

Article

Impact of Guar Gum and Locust Bean Gum Addition on the Pasting, Rheological Properties, and Freeze–Thaw Stability of Rice Starch Gel

Xuejiao Xu ¹, Shuhui Ye ², Xiaobo Zuo ³ and Sheng Fang ^{2,*}¹ College of Biology and Environmental Engineering, Zhejiang Shuren University, Hangzhou 310015, China² School of Food Science and Biotechnology, Zhejiang Gongshang University, Hangzhou 310018, China³ Hangzhou Tea Research Institute, CHINA COOP/Zhejiang Key Laboratory of Transboundary Applied Technology for Tea Resources, Hangzhou 310016, China

* Correspondence: fangsheng@zjgsu.edu.cn; Tel.: +86-130-9375-2831

Abstract: Improving the gel texture and stability of rice starch (RS) by natural hydrocolloids is important for the development of gluten-free starch-based products. In this paper, the effects of guar gum and locust bean gum on the pasting, rheological properties, and freeze–thaw stability of rice starch were investigated by using a rapid visco analyzer, rheometer, and texture analyzer. Both gums can modify the pasting properties, revealed by an increment in the peak, trough, and final viscosities, and prevent the short-term retrogradation tendency of RS. Dynamic viscoelasticity measurements also indicated that the starch–gum system exhibits superior viscoelastic properties compared with starch alone, as revealed by its higher storage modulus (G'). Compared with the control, the hysteresis loop area of the guar gum-containing system and locust bean gum-containing system was reduced by 37.7% and 24.2%, respectively, indicating that the addition of gums could enhance shear resistance and structure recovery properties. The thermodynamic properties indicated that both gums retard short-term retrogradation as well as long-term retrogradation of the RS gels. Interestingly, the textural properties and freeze–thaw stability of the RS gel were significantly improved by the addition of galactomannans ($p < 0.05$), and guar gum was more effective than locust bean gum, which may be due to the different mannose to galactose ratio. The results provide alternatives for gluten-free recipes with improved texture properties and freeze–thaw stability.

Keywords: rice starch; guar gum; locust bean gum; rheological properties; RVA; freeze–thaw stability

Citation: Xu, X.; Ye, S.; Zuo, X.; Fang, S. Impact of Guar Gum and Locust Bean Gum Addition on the Pasting, Rheological Properties, and Freeze–Thaw Stability of Rice Starch Gel. *Foods* **2022**, *11*, 2508. <https://doi.org/10.3390/foods11162508>

Academic Editors: Sidonia Martínez and Javier Carballo

Received: 30 June 2022

Accepted: 16 August 2022

Published: 19 August 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Rice starch (RS) products are common diets in China and Southeast Asian countries due to the consumer-friendly taste, texture, and ease-of-preparation feature [1,2]. RS products are also an important alternative diet for patients with celiac disease [3]. However, RS gels have deficiencies in ductility, elasticity, and extensibility in the processing and texture of products [4,5]. In addition, RS gel is prone to retrograde during storage, especially during freeze–thaw cycles [6]. Hydrocolloids are widely used in the food industry as functional additives [7–9]. Generally, hydrocolloids play critical roles in modifying the rheological and pasting properties of starch, with enhancement in production process efficiency and optimization of texture stability and sensory properties [8,9]. Mechanistic studies have revealed that the incorporation of hydrocolloids in native starches could improve their moisture content and water-holding capacity [10].

Galactomannans are linear hydrocolloids originally found in the endosperm of seeds of various legumes. Studies have revealed that most galactomannans form nonionic structures, which facilitate the formation of functional properties of pH resistance and stability in both ion-enriched solutions and heating processes [11]. In addition, galactomannans are also used as functional ingredients during digestion due to their ability to slow down the

degradation rate of starch [12]. Of all the legume seed galactomannans, guar gum and locust bean gum are the most commonly used in the food industry.

