deep coma, pupil size less than one millimeter and respiratory problem. She had been treated with atropine, 2-PAM and respiratory support. She underwent hemofiltration and hemoperfusion because of insufficient response to treatment. Then she regained consciousness, her pupils were dilated and cholinesterase activity raised [191].

Following new achievement of intravenous lipid emulsion in treatment of intoxicated patient with lipophilic drugs [192], some authors express a hypothesis that the combination of intravenous lipid emulsions and charcoal hemoperfusion can apply to treatment severe OP poisoning [193]. However, some animal studies showed no significant effect of intravenous lipid emulsion against OP toxicity [194].

Magnesium Sulphate

It has been reported that IV administration of magnesium sulfate (4 g) in the first day after admission would decrease hospitalization period and improve outcomes in patients with OP pesticides poisoning [195]. Magnesium sulfate reduces ACh release through blocking calcium channels [196]. It also reduces CNS overstimulation consequential from NMDA receptor activation and reversed the neuroelectrophysiological defects resulted in OP toxicity [197]. In addition, magnesium sulfate has the bronchodilating effect that is evaluated through widely trials in mild to severe asthmatic patients and it could relieve bronchoconstriction in a dose-dependent manner [198]. However, applying magnesium sulfate in NA casualties needs more research. Iranian experiences in treatment of acute OP pesticides poisoning, disclosed that alkalinization of blood with sodium bicarbonate and also administration of magnesium

sulfate may be efficient in recovery of moderate to severe intoxication (Table 3) [1,118].

Antioxidants

OP compounds generate nitric oxide and reactive oxygen radicals, decrease total antioxidant capacity, increase thiobarbituric reactive substances and lipid peroxidation in acute, subchronic or chronic exposure [4,9]. Vitamin E has shown therapeutic effects in OP induced oxidative stress in rat erythrocytes (Table 3) [4].

Bioscavengers

There are three categories of the bioscavengers for the detoxification of OP compounds. (I) Those that stoichiometrically bind to OP compounds. Every organophosphate mole is neutralized by one mole of enzyme and both of them will be inactive. Cholinesterase, carboxylesterase, and other related enzymes are belonging to this category. (II) Some compounds that known as "pseudo catalytic" like those combining AChE and an oxime. Thus, in the presence of an oxime, the catalytic activity of OP-inhibited AChE happens fast and constantly. (III) Natural catalytic hydrolyze OP substances that make them nontoxic like paraoxonase, OP hydrolase and OP anhydrase (Table 3) [199].

Nowadays, researchers try to investigate proteins with biological scavengers' activity on OP compounds which are acceptable to the FDA, and have ability to be stable in circulation for a long time. Through inactivating OPs before they able to inhibit AChE in CNS, could avoid the current antidotes side effects and will reduce the necessity of rapid administration of antidotes [199]. The criteria of an enzyme for applying as an effective in vivo treatment for OP toxicity include: (I) It should be able to react with all kind of OP NAs quickly, specifically, and irreversibly; (II) It should have a constant circulatory half-life (11–15 days) to be effective as a long acting scavenger; (III) The sufficient quantities of this substance should be easily available. (IV) It should not have immunogenic property [199].

Evaluation of BChE purified from human plasma (HuBChE) in animal models proposes that the therapeutic blood concentration of BChE can be kept for at least 4 days after a single dose administration. Its therapeutic index is about 30 and it is safe for human use and has not any tissue toxicity [199]. HuBChE could be stable in lyophilized form at temperatures 4°C to 25°C for 2 years. Immunological response to this enzyme had no interaction with second time pharmacokinetic profile based on animal models [199]. Fetal bovine serum AChE (FBSAChE) protected mice against multiple LD50s of OP NAs [1,4].

Fresh frozen plasma (FFP) and albumin has been recently evaluated for OP toxicity as bioscavenger. In one

clinical trial on 56 OP poisoned patients, efficacy of four packs of FFP at the beginning of treatment was evaluated. The authors reported no significant differences between the two groups on the atropine and 2-PAM dosage, hospitalization length, mortality and clinical course [200]. In another study, administration of FFP, however, increased in pseudocholinesterase level, it made no favorable trends in clinical outcomes [201].

Other new treatments

This interaction between soman and sarin plus beta-cyclodextrin, suggests that it could be a probable antidote against NAs [202,203]. The evaluation of beta-cyclodextrin ability to detoxify various NAs in vitro models, revealed its efficacy in decreasing order of cyclosarin>sarin>tabun>>VX. It could not detoxify VX. A biphasic detoxification reaction was revealed for Sarin; the primary phase, fast reduction of inhibitory potential and the second is a slower phase [204].

Cell migration resulted by cytokine therapy and stem cells engrafting into injured brain tissue of soman-intoxicated mice showed that cell differentiation into functional neurons [205]. However, this method does not ameliorate memory performance in these animals [206]. Cytokine treatment has also enhanced neuronal regeneration in the hippocampus [206].

Encapsulation of drugs or enzymes, as BChE in nanocarriers has been proposed to enhance the blood brain barrier crossing. It is thus hoped that more effective treatments will soon be available for severe neurotoxic effects of human OP pesticides and the NA poisonings [207].

Galantamine is a ChE inhibitor that acts centrally. It also is a nicotinic allosteric potentiating ligand and applied for treat Alzheimer's disease therapy. Galantamine is a safe and effective antidote against intoxication with NAs, including soman. In one study, it was compared with donepezil, rivastigmine, and (\pm)huperzine A, when administered 1–3 hours after soman administration to guinea pigs. Only galantamine could increase survival of the animals [208].

Drug interactions

Medications including morphine, theophylline, aminophylline, reserpine, and phenothiazine-type tranquilizers may have interaction with OP NAs and thus should be avoided. Prescription of drug like procaine and suxamethonium (succinyl choline) that are hydrolyzed by the enzyme ChE should also be avoid [1].

Treatment of High-risk groups

Pregnant women and fetal toxicity

Fetal intoxication may happen because organophosphate NAs cross the placenta [4]. The sensitivity of fetus to

OP compounds and atropine are higher than their mothers [4]. Clinical experience about pregnant women in Sardasht and Halabjah who exposed to sarin in the Iran-Iraq war, and pregnant women poisoned with OPs pesticides, discovered that mortality rate is higher in fetus than in the mothers [1,4]. Fetuses of survived sarin poisoned pregnant women have died within a few hours to a few days [4]. However pregnant women in the second and third trimesters of pregnancy intoxicated with commercial OP compounds have been successfully treated with atropine and 2-PAM and have delivered healthy newborns [209].

In pregnant women administration of atropine and oximes should be with caution and at lower doses. 2-PAM is a pregnancy category C and should be used as clinically necessary [160]. Obstetric consultation is necessary. Removing of dead fetus should be performed immediately after improving the mother clinical condition [4].

Children

As casualties were seen during the Hallabjah massacre, children are more susceptible to organophosphate NAs and suffered higher mortality than adults [4]. Some reasons for this fact include: (I) children have lesser mass and more surface/volume ratio, (II) they have more immature respiratory system, (III) in young children the stratum corneum in the skin is immature that facilitates dermal absorption and (IV) their neurotransmitter systems are immature that makes children more susceptible to an epileptogenic stimulus [102,104].

The clinical manifestation of NAs in children may be quite different from adults. Miosis in OP poisonings of children is not so common as in adults, and also children may have lesser obvious convulsions/seizures [102].

Sensitivity of children to atropine and oximes is higher [104]. Atropine must be administered at least 0.05 mg/kg intramuscular or intravenously, and higher administration dose is up to 0.1 mg/kg in an obvious cholinergic crisis [102]. Atropine administration should be with monitoring of vital signs, especially the pulse rate. Atropine must be adjusted based on heart rate between 140–160 beat/min [1,104]. However, it is showed that young children generally well tolerated atropine overdose [104]. Loading dose of 2-PAM in children should be at 25 mg/kg, that is infused over 15–30 min. It may be followed by 10–20 mg/kg/hr to achieve a plasma concentration of >4 mg/L [4,132]. As half-life of 2-PAM in children is about twice of adults, in small children the initial dose might not be necessary to repeat as frequent as adult dose repetition [102].

