Review Article

Sickling Cells, Cyclic Nucleotides, and Protein Kinases: The Pathophysiology of Urogenital Disorders in Sickle Cell Anemia

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Received 23 January 2012; Revised 16 April 2012; Accepted 22 April 2012

Academic Editor: Solomon F. Ofori-Acquah

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Sickle cell anemia is one of the best studied inherited diseases, and despite being caused by a single point mutation in the *HBB* gene, multiple pleiotropic effects of the abnormal hemoglobin S production range from vaso-occlusive crisis, stroke, and pulmonary hypertension to osteonecrosis and leg ulcers. Urogenital function is not spared, and although priapism is most frequently remembered, other related clinical manifestations have been described, such as nocturia, enuresis, increased frequence of lower urinary tract infections, urinary incontinence, hypogonadism, and testicular infarction. Studies on sickle cell vaso-occlusion and priapism using both *in vitro* and *in vivo* models have shed light on the pathogenesis of some of these events. The authors review what is known about the deleterious effects of sickling on the genitourinary tract and how the role of cyclic nucleotides signaling and protein kinases may help understand the pathophysiology underlying these manifestations and develop novel therapies in the setting of urogenital disorders in sickle cell disease.

1. Introduction

Sickle cell anemia (SCA) has been first described over a century ago [1] and has become one of the best studied inherited human diseases. Despite being caused by a single point mutation in the *HBB* gene, multiple pleiotropic effects of the abnormal hemoglobin S production range from vaso-occlusive crisis, stroke, and pulmonary hypertension to osteonecrosis and leg ulcers [2–4].

Genitourinary tract function is also affected in SCA, and although priapism is most frequently remembered, other related clinical manifestations have been described, such as nocturia, enuresis, increased frequency of lower urinary tract infections, urinary incontinence, hypogonadism, and testicular infarction. Sickle hemoglobin S (HbS) polymerizes when deoxygenated, resulting in a series of cellular alterations in red cell morphology and function that shorten the red cell life span and lead to vascular occlusion. Sickle cell disease (SCD) vaso-occlusion constitutes a complex multifactorial process characterized by oxidative stress and recurrent ischemia-reperfusion injury in a vicious circle contributing to reduced blood flow and results, eventually, in complete obstruction of the microcirculation and organic dysfunction [3–6]. The exact pathogenetic mechanisms that tie genitourinary complications to the fundamental event of HbS polymerization and hemolytic anemia in SCA have just about started to be unraveled.

This paper focuses on how previous, sometimes poorly explained, clinical observations of urogenital disorders in patients with SCD relate to more recent discoveries on the role of cyclic nucleotides and protein kinases in the pathophysiology of sickle vaso-occlusion.

2. Priapism

Priapism is defined as a prolonged and persistent penile erection, unassociated with sexual interest or stimulation, and is one of the complications associated with sickle cell anemia (SCA) since early in 1934 [7]. Priapism reaches a frequency of up to 45% in male patients with SCA, and the rate of resulting erectile dysfunction (ED) exceeds 30% [8–10].

Although this complication has been previously reviewed in depth in this journal [11], the main concepts behind its pathophysiology will be summarized here for better understanding of the mechanisms discussed throughout the paper, but readers are encouraged to read the previous review.

According to the American Urological Association Guidelines on the Management of Priapism, priapism can be subdivided into three categories: ischemic, stuttering, and nonischemic. Ischemic priapism (veno-occlusive, low flow) is a persistent erection marked by rigidity of the corpora cavernosa (CC) and little or no cavernous arterial inflow. In ischemic priapism, there are time-dependent changes in the corporal metabolic environment with progressive hypoxia, hypercarbia, and acidosis that typically generate penile pain. Penile sinusoids are regions prone to red blood cell sickling in SCD men because of blood stasis and slow flow rates, and ischemic priapism is thought to result from prolonged blockage of venous outflow by the vaso-occlusive process. Clinically, there is congestion and tenderness in the CC, sparing the glans and corpus spongiosum, usually with a prolonged course of over 3 hours, and frequently resulting in fibrosis and erectile dysfunction. Stuttering priapism (acute, intermittent, recurrent ischemic priapism) is characterized by a pattern of recurrence, but an increasing frequency or duration of stuttering episodes may herald a major ischemic priapism. Nonischemic priapism (arterial, high flow) is a persistent erection caused by unregulated cavernous arterial inflow. Typically, the corpora are tumescent but not rigid, the penis is not painful and is most frequently associated with trauma [12–16].

Conventional treatments are largely symptomatic, usually administered after the episode of priapism has already occurred, because the etiology and mechanisms involved in the development of priapism are poorly characterized [17, 18]. Preventive interventions have been proposed but, without a clear idea of the molecular mechanisms involved, they remain largely impractical to be applied in a regular basis in the clinic [17]. Due to the difficulty in exploring these mechanisms in patients, the use of animal models of priapism has become of utmost importance to decipher this devastating clinical challenge [19]. Animal models for priapism include dogs [20, 21], rabbits [22], rats [23–27], and mice [28–41].

Molecular biology and genetic engineering have been widely used in animal models to explore gene function in both human physiology and in the study of pathology of human priapism. Four major priapism animal models have been developed and have yielded greater knowledge on the intrinsic mechanisms underlying priapism: the intracorporal opiorphins gene transfer rat model [42–45], the endothelial nitric oxide synthase (eNOS) with or without neuronal NOS (nNOS) knock-out ($eNOS^{-/-} \pm nNOS^{-/-}$) mouse models [28, 29, 31-33], the adenosine deaminase knock-out (Ada^{-/-}) mouse model [35, 36, 40, 41] and the transgenic sickle cell Berkeley mouse model [30, 33, 34, 37-39]. However, the Berkeley mouse is the only well-accepted animal model that displays clinical manifestations similar to those seen in humans with severe forms of SCD, including priapism [30, 34].

Priapism is essentially a derangement of normal erection. Penile erection is a hemodynamic event that is regulated by smooth muscle relaxation/contraction of corpora cavernosa and associated arterioles during sexual stimulation. The penile flaccidity (detumescence state) is mainly maintained by tonic release of norepinephrine through the sympathetic innervations of vascular and cavernosal smooth muscle cells [46]. During penile erection (tumescence state), vascular smooth muscle relaxation decreases vascular resistance, thereby increasing blood flow through cavernous and helicine arteries and filling sinusoids, which are expanded due to the relaxation of smooth muscle cells in the CC [47]. This physiological relaxation of penile smooth muscle is mainly, although not solely, mediated by the neurotransmitter nitric oxide (NO) that is produced by enzymes called NO synthases (NOS). NOSs are subdivided into three isoforms, endothelial NOS (eNOS or NOS3), neural NOS (nNOS or NOS1), and inducible NOS (iNOS or NOS2) [48, 49]. In the penile smooth muscle, NO is released from both nitrergic nerves and the sinusoidal endothelium [46, 50-52]. NO stimulates the soluble guanylyl cyclase (sGC) in the cavernosal smooth muscle, triggering increased synthesis of cyclic GMP (cGMP) that provides the main signal for smooth muscle relaxation [53]. cGMP levels in the CC are regulated by the rate of synthesis determined by sGC and the rate of cGMP hydrolysis mediated by phosphodiesterase type 5 (PDE5) [54, 55]. It has been reported that plasma hemoglobin released by intravascularly hemolysed sickle erythrocytes consumes NO, reducing its bioavailability in the erectile tissue, skewing the normal balance of smooth muscle tone towards vasoconstriction [17, 56, 57]. Champion and collaborators [33] showed that the penile smooth muscle of SCD transgenic mice presents with dysregulated PDE5A expression activity. Moreover, these mice had spontaneous priapism, amplified CC relaxation response mediated by the NO-cGMP signaling pathway, and increased intracavernosal pressure in vivo [37, 38].

Recent evidence has shown that another signaling pathway that may also contribute to the pathophysiology of priapism in SCD involves adenosine regulation. Similarly to NO, adenosine is a potent vasodilator produced by adenine nucleotide degradation. Adenosine is predominantly generated by adenosine monophosphate (AMP) dephosphorylation catalyzed by intracellular 5'-nucleotidase. Hydrolysis of s-adenosyl-homocysteine also contributes to intracellular adenosine formation [58, 59]. Extracellular adenosine may be generated by both adenine nucleotide degradation and dephosphorylation by ectonucleotidases [60]. Adenosine is then catabolized by two enzymes: adenosine kinase (ADK), which phosphorylates adenosine to AMP and is an important regulator of intracellular adenosine levels; and adenosine deaminase (ADA), which catalyzes the irreversible conversion of adenosine to inosine [58].

Several physiological processes may be affected by extracellular adenosine and this is mediated by four different receptors, referred to as A_1 , A_{2A} , A_{2B} , and A_3 . All four subtypes are members of the G protein-coupled receptor (GPCR) superfamily. The activation of the A_1 and A_3 adenosine receptors inhibits adenylyl cyclase activity and also results in increased activity of phospholipase C, while activation of the A_{2A} and A_{2B} subtypes increases adenylyl cyclase activity [58, 61]. Adenosine-induced vasodilation is mediated by increasing intracellular cyclic adenosine monophosphate (cAMP) levels in vascular smooth muscle cells via A_2 receptor signaling [62, 63]. cAMP activates protein kinase A (PKA) resulting in decreased calciumcalmodulin-dependent MLC phosphorylation and enhanced smooth muscle relaxation [64]. Its role in penile erection has been investigated in studies showing that intracavernous injection of adenosine resulted in tumescence and penile erection [36, 61, 65]. In addition, adenosine induces NO synthesis in endothelial cells through A_2 receptor signaling, and adenosine-mediated CC relaxation is partially dependent on endothelium-derived NO [36, 66–70].

A priapic phenotype in Ada^{-/-} mice was identified and led to further investigation of the impact of adenosine in the pathophysiology of priapism [59]. Previous reports showed that high levels of adenosine caused prolonged corporal smooth muscle relaxation in vitro. However, this effect was quickly corrected by intraperitoneal injection of a high dose of polyethylene glycol-ADA (PEG-ADA), which effectively reduces adenosine levels systemically [36, 71]. Moreover, adenosine induced significant increases in cavernosal cAMP levels via A_{2B} receptor activation. This demonstrated that A_{2B} receptor signaling is required for adenosine-mediated stimulation of cAMP production in CC smooth muscle cells [36, 71]. Mi and collaborators [36] have studied adenosine levels in the penis of sickle cell mice and have found a significant increase in adenosine levels, suggesting that overproduction of adenosine may contribute to priapic activity in SCD [71, 72]. Sickle cell mice submitted to PEG-ADA treatment suffered significant reduction of force and duration of relaxation when compared with untreated mice [71]. In addition, increased adenosine levels contributed to the development of penile fibrosis in Ada^{-/-} mice as well as in transgenic sickle cell mice [72]. These findings suggest a general contributory role of elevated adenosine in the pathophysiology of priapism associated with SCD.