Guar gum (*Cyamopsis tetragonoloba*) and locust bean gum (*Ceretonia siliqua*) are classical galactomannans comprising a β -(1 \rightarrow 4) linked mannopyranosyl linear backbone with α -(1 \rightarrow 6) linked D-galactopyranosyl branched chains [13]. Guar gum and locust bean gum have similar molecular structures but differ in the ratio of D-mannosyl: D-galactosyl units, which are 2:1 and 4:1, respectively [14]. Guar gum and/or locust bean gum have been reported to modify the properties of yam starch [15], tapioca starch [16], acorn starch [17], and corn starch [18]. It has been found that guar gum generally exhibits pronounced elastic properties, whereas locust bean gum enhances the viscous properties of tapioca starch owing to the different chain extensions and hydrogen bond numbers [19,20]. Significant improvements in the freeze–thaw stability of corn starch with the incorporation of guar gum rather than that of locust bean gum have been reported, which have contributed to the more frequent interaction between guar gum and leached amylose [15]. Thus, the interactions between hydrocolloids and starch could be crucial for starch-based food products.

It has been reported that the different synergistic effects of starch and hydrocolloids could be related to the physicochemical differences of hydrocolloids such as solubility [21–23], intrinsic viscosity [24], extended conformation [25,26], antioxidant potential [27,28], hydrogen bonding capacity [29], flexibility [30], and temperature of gel formation [24,31,32]. The diversity of hydrocolloid molecular structures, especially for the differences in the side chain, usually contributes to the variations in physicochemical properties [33]. Therefore, comprehensive analyses of hydrocolloids and related starch would be beneficial for the further application of related products. However, to the best of our knowledge, the comprehensive understanding of guar gum and locust bean gum when added to rice starch remains unknown [34].

This study aimed to investigate the effect of two galactomannans (guar gum and locust bean gum) on the pasting, rheological properties, and freeze–thaw stability of RS and the possible interaction mechanism. The results may provide valuable information for the selection of gluten-free recipes with improved textural properties and freeze–thaw stability.

2. Materials and Methods

2.1. Materials

Guar gum (CAS No.: 9000-30-0) and locust bean gum (CAS No.: 9000-40-2) were purchased from Aladdin Industrial Corporation. Rice starch (food-grade) was supplied from Jiangxi Jinnong Co., Ltd. (Yichun, China). The main components of RS were analyzed by standard analytical methods [9]. The total starch content of RS was $90.5 \pm 1.35\%$, the amylose content was $24.67 \pm 0.27\%$, the fat content was $0.05 \pm 0.01\%$, the protein content was $0.90 \pm 0.06\%$, and the ash and moisture content were $0.22 \pm 0.02\%$ and $7.7 \pm 0.03\%$, respectively.

2.2. Pasting Properties

The pasting properties of RS gels were measured using a Rapid Visco Analyzer (RVA Tec Master, Perten instruments, Hägersten, Sweden) following previous methods [35]. RS alone slurries were obtained by dispersing 3 g RS powder in 25 g distilled water. In the case of RS/gum mixtures, weighed amounts of guar gum or locust bean gum powder were dispersed in distilled water firstly to prepare the hydrocolloid solutions (0.1 wt.%), with an hour of magnetic stirring at 80 °C. Then, 3 g RS powder was dispersed in the prepared solution (ca. 25 g) with mild stirring to obtain the mixture. The prepared slurries weighing 28 g were then transferred to aluminum RVA canisters to investigate the pasting properties. The starch slurry was at first held at 50 °C for 1 min, then heated to 95 °C within 3 min 45 s and maintained at 95 °C for 2 min 30 s. It was subsequently cooled to 50 °C within 3 min 45 s and maintained at 50 °C for 1 min 30 s. For the first 10 s before measurement, the speed of the plastic paddle was set as 960 rad/s for complete dispersion and then kept constant at 160 rad/s during measurement. RVA parameters such as peak viscosity (PV),

trough viscosity (TV), final viscosity (FV), breakdown value (BV), pasting temperature (PT), and setback value (SBV) were obtained from the pasting curves. A pan containing the same mass of distilled water was used as a reference. RVA parameters were presented as the mean \pm SD of triplicate experiments.