In May 2003, the Program for Pediatric Preparedness of the National Center for Disaster Preparedness (NCDDP) issued the first recommendations and treatment

guidelines of pediatric disaster and terrorism awareness that is nationally accepted [210]. They recommended the Mark 1 Auto-injector kits that should be applied as first treatment of children with severe and critical NA poisoning, especially when intravenous therapy is impossible or unavailable [104,210]. When an accurate weight-based dosing of antidotes is not possible, symptomatic children less than one year old should be given atropine, while older children should be given atropine and 2-PAM from the Mark 1 kit [104].

Elderly

Another high risk groups are the elderly. The elderly mortality and morbidity related to sarin poisoning in Halabjah and Sardasht during the Iran-Iraq war were higher than the others. In the elderly victims, complications and multiple system failure were more common than the other adults. Drug administration, such as diazepam, oxime and atropine also needs more caution [1].

Experience in management of NA poisoning during the Iraq-Iran war

Majority of exposed people in Majnoon Island died within 30 minutes following respiratory failure, hypersecretion, convulsions, apnea and coma. Advance management of moderate to severe intoxicated patients had been done in medical centers in big cities followed applying first aid therapy and decontamination in the field hospital. Recorded clinical manifestations included hypersecretions, miosis, hypotension, diarrhea, abdominal cramps, nausea, vomiting, pulmonary edema, cyanosis, respiratory depression, muscle twitching, loss of consciousness, and convulsions. Hypotension and bradycardia were more recorded before atropine therapy while hypertension and tachycardia accompany with tongue dryness and mydriasis were more observed after atropinisation. Patients with cyanosis and extreme respiratory distress, who received high doses of atropine, had higher morbidity and mortality rate. In contrast OP pesticide poisoning, intermediate syndrome has not been reported with the NA intoxication.

There is not an exact statistical record of the NAs exposed individuals. It has been expected that in March 1984 more than 2000 patients with NAs intoxication (later on diagnosed as tabun) were managed. Mixed poisoning by tabun and sulfur mustard happened because the Iraqi army had used combination of them. Iraqi army also had used sarin accompanied with sulfur mustard in Halabjah massacre [1].

Return to duty

There are various factors that influence deciding about returning a casualty of NAs to his/her duty. The most

important criteria for person exposed in factory or laboratory is level of RBC-ChE activity. They should not return when activity of their RBC-ChE is less than 90% of their base line and are not symptoms free [13,102].

Deciding in military services is more complicated. Following considerations is important:

- 1) If exposure to NA is repeated, will the soldier be in higher risk due to the previous contact?
- 2) How much can the function of man be well?
- 3) What is the platoon necessary to the fighter? [102]

When measurements of blood AChE is not available, prediction about increasing danger from second NAs exposure of a soldier is difficult. However, the level of RBC-ChE activity is not a very useful criterion in field because

- a) Most of the time it is not available in field.
- b) A man with relatively mild effects (rhinorrhea and miosis) may obviously have AChE inhibition.
- c) The enzyme activity may be restored to near normal if soldier uses oxime (MARK I or ATNAA) and agent is susceptible to oxime [102].

Prophylaxis

The first approach for prophylaxis against NAs, is keeping AChE intact (protection of cholinesterases) [211]. It is possible by simple chemical components like reversible inhibitors (if possible carbamates) that reversibly inhibit AChE [211]. Résistance of AChE to NAs inhibition will be higher after it inhibited by carbamates [96]. When AChE is restored spontaneously, it will act normally [96,211].

Pyridostigmine is a carbamates that binds to the AChE for a few hours [105]. Therefore, pyridostigmine is used as a "pretreatment" for NAs exposure [105]. It is administered 30 mg every 8 hours [96,212]. It does not pass the blood-brain barrier and thus causes no CNS toxicity [1]. However, usage of higher doses present some of the same clinical manifestations of NAs, and the recommended doses caused irritating adverse effects in half of the man in a war zone [1]. The efficacy of pyridostigmine for prophylaxis in soman exposure has been approved. It could show no additional benefit in sarin or VX poisoning [1,105]. If standard therapy is not administered after the NAs exposure pretreatment will be ineffective [1]. Also, usage of carbamates should be discontinued after NAs exposure; otherwise, they will worsen, rather than protect against poisoning [1]. Physostigmine also has this ability, however, it is not the choice drug for pretreatment due to its toxicity at the amounts required [1,213].

The second approach for prophylaxis against NAs is "scavenger" effect. The scavengers are exogenous proteins (enzymes) that NAs bound to them and reduce the level NA in the organism [211,214]. However, this opinion can be considered as a "treatment in advance" [211].

Recombinant DNA-derived AChE is a bioscavenger which is potentially candidate for pre-exposure therapy for OPs toxicity [1]. FBSAChE protected mice from multiple LD50 of NAs [1,207]. HuBuChE was also useful in animal models as a prophylactic antidote against the fatal effects of NAs [207,214,215]. HuBuChE as a pre-treatment has been demonstrated to enhance survival of intoxicated patients.

The third approach is applying antidotes used for NAs treatment [211]. Pretreatment with benactyzine + HI-6 was investigated in rats. It can restore soman induced circulatory and respiratory changes [1]. Due to the limitation of prophylactic effect of Pyridostigmine against most kind of NAs, Czech Army investigated pretreatment with a combination of drug (trihexyphenidyl, pyridostigmine and benactyzine,) have designated as PANPAL tablet, for soman or tabun poisoning [1,102,135,211]. Czech Armed Forces also have designed another prophylactic patch named TRANSANT that is transdermal patch containing HI-6 [216,217].

Prophylactic efficacy of Huperzine A in soman toxicity was compared with physostigmine in mice. The result showed a greater protective ratio for Huperzine A (2 times for Huperzine A and 1.5 for physostigmine) which was more long lasting (6 hour for Huperzine A and 90 min for physostigmine). The protective effect of Huperzine A had been followed a single injection, with no necessary for any post-challenge drug administration [183].

Conclusion

NAs are deadliest CWA that need immediate intervention. Applying first aid kits like MARKI is important to reduce toxicity. However atropine and oximes are the main part of treatment. There are several adjuvant and additional therapies such as magnesium sulfate, sodium bicarbonate, gacyclidine, benactyzine, tezampanel, hemoperfusion, antioxidants and bioscavengers that have recently been used for OP NAs poisoning.

Abbreviations

2-PAMCl: 2-pyridine aldoxime methyl chloride; AChE: Acetyl Choline Esterase; ACh: Acetylcholine; ATNAA: Antidote treatment nerve agent auto-injector; APAF1: Protease-activating factor-1; BChE: Butyrylcholinesterase; CANA: Convulsive Antidote Nerve Agent; CarBE: Carboxylesterases; ChE: Choline esterase; CNS: Central nervous system; EDMPA: Ethyl-dimethylaminophosphoric acid; FBSAChE: Fetal bovine serum Acetyl Choline Esterase; FDA: Food and drug administration; GC: Gas chromatography; GC-FPD: Gas chromatography with flame photometric detection; GC-MS: Gas chromatography-mass spectrometry; HuBChE: Butyrylcholinesterase purified from human plasma; IMS: Intermediate syndrome; IMPA: Isopropyl methylphosphonic acid; LC-MS: Liquid chromatography-mass spectrometry; MALDI-TOF MS: Matrix-assisted laser desorption/ionization time-of-flight mass

spectrometry; MANAA: Medical aerosolized nerve agent antidote; MPA: Methylphosphonic acid; NA: Nerve agent; NMDA: N-methyl-D-aspartate; OP: Organophosphorous; OPCW: Organization for Prohibition of Chemical Weapons; OPIDN: Organophosphate-induced delay neuropathy; PMPA: Pinacolyl methylphosphonic acid; TOCP: Triorthocresyl phosphate.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MM: Drafting the article and revision. EDM: assistance in drafting the article. MBM: Conception and design, supervising and revising the manuscript critically several times. All authors read and approved the final manuscript.

