DOI: 10.1002/rmb2.12594

REVIEW

Reproductive Medicine and Biology

WILEY

Sperm morphology: Evaluating its clinical relevance in contemporary fertility practice

Daniel L. Pelzman 💿 | Jay I. Sandlow

Department of Urology, Medical College of Wisconsin, Milwaukee, Wisconsin, USA

Correspondence

Daniel L. Pelzman, 8701 Watertown Plank Rd., Milwaukee, WI 53226, USA. Email: dpelzman@mcw.edu

Abstract

Background: The World Health Organization (WHO) recommends reporting sperm morphology in a standard semen analysis. However, the clinical utility and prognostic value of morphology is often debated.

Methods: We reviewed and summarized studies that assessed both the benefits and limitations of sperm morphology in the context of natural fertility, assisted reproductive technologies, and recurrent pregnancy loss. We additionally describe possible environmental and anatomical etiologies of teratozoospermia.

Results: Sperm morphology evaluation has continuously evolved since the release of the first WHO manual in 1980. Initially, several large studies reported significant inverse associations between fertility outcomes and teratozoospermia. Most recent studies, however, fail to show an association between sperm morphology and natural or assisted fertility outcomes.

Conclusion: Sperm morphology analysis may have limited diagnostic and prognostic value. Providers should be aware of these limitations when counseling or managing infertile patients.

KEYWORDS

andrology, assisted reproductive technology, sperm physiology

1 | BACKGROUND

Infertility affects approximately 15% of couples, and a male factor is a contributor in about 50% of these couples.¹⁻³ The evaluation of male factor infertility initially consists of two or more semen analyses which are used to guide potential medical and/or surgical therapies.⁴ Morphologic differences in sperm were first identified by van Leeuwenhoek in 1677⁵ but specific morphologic abnormalities and their potential for subfertility were not described until the 1950s.^{6,7} In the 1980s, a series of papers by Kruger et al. found significantly diminished oocyte fertilization rates when sperm morphology (using the "strict" criteria) dropped below 14%,^{8,9} calling more attention to the role of sperm morphology in male fertility. While examination of sperm morphology has been recommended since the 1st edition of the World Health Organization (WHO) manual released in 1980, the criteria for defining spermatozoa as "abnormal" has drastically changed over the past 40 years. Consequently, the most recent 6th edition manual contains the most detailed focus on the systematic assessment of sperm morphology to date.^{10,11} Despite several iterations of the WHO criteria and widespread adoption of morphology as an integral component of a semen analysis, there has been a lack of unanimous support for this test among clinicians due, in part, to its poor analytical reliability and inconclusive prognostic value.^{12,13}

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

© 2024 The Author(s). Reproductive Medicine and Biology published by John Wiley & Sons Australia, Ltd on behalf of Japan Society for Reproductive Medicine.

Reproductive Medicine and Biology

With numerous conflicting studies in the scientific literature, it is critical that clinicians are aware of the limitations and advantages of sperm morphology assessment. The purpose of this review is to describe the factors that impact sperm morphology, report both natural and assisted fertility outcomes in the context of abnormal morphology, and outline future considerations that may improve the usefulness of morphology in the clinical setting.

2 | METHODS

HEY

We conducted a literature search using both PubMed/MEDLINE and Google Scholar databases. The search strategy included relevant key terms and phrases, including "sperm morphology", "male infertility", "natural pregnancy", "spontaneous pregnancy", "assisted reproductive technology", "intrauterine insemination", "in vitro fertilization". Several Boolean searches with various combinations of these terms was performed. Articles were included if they were written in English and if titles or abstracts appeared relevant to this review.

3 | KEY FINDINGS

3.1 | Factors impacting sperm morphology

Sperm morphology criteria have evolved since the introduction of the first WHO manual in 1980. In the 1st and 2nd editions of the WHO handbook, sperm labeled as "morphologically abnormal" needed to have at least one obvious, well-defined abnormality as defined by Macleod and Gold.^{7,12,14} In these manuals, the cutoff for normal forms was 50–80%. The 3rd edition (1992) introduced the Kruger (Tygerberg) strict criteria, which characterized sperm with borderline abnormalities as "morphologically abnormal." Still, in this edition the reference value for normal forms was empirically reported as >30%. The 4th edition (1999) also used the strict criteria, although no precise reference value was reported; rather, a note suggested that a value <15% may be associated with lower IVF rates. The 5th and 6th edition (2010 and 2021, respectively) precisely defined and standardized reporting of morphologic abnormalities, while also decreasing the reference value to 4%.