2.3. Rheological Properties

The rheological behaviors of RS gels were determined by using an AR-G2 rheometer (TA Instruments, New Castle, DE, USA) with 40 mm parallel plates according to previous methods [33]. The gelatinized slurries obtained from RVA analysis were then immediately transferred to the platform of the rheometer. Before testing, all samples were equilibrated at 25 °C for 120 s and then examined by both dynamic viscoelastic and steady flow measurements.

Dynamic viscoelastic tests were conducted in a frequency range from 0.1 to 10 Hz with a constant strain of 1%. The dynamic rheological data storage modulus (G') and loss modulus (G'') vs. angular frequency (ω) were obtained and analyzed by using the following equations [36]:

$$G' = K'(\omega)^{n'} \quad (1)$$

$$G'' = K''(\omega)^{n''} \quad (2)$$

where K' is constant, ω is the angular frequency, and n' is the frequency exponents.

Next, the steady flow tests were performed, and the shear rate as a function of shear stress was obtained by using the Data software. The shear rate ramps from 0.01 to 300 s⁻¹ (upward flow curve) and then decreases from 300 s⁻¹ to 0.01 s⁻¹ after 1 min of equilibration (downward flow curve). Experiment results from the ascending and descending segments of the shear cycle were then fitted using the power-law model to characterize the flow properties [36]:

$$\sigma = K \times \gamma^n \quad (3)$$

where σ is the shear stress (Pa), K is the consistency coefficient (Pa·sⁿ), γ is the shear rate (s⁻¹), and n is the flow behavior index (dimensionless).

2.4. Thermodynamic Properties

Thermodynamic properties of the RS gels in the presence or absence of gums were determined by using a C80 differential scanning calorimeter (Setaram, Lyon, France) following previous methods with some modification [37]. Samples were prepared by dispersing 5 g RS powder in 15 g distilled water or hydrocolloid solutions (0.15 wt.%) and stirred for 2 h. Thereafter, the well-stirred suspensions (ca. 4 g) were transferred to an aluminum crucible and tested. The heating temperature ramped from 30 to 100 °C with an acceleration of 0.5 °C/min, and then the ramp was reversed to 30 °C at a rate of -2 °C/min. The instrument was calibrated by using indium and an empty pan as reference. After gelatinization, samples were cooled down and stored at 4 °C for 0, 3, 5, and 12 days, then heated again to investigate the effect of guar gum and locust bean gum on the retrogradation of RS gels. The retrogradation degree was calculated by the ratio of retrogradation enthalpy in the second run heating (ΔH_2) to the gelatinization enthalpy in the first run test (ΔH_1) [38].

2.5. Texture Properties

The textural properties of the RS in the presence or absence of gums were determined by using a TA-XTplus Texture Analyzer (Stable Micro System Ltd., Godalming, UK). The gel samples were prepared as described in Section 2.1 and stored at 4 °C for 0, 3, and 5 d. After being conditioned at room temperature, the samples were analyzed by using texture profile analysis (TPA). The TPA tests were performed on samples 25 mm in diameter and 20 mm in height. The sample was placed on the text platform and squeezed twice to 15 mm with a 25.0 mm diameter cylinder probe P/25. The speed of the probe was 3 mm/s, and the trigger force was 5 g. The interval between two compressions was 3 s, and the data acquisition rate was 200 pps.