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References

- Balali-Mood M, Balali-Mood K: **Neurotoxic disorders of organophosphorus compounds and their managements.** *Arch Iran Med* 2008, **11**:65-89.
- Gunderson CH, Lehmann CR, Sidell FR, Jabbari B: **Nerve agents: a review.** *Neurology* 1992, **42**:946-950.
- Bajgar J: **Complex view on poisoning with nerve agents and organophosphates.** *Acta Medica (Hradec Kralove)* 2005, **48**:3-21.
- Balali-Mood M, Saber H: **Recent advances in the treatment of organophosphorous poisonings.** *IJMS* 2012, **37**:74-91.
- Newmark J: **Nerve agents.** *Neurologist* 2007, **13**:20-32.
- Leikin JB, Thomas RG, Walter FG, Klein R, Meislin HW: **A review of nerve agent exposure for the critical care physician.** *Crit Care Med* 2002, **30**:2346-2354.
- OPCW: **Report of sixteen session of the scientific advisory board.** In *Book Report of sixteen session of the scientific advisory board*; pp. Available at: www.opcw.org/index.php?elD=dam_frontend_push&docID=14882. City; 4-6 April 2011.
- Stewart CE: *Weapons of mass casualties and terrorism response handbook.* London: Jones and Bartlett; 2006.
- Balali-Mood M, Balali-Mood K, Hosseini Shirazi F: **Recent advances in treatment of acute organophosphorous nerve agents poisoning.** *IJPR* 2006, **5**:79-87.
- Fest C, Schmidt KJ: *The chemistry of organophosphorus pesticides.* 2nd edition. Berlin: Springer-Verlag; 1982.
- Newmark J: **Nerve agents.** *Neurol Clin* 2005, **23**:623-641.
- Dunn MA, Sidell FR: **Progress in Medical defense against nerve agents.** *JAMA* 1998, **262**:649-652.
- Sidell FR, Borak J: **Chemical warfare agents: II. Nerve agents.** *Ann Emerg Med* 1992, **21**:865-871.
- Marrs TC, Maynard RL, Sidell FR: **Organophosphate nerve agents.** In *Chemical warfare agents: toxicology and treatment.* Edited by Marrs TC, Maynard RL, Sidell FR. New York: Wiley; 1996:83-100.
- Balali-Mood M, Shariat M: **Treatment of organophosphate poisoning. Experience of nerve agents and acute pesticide poisoning on the effects of oximes.** *J Physiol Paris* 1998, **92**:375-378.
- Balaf RM, Clarke RJ, Read RW, Reid MT: **Application of gas chromatography-mass spectrometry and gas chromatography-tandem mass spectrometry to the analysis of chemical warfare samples, found to contain residues of the nerve agent sarin, sulphur mustard and their degradation products.** *J Chromatogr A* 1994, **662**:301-321.
- Sidell FR, Maynard RL, Marrs TC: *Chemical warfare agents: toxicology and treatment.* 2 Revisedth edition. London: Wiley-Blackwell; 2007. an imprint of John Wiley & Sons Ltd.
- Delfino RT, Ribeiro TS, Figueroa-Villar JD: **Organophosphorus Compounds as Chemical Warfare Agents: a Review.** *J Braz Chem Soc* 2009, **20**:407-428.

19. Jokanovic M: Current understanding of the mechanisms involved in metabolic detoxification of warfare nerve agents. *Toxicol Lett* 2009, **188**:1–10.
20. Takafuji ET, Kok AB: The chemical warfare threat and the military healthcare provider. In *Textbook of military medicine Part I Warfare, weaponry, and the wasualty medical aspects of chemical and biological warfare*. Edited by Zajtchuk R, Bellamy RF, Sidell FR. Washington, DC: Borden Institute, Walter Reed Medical Center; 1997:111–128.
21. *Material Safety Data Sheet Lethal Nerve Agent Sarin (GB)*. Available at: <http://www.gulfweb.org/bigdoc/report/appgb.html>. Accessed August 13, 2012.
22. Volans AP: Sarin: guidelines on the management of victims of a nerve gas attack. *J Accid Emerg Med* 1996, **13**:202–206.
23. Park SE, Kim ND, Yoo YH: Acetylcholinesterase plays a pivotal role in apoptosome formation. *Cancer Res* 2004, **64**:2652–2655.
24. Jokanovic M: Medical treatment of acute poisoning with organophosphorus and carbamate pesticides. *Toxicol Lett* 2009, **190**:107–115.
25. Mumford H, Troyer JK: Post-exposure therapy with recombinant human BuChE following percutaneous VX challenge in guinea-pigs. *Toxicol Lett* 2011, **206**:29–34.
26. Lenz DE, Yeung D, Smith JR, Sweeney RE, Lumley LA, Cerasoli DM: Stoichiometric and catalytic scavengers as protection against nerve agent toxicity: a mini review. *Toxicology* 2007, **233**:31–39.
27. Worek F, Eyer P, Aurbek N, Szinicz L, Thiermann H: Recent advances in evaluation of oxime efficacy in nerve agent poisoning by in vitro analysis. *Toxicol Appl Pharmacol* 2007, **219**:226–234.
28. Jokanovic M: Biotransformation of organophosphorus compounds. *Toxicology* 2001, **166**:139–160.
29. Worek F, Koller M, Thiermann H, Szinicz L: Diagnostic aspects of organophosphate poisoning. *Toxicology* 2005, **214**:182–189.
30. Jokanovic M, Kosanovic M, Maksimovic M: Interaction of organophosphorus compounds with carboxylesterases in the rat. *Arch Toxicol* 1996, **70**:444–450.
31. Gupta RC, Patterson GT, Dettbarn WD: Acute tabun toxicity; biochemical and histochemical consequences in brain and skeletal muscles of rat. *Toxicology* 1987, **46**:329–341.
32. Bajgar J: Organophosphates/nerve agent poisoning: mechanism of action, diagnosis, prophylaxis, and treatment. *Adv Clin Chem* 2004, **38**:151–216.
33. Talbot BG, Anderson DR, Harris LW, Zarbrough LW, Lennox WJ: A comparison of in vivo and in vitro rates of ageing of soman-inhibited erythrocyte acetylcholinesterase in different animal species. *Drug Chem Toxicol* 1988, **11**. A comparison of in vivo and in vitro rates of ageing of soman-inhibited erythrocyte acetylcholinesterase in different animal species.
34. Jokanovic M: Role of carboxylesterase in soman, sarin and tabun poisoning in rats. *Pharmacol Toxicol* 1989, **65**:181–184.
35. Fonnum F, Sterri SH: Factors modifying the toxicity of organophosphorus compounds including Soman and Sarin. *Fundam Appl Toxicol* 1981, **1**:143–147.