The 6th edition handbook contains several notable recommendations and criteria. First, assessments of sperm morphology should be performed by trained personnel familiar with all criteria used to designate spermatozoa as abnormal. Frequent internal and external quality assessments should be utilized to minimize variability in results. Importantly, a major change in the 6th edition is an increased emphasis on characterizing specific defects in each region of the sperm-head, neck/midpiece, tail, and cytoplasmrather than grouping all defects into a single "abnormal" category. (Table 1) The head should be evaluated for size, shape, and contour; the midpiece for shape, width, length, and alignment; the tail for width and smoothness; and the cytoplasm for size relative to the head.¹¹ Some examples of common morphologic abnormalities are illustrated in Figure 1. After completing the morphologic assessment, the percentage of abnormal forms are tabulated and reported.

3.1.1 | Environmental factors

Sperm morphology may be negatively affected by toxins or environmental/workplace exposures, although most studies examining the magnitude of this impact are retrospective and of lower quality.

Location	Normal appearance	Abnormal	TABLE 1 WHO 6th edition guidelines: classification of sperm morphology.
Head	 Smooth, regular contour Oval Acrosomal region 40%–70% of total area No large vacuoles, no more than two small vacuoles 	 Acrosome <40% or >70% of head area Length-to-width ratio less than 1.5 or larger than 2 Amorphous, asymmetrical, or non-oval Double heads Vacuoles >1/5th head area 	
Midpiece	 Slender About the same length as sperm head Major axis of midpiece aligned with major axis of head 	Irregular shapeIrregular thicknessAsymmetrical, angled, or bent	
Tail	 Uniform caliber ~10 times length of head No sharp angulations 	 Sharply angulated Hairpin bends or coiled Short Irregular width Multiple tails 	
Cytoplasmic residue	 Acceptable for cytoplasmic droplets less than 1/3 normal head size 	 Residual cytoplasm >1/3 normal head size 	

(f)

VILEY

(A) Head defects (b) (c) (d) (e) (a) Vacuolated Tapered Pyriform Round Amorphous Small acrosomal area No Small acrosome (B) Neck and midpiece defects (C) Tail defects (D) Excess residual cytoplasm (i) (1) (q) (h) (j) (k) (m) (n) Thin Bent neck Asymmetrical Thick Short Bent Coiled > one third insertion head

FIGURE 1 WHO 6th edition illustrations of abnormal sperm morphology. Adapted from WHO laboratory manual for the examination and processing of human semen, sixth edition. Geneva: World Health Organization, 2021. License: CC BY-NC-SA 3.0 IGO.

In a meta-analysis of 20 studies with 5865 patients examining the effects of smoking on semen parameters, smokers had a -1.37% difference in normal forms, suggesting a negative association between cigarette smoking and sperm morphology.¹⁵ However, the mean difference for the subgroup analyzed with WHO 1999 criteria was -1.88% (i.e., negative effect on morphology) and with WHO 2010 criteria was 1.36% (i.e., positive effect on morphology). The authors were unable to draw a definitive conclusion due to the confounding factor of semen analysis method. Few studies have addressed the effects of cannabis on sperm morphology. Three larger studies, each with over 200 subjects, had conflicting results with one study demonstrating a negative effect and two showing no association.¹⁶⁻¹⁸ Ultimately, a meta-analysis of these three studies found no association between cannabis usage and teratozoospermia.¹⁹ A more recent prospective single-center study found that cannabis users were two times more likely to have teratozoospermia (strict criteria, normal forms <4%).²⁰ Alcohol use was associated with poor morphology in a meta-analysis of 11 studies and appeared to have a dose-dependent effect, with daily alcohol users having a lower percentage of normal sperm compared with occasional or never users.²¹

Several other environmental exposures have been studied for their effects on sperm morphology. Occupational pesticide exposure showed inadequate evidence of toxicity toward sperm morphology, although decreases in DNA integrity and sperm motility were observed.²² Exposure to air pollution was significantly associated with teratozoospermia in addition to increased rates of DNA fragmentation and asthenozoospermia.²³ Lastly, other groups have studied the effects of cell phone usage on sperm morphology (and other semen parameters). Radiation and heat generated by cell phones, particularly those kept in front pockets, may worsen semen quality, but results thus far have been conflicting.^{24,25}

3.1.2 Anatomic and health-related factors

Many hereditary and acquired illnesses may interfere with normal sperm development, resulting in teratozoospermia. A recent systematic review and meta-analysis of several prospective studies and 1357 patients assessed the outcomes of varicocele repair versus observation in men with any-grade varicocele.²⁶ In the

Reproductive Medicine and Biology

treatment arm, sperm morphology improved by a mean difference in 6.1% after repair, although results were inconsistent among the included studies. Viral and bacterial infections may also induce morphologic changes, both through direct testicular involvement and febrile events (via disruption of testicular thermoregulation). In 1951, Macleod studied three medical students with pneumonia or chickenpox and found reductions in sperm concentration, motility, and morphology in the weeks following infection.²⁷ Other groups have reported similar findings following various febrile illnesses.²⁸ More recently, two small meta-analyses found a minimal, if any, impact of COVID-19 on sperm morphology.^{29,30} The study of the semen microbiome is still in its infancy, but some recent studies have suggested that certain bacteria such as *Ureaplasma urealyticum* or *Mycoplasma hominis* have a detrimental effect on morphology.^{31,32}

Because of the inherent complexities of spermatogenesis, it is challenging to accurately determine the degree to which these exposures affect sperm morphology. Future studies corroborating the existing literature will be needed to more definitively establish causal relationships for each of these factors.