2.6. Freeze–Thaw Stability

The freeze–thaw stability of RS in the presence or absence of gums was determined following the method of Zhai et al. [39] with some modifications. Samples were prepared as described in Section 2.2. The slurry was cooled to 30 °C and then transferred to a preweighed centrifuge tube (10 mL) to record its total weight. The samples were then frozen at −21 °C for 24 h, thawed at 30 °C for 2 h, and centrifuged at 8000 rpm for 20 min. The supernatant was discarded and then weighed. These steps were repeated five times to determine the freeze–thaw stability of samples. The syneresis rate was calculated from the following Equation (4):

$$\text{Syneresis (\%)} = \frac{(M_2 - M_3) \times 100}{M_2 - M_1} \quad (4)$$

where M_1 (g) is the weight of the centrifugal tube, M_2 (g) is the weight of starch paste and centrifuge tube, and M_3 (g) is the weight of starch paste and centrifuge tube after pouring out the supernatant.

2.7. Statistical Analysis

The experimental data were analyzed by variance analysis for a completely random design using the SPSS package (19.0, SPSS Inc., Armonk, NY, USA). Duncan's multiple range tests were conducted to analyze the difference between means with a statistical significance of $p < 0.05$.

3. Results and Discussion

3.1. Pasting Properties

The pasting curves of RS in the presence or absence of guar gum or locust bean gum are shown in Figure 1. According to the results of pasting behaviors (Table 1), a significant increment was determined in peak and trough viscosity in the presence of galactomannans ($p < 0.05$). This observation was interpreted by the thickening properties of galactomannans. Guar gum and locust bean gum would enhance the shearing forces exerted on the starch granules [40] and defer the hydrolysis rate of starch [41], which is related to the higher viscosity. Moreover, the effective starch concentration was increased by the immobilization of the water molecules [11], enhancing a strong entanglement with amylose and hydrocolloids [24]. Particularly, the viscosity increment of the starch system with guar gum was more pronounced than that with locust bean gum. The primary and secondary OH (hydroxyl groups) located at the exterior branch of guar gum preferred to form more hydrogen bonds, which exhibited the highest PV and BV in the starch-related system.

The breakdown viscosity (BV) of the RS mixture ranged between 878.33 and 1137.33 cP. Higher BV was observed in the guar gum-related system, which indicated its relatively lower thermostability and compatibility compared with the locust bean gum. Similar observations have been obtained in that the BV of wheat flour pastes was increased along with an increment in guar gum levels [41].

SBV could be used as an indicator to measure the syneresis level of starch during the cooling process [42]. Starch systems containing both guar gum and locust bean gum exhibited a lower tendency to retrograde, as evidenced by the lower SBV value. The extension of the network structure in the paste system was inhibited due to the interaction between the colloidal molecules and the leached amylose [43]. Meanwhile, the addition of strong hydrophilic colloids could reduce the free water content, thereby hindering the rearrangement of starch.

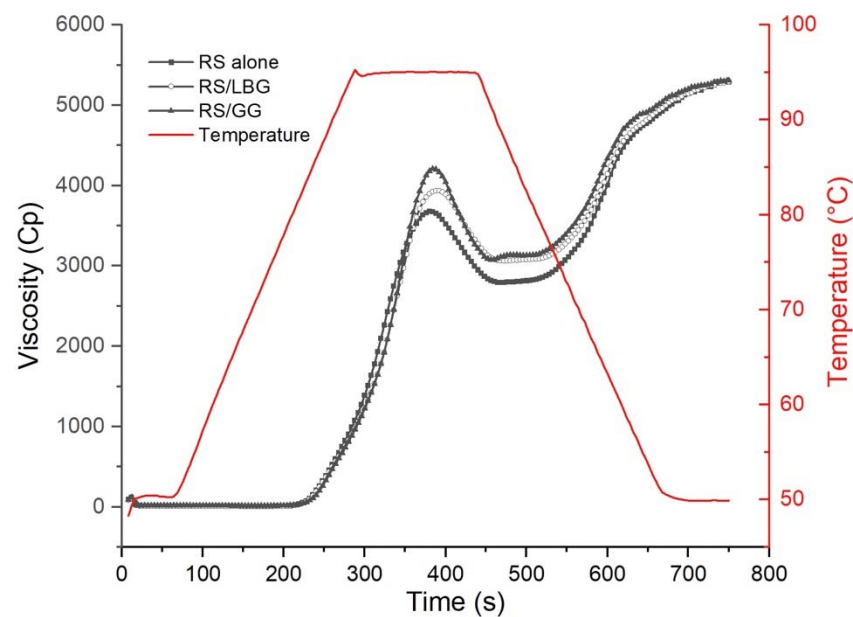


Figure 1. RVA pasting curves of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG).