36. Clement JG, Copeman HT: Soman and sarin induce a long-lasting naloxone-reversible analgesia in mice. *Life Sci* 1984, **34**:1415–1422.
37. Duysen EG, Li B, Xie W, Schopfer LM, Anderson RS, Broomfield CA, Lockridge O: Evidence for nonacetylcholinesterase targets of organophosphorus nerve agent: supersensitivity of acetylcholinesterase knockout mouse to VX lethality. *J Pharmacol Exp Ther* 2001, **299**:528–535.
38. Cowan FM, Shih TM, Lenz DE, Madsen JM, Broomfield CA: Hypothesis for synergistic toxicity of organophosphorus poisoning-induced cholinergic crisis and anaphylactoid reactions. *J Appl Toxicol* 1996, **16**:25–33.
39. Bajgar J: Present views on toxidynamics of soman poisoning. *Acta Med (Hradec Kralove)* 1996, **39**:101–105.
40. Tonkopii V: Oxidative stress in the mechanism of organophosphates neurotoxicity. *Toxicol Lett* 2003, **144**:132.
41. Lotti M: Organophosphorus compounds. In *Experimental and clinical neurotoxicology*. 2nd edition. Edited by Spencer PS, Schaumburg HH. New York: Oxford University Press; 2000:898–925.
42. van Helden HP, Bueters TJ: Protective activity of adenosine receptor agonists in the treatment of organophosphate poisoning. *Trends Pharmacol Sci* 1999, **20**:438–441.
43. Munro N: Toxicity of the organophosphate chemical warfare agents GA, GB, and VX: implications for public protection. *Environ Health Perspect* 1994, **102**:18–38.
44. Cao CJ, Mioduszewski RJ, Menking DE, Valdes JJ, Katz EJ, Eldefrawi ME, Eldefrawi AT: Cytotoxicity of organophosphate anticholinesterases. *In Vitro Cell Dev Biol Anim* 1999, **35**:493–500.
45. Rickell DJ, Glenn JF, Houston WE: Medical defense against nerve agents: New direction. *Milit Med* 1987, **152**:35–41.
46. van der Schans MJ, Polhuijs M, van Dijk C, Degenhardt CE, Pleijsier K, Langenberg JP, Benschop HP: Retrospective detection of exposure to nerve agents: analysis of phosphofluoridates originating from fluoride-induced reactivation of phosphorylated BuChE. *Arch Toxicol* 2004, **78**:508–524.
47. Minami M, Hui DM, Katsumata M, Inagaki H, Boulet CA: Method for the analysis of the methylphosphonic acid metabolites of sarin and its ethanol-substituted analogue in urine as applied to the victims of the Tokyo sarin disaster. *J Chromatogr B Biomed Sci Appl* 1997, **695**:237–244.
48. van der Schans MJ, Lander BJ, van der Wiel H, Langenberg JP, Benschop HP: Toxicokinetics of the nerve agent (+/-)-VX in anesthetized and atropinized hairless guinea pigs and marmosets after intravenous and percutaneous administration. *Toxicol Appl Pharmacol* 2003, **191**:48–62.
49. U S Department of the Army and U S (Ed): *Military chemistry and chemical compounds. Field manual*. Army. Washington, DC: Department of the Air Force; 1975.
50. Munro NB, Talmage SS, Griffin GD, Waters LC, Watson AP, King JF, Hauschild V: The sources, fate, and toxicity of chemical warfare agent degradation products. *Environ Health Perspect* 1999, **107**:933–974.
51. Reynolds ML, Little PJ, Thomas BF, Bagley RB, Martin BR: Relationship between the biodisposition of [3H]soman and its pharmacological effects in mice. *Toxicol Appl Pharmacol* 1985, **80**:409–420.
52. Polak RL, Cohen EM: The binding of sarin in the blood plasma of the rat. *Biochem Pharmacol* 1970, **19**:877–881.
53. Polak RL, Cohen EM: The influence of triorthocresylphosphate on the distribution of 32P in the body of the rat after the injection of 32P-sarin. *Biochem Pharmacol* 1969, **18**:813–820.
54. Little PJ, Reynolds ML, Bowman ER, Martin BR: Tissue disposition of [3H] sarin and its metabolites in mice. *Toxicol Appl Pharmacol* 1986, **83**:412–419.
55. Little PJ, Scimeca JA, Martin BR: Distribution of [3H] diisopropyl fluorophosphate, [3H]soman, [3H]sarin, and their metabolites in mouse brain. *Drug Metab Dispos* 1988, **16**:515–520.
56. Nakajima T, Sasaki K, Ozawa H, Sekijima Y, Morita H, Fukushima Y, Yanagisawa N: Urinary metabolites of sarin in a patient of the Matsumoto sarin incident. *Arch Toxicol* 1998, **72**:601–603.
57. Allert M, Rizk SS, Looger LL, Hellinga HW: Computational design of receptors for an organophosphate surrogate of the nerve agent soman. *Proc Natl Acad Sci U S A* 2004, **101**:7907–7912.
58. Tsuchihashi H, Katagi M, Nishikawa M, Tatsuno M: Identification of metabolites of nerve agent VX in serum collected from a victim. *J Anal Toxicol* 1998, **22**:383–388.
59. Noort D, Hulst AG, Platenburg DH, Polhuijs M, Benschop HP: Quantitative analysis of O-isopropyl methylphosphonic acid in serum samples of Japanese citizens allegedly exposed to sarin: estimation of internal dosage. *Arch Toxicol* 1998, **72**:671–675.
60. Noort D, Benschop HP, Black RM: Biomonitoring of exposure to chemical warfare agents: a review. *Toxicol Appl Pharmacol* 2002, **184**:116–126.
61. Wang J, Timchalk C, Lin Y: Carbon nanotube-based electrochemical sensor for assay of salivary cholinesterase enzyme activity: an exposure biomarker of organophosphate pesticides and nerve agents. *Environ Sci Technol* 2008, **42**:2688–2693.
62. Marsillach J, Richter RJ, Kim JH, Stevens RC, MacCoss MJ, Tomazela D, Suzuki SM, Schopfer LM, Lockridge O, Furlong CE: Biomarkers of organophosphorus (OP) exposures in humans. *Neurotoxicology* 2011, **32**:656–660.
63. John H, Breyer F, Thumfart JO, Hochstetter H, Thiermann H: Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) for detection and identification of albumin phosphorylation by organophosphorus pesticides and G- and V-type nerve agents. *Anal Bioanal Chem* 2010, **398**:2677–2691.
64. Jakubowski M, Heykamp LS, Durst HD, Thomson SA: Preliminary studies in the formation of ethyl methylphosphonofluoridate from rat and human