3.2 | Effect of sperm morphology on natural fertilization outcomes

There is sparse data assessing the prognostic value of sperm morphology on natural conception. In the LIFE study of 501 couples who recently discontinued contraception, percent abnormal morphology by both strict and traditional criteria was associated with a small but statistically significant increase in time to pregnancy.³³ However, after controlling for other semen parameters such as sperm count or concentration, this association was not retained, suggesting that sperm morphology is not an independent predictor of fecundity. In a retrospective analysis of 24 patients with 0% normal forms, 29% of patients were able to conceive without assisted reproductive technologies (ART) compared with 56% of controls.³⁴ In addition, all men with 0% normal forms who conceived naturally went on to have another child also via natural conception. The authors concluded that morphology alone should not be used to predict fertilization, pregnancy, or live birth potential.

3.3 | Effect of sperm morphology on assisted reproductive technology outcomes

The association between ART outcomes and sperm morphology have been rigorously studied, although evolution of sperm morphology assessment over time and advances in ART have confounded direct comparison between studies of various eras. Sperm morphology has been considered in both the context of intrauterine insemination (IUI) and in vitro fertilization (IVF) with or without intracytoplasmic sperm injection (ICSI).

3.3.1 | Intrauterine insemination

While it is commonly accepted that total motile sperm counts $>10 \times 10^6$ correlate positively with IUI success, ³⁵ similar conclusions cannot be drawn for sperm morphology. In theory, morphologically abnormal sperm may result in poor capacity to bind to the zona pellucida, ineffective or absent acrosome reactions, or impaired intracellular calcium regulation.³⁶ We identified 21 studies from 1995 to 2023 which examined the relationship between sperm morphology and IUI outcomes, including clinical pregnancy rates and live birth rates.

Earlier studies from the mid-late 1990s and early 2000s were highly variable in both their assessments of morphology and outcomes. While few studies reported no association between teratozoospermia and IUI outcomes,³⁷⁻³⁹ most studies from this time period demonstrated a significant decrease in IUI success when teratozoospermia was present.⁴⁰⁻⁴⁶ In studies that showed an association, the clinical pregnancy rates for couples with normal morphology were three to 40 times those of couples with poor morphology. However, it should be noted that the definition of abnormal morphology was inconsistent and ranged from <4% to <14% normal forms. Given the significant decrease in success, some authors advocated forgoing IUI in favor of IVF +/- ICSI if morphology <4% was seen.^{43,44}

More recent studies, particularly those which came after the release of the 5th Edition WHO manual in 2010, have largely been consistent in finding no association between sperm morphology and IUI outcomes. Of the 10 studies we identified which were published after 2005, eight found no association⁴⁷⁻⁵⁴ while two reported differences in outcomes for couples with abnormal morphology.^{55,56} In a study of 984 IUI procedures, Patel et al. additionally examined rates of birth abnormalities or spontaneous abortions and found no relationship to teratozoospermia.⁵²

Kohn et al. conducted a large systematic review and metaanalysis in 2018 comparing results from 20 observational studies and 21018 IUI cycles.⁵⁷ No significant difference in ultrasoundverified pregnancy rates was found when using either 4% or 1% normal forms threshold. Pregnancy rates per cycle were 14.2% versus 12.1% in the \geq 4% and <4% morphology groups and 14.0% versus 13.9% in the \geq 1% and <1% morphology groups, respectively. Interestingly, after stratifying by WHO manual era (3rd edition, pre-1999; 4th edition, 2000–2010; 5th edition, 2011–2018), a significant difference in pregnancy rate was observed for studies in the 3rd edition era (13.1% vs. 7.3%) but not for later eras.

In conclusion, an analysis of the entire body of literature assessing the effects of morphology on IUI outcomes is difficult due to the varied results from many studies over the last 30 years. It is impossible to determine whether the recent shift is purely due to changes in our assessment of sperm morphology or improvement in IUI techniques and/or patient selection. Ultimately, the most recent data, including a large meta-analysis, do not show any correlation between teratozoospermia and pregnancy rates after IUI.