Table 1. Pasting parameters of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG) *.

Sample	PV (cP)	TV (cP)	BV (cP)	FV (cP)	SBV (cP)	PT (°C)
RS alone	3679.67 ± 17.47 ^c	2790.67 ± 8.33 ^b	889.00 ± 19.29 ^b	5287.33 ± 25.89 ^a	2496.67 ± 26.50 ^a	82.43 ± 0.42 ^b
RS/LBG	3940.33 ± 4.04 ^b	3062.00 ± 23.90 ^a	878.33 ± 21.03 ^b	5291.67 ± 7.23 ^a	2229.67 ± 24.17 ^b	82.42 ± 0.49 ^b
RS/GG	4210.00 ± 36.29 ^a	3072.67 ± 43.68 ^a	1137.33 ± 22.81 ^a	5308.67 ± 10.69 ^a	2236.00 ± 39.00 ^b	83.25 ± 0.43 ^a

* Each value represents the mean ± SD of triplicate experiments. Different superscripts with the same columns are significantly different ($p < 0.05$ by Duncan's multiple range test).

3.2. Dynamic Viscoelastic Properties

The storage modulus (G') and loss modulus (G'') of RS gels in the presence or absence of gums are shown in Figure 2. It shows that the values of G' and G'' increased with an increasing angular frequency for all samples. Similar results have also been reported with other starch-related systems [44]. In addition, a significant increment in G' and G'' was determined with the addition of galactomannans. For example, the G' values of the guar gum-containing system increased from 356.9 to 459.9 Pa with respect to the control. The viscoelastic properties of starch-related systems were enhanced owing to the thickening properties of galactomannans [45].

Interestingly, it seemed different hydrocolloids modified the elastic and viscous properties of starch systems variously. For the guar gum-containing system, the increasing rate of the G' value was much greater than that of the G'' value, indicating remarkable elastic properties that could be considered an enhancement of the weak gel network due to more OH groups on the galactose chain. It has been reported that more galactose substitutions of guar gum would hinder intramolecular hydrogen bond formation, thereby exhibiting a more extended conformation than locust bean gum [20]. It is known that noncovalent intermolecular interactions such as hydrogen bonding are the most common and important interaction types between biopolymers [46,47]. It is predicted that the extended chain form promoted the interaction between amylose and guar gum via a noncovalent bond, thereby enhancing the elasticity and pseudo-plasticity of the system [39,48]. On the contrary, the locust bean gum-containing system exhibited pronounced viscous properties, as evidenced by the higher $\tan(\delta)$ and G'' values (Table 2). A similar trend has also been reported with a corn starch–galactomannan system [17].