- serum exposed to VX and treated with fluoride ion. *Anal Lett* 2001, **4**:727-747.
65. Black RM: **History and perspectives of bioanalytical methods for chemical warfare agent detection.** *J Chromatogr B Analyt Technol Biomed Life Sci* 2010, **878**:1207-1215.
 66. Polhuijs M, Langenberg JP, Benschop HP: **New method for retrospective detection of exposure to organophosphorus anticholinesterases: application to alleged sarin victims of Japanese terrorists.** *Toxicol Appl Pharmacol* 1997, **146**:156-161.
 67. Matsuda Y, Nagao M, Takatori T, Nijijima H, Nakajima M, Iwase H, Kobayashi M, Iwadate K: **Detection of the sarin hydrolysis product in formalin-fixed brain tissues of victims of the Tokyo subway terrorist attack.** *Toxicol Appl Pharmacol* 1998, **150**:310-320.
 68. Fidder A, Hulst AG, Noort D, de Ruiter R, van der Schans MJ, Benschop HP, Langenberg JP: **Retrospective detection of exposure to organophosphorus anti-cholinesterases: mass spectrometric analysis of phosphorylated human butyrylcholinesterase.** *Chem Res Toxicol* 2002, **15**:582-590.
 69. Balali-Mood M, Hefazi M: **Comparison of early and late toxic effects of sulfur mustard in Iranian veterans.** *Basic Clin Pharmacol Toxicol* 2006, **99**:273-282.
 70. DM P: *Environment committee armed forces epidemiological board long-term health effects associated with sub-clinical exposures to GB and mustard.* 1996. Gulf link: Office of the special assistant for Gulf War Illnesses, Available at: www.gul_ink.osd.mil/agent.html.
 71. Rickett DL, Glenn JF, Beers ET: **Central respiratory effects versus neuromuscular actions of nerve agents.** *Neurotoxicology* 1986, **7**:225-236.
 72. Worek F, Kleine A, Falke K, Szinicz L: **Arrhythmias in organophosphate poisoning: effect of atropine and bispyridinium oximes.** *Arch Int Pharmacodyn Ther* 1995, **329**:418-435.
 73. Sidell FR: **Soman and sarin: clinical manifestations and treatment of accidental poisoning by organophosphates.** *Clin Toxicol* 1974, **7**:1-17.
 74. Rengstorff RH: **Accidental exposure to sarin: vision effects.** *Arch Toxicol* 1985, **56**:201-203.
 75. Nozaki H, Hori S, Shinozawa Y, Fujishima S, Takuma K, Sagoh M, Kimura H, Ohki T, Suzuki M, Aikawa N: **Secondary exposure of medical staff to sarin vapor in the emergency room.** *Intensive Care Med* 1995, **21**:1032-1035.
 76. Marrs TC, Maynard RL, Sidell FR: *Chemical warfare agents. Toxicology and treatment.* Toronto: J. Wiley and Sons; 1996.
 77. US Army Medical Research Institute of Chemical Defense: **Nerve agents.** In *Medical management of chemical casualties handbook.* 3rd edition. Edited by (USAMRICD) UAmriocdCccd. Edgewood: Aberdeen Proving Ground; 1998.
 78. Christensen MK, Crethull P, Crook JW, Oberst FW, Ross RS, Umland CW 2nd: **Resuscitation of dogs poisoned by inhalation of the nerve gas GB.** *Mil Med* 1956, **119**:377-386.
 79. Ludomirsky A, Klein HO, Sarelli P, Becker B, Hoffman S, Taitelman U, Barzilai J, Lang R, David D, DiSegni E, Kaplinsky E: **Q-T prolongation and polymorphous ("torsade de pointes") ventricular arrhythmias associated with organophosphorus insecticide poisoning.** *Am J Cardiol* 1982, **49**:1654-1658.
 80. Singer AW, Jaax NK, Graham JS, McLeod CG Jr: **Cardiomyopathy in Soman and Sarin intoxicated rats.** *Toxicol Lett* 1987, **36**:243-249.
 81. Jalali N, Balali-Mood M, Jalali I, Shakeri MT: **Electrophysiological changes in patients with acute organophosphorous pesticide poisoning.** *Basic Clin Pharmacol Toxicol* 2012, **108**:251-255.
 82. Sivam SP, Hoskins B, Ho IK: **An assessment of comparative acute toxicity of diisopropyl-fluorophosphate, tabun, sarin, and soman in relation to cholinergic and GABAergic enzyme activities in rats.** *Fundam Appl Toxicol* 1984, **4**:531-538.
 83. Balali-Mood M, Balali-Mood K: **Nerve agents.** In *Critical Care Toxicology.* Edited by Brent J. Philadelphia, USA: Elsevier Mosby; 2005:1379-1393.
 84. Heide EA: *Cholinesterase inhibitors: Including insecticides and chemical warfare nerve agents Part 5: The intermediate syndrome.* 2012th edition. 2012. Agency for toxic substances and disease registry (ATSDR), Available at: <http://www.atsdr.cdc.gov/csem/csem.asp?csem=11&po=28>.
 85. Rickett D, Glenn J, Beers ET: **Central respiratory effects versus neuromuscular actions of nerve agents.** *Neurotoxicology* 1953, **8**:466-475.
 86. Schecter WP: **Cholinergic symptoms due to nerve agent attack: a strategy for management.** *Anesthesiol Clin North America* 2004, **22**:579-590. viii.
 87. Abdollahi M, Karami-Mohajeri S: **A comprehensive review on experimental and clinical findings in intermediate syndrome caused by organophosphate poisoning.** *Toxicol Appl Pharmacol* 2012, **258**:309-314.
 88. Avasthi G, Singh G: **Serial neuro-electrophysiological studies in acute organophosphate poisoning—correlation with clinical findings, serum cholinesterase levels and atropine dosages.** *J Assoc Physicians India* 2000, **48**:794-799.
 89. Smith WJ, Clark MWG, Talbot TB, Caple PA, Sidell FR, Hurst CG: **Chapter 9, Long-term health effects of chemical threat agents.** In *Textbooks of military medicine, medical aspects of chemical warfare.* Edited by Lenhart MK, Tuorinsky SD. Washington, DC: The Office of the Surgeon General at TMM Publications; 2008:311-338.
 90. Karaliedde L, Wheeler H, Maclehorse R, Murray V: **Possible immediate and long-term health effects following exposure to chemical warfare agents.** *Public Health* 2000, **114**:238-248.
 91. Emerick GL, Peccinini RG, de Oliveira GH: **Organophosphorus-induced delayed neuropathy: a simple and efficient therapeutic strategy.** *Toxicol Lett* 2010, **192**:238-244.
 92. Jokanovic M, Kosanovic M, Brkic D, Vukomanovic P: **Organophosphate induced delayed polyneuropathy in man: an overview.** *Clin Neurol Neurosurg* 2011, **113**:7-10.
 93. Darchini-Maragheh E, Nemati-Karimooy H, Hasanabadi H, Balali-Mood M: **Delayed Neurological Complications of Sulphur Mustard and Tabun Poisoning in 43 Iranian Veterans.** *Basic Clin Pharmacol Toxicol* 2012, **111**:426-432.
 94. Balali-Mood M, Navaeian A: **Clinical and paraclinical findings in 233 patients with sulfur mustard poisoning.** In *Proceedings of the second world congress on new compounds in biological and chemical warfare, toxicological evaluation.* Edited by Heyndrickx B. Ghent, Belgium: Ghent University Press; 1986:464-473.
 95. Engel CC, Jaffer A, Adkins J, Riddle JR, Gibson R: **Can we prevent a second 'Gulf War syndrome'? Population-based healthcare for chronic idiopathic pain and fatigue after war.** *Adv Psychosom Med* 2004, **25**:102-122.
 96. Masson P: **Evolution of and perspectives on therapeutic approaches to nerve agent poisoning.** *Toxicol Lett* 2011, **206**:5-13.
 97. Tang FR, Loke WK, Ling EA: **Comparison of status epilepticus models induced by pilocarpine and nerve agents - a systematic review of the underlying aetiology and adopted therapeutic approaches.** *Curr Med Chem* 2011, **18**:886-899.
 98. Nakajima T, Sato S, Morita H, Yanagisawa N: **Sarin poisoning of a rescue team in the Matsumoto sarin incident in Japan.** *Occup Environ Med* 1997, **54**:697-701.
 99. Fullerton CS, Ursano RJ: **Behavioral and psychological responses to chemical and biological warfare.** *Mil Med* 1990, **155**:54-59.
 100. Page WF: **Long-term health effects of exposure to sarin and other anticholinesterase chemical warfare agents.** *Mil Med* 2003, **168**:239-245.
 101. Grauer E, Chapman S, Rabinovitz I, Raveh L, Weissman BA, Kadar T, Allon N: **Single whole-body exposure to sarin vapor in rats: long-term neuronal and behavioral deficits.** *Toxicol Appl Pharmacol* 2008, **227**:265-274.
 102. Sidell FR, Newmark J, McDonough JH: In *Textbooks of military medicine, medical aspects of chemical warfare.* Edited by Lenhart MK, Tuorinsky SD. Washington D.C: Department of the Army, United States of America; 2008:155-219.
 103. Oberst FW, Koon WS, Christensen MK, Crook JW, Cresthull P, Freeman G: **Retention of inhaled sarin vapor and its effect on red blood cell cholinesterase activity in man.** *Clin Pharmacol Ther* 1968, **9**:421-427.
 104. Baker MD: **Antidotes for nerve agent poisoning: should we differentiate children from adults?** *Curr Opin Pediatr* 2007, **19**:211-215.
 105. Tang SYH, Chan JTS: **A review article on nerve agents.** *Hong Kong J Emergmed* 2002, **9**:83-89.
 106. *Drug information online (drug.com).* 2012. ATNA. <http://www.drugs.com/pro/atnaa.html>.
 107. Marrs TC, Rice P, Vale JA: **The role of oximes in the treatment of nerve agent poisoning in civilian casualties.** *Toxicol Rev* 2006, **25**:297-323.
 108. Yergler M: **Nerve gas attack.** *Am J Nurs* 2002, **102**:57-60.
 109. Brown MA, Brix KA: **Review of health consequences from high-, intermediate- and low-level exposure to organophosphorus nerve agents.** *J Appl Toxicol* 1998, **18**:393-408.
 110. ATSDR: *Medical management guidelines for Nerve Agents: Tabun (GA); Sarin (GB); Soman (GD); and VX.* 2012. Agency for toxic substances and disease registry <http://www.atsdr.cdc.gov/MHML/mmg166.pdf>.