WILEY

3.3.2 | In vitro fertilization

The prognostic value of sperm morphology for IVF±ICSI outcomes has also evolved over the past 4 decades. The seminal study by Kruger et al. from 1986 reported decreased fertilization success for patients with <14% versus >14% normal forms, and a follow-up study reported lower fertilization rates for patients with <4% versus 4-14% normal forms.^{8,9} Subsequent work in 1994 by Ombelet et al. echoed these findings, albeit with different thresholds of 5% and 9%.⁵⁸ In this study, no couple with "severe" teratozoospermia (<5% normal forms) achieved a successful pregnancy, while fertilization rates were similar between control and "good prognosis" teratozoospermia patients (5%-8%). However, pregnancy rate per transfer, baby take-home rate, pregnancy rate per cycle, and pregnancy rate per embryo were all significantly decreased in men with normal forms <9% compared with control. A large, structured review of 18 studies written in 1998 echoed these findings.⁵⁹ In this review analyzing more than 25000 cycles. fertilization rates were 59.3% versus 77.6% for men with ≤4% and >4% normal forms, respectively. When using a 14% threshold, fertilization rates were 72.7% versus 83.6% for ≤14% and >14% normal forms, respectively.

Starting in the mid-1990s, the results of these landmark papers were challenged as newer methodologies for morphology analysis began to increase in popularity. Nagy et al.⁶⁰ reported in 1995 that neither total sperm count, sperm motility, nor sperm morphology were predictive of ICSI outcome. Similarly, in a series of 354 consecutive ICSI cycles, Svalander et al.⁶¹ found no difference in fertilization, pregnancy, or implantation rates in men with morphology >14%, 4%-14%, and <4%. Even in men with complete teratozoospermia (i.e., 0% normal forms), pregnancy rates of 39% per cycle have been reported, leading the authors to conclude that use of donor sperm should not be recommended as the sole fertility option in this population.⁶² Similar findings were reported in a small series of 75 fresh TESE IVF+ICSI cycles, of which 17 cycles used no morphologically normal sperm. Of these, a 70% fertilization rate and 65% pregnancy rate was observed, similar to rates seen in the normal morphology group.63

Overall, there is no clear consensus for whether sperm morphology is a valuable prognostic indicator for IVF \pm ICSI cycle success. Like the literature regarding IUI success, studies exploring the relationship of morphology and IVF are plagued by inconsistent results and continuously evolving assessments of morphology over the past 30 years. However, most recent studies have robustly and reproducibly shown that using morphologically abnormal sperm leads to similar pregnancy outcomes in couples undergoing IVF.

3.4 | Effect of sperm morphology on recurrent pregnancy loss

Few studies examine the role of sperm morphology in couples with recurrent pregnancy loss (defined as either ≥2 or ≥3 recurrent

abortions before the 20th week of gestation, depending on society guidelines). Much of this literature has focused on genetic abnormalities within sperm (i.e. DNA fragmentation or chromosomal aneuploidies) as a primary outcome; the importance of these abnormalities remains poorly understood and is beyond the scope of this review. However, many of these studies reported morphologic assessment in their results or as a secondary outcome even though they were not designed to analyze morphology directly.

The causal relationship between DFI and teratozoospermia is unclear and, thus, it is important to understand that DFI may be a confounding factor if we look at morphology in isolation.^{64,65} Sperm with high DNA fragmentation may have normal morphology and sperm with abnormal morphology may have low rates of DNA fragmentation. A review and meta-analysis of 15 studies which examined DFI and RPL found that eight of the 15 studies reported lower morphology in men with RPL compared to control, but these results were not expounded upon.⁶⁶ Three recent studies reported significant increases in morphologic abnormalities for men with RPL versus controls,^{67–69} although none of these used the 5th or 6th edition WHO morphology criteria. Moderately large studies by Busnelli et al.^{70,71} and Eisenberg et al. demonstrated no difference in sperm morphology between RPL couples and control despite significant differences in DFI.

3.5 | Known morphologic abnormalities that affect fertility

Most studies that investigate sperm morphology tend to report abnormal forms as one all-encompassing variable (usually a percentage), with few specifying the predominant location or character of the abnormality. For many abnormal forms, the cause—whether genetic environmental, anatomic, etc—is unknown. However, it is well established that some genetic conditions produce characteristic morphologic defects and, in turn, require specific counseling and management.⁷²

First described in 1971, globozoospermia is a rare condition associated with round-headed spermatozoa which lack an acrosome. It typically occurs in families and leads to infertility due to inability of the spermatozoa to bind or penetrate the zona pellucida.⁷³ For this reason, patients with globozoospermia should be counseled about the very low likelihood of success with natural pregnancy, IUI, or IVF without ICSI. Accordingly, ICSI with or without artificial oocyte activation should be recommended as a treatment option for these patients.