Although the penile vascular endothelium and smooth muscle cells are sources of vasodilation factors such as NO and adenosine, there are vasoconstriction pathways important to the penile hemodynamics, such as the Rho-kinase (ROCK) pathway. The RhoA/ROCK signal transduction pathway has been shown to influence erectile function in vivo through an array of mechanisms, including vasoconstriction of the penile vasculature via smooth muscle contraction and regulation of eNOS [73-76]. This pathway is involved in the regulation of smooth muscle tone by modulating the sensitivity of contractile proteins to Ca²⁺ [77]. RhoA regulates smooth muscle contraction by cycling between a GDP-bound inactive form (coupled to a guanine dissociation inhibitor, RhoGDI) and a GTP-bound active form [78-80]. Upstream activation of heterotrimeric G proteins leads to the exchange of GDP for GTP, an event carried out by the guanine exchange factors (GEFs) p115RhoGEF [81], PDZ-RhoGEF [82], and LARG (Leukemia-associated RhoGEF) [83], which are able to transduce signals from G proteincoupled receptors to RhoA [84-86]. ROCK is activated

by RhoA and inhibits myosin phosphatase through the phosphorylation of its myosin-binding subunit, leading to an increase in Ca²⁺ sensitivity. The RhoA/ROCK Ca²⁺ sensitization pathway has been implicated in the regulation of penile smooth muscle contraction and tone both in humans and animals [77, 87]. ROCK exerts contractile effects in the penis by Ca²⁺-independent promotion of myosin light chain (MLC) kinase or the attenuation of MLC phosphatase activity and reduction in endothelial-derived NO production [88]. RhoA activation, ROCK2 protein expression, as well as total ROCK activity decline in penile of SCD transgenic mice, highlighting that the molecular mechanism of priapism in SCD is associated with decreased vasoconstrictor activity in the penis [39]. Therefore, should impaired RhoA/ROCKmediated vasoconstriction contribute to SCD-associated priapism, this pathway may become a novel therapeutic target in the management of this complication.

There has been no definite advance in the management of sickle cell-associated acute, severe priapism. Penile aspiration with or without saline intracavernosal injection and eventually performing surgical shunts remains mainstays of care, with no evident benefit of more common approaches, such as intravenous hydration, blood transfusions, and urinary alkalinization [89, 90]. Pharmacological interventions in such cases have been limited to intracavernosal use of sympathomimetic drugs, such as epinephrine, norepinephrine, and etilefrine, but there are anecdotal reports of acute use of PDE5 inhibitor sildenafil [91].

Nonetheless, most attempts to control SCD priapism have focused on its recurrent, stuttering form. Small case series of hormonal manipulation with diethylstilbestrol [92], gonadotropin-releasing hormone (GnRH) analogues [93], and finasteride [94] have been reported to successfully manage recurrent priapism. Increasing smooth muscle tone with oral α -agonist etilefrine has also yielded only anecdotal evidence of benefit [95]. Unfortunately, a prospective study comparing etilefrine and ephedrine failed to demonstrate superiority or equivalence of both drugs in preventing recurrent priapism due to poor compliance and low recruitment reducing statistical power, but some evidence was obtained reassuring safety of the use of such strategies, and possibly indicating a lower severity of priapism attacks among compliant patients [96]. This favors off-label use of pseudoephedrine at bedtime advocated by some experts [57, 90]. Hydroxyurea has also been effective in preventing priapism recurrence in SCD in a small number of cases [97, 98]. Based on current knowledge of NO-dependent pathways, the use of PDE5 inhibitors has been studied. One clinical trial testing tadalafil in SCD patients has been terminated, but no outcome data have yet been published (ClinicalTrials.gov NCT00538564), and one ongoing trial aims at the effect of sildenafil in the same setting (ClinicalTrials.gov NCT00940901). Despite these efforts, scientists have become less optimistic concerning the tolerability of this approach, ever since the premature termination of the sildenafil trial for pulmonary hypertension in SCD patients, in which subjects on PDE5 inhibitor were more likely to have severe pain crises requiring hospitalization [99]. Therefore, novel therapies for preventing and treating priapism in SCD

are still warranted if the incidence of impotence among these patients is expected to be reduced in the long term.

3. Infertility

Progress in the therapy of SCD, particularly the use of hydroxyurea, has considerably improved the prognosis of patients with SCD [100, 101], with their mean life expectancy reaching much over 40 years [102–104], rendering infertility an important issue. Nevertheless, long before hydroxyurea became a standard of care in SCD, seminal fluid parameters of SCD males had been reported to fall within the subfertile range due to decreased sperm concentration, total count, motility, and altered morphology [105–107], and a more recent study reported over 90% of patients had at least one abnormal sperm parameter [108].

Hydroxyurea (HU) has been reported to impair spermatogenesis, causing testicular atrophy, reversible decrease in sperm count, as well as abnormal sperm morphology and motility [108–114], and its current or previous use should be among the first probable causes to be considered in SCD patients complaining of infertility. Moreover, sperm abnormalities prior to HU have been attributed to variable effects of hypogonadism induced by SCD itself, and lack of appropriate testosterone production seems to be exacerbated by HU use in a mouse SCD model [115].

Considering that male fertility does not rely solely on the quality of the seminal fluid, other causes that may also render male patients with SCD prone to suffer from infertility include sexual problems, such as loss of libido, premature ejaculation, frequent priapism, and priapismrelated impotence [105–107, 116–121].

Finding a single main cause for male infertility in a particular SCD patient is highly unlikely and probably will involve some degree of endocrinological impairment. A broader understanding of how hypogonadism takes place in SCD is necessary to explain fertility problems and requires knowledge of the complexity of sex hormone production regulation.

4. Hypogonadism

The etiology of hypogonadism in SCD patients is multifactorial, as several mechanisms have been suggested to contribute to its occurrence, such as primary gonadal failure [117, 122, 123], associated with or caused by repeated testicular infarction [124], zinc deficiency [125, 126], and partial hypothalamic hypogonadism [127].

Physical and sexual development are affected in both male and female SCD patients, with onset of puberty (menarche) and appearance of secondary sexual characteristics (pubic and axillary hair and beard) being usually delayed. The delay is greater in homozygous SCA and S- β^0 -thalassemia than in SC disease and S- β^+ -thalassemia [128–130]. Moreover, studies in male patients with SCD reported reduction of ejaculate volume, spermatozoa count, motility, and abnormal sperm morphology [106, 116].

Biochemical analyses have demonstrated low levels of testosterone and dihydrotestosterone and variable levels of follicle-stimulating hormone (FSH) and luteinizing hormone (LH) in patients with SCD [105–107, 118, 119, 121, 131]. The comparison between patients and controls matched according to stage of development of secondary sexual characteristics showed higher levels of LH in sickle cell disease, favoring some role for hypergonadotropic hypogonadism.

Leydig cells of the testes and other steroidogenic tissues produce hormones by a multienzymatic process, in which free cholesterol from intracellular stores is transferred to the outer and then to the inner mitochondrial membrane. Leydig cells produce androgens under the control of LH or its placental counterpart human chorionic gonadotropin (hCG), as well as in response to numerous intratesticular factors [114, 132]. LH/hCG receptors belong to the sGC-coupled seventransmembrane-domain receptor family, whose activation leads to stimulation of adenylyl cyclase [133]. The resulting accumulation of intracellular cyclic adenosine monophosphate (cAMP) levels and the concomitant activation of the cAMP-dependent protein kinase (PKA) lead to the phosphorylation of numerous proteins, including the steroidogenic acute regulatory (StAR) protein [134, 135]. StAR localizes predominantly to steroid hormone-producing tissues and consists of a 37 kDa precursor containing an NH2-terminal mitochondrial targeting sequence and several isoelectric 30 kDa mature protein forms [136–138]. Steroid production in gonadal and adrenal cells requires both de novo synthesis and PKA-dependent phosphorylation of StAR-37 protein [139]. The newly synthesized StAR is functional and plays a critical role in the transfer of cholesterol from the outer to the inner mitochondrial membrane, whereas mitochondrial import and processing to 30 kDa StAR protein terminate this action [140-142].

HbS polymerization is mediated by upstream activation of adenosine receptor $A_{2B}R$ by hypoxia, and hemolysis of irreversibly sickled red blood cells increases adenosine bioavailability through conversion of ATP by ectonucleotidases CD39 and CD73, thus predisposing patients with SCD to sustained high levels of cAMP [143, 144]. From this point of view, steroidogenesis could be expected to be increased in these patients.

Although Leydig cell steroidogenesis is predominantly regulated by cAMP/PKA, other pathways also influence this process [145], including the NO-cGMP signaling pathway [146]. NO promotes a biphasic modulation in the androgen production, stimulatory at low concentrations, and inhibitory at high concentrations [49, 147, 148]. SCA causes NO depletion, and in low levels, NO stimulates Leydig cell steroidogenesis by activating sGC [48, 49, 149] and promotes the formation of low levels of cGMP, albeit enough to activate the cGMP-dependent protein kinase (PKG) and phosphorylate StAR [49, 150]. This signaling is controlled by phosphodiesterases (PDEs) [151] and active transport systems that export cyclic nucleotides (multidrugresistance proteins) from the cell [152]. In zona glomerulosa cells, activation of PKG II by cGMP regulates basal levels of aldosterone production and phosphorylation of StAR



FIGURE 1: Schematic pathophysiology of hypogonadism and testicular infarction in sickle cell disease. The dashed arrow represents the blocking effect of gonadal failure over cyclic nucleotide-stimulated androgen production.

protein [150], but whether there is a role for cGMP in the zona reticularis, where adrenal androgenesis takes place, is unknown.

Hypogonadism observed in patients with SCD with lower circulating testosterone and higher LH levels suggests that, at least in this setting, despite the reduced cGMP- and elevated cAMP-mediated stimuli on androgen production, gonadal failure with Leydig cell impairment predominates in sex hormone production dysfunction (Figure 1). This further highlights that primary hypogonadism is possibly largely underdiagnosed and elicits more studies on the pathogenesis of testicular infarction.