111. Robinson S, Magenis TP, Minter DI, Harper H: **The effects of varying doses of atropine on temperature regulation of men and dogs.** In *The Physiological Effects of Atropine and Potential Atropine Substitutes*. Edited by Robinson S. Edgewood Arsenal, MD: Medical Research Laboratories; 1953.
112. Pangi B: **Consequence management in the 1995 Sarin attacks on the Japanese subway system.** *Stud Conflit Terrorri* 2002, **25**:421–448.
113. Anonymus: *ACT FAST, Agent Characteristics and Toxicology, First Aid and Special Treatment, Student guideline*. U.S. Department of the Army; 2007.
114. Hurst CG: **Decontamination.** In *Textbooks of military medicine, medical aspects of chemical warfare*. Edited by Lenhart MK, Tuorinsky SD. Washington D.C: Department of the Army, United States of America; 2008:351–359.
115. Zheng X, Okolotowicz K, Wang B, Macdonald M, Cashman JR, Zhang J: **Direct detection of the hydrolysis of nerve agent model compounds using a fluorescent probe.** *Chem Biol Interact* 2010, **187**:330–334.
116. Dawson RM, Pantelidis S, Rose HR, Kotsonis SE: **Degradation of nerve agents by an organophosphate-degrading agent (OpdA).** *J Hazard Mater* 2008, **157**:308–314.
117. Balali Mood M, Balali Mood B, Moshiri M: **Sulfur mustard.** In *Encyclopedia of Toxicology*. Edited by Wexler P, Greim H, Moser V, Wiegand TJ, Lafarga JVT, Peyster A, Harper S, Abdollahi M, Gad SC, Ray SD. Elsevier; 2012. in press.
118. Balali-Mood M, Ayati MH, Ali-Akbarian H: **Effect of high doses of sodium bicarbonate in acute organophosphorous pesticide poisoning.** *Clin Toxicol (Phila)* 2005, **43**:571–574.
119. Balali-Mood K, Bond PJ, Sansom MS: **Interaction of monotopic membrane enzymes with a lipid bilayer: a coarse-grained MD simulation study.** *Biochemistry* 2009, **48**:2135–2145.
120. Moody RP, Maibach HI: **Skin decontamination: Importance of the wash-in effect.** *Food Chem Toxicol* 2006, **44**:1783–1788.
121. Seto Y: **Decontamination of chemical and biological warfare agents.** *Yakugaku Zasshi* 2009, **129**:53–69.
122. Ishoj T, Jensen K: **Chemical warfare poisoning in time of peace.** *Ugeskr Laeger* 1999, **161**:808–810.
123. Clarkon ED, Schulz SM, Railer RF, Smith KH: **Median lethal dose determination for percutaneous exposure to soman and VX in guinea pigs and the effectiveness of decontamination with M291 SDK or SANDIA foam.** *Toxicolo Lette* 2012, **212**:282–287.
124. Bismuth C, Borron SW, Baud FJ, Barriot P: **Chemical weapons: documented use and compounds on the horizon.** *Toxicolo Lette* 2004, **149**:11–18.
125. Cheng TC, DeFrank JJ, Rastogi VK: **Alteromonas prolidase for organophosphorus G-agent decontamination.** *Chem Biol Interact* 1999, **119**–120:455–462.
126. Cheng TC, Calomiris JJ: **A cloned bacterial enzyme for nerve agent decontamination.** *Enzy Microb Technolo* 1996, **18**:597–601.
127. Ghanem E, Raushel FM: **Detoxification of organophosphate nerve agents by bacterial phosphotriesterase.** *Toxicol Appl Pharmacol* 2005, **207**:459–470.
128. Gordon RK, Feaster SR, Russell AJ, LeJeune KE, Maxwell DM, Lenz DE, Ross MC, Doctor BP: **Organophosphate skin decontamination using immobilized enzymes.** *Chem Biol Interact* 1999, **119**–120:463–470.
129. Wang RF, Wang TL: **Nerve Agents.** *Disaster Med* 2005, **4**(Suppl 1):S29–S34.
130. ATSDR: **Nerve agents (GA, GB, GD, VX).** In *Agency for toxic substances and disease registry, division of toxicology*. 2012. <http://www.atsdr.cdc.gov/toxfaqs/tfacts166.pdf>.
131. Hrobak PK: **Nerve agents: implications for anesthesia providers.** *AANA J* 2008, **76**:95–97.
132. Barthold CL, Schier JG: **Organic phosphorus compounds—nerve agents.** *Crit Care Clin* 2005, **21**:673–689. v-vi.
133. Newmark J: **The birth of nerve agent warfare: lessons from Syed Abbas Foroutan.** *Neurology* 2004, **62**:1590–1596.
134. Eddleston M, Buckley NA, Cheeketts H, Senarathna L, Mohamed F, Sheriff MH, Dawson A: **Speed of initial atropinisation in significant organophosphorus pesticide poisoning—a systematic comparison of recommended regimens.** *J Toxicol Clin Toxicol* 2004, **42**:865–875.
135. Kassa J: **Therapeutic and neuroprotective efficacy of pharmacological pretreatment and antidotal treatment of acute tabun or soman poisoning with the emphasis on pretreatment drug PANPAL.** *Arh Hig Rada Toksikol* 2006, **57**:427–434.
136. McLaughlin B, Rickels K, Abidi M, Toro R: **Meprobamate-benactyzine (Deprol) and placebo in two depressed outpatient populations.** *Psychosomatics* 1969, **10**:73–81.
137. Herz A, Teschemacher H, Hofstetter A, Kurz H: **The Importance of Lipid-Solubility for the Central Action of Cholinolytic Drugs.** *Int J Neuropharmacol* 1965, **4**:207–218.
138. McDonough JH Jr, Zoeffel LD, McMonagle J, Copeland TL, Smith CD, Shih TM: **Anticonvulsant treatment of nerve agent seizures: anticholinergics versus diazepam in soman-intoxicated guinea pigs.** *Epilepsy Res* 2000, **38**:1–14.
139. Eyer P: **The role of oximes in the management of organophosphorus pesticide poisoning.** *Toxicol Rev* 2003, **22**:165–190.
140. Kovacic P: **Mechanism of organophosphates (nerve gases and pesticides) and antidotes: electron transfer and oxidative stress.** *Curr Med Chem* 2003, **10**:2705–2709.
141. McDonough JH Jr, Shih T-M: **Neuropharmacological Mechanisms of Nerve Agent-induced Seizure and Neuropathology.** *Neurosci Biobehav Rev* 1997, **21**:559–579.
142. Kassa J: **Review of oximes in the antidotal treatment of poisoning by organophosphorus nerve agents.** *J Toxicol Clin Toxicol* 2002, **40**:803–816.
143. Kassa J, Kuca K, Karasova J, Musilek K: **The development of new oximes and the evaluation of their reactivating, therapeutic and neuroprotective efficacy against tabun.** *Mini Rev Med Chem* 2008, **8**:1134–1143.
144. Mercey G, Verdelet T, Renou J, Kliachyna M, Baati R, Nachon F, Jean L, Renard PY: **Reactivators of acetylcholinesterase inhibited by organophosphorus nerve agents.** *Acc Chem Res* 2012, **45**:756–766.
145. Rahimi R, Nikfar S, Abdollahi M: **Increased morbidity and mortality in acute human organophosphate-poisoned patients treated by oximes: a meta-analysis of clinical trials.** *Hum Exp Toxicol* 2006, **25**:157–162.
146. Boskovic B, Kovacevic V, Jovanovic D: **PAM-2 Cl, HI-6, and HGG-12 in soman and tabun poisoning.** *Fundam Appl Toxicol* 1984, **4**:S106–115.
147. Kuca K, Cabal J, Kassa J, Jun D, Hrabnova M: **A comparison of the potency of the oxime HLo-7 and currently used oximes (HI-6, pralidoxime, obidoxime) to reactivate nerve agent-inhibited rat brain acetylcholinesterase by in vitro methods.** *Acta Medica (Hradec Kralove)* 2005, **48**:81–86.
148. Kuca K, Cabal J, Kassa J, Jun D, Hrabnova M: **In vitro potency of H oximes (HI-6, HLo-7), the oxime BI-6, and currently used oximes (pralidoxime, obidoxime, trimedoxime) to reactivate nerve agent-inhibited rat brain acetylcholinesterase.** *J Toxicol Environ Health A* 2006, **69**:1431–1440.
149. Fusek J, Bajgar J: **Treatment of intoxication with GV compound in laboratory rats.** *Sb Ved Pr Lek Fak Karlovy Univerzity Hradci Kralove* 1994, **37**:57–62.
150. Kassa J, Humlicek V: **A comparison of the potency of newly developed oximes (K074, K075) and currently available oximes (obidoxime, trimedoxime, HI-6) to counteract acute toxic effects of tabun and cyclosarin in mice.** *Drug Chem Toxicol* 2008, **31**:127–135.
151. Kassa J, Jun D, Kuca K, Bajgar J: **Comparison of reactivating and therapeutic efficacy of two salts of the oxime HI-6 against tabun, soman and cyclosarin in rats.** *Basic Clin Pharmacol Toxicol* 2007, **101**:328–332.
152. Kassa J, Jun D, Kuca K: **A comparison of reactivating efficacy of newly developed oximes (K074, K075) and currently available oximes (obidoxime, HI-6) in cyclosarin-and tabun-poisoned rats.** *J Enzyme Inhib Med Chem* 2007, **22**:297–300.
153. Kassa J, Jun D, Karasova J, Bajgar J, Kuca K: **A comparison of reactivating efficacy of newly developed oximes (K074, K075) and currently available oximes (obidoxime, HI-6) in soman, cyclosarin and tabun-poisoned rats.** *Chem Biol Interact* 2008, **175**:425–427.
154. Bajgar J, Hajek P, Zdarova JK, Kassa J, Paseka A, Slizova D, Krs O, Kuca K, Jun D, Fusek J, Capek L: **A comparison of tabun-inhibited rat brain acetylcholinesterase reactivation by three oximes (HI-6, obidoxime, and K048) in vivo detected by biochemical and histochemical techniques.** *J Enzyme Inhib Med Chem* 2010, **25**:790–797.
155. Kuca K, Cabal J, Jung YS, Musilek K, Soukup O, Jun D, Pohanka M, Musilova L, Karasova J, Novotny L, Hrabnova M: **Reactivation of human brain homogenate cholinesterases inhibited by Tabun using newly developed oximes K117 and K127.** *Basic Clin Pharmacol Toxicol* 2009, **105**:207–210.
156. Melchers BP, Philippens IH, Wolthuis OL: **Efficacy of HI-6 and HLo-7 in preventing incapacitation following nerve agent poisoning.** *Pharmacol Biochem Behav* 1994, **49**:781–788.
157. Cassel G, Karlsson L, Waara L, Ang KW, Goransson-Nyberg A: **Pharmacokinetics and effects of HI 6 in blood and brain of soman-intoxicated rats: a microdialysis study.** *Eur J Pharmacol* 1997, **332**:43–52.