Macrozoospermia, also known as large-headed multiflagellar spermatozoa, causes sperm with irregular-appearing, oversized heads and often multiple flagella.^{72,74} In most cases, this is due to a deletion within the Aurora kinase C (*AURKC*) gene. Interestingly, patients with this mutation have a 100% (or near 100%) chance of having only aneuploid sperm present, resulting in no chance of a successful term pregnancy. In these cases, IVF+ICSI should not be attempted, as even successful fertilization would produce an aneuploid embryo. 6 of 8

Reproductive Medicine and Biology

Patients with macrozoospermia who do not have the *AURKC* mutation can produce offspring, although pregnancies should be closely followed for aneuploidy. Consequently, genetic testing should be performed in all patients with macrozoospermia in order to appropriately counsel patients about reproductive options.⁷⁴

These two examples illustrate the importance of associating specific morphologies to respective pathologies within the male reproductive tract to assist with counseling and ART patient selection. Focusing solely on the percentage of abnormal sperm (and not the abnormalities themselves) may limit our ability to optimally manage patients with these genetic conditions.

4 | CONCLUSIONS

II FV

The value of sperm morphology assessment has often been debated since the release of the WHO manual's 1st edition in 1980. The role of morphology in clinical practice remains largely controversial, even despite 4 decades of study. Unfortunately, due to advancements in ART techniques and changes in morphology criteria, it is nearly impossible to directly compare studies from different eras. Even in contemporary studies, semen analyses are still routinely performed by humans, not machines, and consequently are subject to interand intra-laboratory variability. In addition, most modern studies have not associated teratozoospermia to inferior pregnancy rates or outcomes.

In conclusion, based on the most recent literature which includes several large meta-analyses, it seems that morphology may have a limited impact on fertility potential than previously stated. Clinicians, researchers, and laboratory staff should be aware of these limitations, particularly as they relate to management and counseling.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

ORCID

Daniel L. Pelzman 🔟 https://orcid.org/0000-0003-1275-1461

REFERENCES

- 1. de Kretser D. Male infertility. Lancet. 1997;349(9054):787-90.
- 2. Mosher WD, Pratt WF. Fecundity and infertility in the United States: incidence and trends. Fertil Steril. 1991;56(2):192–3.
- Thoma ME, McLain AC, Louis JF, King RB, Trumble AC, Sundaram R, et al. Prevalence of infertility in the United States as estimated by the current duration approach and a traditional constructed approach. Fertil Steril. 2013;99(5):1324–31.e1.
- Schlegel PN, Sigman M, Collura B, Jonge CJD, Eisenberg ML, Lamb DJ, et al. Diagnosis and treatment of infertility in men: AUA/ASRM guideline part I. Fertil Steril. 2021;115(1):54–61.
- Leeuwenhoek AV, Observationes D. Anthonii Lewenhoeck, de natis'e semine genitali animalculis. Phil Trans R Soc. 1677;12(142):1040–6.
- Page EW, Houlding F. The clinical interpretation of 1000 semen analyses among applicants for sterility studies. Fertil Steril. 1951;2(2):140–51.