5. Testicular Infarction

Segmental testicular infarction is an infrequent cause of acute scrotum and is rarely reported, with fewer than 40 cases published at the time of this paper. Its etiology is not always well defined, and it may be, at first, clinically mistaken for a testicular tumour [153, 154]. Common causes for testicular infarction are torsion of the spermatic cord, incarcerated hernia, infection, trauma, and vasculitis [131]. The usual presentation is a painful testicular mass unresponsive to antibiotics [155]. This testicular disorder has been associated with epididymitis, hypersensitivity angiitis, intimal fibroplasia of the spermatic cord arteries, polycythemia, anticoagulant use, benign testicular tumors and, in the interest of this review, sickle cell trait and sickle cell disease [124, 131, 155–158].

Testicular infarction related to sickling has been very rarely reported with only five individual cases found retrospectively, three associated with sickle cell disease and two with sickle cell trait [124, 155–157, 159]. Holmes and Kane reported the first testicular infarction in a patient with SCD who presented with testicular swelling unresponsive to antibiotics. Physical examination revealed that a lesion suspicious for malignancy and ultrasonography demonstrated a hyperechoic mass with an anechoid rim and normal blood flow in the surrounding parenchyma. Radical orchiectomy revealed hemorrhagic infarction with sickle blood red cells. In another case report, SCA patient presented with acute scrotum and history of acute chest syndrome, splenic infarction, osteomyelitis, and hemolysis. Physical examination demonstrated an erythematous, tender, swollen testicle and ultrasound once again revealed normal echotexture and blood flow. Surgical exploration and pathological examination diagnosed segmental testicular infarction with vascular congestion and sickled red blood cells [124]. In the last testicular infarction case report in a patient with SCD presented with increased testicular volume, scrotal ultrasonography showed both echogenic and hypoechogenic regions and Doppler ultrasonography revealed vascular changes compatible with testicular infarction. Radical orchiectomy was performed 10 days after the initial presentation and microscopic evaluation showed necrotic seminiferous tubules devoid of nuclear debris, congestion, or acute inflammatory infiltrate, consistent with coagulative necrosis of ischemic origin [131].

Testicular blood flow is dependent on the internal spermatic, cremasteric, and deferential arteries. Obstruction of venous outflow may create venous thrombosis, testicular engorgement, and subsequent hemorrhagic infarction. In SCD, low oxygen tensions in erythrocytes lead to sickling cells that lose pliability in the microcirculation. Consequently, capillary flow becomes obstructed, worsening local tissue hypoxia, perpetuating the cycle of sickling, and promoting testicular infarction [124, 131, 157].

The cyclic nucleotides and protein kinases may play an important role in the pathophysiology of testicular infarction

in SCD. Enhanced hemolysis and oxidative stress contribute to a reduction in nitric oxide (NO) bioavailability due to NO scavenging by free hemoglobin and reactive oxygen species (ROS) generation [160, 161]. As mentioned before, testicular NO signaling pathway is involved in the regulation of Leydig cell steroidogenesis [48, 49, 147–149, 162–164] but may also influence testicular circulation. We suggest that the reduction of NO bioavailability and consequent reduction of GMPc levels and of activity of PKG may decrease the vasodilation process in the testes. Moreover, reduced NO levels in patients with sickle cell disease contribute to the development of thrombus formation in the vascular system and could further enhance local ischemia [165, 166]. Furthermore, the cGMP-dependent protein kinase signaling pathway would normally inhibit RhoA-induced Ca2+ sensitization, RhoA/ROCK signaling, and protein kinase C (PKC) activity that mediate contraction in vascular smooth muscle [167–171]. Thus, reduced NO levels may decrease cGMPdependent protein kinase activity and promote increasing RhoA-induced Ca²⁺ sensitization and PKC activity, favoring vasoconstriction in the testes. Therefore, tissue hypoxia, sickling of red blood cells, reduced levels of NO, possible thrombus formation, increased RhoA-induced Ca²⁺ sensitization, and PKC activity may all lead to capillary and venous flow obstruction promoting testicular infarction (Figure 1).

Although testicular infarction in SCD has been very rarely reported, it has been speculated that silent testicular infarctions are much more common but generally overlooked clinically. Testicular biopsy in patients is rarely performed and additional studies are necessary to establish the true incidence of testicular infarction in patients with SCD or even sickle cell trait.

6. Urinary Bladder Dysfunction

The urinary bladder has two important functions: urine storage and emptying. Urine storage occurs at low pressure, implying that the bladder relaxes during the filling phase. Disturbances of the storage function may result in lower urinary tract symptoms (LUTSs), such as urgency, increased frequency, and urge incontinence, the components of the hypoactive or overactive bladder syndromes [172, 173]. The passive phase of bladder filling allows an increase in volume at a low intravesical pressure. The bladder neck and urethra remain in a tonic state to prevent leakage, thus maintaining urinary continence. Bladder emptying is accompanied by a reversal of function in which detrusor smooth muscle (DSM) contraction predominates in the bladder body that is accompanied by a concomitant reduction in outlet resistance of the bladder neck and urethra [174–176]. The bladder filling and emptying are regulated by interactions of norepinephrine (sympathetic component released by hypogastric nerve stimulation), acetylcholine and ATP (parasympathetic components released by pelvic nerve stimulation) with activation of adrenergic, muscarinic, and purinergic receptors, respectively [175].

Urinary bladder dysfunction is rarely spontaneously reported by SCD patients to their caregivers. With increasing survival of these patients, physicians may expect that urinary complaints increase in association with classical urological disorders associated with advanced age, such as urinary stress incontinence in multiparous women and benign prostatic hyperplasia in men. Nonetheless, clinical observations of medical complaints involving the urinary bladder start as early as childhood, with enuresis, and continue onto adulthood with nocturia and urinary tract infections, to name a few, although frequently neglected.

Nocturia has long been attributed to constant increased urinary volumes in SCD. As part of the renal complications of sickling, renal medullary infarcts lead to decreased ability to concentrate urine, yielding higher daily urinary volumes [177], compensatory polydipsia, and eventually, the need for nocturnal bladder voiding.

For comparison, the effects of polyuria on bladder function have been better characterized in diabetic bladder dysfunction (DBD). Both SCD and diabetes mellitus cause increased urinary volume and, to some extent, the two diseases involve cellular damage by oxidative stress mediators; so data from previous studies on DBD may help shed some light on preliminary data on bladder function in SCD animal models by understanding a known model of bladder dysfunction.

It has been suggested that DBD comprehends so-called early and late phases of the disease, owing to cumulative effects of initial polyuria secondary to hyperglycemia, complicated by oxidative stress influence on the urothelium and nervous damage in the long term of the natural history of diabetes mellitus. In the early phase of DBD, the bladder is hyperactive, leading to LUTS comprised mainly by nocturia and urge incontinence. Later in the course of the disease, the detrusor smooth muscle becomes atonic, abnormally distended, and incontinence is mainly by overflow associated with a poor control of urethral sphincters, and voiding problems take over [178].

DSM physiology also involves cyclic nucleotides and activation of protein kinases. DSM contractions are a consequence of cholinergic-mediated contractions and decreased β -adrenoceptor-mediated relaxations [179]. DSM contains a heterogeneous population of muscarinic receptor subtypes [180, 181], with a predominance of the M2 subtype and a smaller population of M3 receptors. However, functional studies showed that M3 receptors are responsible for promotion of contraction in the DSM of several animal models [182-185] and in humans [186, 187]. Activation of M3 muscarinic receptors in the DSM promotes stimulation of phospholipase C, activates PKC, and increases formation of inositol trisphosphate (IP₃) and diacylglycerol (DAG) to release calcium from intracellular stores, leading to DSM contraction [87]. Moreover, activation of M2 receptors also induces a DSM contraction indirectly by inhibiting the production of cAMP, reducing PKA activity, and reversing the relaxation induced by β -adrenoceptors [179]. Hence, both mechanisms promote urinary bladder emptying.

There is evidence that the Ca²⁺-independent RhoA/ ROCK pathway is involved in the regulation of smooth muscle tone by altering the sensitivity of contractile proteins to Ca^{2+} [77]. This pathway has been shown to influence erectile function in vivo through an array of mechanisms, including phosphorylation of the myosin-binding subunit of MLC phosphatase, resulting in increased myosin phosphorylation. RhoA, a member of the Ras (Rat Sarcoma) low molecular weight of GTP-binding proteins, mediates agonist-induced activation of ROCK. The exchange of GDP for GTP on RhoA and translocation of RhoA from the cytosol to the membrane are markers of its activation and enable the downstream stimulation of various effectors such as ROCK, protein kinase N, phosphatidylinositol 3-kinase, and tyrosine phosphorylation [77]. The RhoA/ROCK Ca²⁺ sensitization pathway has been implicated in the regulation of bladder smooth muscle contraction and tone in humans and animals [77, 188-191]. Thus, alterations in the contraction or relaxation mechanisms of DSM during the filling and emptying phases may contribute to urinary bladder dysfunction. Patients with SCD have not been evaluated for bladder dysfunction in a systematic manner, but preliminary data have shown that Berkeley mice (homozygous SS) exhibit hypocontractile DSM ex vivo, due to a significant decrease of contractile responses to muscarinic agonist carbachol and electrical field stimulation [192]. This bladder dysfunction may contribute to the increased risk of urinary tract infections observed in SCD patients.

In an epidemiological study of 321 children with SCD, 7% had a documented urinary tract infection (UTI), onethird had recurrent infections, and two-thirds had had a febrile UTI [193]. As in normal children, there was a strong predominance of females, and gram-negative organisms, particularly Escherichia coli, were usually cultured. Most episodes of gram-negative septicemia in SCD are secondary to UTI [194]. Moreover, UTIs are more frequent during pregnancy in women with SCA or sickle cell trait [195-197]. The prevalence of UTI in women with SCA is nearly twofold that of unaffected black American women. This association appears to be directly related to HbS levels, since patients with sickle trait have an increased prevalence of bacteriuria, but to a lesser degree than those with SCA. More recently, a study detected that a group of SCD children and adolescents had more symptoms of overactive bladder than a control group [198]. This could be a first documentation of a clinically evident of an early phase of sickle cell bladder dysfunction, but whether there is a late, hypotonic bladder phase in older sickle cell adults remains to be demonstrated.

The presence of increased intracavernosal pressure associated with the amplified corpus cavernosum relaxation response (priapism) mediated by NO-cGMP signaling pathway, the lack of RhoA/ROCK-mediated vasoconstriction in sickle cell transgenic Berkeley mice, and the association of priapism with genitourinary infections and urinary retention further suggest the possibility that changes in the DSM reactivity may contribute to urogenital complications in SCD [36, 38–40, 192]. Despite advances in the understanding of urogenital disorders in the SCD, further studies should clarify the pathophysiological mechanisms that underlie genitourinary manifestations of SCD.