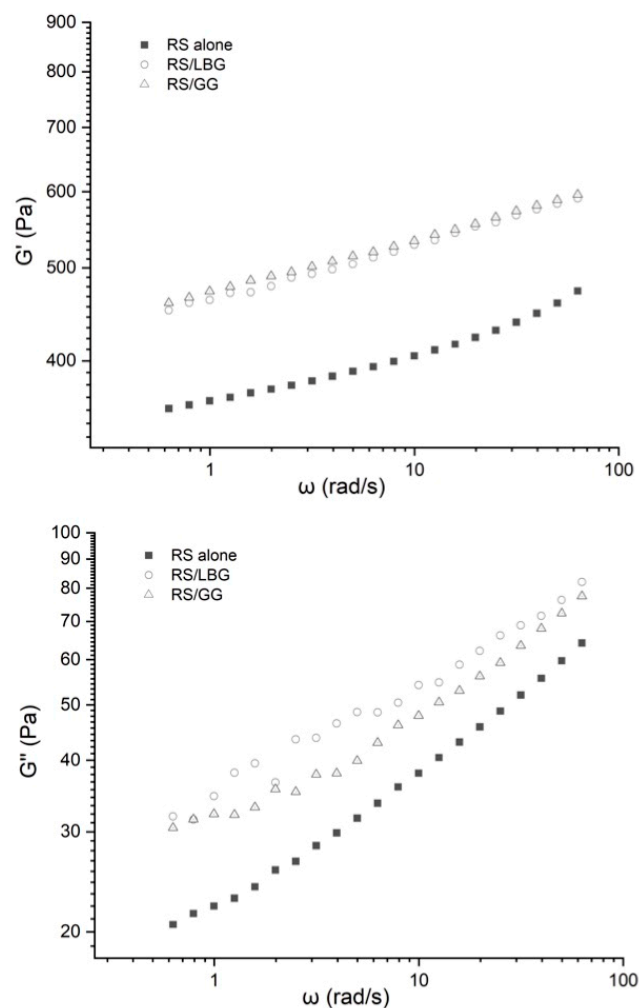


Figure 2. Storage modulus and loss modulus of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG).

Table 2. Storage (G'), loss moduli (G''), and loss tangent ($\tan \delta$) at 6.28 rad s^{-1} for RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG) *.

Sample	G' (Pa)	G'' (Pa)	$\tan(\delta)$
RS alone	394.47 ± 21.27^b	33.65 ± 1.35^c	0.085 ± 0.003^b
RS/LBG	512.77 ± 27.98^a	48.51 ± 0.41^a	0.095 ± 0.006^a
RS/GG	519.60 ± 30.47^a	42.95 ± 1.63^b	0.083 ± 0.005^b

* Mean \pm SD. Different superscripts with the same columns are significantly different ($p < 0.05$ by Duncan's multiple range test).

For all RS-related systems, $\ln(G', G'')$ as a function of $\ln \omega$ were conducted using linear regression and are summarized in Table 3. It was observed that each starch-related system exhibited weak gel-like behavior with positive slopes (n' and n'') [48]. The value of K' and K'' increased significantly after the addition of galactomannans, indicating enhanced viscoelasticity of the continuous phase caused by the thickening properties of gums. Such observation is in agreement with maize starch–guar gum mixtures [45] and rice starch–xanthan gum mixtures [34]. The presence of galactomannans facilitated the associations of ordered chain segments, thereby enhancing the weak three-dimensional network of RS–gum mixtures [49].

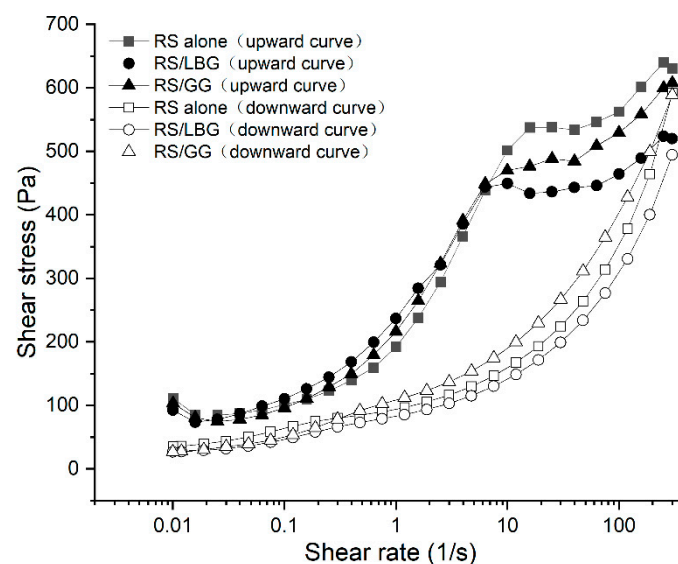
Table 3. Parameters of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG) at 25 °C as determined from Equations (1) and (2) *.