158. Clement JG, Hansen AS, Boulet CA: **Efficacy of HLO-7 and pyrimidoxime as antidotes of nerve agent poisoning in mice.** *Arch Toxicol* 1992, **66**:216–219.
159. Shih TM, Guarisco JA, Myers TM, Kan RK, McDonough JH: **The oxime pro-2-PAM provides minimal protection against the CNS effects of the nerve agents sarin, cyclosarin, and VX in guinea pigs.** *Toxicol Mech Methods* 2011, **21**:53–62.
160. Howland MA: **Pyridoxine.** In *Goldfrank's toxicologic emergencies*. ninthth edition. Edited by Nelson L, Lewin N, Howland MA, Hoffman R, Goldfrank L, Flomenbaum NE. New York: McGroHill; 2011.
161. Holstege CP, Dobmeier SG, Bechtel LK: **Critical care toxicology.** *Emerg Med Clin Nor Am* 2008, **26**:715–739.
162. Greaves I, Hunt P: **Chapter 5: Chemical agents.** In *Responding to terrorism*. Edited by Imre R, Mooney TB, Clarke B. Edinburgh: Churchill Livingstone; 2010:233–344.
163. Mongan PD, Winkley J, Joseph HMI: **Nerve and chemical agents.** In *Hospital preparation for bioterror.* Burlington: Academic Press; 2006:73–88.
164. Kayouka M, Houz e P, Debray M, Baud FJ: **Acute renal failure enhances the antidotal activity of pralidoxime towards paraoxon-induced respiratory toxicity.** *Toxicol Lett* 2009, **189**:48–56.
165. Kayouka M, Houz e P, Ris ede P, Debray M, Baud FJ: **Acute renal failure alters the kinetics of pralidoxime in rats.** *Toxicol Lett* 2009, **184**:61–66.
166. Kassa J: **Effect of diazepam on the effectiveness of antidote therapy in eliminating the acute lethal effects of soman in mice.** *Cas Lek Cesk* 2001, **140**:497–499.
167. Shih TM, Koviak TA, Capacio BR: **Anticonvulsants for poisoning by the organophosphorus compound soman: pharmacological mechanisms.** *Neurosci Biobehav Rev* 1991, **15**:349–362.
168. Shih TM, Rowland TC, McDonough JH: **Anticonvulsants for nerve agent-induced seizures: The influence of the therapeutic dose of atropine.** *J Pharmacol Exp Ther* 2007, **320**:154–161.
169. Joosen MJ, van der Schans MJ, van Helden HP: **Percutaneous exposure to the nerve agent VX: Efficacy of combined atropine, obidoxime and diazepam treatment.** *Chem Biol Interact* 2010, **188**:255–263.
170. McDonough JH: **Midazolam: An Improved Anticonvulsant Treatment for Nerve Agent-Induced Seizures.** 2002.
171. McDonough JH Jr, McMonagle J, Copeland T, Zoefel D, Shih TM: **Comparative evaluation of benzodiazepines for control of soman-induced seizures.** *Arch Toxicol* 1999, **73**:473–478.
172. Myhrer T, Enger S, Aas P: **Anticonvulsant efficacy of drugs with cholinergic and/or glutamatergic antagonism microinfused into area tempestas of rats exposed to soman.** *Neurochem Res* 2008, **33**:348–354.
173. Lallement G, Baubichon D, Clarencon D, Galonnier M, Peoc'h M, Carpentier P: **Review of the value of gacyclidine (GK-11) as adjuvant medication to conventional treatments of organophosphate poisoning: primate experiments mimicking various scenarios of military or terrorist attack by soman.** *Neurotoxicology* 1999, **20**:675–684.
174. Smith JS, Fulop ZL, Levinsohn SA, Darrell RS, Stein DG: **Effects of the novel NMDA receptor antagonist gacyclidine on recovery from medial frontal cortex contusion injury in rats.** *Neural Plast* 2000, **7**:73–91.
175. Hirbec H, Gavrira M, Vignon J: **Gacyclidine: a new neuroprotective agent acting at the N-methyl-D-aspartate receptor.** *CNS Drug Rev* 2001, **7**:172–198.
176. Figueiredo TH, Qashu F, Apland JP, Aroniadou-Anderjaska V, Souza AP, Braga MF: **The GluK1 (GluR5) Kainate/[alpha]-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor antagonist LY293558 reduces soman-induced seizures and neuropathology.** *J Pharmacol Exp Ther* 2011, **336**:303–312.
177. Dorandeu F, Carpentier P, Baubichon D, Four E, Bernabe D, Burckhart MF, Lallement G: **Efficacy of the ketamine-atropine combination in the delayed treatment of soman-induced status epilepticus.** *Brain Res* 2005, **1051**:164–175.
178. Dhote F, Carpentier P, Barbier L, Peinnequin A, Baille V, Pernot F, Testylier G, Beauc C, Foquin A, Dorandeu F: **Combinations of ketamine and atropine are neuroprotective and reduce neuroinflammation after a toxic status epilepticus in mice.** *Toxicol Appl Pharmacol* 2012, **259**:195–209.
179. Zhi QX, Yi FH, Xi CT: **Huperzine A ameliorates the spatial working memory impairments induced by AF64A.** *Neurorep* 1995, **6**:2221–2224.
180. Zhang JM, Hu GY: **Huperzine A, a nootropic alkaloid, inhibits N-methyl-D-aspartate-induced current in rat dissociated hippocampal neurons.** *Neurosci* 2001, **105**:663–669.
181. Zangara A: **The psychopharmacology of huperzine A: an alkaloid with cognitive enhancing and neuroprotective properties of interest in the treatment of Alzheimer's disease.** *Pharmacol Biochem Behav* 2003, **75**:675–686.
182. Coleman BR, Ratcliffe RH, Oguntayo SA, Shi X, Doctor BP, Gordon RK, Nambiar MP: **[+]-Huperzine A treatment protects against N-methyl-D-aspartate-induced seizure/status epilepticus in rats.** *Chem Biol Interact* 2008, **175**:387–395.
183. Grunwald J, Raveh L, Doctor BP, Ashani Y: **Huperzine A as a pretreatment candidate drug against nerve agent toxicity.** *Life Sci* 1994, **54**:991–997.
184. Garcia GE, Vernon A, Moorad-Doctor D, Ratcliffe RH: **(-)-Huperzine A, replacement for pyridostigmine bromide as nerve agent pretreatment, measured in Guinea Pig plasma by a new ultrahigh-pressure liquid chromatography (UHPLC)-MS method.** *FASEBJ* 2009, **23**. Suppl.
185. Nurulain SN: **Different approaches to acute organophosphorus poison treatment.** *J Pak Med Assoc* 2012, **62**:712–717.
186. Antonijivic B, Stefanovic D, Milovanovic ZA, Stojiljkovic MP, Bokonjic D, Dukic M: **Standard antidotes along with sodium bicarbonate in organophosphate poisoning.** In *Proceedings of chemical and biological medical treatment symposium (CBMTS): Spiez, Switzerland.* 2002. April 28 - May 3.
187. Koller M, Becker C, Thiermann H, Worek F: **GC-MS and LC-MS analysis of nerve agents in body fluids: intra-laboratory verification test using spiked plasma and urine samples.** *J Chromatogr B Analyt Technol Biomed Life Sci* 2010, **878**:1226–1233.
188. Cole JB, Stellpflug SJ, Gross EA, Smith SW: **Wide complex tachycardia in a pediatric diphenhydramine overdose treated with sodium bicarbonate.** *Pediatr Emerg Care* 2011, **27**:1175–1177.
189. Valero A, Golan D: **Accidental organic phosphorus poisoning: the use of propranolol to counteract vagolytic cardiac effects of atropine.** *Isr J Med Sci* 1967, **3**:582–584.
190. Monhart V, Fusek J, Brndiar M, Tlustakova M: **Use of hemoperfusion in experimental intoxication with nerve agents.** *Artif Organs* 1994, **18**:770–772.
191. Yokoyama K, Ogura Y, Kishimoto M, Hinochita F, Hara S, Yamada A, Mimura N, Seki A, Sakai O: **Blood purification for severe sarin poisoning after the Tokyo subway attack.** *JAMA* 1995, **274**:379.
192. Rothschild L, Bern S, Oswald S, Weinberg G: **Intravenous lipid emulsion in clinical toxicology.** *Scand J Trauma Resusc Emerg Med* 2010, **18**:51.
193. Zhou Y, Zhan C, Li Y, Zhong Q, Pan H, Yang G: **Intravenous lipid emulsions combine extracorporeal blood purification: a novel therapeutic strategy for severe organophosphate poisoning.** *Med Hypotheses* 2010, **74**:309–311.
194. Bania TC, Chu J, Stolbach A: **The Effect of Intralipid on organophosphate toxicity in mice.** *Acad Emerg Med* 2005, **12**(Suppl. 1):7–12.
195. Pajoumand A, Shadnia S, Rezaie A, Abdi M, Abdollahi M: **Benefits of magnesium sulfate in the management of acute human poisoning by organophosphorus insecticides.** *Hum Exp Toxicol* 2004, **23**:565–569.
196. Fuchs-Buder T, Tassonyi E: **Magnesium sulphate enhances residual neuromuscular block induced by vecuronium.** *Br J Anaesth* 1996, **76**:565–566.
197. Eddleston M, Buckley NA, Eyer P, Dawson AH: **Management of acute organophosphorus pesticide poisoning.** *Lancet* 2008, **371**:597–607.
198. Okayama H, Aikawa T, Okayama M, Sasaki H, Mue S, Takishima T: **Bronchodilating effect of intravenous magnesium sulfate in bronchial asthma.** *JAMA* 1987, **257**:1076–1078.
199. Ross MC, Broomfield CA, Cerasoli DM, Doctor BP, Lenz DE, Maxwell DM, Saxena A: **Chapter 7: Nerve agent bioscavenger: Development of a new approach to protect against organophosphorus exposure.** In *Textbooks of military medicine, medical aspects of chemical warfare.* Edited by Lenhart MK, Tuorinsky SD. Washington, DC: The Office of the Surgeon General at TMM Publications; 2008:243–259.
200. Pazooki S, Solhi H, Vishteh HR, Shadnia S, Beigi MJ: **Effectiveness of fresh frozen plasma as supplementary treatment in organophosphate poisoning.** *Med J Malaysia* 2011, **66**:342–345.
201. Pichamuthu K, Jerobin J, Nair A, John G, Kamalesh J, Thomas K, Jose A, Fleming JJ, Zachariah A, David SS, et al: **Bioscavenger therapy for organophosphate poisoning - an open-labeled pilot randomized trial comparing fresh frozen plasma or albumin with saline in acute organophosphate poisoning in humans.** *Clin Toxicol (Phila)* 2010, **48**:813–819.