7. Conclusions

Urogenital disorders in SCD are the result of pleotropic effects of the production of the abnormal sickling hemoglobin S. While priapism still stands out as the most frequently encountered, current knowledge of the effects of cyclic nucleotide production and activation of protein kinases allows to suspect underdiagnosis of bladder dysfunction and hypogonadism secondary to testicular failure. Moreover, despite our growing understanding of these complications, adequate, efficacious, and well-tolerated treatments are still unavailable, and male patients continue to suffer from infertility and erectile dysfunction. Further work in, both clinical assessments and experimental studies in this field are promising and should help increase physicians' awareness of the importance of more accurate diagnoses, design improved therapeutic strategies, and eventually, achieve better quality of life for SCD patients.

Abbreviations

- ROS: Reactive oxygen species
- NO: Nitric oxide
- cAMP: Cyclic adenosine monophosphate
- PKA: Cyclic adenosine monophosphate-dependent protein kinase
- cGMP: Cyclic Guanosine monophosphate;
- PKG: Cyclic Guanosine monophosphate protein kinase;
- PKC: Protein kinase C.

References

- C. J. Herrick, "The evolution of intelligence and its organs," Science, vol. 31, no. 784, pp. 7–18, 1910.
- [2] M. H. Steinberg, "Management of sickle cell disease," *The New England Journal of Medicine*, vol. 340, no. 13, pp. 1021–1030, 1999.
- [3] G. J. Kato and M. T. Gladwin, "Evolution of novel smallmolecule therapeutics targeting sickle cell vasculopathy," *Journal of the American Medical Association*, vol. 300, no. 22, pp. 2638–2646, 2008.
- [4] N. Conran, C. F. Franco-Penteado, and F. F. Costa, "Newer aspects of the pathophysiology of sickle cell disease vasoocclusion," *Hemoglobin*, vol. 33, no. 1, pp. 1–16, 2009.
- [5] R. P. Hebbel, M. A. B. Boogaerts, J. W. Eaton, and M. H. Steinberg, "Erythrocyte adherence to endothelium in sicklecell anemia. A possible determinant of disease severity," *The New England Journal of Medicine*, vol. 302, no. 18, pp. 992– 995, 1980.
- [6] R. B. Francis Jr. and C. S. Johnson, "Vascular occlusion in sickle cell disease: current concepts and unanswered questions," *Blood*, vol. 77, no. 7, pp. 1405–1414, 1991.
- [7] L. W. Diggs and R. E. Ching, "Pathology of sickle cell anemia," *Southern Medical Journal*, vol. 27, pp. 839–845, 1934.
- [8] A. B. Adeyoju, A. B. K. Olujohungbe, J. Morris et al., "Priapism in sickle-cell disease; incidence, risk factors and complications—an international multicentre study," *BJU International*, vol. 90, no. 9, pp. 898–902, 2002.
- [9] V. G. Nolan, D. F. Wyszynski, L. A. Farrer, and M. H. Steinberg, "Hemolysis-associated priapism in sickle cell disease," *Blood*, vol. 106, no. 9, pp. 3264–3267, 2005.

- [10] T. J. Bivalacqua and A. L. Burnett, "Priapism: new concepts in the pathophysiology and new treatment strategies," *Current Urology Reports*, vol. 7, no. 6, pp. 497–502, 2006.
- [11] G. M. Crane and N. E. Bennett Jr., "Priapism in sickle cell anemia: emerging mechanistic understanding and better preventative strategies," *Anemia*, vol. 2011, Article ID 297364, 6 pages, 2011.
- [12] American Foundation for Urologic Disease, "Thought leader panel on evaluation and treatment of priapism. Report of the American Foundation for Urologic Disease (AFUD) thought leader panel for evaluation and treatment of priapism," *International Journal of Impotence Research*, vol. 15, supplement, pp. S39–S43, 2001.
- [13] F. Numan, M. Cantasdemir, M. Ozbayrak et al., "Posttraumatic nonischemic priapism treated with autologous blood clot embolization," *Journal of Sexual Medicine*, vol. 5, no. 1, pp. 173–179, 2008.
- [14] A. L. Burnett and T. J. Bivalacqua, "Glucose-6-phosphate dehydrogenase deficiency: an etiology for idiopathic priapism?" *Journal of Sexual Medicine*, vol. 5, no. 1, pp. 237–240, 2008.
- [15] D. S. Finley, "Glucose-6-phosphate dehydrogenase deficiency associated stuttering priapism: report of a case," *Journal of Sexual Medicine*, vol. 5, no. 12, pp. 2963–2966, 2008.
- [16] Y. C. Jin, S. C. Gam, J. H. Jung, J. S. Hyun, K. C. Chang, and J. S. Hyun, "Expression and activity of heme oxygenase-1 in artificially induced low-flow priapism in rat penile tissues," *Journal of Sexual Medicine*, vol. 5, no. 8, pp. 1876–1882, 2008.
- [17] A. L. Burnett, "Pathophysiology of priapism: dysregulatory erection physiology thesis," *Journal of Urology*, vol. 170, no. 1, pp. 26–34, 2003.
- [18] T. J. Bivalacqua, B. Musicki, O. Kutlu, and A. L. Burnett, "New insights into the pathophysiology of sickle cell diseaseassociated priapism," *Journal of Sexual Medicine*, vol. 9, pp. 79–87, 2011.
- [19] Q. Dong, S. Deng, R. Wang, and J. Yuan, "In vitro and in vivo animal models in priapism research," *Journal of Sexual Medicine*, vol. 8, no. 2, pp. 347–359, 2011.
- [20] K. K. Chen, J. Y. Chan, L. S. Chang, M. T. Chen, and S. H. Chan, "Intracavernous pressure as an experimental index in a rat model for the evaluation of penile erection," *Journal of Urology*, vol. 147, no. 4, pp. 1124–1128, 1992.
- [21] M. Ul-Hasan, A. I. El-Sakka, C. Lee, T. S. Yen, R. Dahiya, and T. F. Lue, "Expression of TGF-beta-1 mRNA and ultrastructural alterations in pharmacologically induced prolonged penile erection in a canine model," *The Journal of Urology*, vol. 160, no. 6, pp. 2263–2266, 1998.
- [22] R. Munarriz, K. Park, Y. H. Huang et al., "Reperfusion of ischemic corporal tissue: physiologic and biochemical changes in an animal model of ischemic priapism," *Urology*, vol. 62, no. 4, pp. 760–764, 2003.
- [23] Y. Evliyaoglu, L. Kayrin, and B. Kaya, "Effect of allopurinol on lipid peroxidation induced in corporeal tissue by venoocclusive priapism in a rat model," *British Journal of Urology*, vol. 80, no. 3, pp. 476–479, 1997.
- [24] Y. Evliyaoğlu, L. Kayrin, and B. Kaya, "Effect of pentoxifylline on veno-occlusive priapism-induced corporeal tissule lipid peroxidation in a rat model," *Urological Research*, vol. 25, no. 2, pp. 143–147, 1997.
- [25] O. Sanli, A. Armagan, E. Kandirali et al., "TGF-β1 neutralizing antibodies decrease the fibrotic effects of ischemic priapism," *International Journal of Impotence Research*, vol. 16, no. 6, pp. 492–497, 2004.

- [26] Y. C. Jin, S. C. Gam, J. H. Jung, J. S. Hyun, K. C. Chang, and J. S. Hyun, "Expression and activity of heme oxygenase-1 in artificially induced low-flow priapism in rat penile tissues," *Journal of Sexual Medicine*, vol. 5, no. 8, pp. 1876–1882, 2008.
- [27] N. Uluocak, D. AtIlgan, F. Erdemir et al., "An animal model of ischemic priapism and the effects of melatonin on antioxidant enzymes and oxidative injury parameters in rat penis," *International Urology and Nephrology*, vol. 42, no. 4, pp. 889–895, 2010.
- [28] P. L. Huang, T. M. Dawson, D. S. Bredt, S. H. Snyder, and M. C. Fishman, "Targeted disruption of the neuronal nitric oxide synthase gene," *Cell*, vol. 75, no. 7, pp. 1273–1286, 1993.
- [29] P. L. Huang, Z. Huang, H. Mashimo et al., "Hypertension in mice lacking the gene for endothelial nitric oxide synthase," *Nature*, vol. 377, no. 6546, pp. 239–242, 1995.
- [30] C. Pászty, C. M. Brion, E. Manci et al., "Transgenic knockout mice with exclusively human sickle hemoglobin and sickle cell disease," *Science*, vol. 278, no. 5339, pp. 876–878, 1997.
- [31] P. L. Huang, "Lessons learned from nitric oxide synthase knockout animals," *Seminars in Perinatology*, vol. 24, no. 1, pp. 87–90, 2000.
- [32] L. A. Barouch, R. W. Harrison, M. W. Skaf et al., "Nitric oxide regulates the heart by spatial confinement of nitric oxide synthase isoforms," *Nature*, vol. 416, no. 6878, pp. 337–340, 2002.
- [33] H. C. Champion, T. J. Bivalacqua, E. Takimoto, D. A. Kass, and A. L. Burnett, "Phosphodiesterase-5A dysregulation in penile erectile tissue is a mechanism of priapism," *Proceedings* of the National Academy of Sciences of the United States of America, vol. 102, no. 5, pp. 1661–1666, 2005.
- [34] Hsu, "Hemolysis in sickle cell mice causes pulmonary hypertension due to global impairment in nitric oxide bioavailability," *Blood*, vol. 109, no. 7, pp. 3088–3098, 2007.
- [35] J. H. Yuan, J. L. Chunn, T. J. Mi et al., "Adenosine deaminase knockout in mice induces priapism via A2b receptor," *Journal* of Urology, vol. 177, supplement, p. 227, 2007.
- [36] T. Mi, S. Abbasi, H. Zhang et al., "Excess adenosine in murine penile erectile tissues contributes to priapism via A2B adenosine receptor signaling," *Journal of Clinical Investigation*, vol. 118, no. 4, pp. 1491–1501, 2008.
- [37] T. J. Bivalacqua, B. Musicki, L. L. Hsu, M. T. Gladwin, A. L. Burnett, and H. C. Champion, "Establishment of a transgenic sickle-cell mouse model to study the pathophysiology of priapism," *Journal of Sexual Medicine*, vol. 6, no. 9, pp. 2494– 2504, 2009.
- [38] M. A. Claudino, C. F. Franco-penteado, M. A. F. Corat et al., "Increased cavernosal relaxations in sickle cell mice priapism are associated with alterations in the NO-cGMP signaling pathway," *Journal of Sexual Medicine*, vol. 6, no. 8, pp. 2187– 2196, 2009.
- [39] T. J. Bivalacqua, A. E. Ross, T. D. Strong et al., "Attenuated rhoA/rho-kinase signaling in penis of transgenic sickle cell mice," *Urology*, vol. 76, no. 2, pp. 510.e7–510.e12, 2010.
- [40] J. Wen, X. Jiang, Y. Dai et al., "Adenosine deaminase enzyme therapy prevents and reverses the heightened cavernosal relaxation in priapism," *Journal of Sexual Medicine*, vol. 7, no. 9, pp. 3011–3022, 2010.
- [41] J. Wen, X. Jiang, Y. Dai et al., "Increased adenosine contributes to penile fibrosis, a dangerous feature of priapism, via A2B adenosine receptor signaling," *The FASEB Journal*, vol. 24, no. 3, pp. 740–749, 2010.
- [42] Y. Tong, M. Tar, F. Davelman, G. Christ, A. Melman, and K. P. Davies, "Variable coding sequence protein A1 as a marker

for erectile dysfunction," *BJU International*, vol. 98, no. 2, pp. 396–401, 2006.