- MacLeod J, Gold RZ. The male factor in fertility and infertility. IV. Sperm morphology in fertile and infertile marriage. Fertil Steril. 1951;2(5):394–414.
- Kruger TF, Acosta AA, Simmons KF, Swanson RJ, Matta JF, Oehninger S. Predictive value of abnormal sperm morphology in in vitro fertilization. Fertil Steril. 1988;49(1):112–7.
- Kruger TF, Menkveld R, Stander FSH, Lombard CJ, Van der Merwe JP, van Zyl JA, et al. Sperm morphologic features as a prognostic factor in in vitro fertilization. Fertil Steril. 1986;46(6):1118–23.
- Björndahl L. A paradigmatic shift in the care of male factor infertility: how can the recommendations for basic semen examination in the sixth edition of the WHO manual and the ISO 23162:2021 standard help? Reprod Biomed Online. 2022;45(4):731-6.
- 11. WHO. Laboratory manual for the examination and processing of human semen. 6th ed. Geneva: World Health Organization; 2021.
- Gatimel N, Moreau J, Parinaud J, Léandri RD. Sperm morphology: assessment, pathophysiology, clinical relevance, and state of the art in 2017. Andrology. 2017;5(5):845–62.
- Danis RB, Samplaski MK. Sperm morphology: history, challenges, and impact on natural and assisted fertility. Curr Urol Rep. 2019;20(8):43.
- Wang C, Mbizvo M, Festin MP, Björndahl L, Toskin I. Other editorial board members of the WHO laboratory manual for the examination and processing of human semen. Evolution of the WHO "semen" processing manual from the first (1980) to the sixth edition (2021). Fertil Steril. 2022;117(2):237–45.
- Sharma R, Harlev A, Agarwal A, Esteves SC. Cigarette smoking and semen quality: a new meta-analysis examining the effect of the 2010 World Health Organization Laboratory methods for the examination of human semen. Eur Urol. 2016;70(4):635–45.
- Gundersen TD, Jørgensen N, Andersson AM, Bang AK, Nordkap L, Skakkebæk NE, et al. Association between use of marijuana and male reproductive hormones and semen quality: a study among 1,215 healthy young men. Am J Epidemiol. 2015;182(6):473-81.
- Pacey AA, Povey AC, Clyma JA, McNamee R, Moore HD, Baillie H, et al. Modifiable and non-modifiable risk factors for poor sperm morphology. Hum Reprod. 2014;29(8):1629–36.
- Carroll K, Pottinger AM, Wynter S, DaCosta V. Marijuana use and its influence on sperm morphology and motility: identified risk for fertility among Jamaican men. Andrology. 2020;8(1):136–42.
- Belladelli F, Del Giudice F, Kasman A, Kold Jensen T, Jørgensen N, Salonia A, et al. The association between cannabis use and testicular function in men: a systematic review and meta-analysis. Andrology. 2021;9(2):503–10.
- Hehemann MC, Raheem OA, Rajanahally S, Holt S, Chen T, Fustok JN, et al. Evaluation of the impact of marijuana use on semen quality: a prospective analysis. Ther Adv Urol. 2021;13:17562872211032484.
- Ricci E, Al Beitawi S, Cipriani S, Candiani M, Chiaffarino F, Viganò P, et al. Semen quality and alcohol intake: a systematic review and meta-analysis. Reprod Biomed Online. 2017;34(1):38–47.
- Knapke ET, de Magalhaes DP, Dalvie MA, Mandrioli D, Perry MJ. Environmental and occupational pesticide exposure and human sperm parameters: a navigation guide review. Toxicology. 2022;465:153017.
- 23. Lafuente R, García-Blàquez N, Jacquemin B, Checa MA. Outdoor air pollution and sperm quality. Fertil Steril. 2016;106(4):880–96.
- Rahban R, Senn A, Nef S, Röösli M. Association between selfreported mobile phone use and the semen quality of young men. Fertil Steril. 2023;120(6):1181–92.
- Agarwal A, Deepinder F, Sharma RK, Ranga G, Li J. Effect of cell phone usage on semen analysis in men attending infertility clinic: an observational study. Fertil Steril. 2008;89(1):124–8.
- 26. Fallara G, Capogrosso P, Pozzi E, Belladelli F, Corsini C, Boeri L, et al. The effect of varicocele treatment on fertility in adults: a

systematic review and meta-analysis of published prospective trials. Eur Urol Focus. 2023;9(1):154-61.

- Macleod J. Effect of chickenpox and of pneumonia on semen quality. Fertil Steril. 1951;2(6):523–33.
- Carlsen E, Andersson AM, Petersen JH, Skakkebaek NE. History of febrile illness and variation in semen quality. Hum Reprod. 2003;18(10):2089–92.
- Edele Santos D, Colonetti T, Rodrigues Uggioni ML, Rech P, Marcelino Baptista M, Medeiros LR, et al. Effects of COVID-19 or vaccines for SARS-COV-2 on sperm parameters: a systematic review and meta-analysis. J Reprod Immunol. 2023;160:104140.
- Kloping YP, Hidayatullah F, Rahman ZA, Chung E, Hakim L. Male reproductive tract involvement and sperm parameters in SARS-CoV-2 patients: a systematic review and meta-analysis. World J Mens Health. 2023;41(3):538–57.
- Farahani L, Tharakan T, Yap T, Ramsay JW, Jayasena CN, Minhas S. The semen microbiome and its impact on sperm function and male fertility: a systematic review and meta-analysis. Andrology. 2021;9(1):115-44.
- Aghazarian A, Huf W, Klingler HC, Klatte T. The effect of seminal pathogens on standard semen parameters, sperm kinematics and seminal inflammatory markers. J Reprod Immunol. 2023;161:104183.
- Buck Louis GM, Sundaram R, Schisterman EF, Sweeney A, Lynch CD, Kim S, et al. Semen quality and time-to-pregnancy, the LIFE study. Fertil Steril. 2014;101(2):453–62.
- Kovac JR, Smith RP, Cajipe M, Lamb DJ, Lipshultz LI. Men with a complete absence of normal sperm morphology exhibit high rates of success without assisted reproduction. Asian J Androl. 2017;19(1):39-42.
- Muthigi A, Jahandideh S, Bishop LA, Naeemi FK, Shipley SK, O'Brien JE, et al. Clarifying the relationship between total motile sperm counts and intrauterine insemination pregnancy rates. Fertil Steril. 2021;115(6):1454–60.
- Grow D, Oehninger S. Strict criteria for the evaluation of human sperm morphology and its impact on assisted reproduction. Andrologia. 1995;27(6):325–33.
- Matorras R, Corcóstegui B, Perez C, Mandiola M, Mendoza R, Rodríguez-Escudero FJ. Sperm morphology analysis (strict criteria) in male infertility is not a prognostic factor in intrauterine insemination with husband's sperm. Fertil Steril. 1995;63(3):608-11.
- Karabinus DS, Gelety TJ. The impact of sperm morphology evaluated by strict criteria on intrauterine insemination success. Fertil Steril. 1997;67(3):536-41.
- Check ML, Bollendorf A, Check JH, Katsoff D. Reevaluation of the clinical importance of evaluating sperm morphology using strict criteria. Arch Androl. 2002;48(1):1–3.
- Toner JP, Mossad H, Grow DR, Morshedi M, Swanson RJ, Oehninger S. Value of sperm morphology assessed by strict criteria for prediction of the outcome of artificial (intrauterine) insemination. Andrologia. 1995;27(3):143–8.
- 41. Burr RW, Siegberg R, Flaherty SP, Wang XJ, Matthews CD. The influence of sperm morphology and the number of motile sperm inseminated on the outcome of intrauterine insemination combined with mild ovarian stimulation. Fertil Steril. 1996;65(1):127–32.
- 42. Lindheim SR, Barad DH, Zinger M, Witt B, Amin H, Cohen B, et al. Abnormal sperm morphology is highly predictive of pregnancy outcome during controlled ovarian hyperstimulation and intrauterine insemination. J Assist Reprod Genet. 1996;13(7):569-72.
- 43. Hauser R, Yogev L, Botchan A, Lessing JB, Paz G, Yavetz H. Intrauterine insemination in male factor subfertility: significance of sperm motility and morphology assessed by strict criteria. Andrologia. 2001;33(1):13–7.
- 44. Lee RKK, Hou JW, Ho HY, Hwu YM, Lin MH, Tsai YC, et al. Sperm morphology analysis using strict criteria as a prognostic factor in intrauterine insemination. Int J Androl. 2002;25(5):277–80.