202. Desire B, Saint-Andre S: **Interaction of soman with beta-cyclodextrin.** *Fundam Appl Toxicol* 1986, **7**:646–657.
203. Desire B, Saint-Andre S: **Inactivation of sarin and soman by cyclodextrins in vitro.** *Experientia* 1987, **43**:395–397.
204. Wille T, Tenberken O, Reiter G, Muller S, Le Provost R, Lafont O, Estour F, Thiermann H, Worek F: **Detoxification of nerve agents by a substituted beta-cyclodextrin: application of a modified biological assay.** *Toxicology* 2009, **265**:96–100.
205. Collombet JM, Four E, Burckhart MF, Masqueliez C, Bernabe D, Baubichon D, Herodin F, Lallement G: **Effect of cytokine treatment on the neurogenesis process in the brain of soman-poisoned mice.** *Toxicology* 2005, **210**:9–23.
206. Collombet JM, Beracochea D, Liscia P, Pierard C, Lallement G, Filliat P: **Long-term effects of cytokine treatment on cognitive behavioral recovery and neuronal regeneration in soman-poisoned mice.** *Behav Brain Res* 2011, **221**:261–270.
207. Gaydess A, Duysen E, Li Y, Gilman V, Kabanov A, Lockridge O, Bronich T: **Visualization of exogenous delivery of nanoformulated butyrylcholinesterase to the central nervous system.** *Chem Biol Interact* 2010, **187**:295–298.
208. Aracava Y, Pereira EF, Akkerman M, Adler M, Albuquerque EX: **Effectiveness of donepezil, rivastigmine, and (+/-)huperzine A in counteracting the acute toxicity of organophosphorus nerve agents: comparison with galantamine.** *J Pharmacol Exp Ther* 2009, **331**:1014–1024.
209. Teran-Maclver M, Larson K: **Implication of chemical biological terrorist event for children and pregnant women.** *MCN* 2008, **33**:224–232.
210. Pediatric Expert Advisory Panel Participants: **Atropine use in children after nerve gas exposure.** In *Pediatric Expert Advisory Panel (PEAP) Addressing terrorism, Disaster and public health emergency.* New York: Columbia University Maliman School of Public Health, National Center for Disaster Preparedness; 2004:1–8.
211. Bajgar J, Fusek J, Kassa J, Kuca K, Jun D: **Chemical aspects of pharmacological prophylaxis against nerve agent poisoning.** *Curr Med Chem* 2009, **16**:2977–2986.
212. Weinbroum AA, Rudick V, Paret G, Kluger Y, Ben Abraham R: **Anaesthesia and critical care considerations in nerve agent warfare trauma casualties.** *Resuscitation* 2000, **47**:113–123.
213. Sogorb MA, Vilanova E, Carrera V: **Future applications of phosphotriesterases in the prophylaxis and treatment of organophosphorus insecticide and nerve agent poisonings.** *Toxicol Lett* 2004, **151**:219–233.
214. Wolfe AD, Rush RS, Doctor BP, Koplovitz I, Jones D: **Acetylcholinesterase prophylaxis against organophosphate toxicity.** *Fundam Appl Toxicol* 1987, **9**:266–270.
215. Broomfield CA, Lockridge O, Millard CB: **Protein engineering of a human enzyme that hydrolyzes V and G nerve agents: design, construction and characterization.** *Chem Biol Interact* 1999, **119–120**:413–418.
216. Bajgar J, Kuca K, Fusek J, Jun D, Bartosova L: **Cholinesterase reactivators as prophylactics against nerve agents.** *Curr Bioact Compd* 2010, **6**:2–8.
217. Bajgar J, Fusek J, Sevelova L, Kassa J: **Original transdermal prophylactic antidote against nerve agents-TRANSANT.** In *Chemical and biological medical treatment symposium; Spiez, Switzerland.* 2004:14. 25–30 April.

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