- [43] Y. Tong, M. Tar, V. Monrose, M. DiSanto, A. Melman, and K. P. Davies, "hSMR3A as a marker for patients with erectile dysfunction," *Journal of Urology*, vol. 178, no. 1, pp. 338–343, 2007.
- [44] Y. Tong, M. Tar, A. Melman, and K. Davies, "The opiorphin gene (ProL1) and its homologues function in erectile physiology," *BJU International*, vol. 102, no. 6, pp. 736–740, 2008.
- [45] N. D. Kanika, M. Tar, Y. Tong, D. S. R. Kuppam, A. Melman, and K. P. Davies, "The mechanism of opiorphininduced experimental priapism in rats involves activation of the polyamine synthetic pathway," *American Journal of Physiology*, vol. 297, no. 4, pp. C916–C927, 2009.
- [46] K. E. Andersson, "Pharmacology of penile erection," *Pharmacological Reviews*, vol. 53, no. 3, pp. 417–450, 2001.
- [47] P. V. Phatarpekar, J. Wen, and Y. Xia, "Role of adenosine signaling in penile erection and erectile disorders," *Journal of Sexual Medicine*, vol. 7, no. 11, pp. 3553–3564, 2010.
- [48] M. S. Davidoff, R. Middendorff, B. Mayer, J. DeVente, D. Koesling, and A. F. Holstein, "Nitric oxide/cGMP pathway components in the Leydig cells of the human testis," *Cell and Tissue Research*, vol. 287, no. 1, pp. 161–170, 1997.
- [49] S. A. Andric, M. M. Janjic, N. J. Stojkov, and T. S. Kostic, "Protein kinase G-mediated stimulation of basal Leydig cell steroidogenesis," *American Journal of Physiology*, vol. 293, no. 5, pp. E1399–E1408, 2007.
- [50] A. L. Burnett, C. J. Lowenstein, D. S. Bredt, T. S. K. Chang, and S. H. Snyder, "Nitric oxide: a physiologic mediator of penile erection," *Science*, vol. 257, no. 5068, pp. 401–403, 1992.
- [51] K. E. Andersson and G. Wagner, "Physiology of penile erection," *Physiological Reviews*, vol. 75, no. 1, pp. 191–236, 1995.
- [52] T. F. Lue, "Erectile dysfunction," The New England Journal of Medicine, vol. 342, pp. 1802–1813, 2000.
- [53] K. A. Lucas, G. M. Pitari, S. Kazerounian et al., "Guanylyl cyclases and signaling by cyclic GMP," *Pharmacological Reviews*, vol. 52, no. 3, pp. 375–414, 2000.
- [54] M. Boolell, M. J. Allen, S. A. Ballard et al., "Sildenafil: an orally active type 5 cyclic GMP-specific phosphodiesterase inhibitor for the treatment of penile erectile dysfunction," *International Journal of Impotence Research*, vol. 8, no. 2, pp. 47–52, 1996.
- [55] V. K. Gopal, S. H. Francis, and J. D. Corbin, "Allosteric sites of phosphodiesterase-5 (PDE5). A potential role in negative feedback regulation of cGMP signaling in corpus cavernosum," *European Journal of Biochemistry*, vol. 268, no. 11, pp. 3304–3312, 2001.
- [56] K. Ohene-Frempong and M. H. Steinberg, "Clinical aspects of sickle cell anemia in adults and children," in *Disorders* of Hemoglobin: Genetics, Pathophysiology and Clinical Management, M. H. Steinberg, B. G. Forget, D. R. Higgs, and R. L. Nagel, Eds., pp. 611–670, Cambridge University Press, Cambridge, UK, 2001.
- [57] Z. R. Rogers, "Priapism in sickle cell disease," *Hematology* Oncology Clinics of North America, vol. 19, pp. 917–928, 2005.
- [58] B. B. Fredholm, A. P. Ijzerman, K. A. Jacobson, K. N. Klotz, and J. Linden, "International Union of Pharmacology. XXV. Nomenclature and classification of adenosine receptors," *Pharmacological Reviews*, vol. 53, no. 4, pp. 527–552, 2001.
- [59] P. V. Phatarpekar, J. Wen, and Y. Xia, "Role of adenosine signaling in penile erection and erectile disorders," *Journal of Sexual Medicine*, vol. 7, no. 11, pp. 3553–3564, 2010.

- [60] S. P. Colgan, H. K. Eltzschig, T. Eckle, and L. F. Thompson, "Physiological roles for ecto-5'-nucleotidase (CD73)," *Purinergic Signalling*, vol. 2, no. 2, pp. 351–360, 2006.
- [61] R. C. Tostes, F. R. C. Giachini, F. S. Carneiro, R. Leite, E. W. Inscho, and R. C. Webb, "Determination of adenosine effects and adenosine receptors in murine corpus cavernosum," *Journal of Pharmacology and Experimental Therapeutics*, vol. 322, no. 2, pp. 678–685, 2007.
- [62] R. A. Olsson and J. D. Pearson, "Cardiovascular purinoceptors," *Physiological Reviews*, vol. 70, no. 3, pp. 761–845, 1990.
- [63] A. M. Tager, P. LaCamera, B. S. Shea et al., "The lysophosphatidic acid receptor LPA1 links pulmonary fibrosis to lung injury by mediating fibroblast recruitment and vascular leak," *Nature Medicine*, vol. 14, no. 1, pp. 45–54, 2008.
- [64] C. S. Lin, G. Lin, and T. F. Lue, "Cyclic nucleotide signaling in cavernous smooth muscle," *Journal of Sexual Medicine*, vol. 2, no. 4, pp. 478–491, 2005.
- [65] D. Prieto, "Physiological regulation of penile arteries and veins," *International Journal of Impotence Research*, vol. 20, no. 1, pp. 17–29, 2008.
- [66] A. Vials and G. Burnstock, "A₂-purinoceptor-mediated relaxation in the guinea-pig coronary vasculature: a role for nitric oxide," *British Journal of Pharmacology*, vol. 109, no. 2, pp. 424–429, 1993.
- [67] L. Sobrevia, D. L. Yudilevich, and G. E. Mann, "Activation of A₂-purinoceptors by adenosine stimulates L-arginine transport (system y⁺) and nitric oxide synthesis in human fetal endothelial cells," *Journal of Physiology*, vol. 499, no. 1, pp. 135–140, 1997.
- [68] J. M. Li, R. A. Fenton, H. B. Wheeler et al., "Adenosine A_{2a} receptors increase arterial endothelial cell nritric oxide," *Journal of Surgical Research*, vol. 80, no. 2, pp. 357–364, 1998.
- [69] P. H. Chiang, S. N. Wu, E. M. Tsai et al., "Adenosine modulation of neurotransmission in penile erection," *British Journal of Clinical Pharmacology*, vol. 38, no. 4, pp. 357–362, 1994.
- [70] M. Faria, T. Magalhães-Cardoso, J. M. Lafuente-De-Carvalho, and P. Correia-De-Sá, "Corpus cavernosum from men with vasculogenic impotence is partially resistant to adenosine relaxation due to endothelial A_{2B} receptor dysfunction," *Journal of Pharmacology and Experimental Therapeutics*, vol. 319, no. 1, pp. 405–413, 2006.
- [71] Y. Dai, Y. Zhang, P. Phatarpekar et al., "Adenosine signaling, priapism and novel therapies," *Journal of Sexual Medicine*, vol. 6, no. 3, supplement, pp. 292–301, 2009.
- [72] J. Wen, X. Jiang, Y. Dai et al., "Increased adenosine contributes to penile fibrosis, a dangerous feature of priapism, via A_{2B} adenosine receptor signaling," *The FASEB Journal*, vol. 24, no. 3, pp. 740–749, 2010.
- [73] K. Chitaley, C. J. Wingard, R. Clinton Webb et al., "Antagonism of Rho-kinase stimulates rat penile erection via a nitric oxide-independent pathway," *Nature Medicine*, vol. 7, no. 1, pp. 119–122, 2001.
- [74] T. M. Mills, K. Chitaley, C. J. Wingard, R. W. Lewis, and R. C. Webb, "Effect of rho-kinase inhibition on vasoconstriction in the penile circulation," *Journal of Applied Physiology*, vol. 91, no. 3, pp. 1269–1273, 2001.
- [75] T. J. Bivalacqua, H. C. Champion, M. F. Usta et al., "RhoA/Rho-kinase suppresses endothelial nitric oxide synthase in the penis: a mechanism for diabetes-associated erectile dysfunction," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 24, pp. 9121–9126, 2004.