- Spiessens C, Vanderschueren D, Meuleman C, D'Hooghe T. Isolated teratozoospermia and intrauterine insemination. Fertil Steril. 2003;80(5):1185-9.
- Grigoriou O, Pantos K, Makrakis E, Hassiakos D, Konidaris S, Creatsas G. Impact of isolated teratozoospermia on the outcome of intrauterine insemination. Fertil Steril. 2005;83(3):773–5.
- Sun Y, Li B, Fan LQ, Zhu WB, Chen XJ, Feng JH, et al. Does sperm morphology affect the outcome of intrauterine insemination in patients with normal sperm concentration and motility? Andrologia. 2012;44(5):299–304.
- Deveneau NE, Sinno O, Krause M, Eastwood D, Sandlow JI, Robb P, et al. Impact of sperm morphology on the likelihood of pregnancy after intrauterine insemination. Fertil Steril. 2014;102(6):1584–90. e2.
- Lockwood GM, Deveneau NE, Shridharani AN, Strawn EY, Sandlow JI. Isolated abnormal strict morphology is not a contraindication for intrauterine insemination. Andrology. 2015;3(6):1088–93.
- Erdem M, Erdem A, Mutlu MF, Ozisik S, Yildiz S, Guler I, et al. The impact of sperm morphology on the outcome of intrauterine insemination cycles with gonadotropins in unexplained and male subfertility. Eur J Obstet Gynecol Reprod Biol. 2016;197:120–4.
- Lemmens L, Kos S, Beijer C, Brinkman JW, van der Horst FAL, van den Hoven L, et al. Predictive value of sperm morphology and progressively motile sperm count for pregnancy outcomes in intrauterine insemination. Fertil Steril. 2016;105(6):1462–8.
- Patel P, Carrasquillo R, Madhusoodanan V, Dadoun S, Patel A, Smith N, et al. Impact of abnormal sperm morphology on live birth rates following intrauterine insemination. J Urol. 2019;202(4):801–5.
- Fuentes Ávila A, Blasco Sanz R, Cortés Alaguero C. Effect of sperm morphology in intrauterine insemination: analysis of 115 cycles and literature review. Obstet Gynecol Surv. 2021;76(3):170–4.
- Stanhiser J, Mersereau JE, Dock D, Boylan C, Caprell H, Coward RM, et al. Sperm morphology from the actual inseminated sample does not predict clinical pregnancy following intrauterine insemination. F S Rep. 2021;2(1):16–21.
- 55. Nikbakht R, Saharkhiz N. The influence of sperm morphology, total motile sperm count of semen and the number of motile sperm inseminated in sperm samples on the success of intrauterine insemination. Int J Fertil Steril. 2011;5(3):168–73.
- Ozcan P, Takmaz T. Identification of predictive factors for the probability of pregnancy following ovulation stimulation-intra-uterine insemination cycles in terms of female and male. J Obstet Gynaecol Res. 2021;47(3):893–9.
- 57. Kohn TP, Kohn JR, Ramasamy R. Effect of sperm morphology on pregnancy success via intrauterine insemination: a systematic review and meta-analysis. J Urol. 2018;199(3):812–22.
- Ombelet W, Fourie FL, Vandeput H, Bosmans E, Cox A, Janssen M, et al. Teratozoospermia and in-vitro fertilization: a randomized prospective study. Hum Reprod. 1994;9(8):1479–84.
- Coetzee K, Kruge T, Lombard C. Predictive value of normal sperm morphology: a structured literature review. Hum Reprod Update. 1998;4(1):73–82.
- Nagy ZP, Liu J, Joris H, Verheyen G, Tournaye H, Camus M, et al. The result of intracytoplasmic sperm injection is not related to any of the three basic sperm parameters. Hum Reprod. 1995;10(5):1123–9.
- Svalander P, Jakobsson AH, Forsberg AS, Bengtsson AC, Wikland M. The outcome of intracytoplasmic sperm injection is unrelated to 'strict criteria' sperm morphology. Hum Reprod. 1996;11(5):1019–22.
- McKenzie LJ, Kovanci E, Amato P, Cisneros P, Lamb D, Carson SA. Pregnancy outcome of in vitro fertilization/intracytoplasmic sperm injection with profound teratospermia. Fertil Steril. 2004;82(4):847–9.
- Schiff JD, Luna M, Barritt J, Duke M, Copperman A, Bar-Chama N. The morphology of extracted testicular sperm correlates with fertilization but not pregnancy rates. BJU Int. 2007;100(6):1326–9.