- [76] B. Musicki, A. E. Ross, H. C. Champion, A. L. Burnett, and T. J. Bivalacqua, "Posttranslational modification of constitutive nitric oxide synthase in the penis," *Journal of Andrology*, vol. 30, no. 4, pp. 352–362, 2009.
- [77] A. P. Somlyo and A. V. Somlyo, "Ca²⁺ sensitivity of smooth muscle and nonmuscle myosin II: modulated by G proteins, kinases, and myosin phosphatase," *Physiological Reviews*, vol. 83, no. 4, pp. 1325–1358, 2003.
- [78] N. Wettschureck and S. Offermanns, "Rho/Rho-kinase mediated signaling in physiology and pathophysiology," *Journal of Molecular Medicine*, vol. 80, no. 10, pp. 629–638, 2002.
- [79] K. Riento and A. J. Ridley, "Rocks: multifunctional kinases in cell behaviour," *Nature Reviews Molecular Cell Biology*, vol. 4, no. 6, pp. 446–456, 2003.
- [80] M. Bhattacharya, A. V. Babwah, and S. S. G. Ferguson, "Small GTP-binding protein-coupled receptors," *Biochemical Society Transactions*, vol. 32, no. 6, pp. 1040–1044, 2004.
- [81] M. J. Hart, S. Sharma, N. Elmasry et al., "Identification of a novel guanine nucleotide exchange factor for the Rho GTPase," *Journal of Biological Chemistry*, vol. 271, no. 41, pp. 25452–25458, 1996.
- [82] S. Fukuhara, C. Murga, M. Zohar, T. Igishi, and J. S. Gutkind, "A novel PDZ domain containing guanine nucleotide exchange factor links heterotrimeric G proteins to Rho," *Journal of Biological Chemistry*, vol. 274, no. 9, pp. 5868– 5879, 1999.
- [83] P. J. Kourlas, M. P. Strout, B. Becknell et al., "Identification of a gene at 11q23 encoding a guanine nucleotide exchange factor: evidence for its fusion with MLL in acute myeloid leukemia," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 5, pp. 2145–2150, 2000.
- [84] E. M. Ross and T. M. Wilkie, "GTPase-activating proteins for heterotrimeric G proteins: regulators of G Protein Signaling (RGS) and RGS-like proteins," *Annual Review of Biochemistry*, vol. 69, pp. 795–827, 2000.
- [85] S. Fukuharaa, H. Chikumi, and J. Silvio Gutkind, "RGScontaining RhoGEFs: the missing link between transforming G proteins and Rho?" *Oncogene*, vol. 20, no. 13, pp. 1661– 1668, 2001.
- [86] A. Schmidt and A. Hall, "Guanine nucleotide exchange factors for Rho GTPases: turning on the switch," *Genes and Development*, vol. 16, no. 13, pp. 1587–1609, 2002.
- [87] C. E. Teixeira, F. B. M. Priviero, and R. C. Webb, "Effects of 5cyclopropyl-2-[1-(2-fluoro-benzyl)-1H-pyrazolo[3,4-b]pyridine-3-yl]pyrimidin-4-ylamine (BAY 41-2272) on smooth muscle tone, soluble guanylyl cyclase activity, and NADPH oxidase activity/expression in corpus cavernosum from wild-type, neuronal, and endothelial nitric-oxide synthase null mice," *Journal of Pharmacology and Experimental Therapeutics*, vol. 322, no. 3, pp. 1093–1102, 2007.
- [88] A. V. Somlyo, "New roads leading to Ca²⁺ sensitization," *Circulation Research*, vol. 91, no. 2, pp. 83–84, 2002.
- [89] E. Mantadakis, D. H. Ewalt, J. D. Cavender, Z. R. Rogers, and G. R. Buchanan, "Outpatient penile aspiration and epinephrine irrigation for young patients with sickle cell anemia and prolonged priapism," *Blood*, vol. 95, no. 1, pp. 78–82, 2000.
- [90] G. J. Kato, "Priapism in sickle-cell disease: a hematologist's perspective," *The Journal of Sexual Medicine*, vol. 9, no. 1, pp. 70–78, 2012.
- [91] E. S. Bialecki and K. R. Bridges, "Sildenafil relieves priapism in patients with sickle cell disease," *American Journal of Medicine*, vol. 113, no. 3, p. 252, 2002.

- [92] G. R. Serjeant, K. de Ceulaer, and G. H. Maude, "Stilboestrol and stuttering priapism in homozygous sickle-cell disease," *The Lancet*, vol. 2, no. 8467, pp. 1274–1276, 1985.
- [93] L. A. Levine and S. P. Guss, "Gonadotropin-releasing hormone analogues in the treatment of sickle cell anemiaassociated priapism," *Journal of Urology*, vol. 150, no. 2, pp. 475–477, 1993.
- [94] D. Rachid-Filho, A. G. Cavalcanti, L. A. Favorito, W. S. Costa, and F. J. B. Sampaio, "Treatment of recurrent priapism in sickle cell anemia with finasteride: a new approach," *Urology*, vol. 74, no. 5, pp. 1054–1057, 2009.
- [95] I. Okpala, N. Westerdale, T. Jegede, and B. Cheung, "Etilefrine for the prevention of priapism in adult sickle cell disease," *British Journal of Haematology*, vol. 118, no. 3, pp. 918–921, 2002.
- [96] A. D. Seftel, "A prospective diary study of stuttering priapism in adolescents and young men with sickle cell anemia: report of an international randomized control trial; The priapism in sickle cell study (PISCES study)," *Journal of Urology*, vol. 185, no. 5, pp. 1837–1838, 2011.
- [97] S. T. O. Saad, C. Lajolo, S. Gilli et al., "Follow-up of sickle cell disease patients with priapism treated by hydroxyurea," *American Journal of Hematology*, vol. 77, no. 1, pp. 45–49, 2004.
- [98] A. Hassan, A. Jam'a, and I. A. Al Dabbous, "Hydroxyurea in the treatment of sickle cell associated priapism," *Journal of Urology*, vol. 159, no. 5, p. 1642, 1998.
- [99] R. F. Machado, R. J. Barst, N. A. Yovetich et al., "Hospitalization for pain in patients with sickle cell disease treated with sildenafil for elevated TRV and low exercise capacity," *Blood*, vol. 118, no. 4, pp. 855–864, 2011.
- [100] S. Charache, M. L. Terrin, R. D. Moore et al., "Effect of hydroxyurea on the frequency of painful crises in Sickle cell anemia," *The New England Journal of Medicine*, vol. 332, no. 20, pp. 1317–1322, 1995.
- [101] S. M. Bakanay, E. Dainer, B. Clair et al., "Mortality in sickle cell patients on hydroxyurea therapy," *Blood*, vol. 105, no. 2, pp. 545–547, 2005.
- [102] O. S. Platt, D. J. Brambilla, W. F. Rosse et al., "Mortality in sickle cell disease—life expectancy and risk factors for early death," *The New England Journal of Medicine*, vol. 330, no. 23, pp. 1639–1644, 1994.
- [103] D. R. Powars, L. S. Chan, A. Hiti, E. Ramicone, and C. Johnson, "Outcome of sickle cell anemia: a 4-decade observational study of 1056 patients," *Medicine*, vol. 84, no. 6, pp. 363–376, 2005.
- [104] C. D. Fitzhugh, N. Lauder, J. C. Jonassaint et al., "Cardiopulmonary complications leading to premature deaths in adult patients with sickle cell disease," *American Journal of Hematology*, vol. 85, no. 1, pp. 36–40, 2010.
- [105] C. R. D. Nahoum, E. A. Fontes, and F. R. Freire, "Semen analysis in sickle cell disease," *Andrologia*, vol. 12, no. 6, pp. 542–545, 1980.
- [106] D. N. Osegbe, O. Akinyanju, and E. O. Amaku, "Fertility in males with sickle cell disease," *The Lancet*, vol. 2, no. 8241, pp. 275–276, 1981.
- [107] V. O. Agbaraji, R. B. Scott, S. Leto, and L. W. Kingslow, "Fertility studies in sickle cell disease: semen analysis in adult male patients," *International Journal of Fertility*, vol. 33, no. 5, pp. 347–352, 1988.
- [108] I. Berthaut, G. Guignedoux, F. Kirsch-Noir et al., "Influence of sickle cell disease and treatment with hydroxyurea on sperm parameters and fertility of human males," *Haematologica*, vol. 93, no. 7, pp. 988–993, 2008.

- [109] C. C. Lu and M. L. Meistrich, "Cytotoxic effects of chemotherapeutic drugs on mouse testis cells," *Cancer Research*, vol. 39, no. 9, pp. 3575–3582, 1979.
- [110] G. Ficsor and L. C. Ginsberg, "The effect of hydroxyurea and mitomycin C on sperm motility in mice," *Mutation Research*, vol. 70, no. 3, pp. 383–387, 1980.
- [111] H. Singh and C. Taylor, "Effects of Thio-TEPA and hydroxyurea on sperm production in Lakeview hamsters," *Journal* of *Toxicology and Environmental Health*, vol. 8, no. 1-2, pp. 307–316, 1981.
- [112] D. P. Evenson and L. K. Jost, "Hydroxyurea exposure alters mouse testicular kinetics and sperm chromatin structure," *Cell Proliferation*, vol. 26, no. 2, pp. 147–159, 1993.
- [113] R. Wiger, J. K. Hongslo, D. P. Evenson, P. De Angelis, P. E. Schwarze, and J. A. Holme, "Effects of acetaminophen and hydroxyurea on spermatogenesis and sperm chromatin structure in laboratory mice," *Reproductive Toxicology*, vol. 9, no. 1, pp. 21–33, 1995.
- [114] J. M. Saez, "Leydig cells: endocrine, paracrine, and autocrine regulation," *Endocrine Reviews*, vol. 15, no. 5, pp. 574–626, 1994.
- [115] K. M. Jones, M. S. Niaz, C. M. Brooks et al., "Adverse effects of a clinically relevant dose of hydroxyurea used for the treatment of sickle cell disease on male fertility endpoints," *International Journal of Environmental Research and Public Health*, vol. 6, no. 3, pp. 1124–1144, 2009.
- [116] G. Friedman, R. Freeman, and R. Bookchin, "Testicular function in sickel cell disease," *Fertility and Sterility*, vol. 25, no. 12, pp. 1018–1021, 1974.
- [117] A. A. Abbasi, A. S. Prasad, and J. Ortega, "Gonadal function abnormalities in sickle cell anemia; studies in male adult patients," *Annals of Internal Medicine*, vol. 85, no. 5, pp. 601– 605, 1976.
- [118] O. Modebe and U. O. Ezeh, "Effect of age on testicular function in adult males with sickle cell anemia," *Fertility and Sterility*, vol. 63, no. 4, pp. 907–912, 1995.
- [119] O. A. Dada and E. U. Nduka, "Endocrine function and hemoglobinopathies: relation between the sickle cell gene and circulating plasma levels of testosterone, luteinizing hormone (LH) and follicle stimulating hormone (FSH) in adult males," *Clinica Chimica Acta*, vol. 105, no. 2, pp. 269– 273, 1980.
- [120] M. A. F. El-Hazmi, H. M. Bahakim, and I. Al-Fawaz, "Endocrine functions in sickle cell anaemia patients," *Journal* of Tropical Pediatrics, vol. 38, no. 6, pp. 307–313, 1992.
- [121] E. K. Abudu, S. A. Akanmu, O. O. Soriyan et al., "Serum testosterone levels of HbSS (sickle cell disease) male subjects in Lagos, Nigeria," *BMC Research Notes*, vol. 17, no. 4, p. 298, 2011.
- [122] D. N. Osegbe and O. O. Akinyanju, "Testicular dysfunction in men with sickle cell disease," *Postgraduate Medical Journal*, vol. 63, no. 736, pp. 95–98, 1987.
- [123] O. O. Abdulwaheed, A. A. Abdulrasaq, A. K. Sulaiman et al., "The hormonal assessment of the infertile male in Ilorin, Nigeria," *African Journal of Clinical Endocrinology & Metabolism*, vol. 3, pp. 62–64, 2002.
- [124] O. N. Gofrit, D. Rund, A. Shapiro, O. Pappo, E. H. Landau, and D. Pode, "Segmental testicular infarction due to sickle cell disease," *Journal of Urology*, vol. 160, no. 3, part 1, pp. 835–836, 1998.
- [125] A. S. Prasad, E. B. Schoomaker, and J. Ortega, "Zinc deficiency in sickle cell disease," *Clinical Chemistry*, vol. 21, no. 4, pp. 582–587, 1975.

- [126] A. S. Prasad and Z. T. Cossack, "Zinc supplementation and growth in sickle cell disease," *Annals of Internal Medicine*, vol. 100, no. 3, pp. 367–371, 1984.
- [127] C. S. Landefeld, M. Schambelan, S. L. Kaplan, and S. H. Embury, "Clomiphene-responsive hypogonadism in sickle cell anemia," *Annals of Internal Medicine*, vol. 99, no. 4, pp. 480–483, 1983.
- [128] C. T. Jiminez, R. B. Scott, W. L. Henry et al., "Studies in sickle cell anemia. XXVI. The effect of homozygous sickle cell disease on the onset of menarche, pregnancy, fertility, pubescent changes and body growth in Negro subjects," *American Journal Of Diseases Of Children*, vol. 111, pp. 497– 503, 1966.
- [129] O. S. Platt, W. Rosenstock, and M. A. Espeland, "Influence of sickle hemoglobinopathies on growth and development," *The New England Journal of Medicine*, vol. 311, no. 1, pp. 7– 12, 1984.
- [130] M. A. Zago, J. Kerbauy, H. M. Souza et al., "Growth and sexual maturation of Brazilian patients with sickle cell diseases," *Tropical and Geographical Medicine*, vol. 44, no. 4, pp. 317–321, 1992.
- [131] M. Li, J. Fogarty, K. D. Whitney, and P. Stone, "Repeated testicular infarction in a patient with sickle cell disease: a possible mechanism for testicular failure," *Urology*, vol. 62, no. 3, p. 551, 2003.
- [132] M. L. Dufau, "The luteinizing hormone receptor," Annual Review of Physiology, vol. 60, pp. 461–496, 1998.
- [133] M. Ascoli, F. Fanelli, and D. L. Segaloff, "The lutropin/choriogonadotropin receptor, a 2002 perspective," *Endocrine Reviews*, vol. 23, no. 2, pp. 141–174, 2002.
- [134] D. M. Stocco, "StAR protein and the regulation of steroid hormone biosynthesis," *Annual Review of Physiology*, vol. 63, pp. 193–213, 2001.
- [135] J. J. Tremblay, F. Hamel, and R. S. Viger, "Protein kinase A-dependent cooperation between GATA and CCAAT/enhancer-binding protein transcription factors regulates steroidogenic acute regulatory protein promoter activity," *Endocrinology*, vol. 143, no. 10, pp. 3935–3945, 2002.
- [136] L. F. Epstein and N. R. Orme-Johnson, "Acute action of luteinizing hormone on mouse Leydig cells: accumulation of mitochondrial phosphoproteins and stimulation of testosterone synthesis," *Molecular and Cellular Endocrinology*, vol. 81, no. 1–3, pp. 113–126, 1991.
- [137] L. F. Epstein and N. R. Orme-Johnson, "Regulation of steroid hormone biosynthesis: identification of precursors of a phosphoprotein targeted to the mitochondrion in stimulated rat adrenal cortex cells," *Journal of Biological Chemistry*, vol. 266, no. 29, pp. 19739–19745, 1991.
- [138] T. Seebacher, E. Beitz, H. Kumagami, K. Wild, J. P. Ruppersberg, and J. E. Schultz, "Expression of membrane-bound and cytosolic guanylyl cyclases in the rat inner ear," *Hearing Research*, vol. 127, no. 1-2, pp. 95–102, 1999.
- [139] F. Arakane, S. R. King, Y. Du et al., "Phosphorylation of steroidogenic acute regulatory protein (StAR) modulates its steroidogenic activity," *Journal of Biological Chemistry*, vol. 272, no. 51, pp. 32656–32662, 1997.
- [140] I. P. Artemenko, D. Zhao, D. B. Hales, K. H. Hales, and C. R. Jefcoate, "Mitochondrial processing of newly synthesized steroidogenic acute regulatory protein (StAR), but not total StAR, mediates cholesterol transfer to cytochrome P450 side chain cleavage enzyme in adrenal cells," *Journal of Biological Chemistry*, vol. 276, no. 49, pp. 46583–46596, 2001.

- [141] C. Jefcoate, "High-flux mitochondrial cholesterol trafficking, a specialized function of the adrenal cortex," *Journal of Clinical Investigation*, vol. 110, no. 7, pp. 881–890, 2002.
- [142] J. Liu, M. B. Rone, and V. Papadopoulos, "Protein-protein interactions mediate mitochondrial cholesterol transport and steroid biosynthesis," *Journal of Biological Chemistry*, vol. 281, no. 50, pp. 38879–38893, 2006.
- [143] H. K. Eltzschig, J. C. Ibla, G. T. Furuta et al., "Coordinated adenine nucleotide phosphohydrolysis and nucleoside signaling in posthypoxic endothelium: role of ectonucleotidases and adenosine A_{2B} receptors," *Journal of Experimental Medicine*, vol. 198, no. 5, pp. 783–796, 2003.
- [144] Y. Zhang, Y. Dai, J. Wen et al., "Detrimental effects of adenosine signaling in sickle cell disease," *Nature Medicine*, vol. 17, no. 1, pp. 79–86, 2011.
- [145] D. M. Stocco, X. Wang, Y. Jo, and P. R. Manna, "Multiple signaling pathways regulating steroidogenesis and steroidogenic acute regulatory protein expression: more complicated than we thought," *Molecular Endocrinology*, vol. 19, no. 11, pp. 2647–2659, 2005.
- [146] M. L. Khurana and K. N. Pandey, "Receptor-mediated stimulatory effect of atrial natriuretic factor, brain natriuretic peptide, and C- type natriuretic peptide on testosterone production in purified mouse Leydig cells: activation of cholesterol side- chain cleavage enzyme," *Endocrinology*, vol. 133, no. 5, pp. 2141–2149, 1993.
- [147] K. Del Punta, E. H. Charreau, and O. P. Pignataro, "Nitric oxide inhibits leydig cell steroidogenesis," *Endocrinology*, vol. 137, no. 12, pp. 5337–5343, 1996.
- [148] J. G. Drewett, R. L. Adams-Hays, B. Y. Ho, and D. J. Hegge, "Nitric oxide potently inhibits the rate-limiting enzymatic step in steroidogenesis," *Molecular and Cellular Endocrinology*, vol. 194, no. 1-2, pp. 39–45, 2002.
- [149] S. Valenti, C. M. Cuttica, L. Fazzuoli, G. Giordano, and M. Giusti, "Biphasic effect of nitric oxide on testosterone and cyclic GMP production by purified rat Leydig cells cultured in vitro," *International Journal of Andrology*, vol. 22, no. 5, pp. 336–341, 1999.
- [150] S. Gambaryan, E. Butt, K. Marcus et al., "cGMP-dependent protein kinase type II regulates basal level of aldosterone production by zona glomerulosa cells without increasing expression of the steroidogenic acute regulatory protein gene," *Journal of Biological Chemistry*, vol. 278, no. 32, pp. 29640–29648, 2003.
- [151] D. A. Kass, H. C. Champion, and J. A. Beavo, "Phosphodiesterase type 5: expanding roles in cardiovascular regulation," *Circulation Research*, vol. 101, no. 11, pp. 1084–1095, 2007.
- [152] S. A. Andric, T. S. Kostic, and S. S. Stojilkovic, "Contribution of multidrug resistance protein MRP5 in control of cyclic guanosine 5'-monophosphate intracellular signaling in anterior pituitary cells," *Endocrinology*, vol. 147, no. 7, pp. 3435– 3445, 2006.
- [153] G. C. Fernández-Pérez, F. M. Tardáguila, M. Velasco et al., "Radiologic findings of segmental testicular infarction," *American Journal of Roentgenology*, vol. 184, no. 5, pp. 1587– 1593, 2005.
- [154] S. Madaan, S. Joniau, K. Klockaerts et al., "Segmental testicular infarction: conservative management is feasible and safe," *European Urology*, vol. 53, no. 2, pp. 441–445, 2008.
- [155] D. P. Han, R. R. Dmochowski, M. H. Blasser, and J. R. Auman, "Segmental infarction of the testicle: atypical presentation of a testicular mass," *Journal of Urology*, vol. 151, no. 1, pp. 159– 160, 1994.