-WILEY Reproductive Medic

- 64. Bhattacharya SM. Association of various sperm parameters with unexplained repeated early pregnancy loss—which is most important? Int Urol Nephrol. 2008;40(2):391–5.
- Brahem S, Mehdi M, Landolsi H, Mougou S, Elghezal H, Saad A. Semen parameters and sperm DNA fragmentation as causes of recurrent pregnancy loss. Urology. 2011;78(4):792–6.
- McQueen DB, Zhang J, Robins JC. Sperm DNA fragmentation and recurrent pregnancy loss: a systematic review and meta-analysis. Fertil Steril. 2019;112(1):54–60.e3.
- 67. Esquerré-Lamare C, Walschaerts M, Chansel Debordeaux L, Moreau J, Bretelle F, Isus F, et al. Sperm aneuploidy and DNA fragmentation in unexplained recurrent pregnancy loss: a multicenter case-control study. Basic Clin Androl. 2018;28:4.
- Zidi-Jrah I, Hajlaoui A, Mougou-Zerelli S, Kammoun M, Meniaoui I, Sallem A, et al. Relationship between sperm aneuploidy, sperm DNA integrity, chromatin packaging, traditional semen parameters, and recurrent pregnancy loss. Fertil Steril. 2016;105(1):58–64.
- Zhang L, Wang L, Zhang X, Xu G, Zhang W, Wang K, et al. Sperm chromatin integrity may predict future fertility for unexplained recurrent spontaneous abortion patients. Int J Androl. 2012;35(5):752–7.
- Eisenberg ML, Sapra KJ, Kim SD, Chen Z, Buck Louis GM. Semen quality and pregnancy loss in a contemporary cohort of couples recruited before conception: data from the Longitudinal Investigation of Fertility and the Environment (LIFE) study. Fertil Steril. 2017;108(4):613–9.

- Busnelli A, Garolla A, Di Credico E, D'Ippolito S, Merola AM, Milardi D, et al. Sperm DNA fragmentation and idiopathic recurrent pregnancy loss: results from a multicenter case-control study. Andrology. 2023;11(8):1673–81.
- Ray PF, Toure A, Metzler-Guillemain C, Mitchell MJ, Arnoult C, Coutton C. Genetic abnormalities leading to qualitative defects of sperm morphology or function. Clin Genet. 2017;91(2):217–32.
- 73. Fesahat F, Henkel R, Agarwal A. Globozoospermia syndrome: an update. Andrologia. 2020;52(2):e13459.
- Carmignac V, Dupont JM, Fierro RC, Barberet J, Bruno C, Lieury N, et al. Diagnostic genetic screening for assisted reproductive technologies patients with macrozoospermia. Andrology. 2017;5(2):370-80.

How to cite this article: Pelzman DL, Sandlow JI. Sperm morphology: Evaluating its clinical relevance in contemporary fertility practice. Reprod Med Biol. 2024;23:e12594. https://doi.org/10.1002/rmb2.12594