- [156] G. H. Urwin, N. Kehoe, S. Dundas, and M. Fox, "Testicular infarction in a patient with sickle cell trait," *British Journal of Urology*, vol. 58, no. 3, pp. 340–341, 1986.
- [157] N. M. Holmes and C. J. Kane, "Testicular infarction associated with sickle cell disease," *Journal of Urology*, vol. 160, no. 1, p. 130, 1998.
- [158] D. Bruno, D. R. Wigfall, S. A. Zimmerman, P. M. Rosoff, and J. S. Wiener, "Genitourinary complications of sickle cell disease," *Journal of Urology*, vol. 166, no. 3, pp. 803–811, 2001.
- [159] P. S. Sarma, "Testis involvement in sickle cell trait," *The Journal of the Association of Physicians of India*, vol. 35, no. 4, p. 321, 1987.
- [160] K. J. Hurt, B. Musicki, M. A. Palese et al., "Akt-dependent phosphorylation of endothelial nitric-oxide synthase mediates penile erection," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 6, pp. 4061–4066, 2002.
- [161] K. C. Wood, L. L. Hsu, and M. T. Gladwin, "Sickle cell disease vasculopathy: a state of nitric oxide resistance," *Free Radical Biology and Medicine*, vol. 44, no. 8, pp. 1506–1528, 2008.
- [162] M. L. Adams, E. R. Meyer, B. N. Sewing, and T. J. Cicero, "Effects of nitric oxide-related agents on rat testicular function," *Journal of Pharmacology and Experimental Therapeutics*, vol. 269, no. 1, pp. 230–237, 1994.
- [163] D. B. Hales, "Testicular macrophage modulation of Leydig cell steroidogenesis," *Journal of Reproductive Immunology*, vol. 57, no. 1-2, pp. 3–18, 2002.
- [164] C. Mondillo, R. M. Pagotto, B. Piotrkowski et al., "Involvement of nitric oxide synthase in the mechanism of histamineinduced inhibition of leydig cell steroidogenesis via histamine receptor subtypes in sprague-dawley rats," *Biology of Reproduction*, vol. 80, no. 1, pp. 144–152, 2009.
- [165] A. Solovey, R. Kollander, L. C. Milbauer et al., "Endothelial nitric oxide synthase and nitric oxide regulate endothelial tissue factor expression in vivo in the sickle transgenic mouse," *American Journal of Hematology*, vol. 85, no. 1, pp. 41–45, 2010.
- [166] L. De Franceschi, M. D. Cappellini, and O. Olivieri, "Thrombosis and sickle cell disease," *Seminars in Thrombosis and Hemostasis*, vol. 37, no. 3, pp. 226–236, 2011.
- [167] R. Gopalakrishna, Zhen Hai Chen, and U. Gundimeda, "Nitric oxide and nitric oxide-generating agents induce a reversible inactivation of protein kinase C activity and phorbol ester binding," *Journal of Biological Chemistry*, vol. 268, no. 36, pp. 27180–27185, 1993.
- [168] V. Sauzeau, H. Le Jeune, C. Cario-Toumaniantz et al., "Cyclic GMP-dependent protein kinase signaling pathway inhibits RhoA- induced Ca²⁺ sensitization of contraction in vascular smooth muscle," *Journal of Biological Chemistry*, vol. 275, no. 28, pp. 21722–91729, 2000.
- [169] N. Sawada, H. Itoh, J. Yamashita et al., "cGMP-dependent protein kinase phosphorylates and inactivates RhoA," *Biochemical and Biophysical Research Communications*, vol. 280, no. 3, pp. 798–805, 2001.
- [170] N. L. Jernigan, B. R. Walker, and T. C. Resta, "Chronic hypoxia augments protein kinase G-mediated Ca²⁺ desensitization in pulmonary vascular smooth muscle through inhibition of RhoA/Rho kinase signaling," *American Journal* of *Physiology*, vol. 287, no. 6, pp. L1220–L1229, 2004.
- [171] F. B. M. Priviero, L. M. Jin, Z. Ying, C. E. Teixeira, and R. C. Webb, "Up-regulation of the RhoA/Rho-kinase signaling pathway in corpus cavernosum from endothelial Nitric-Oxide Synthase (NOS), but not neuronal NOS, null mice,"

Journal of Pharmacology and Experimental Therapeutics, vol. 333, no. 2, pp. 184–192, 2010.

- [172] P. Abrams, "Describing bladder storage function: overactive bladder syndrome and detrusor overactivity," *Urology*, vol. 62, no. 5, supplement 2, pp. 28–37, 2003.
- [173] M. C. Michel and M. M. Barendrecht, "Physiological and pathological regulation of the autonomic control of urinary bladder contractility," *Pharmacology and Therapeutics*, vol. 117, no. 3, pp. 297–312, 2008.
- [174] K. E. Andersson, P. Hedlund, A. J. Wein, R. R. Dmochowski, and D. R. Staskin, "Pharmacologic perspective on the physiology of the lower urinary tract," *Urology*, vol. 60, no. 5, pp. 13–20, 2002.
- [175] K. E. Andersson and A. Arner, "Urinary bladder contraction and relaxation: physiology and pathophysiology," *Physiological Reviews*, vol. 84, no. 3, pp. 935–986, 2004.
- [176] S. L. M. Peters, M. Schmidt, and M. C. Michel, "Rho kinase: a target for treating urinary bladder dysfunction?" *Trends in Pharmacological Sciences*, vol. 27, no. 9, pp. 492–497, 2006.
- [177] K. I. Ataga and E. P. Orringer, "Renal abnormalities in sickle cell disease," *American Journal of Hematology*, vol. 63, no. 4, pp. 205–211, 2000.
- [178] F. Daneshgari, G. Liu, L. Birder, A. T. Hanna-Mitchell, and S. Chacko, "Diabetic bladder dysfunction: current translational knowledge," *Journal of Urology*, vol. 182, no. 6, pp. S18–S26, 2009.
- [179] C. R. Chapple, T. Yamanishi, R. Chess-Williams, J. G. Ouslander, J. P. Weiss, and K. E. Andersson, "Muscarinic receptor subtypes and management of the overactive bladder," *Urology*, vol. 60, no. 5, pp. 82–89, 2002.
- [180] S. S. Hegde and R. M. Eglen, "Muscarinic receptor subtypes modulating smooth muscle contractility in the urinary bladder," *Life Sciences*, vol. 64, no. 6-7, pp. 419–428, 1999.
- [181] S. S. Hegde, A. Choppin, D. Bonhaus et al., "Functional role of M2 and M3 muscarinic receptors in the urinary bladder of rats in vitro and in vivo," *British Journal of Pharmacology*, vol. 120, no. 8, pp. 1409–1418, 1997.
- [182] P. A. Longhurst, R. E. Leggett, and J. A. K. Briscoe, "Characterization of the functional muscarinic receptors in the rat urinary bladder," *British Journal of Pharmacology*, vol. 116, no. 4, pp. 2279–2285, 1995.
- [183] A. Choppin, R. M. Eglen, and S. S. Hegde, "Pharmacological characterization of muscarinic receptors in rabbit isolated iris sphincter muscle and urinary bladder smooth muscle," *British Journal of Pharmacology*, vol. 124, no. 5, pp. 883–888, 1998.
- [184] S. Mutoh, J. Latifpour, M. Saito, and R. M. Weiss, "Evidence for the presence of regional differences in the subtype specificity of muscarinic receptors in rabbit lower urinary tract," *Journal of Urology*, vol. 157, no. 2, pp. 717–721, 1997.
- [185] D. J. Sellers, T. Yamanishi, C. R. Chapple, C. Couldwell, K. Yasuda, and R. Chess-Williams, "M3 muscarinic receptors but not M2 mediate contraction of the porcine detrusor muscle in vitro," *Journal of Autonomic Pharmacology*, vol. 20, no. 3, pp. 171–176, 2000.
- [186] G. D'Agostino, M. L. Bolognesi, A. Lucchelli et al., "Prejunctional muscarinic inhibitory control of acetylcholine release in the human isolated detrusor: involvement of the M4 receptor subtype," *British Journal of Pharmacology*, vol. 129, no. 3, pp. 493–500, 2000.
- [187] R. Chess-Williams, C. R. Chapple, T. Yamanishi, K. Yasuda, and D. J. Sellers, "The minor population of M3-receptors mediate contraction of human detrusor muscle in vitro,"

Journal of Autonomic Pharmacology, vol. 21, no. 5, pp. 243–248, 2001.

- [188] L. O. S. Leiria, F. Z. T. Mõnica, F. D. G. F. Carvalho et al., "Functional, morphological and molecular characterization of bladder dysfunction in streptozotocin-induced diabetic mice: eidence of a role for L-type voltage-operated Ca²⁺ channels," *British Journal of Pharmacology*, vol. 163, no. 6, pp. 1276–1288, 2011.
- [189] A. C. Ramos-Filho, F. Z. Mónica, C. F. Franco-Penteado et al., "Characterization of the urinary bladder dysfunction in renovascular hypertensive rats," *Neurourology and Urodynamics*, vol. 30, no. 7, pp. 1392–402, 2011.
- [190] K. Nakanishi, T. Kamai, T. Mizuno, K. Arai, and T. Yamanishi, "Expression of RhoA mRNA and activated RhoA in urothelium and smooth muscle, and effects of a Rho-kinase inhibitor on contraction of the porcine urinary bladder," *Neurourology and Urodynamics*, vol. 28, no. 6, pp. 521–528, 2009.
- [191] L. Boberg, M. Poljakovic, A. Rahman, R. Eccles, and A. Arner, "Role of Rho-kinase and protein kinase C during contraction of hypertrophic detrusor in mice with partial urinary bladder outlet obstruction," *BJU International*, vol. 109, no. 1, pp. 132–140, 2012.
- [192] M. A. Claudino, C. F. Franco-Penteado, M. A. F. Corat et al., "Reduction of urinary bladder activity in transgenic sickle cell disease mice," *Blood*, vol. 114, abstract 2580, 2009, (ASH Annual Meeting Abstracts).
- [193] W. F. Tarry, J. W. Duckett, and M. I. H. Snyder, "Urological complications of sickle cell disease in a pediatric population," *Journal of Urology*, vol. 138, no. 3, pp. 592–594, 1987.
- [194] H. S. Zarkowsky, D. Gallagher, and F. M. Gill, "Bacteremia in sickle hemoglobinopathies," *Journal of Pediatrics*, vol. 109, no. 4, pp. 579–585, 1986.
- [195] J. M. Miller Jr., "Sickle cell trait in pregnancy," Southern Medical Journal, vol. 76, no. 8, pp. 962–963, 1983.
- [196] I. C. Baill and F. R. Witter, "Sickle trait and its association with birthweight and urinary tract infections in pregnancy," *International Journal of Gynecology and Obstetrics*, vol. 33, no. 1, pp. 19–21, 1990.
- [197] L. M. Pasture, D. A. Savitz, and J. M. Thorp, "Predictors of urinary tract infection at the first prenatal visit," *Epidemiol*ogy, vol. 10, no. 3, pp. 282–287, 1999.
- [198] M. L. Portocarrero, M. L. Portocarrero, M. M. Sobral, I. Lyra, P. Lordêlo, and U. Barroso Jr., "Prevalence of enuresis and daytime urinary incontinence in children and adolescents with sickle cell disease," *Journal of Urology*, vol. 187, no. 3, pp. 1037–1040, 2012.