Sample	Storage Modulus G'			Loss Modulus G''		
	$k'(\text{pa}\cdot\text{s}^n)$	n''	R^2	$k''(\text{pa}\cdot\text{s}^n)$	n''	R^2
RS alone	358.76 ± 20.01^b	0.06 ± 0.00^a	0.970	21.25 ± 0.66^c	0.26 ± 0.01^a	0.996
RS/LBG	461.77 ± 28.63^a	0.06 ± 0.00^a	0.997	34.29 ± 0.42^a	0.20 ± 0.00^b	0.988
RS/GG	470.98 ± 33.50^a	0.06 ± 0.01^a	0.998	29.69 ± 1.40^b	0.22 ± 0.01^c	0.974

* Mean \pm SD. Different superscripts with the same columns are significantly different ($p < 0.05$ by Duncan's multiple range test).

3.3. Steady Shear Properties

The effects of gums on the steady shear properties of RS gels are shown in Figure 3. Thixotropic behaviors of all samples were observed within the range of shear rates ($0.01\text{--}300\text{ s}^{-1}$). The first peak that appeared in the upward flow curves was related to the stress required to break the gel structure and caused the solution to recover. Similar results have been reported in the RS–glucans system [38]. The collected data showed a good fit to the power-law model with R^2 between 0.976 and 0.997. For all samples, the consistency coefficient (K), the yield stresses (σ), flow behavior indices (n), and the apparent viscosity at 300 s^{-1} ($\eta_{a,300}$), as well as hysteresis loop areas between the upward and downward curves are summarized in Table 4. Obvious hysteresis loop areas were observed in all starch-related systems, which could be explained by the structural breakdown of the shear field to change the original structure or build up a new one, which then maintained the shear-thinning properties in subsequent shear sweeps [50]. The decomposition of the original structures was observed during gel shearing, as evidenced by $n < 1$. After shearing, the gel structure could only be partially recovered, which is related to the lower $\eta_{a,300}$ values of the downward curves compared with the upward ones [51]. For the downward curve, it showed that the addition of both gums markedly increased the K values, which reflects that the gums mainly enhanced viscoelastic properties, especially for guar gum, owing to the thickening effect.

**Figure 3.** Steady flow curves of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG).

Compared with the control, the gum-containing systems exhibited more pseudoplastic properties, as evidenced by the higher K values (Table 4). Specifically, this effect was more pronounced for the RS–guar gum systems than the RS–locust bean gum systems. This

observation is in agreement with the creep recovery results, showing that the guar gum-containing system exhibited higher shear resistance (please see Supplementary Materials Figure S1).

Table 4. The steady flow fitting parameters of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG) *.

Sample	Hla (Pa·s)	Upward Curve				Downward Curve			
		K (Pa·s ⁿ)	n	$\eta_{a,300}$ (Pa·s)	R ²	K (Pa·s ⁿ)	n	$\eta_{a,300}$ (Pa·s)	R ²
RS alone	53,245	236.57 ± 2.41 ^a	0.187 ± 0.004 ^{ab}	2.15 ± 0.05 ^a	0.909	81.84 ± 1.6 ^c	0.334 ± 0.002 ^a	2.02 ± 0.03 ^a	0.976
RS/LBG	40,385	236.51 ± 32.32 ^a	0.152 ± 0.056 ^b	2.23 ± 0.03 ^a	0.915	86.64 ± 6.6 ^b	0.328 ± 0.004 ^b	2.15 ± 0.02 ^a	0.987
RS/GG	33,150	237.62 ± 16.60 ^a	0.180 ± 0.033 ^{ab}	2.02 ± 0.56 ^a	0.951	101.9 ± 12.6 ^a	0.312 ± 0.002 ^c	1.96 ± 0.54 ^a	0.997

* Hla: hysteresis loop area (Pa·s); power-law parameters: K, consistency coefficient; n, flow behavior index; $\eta_{a,300}$, the apparent viscosity at 300 s⁻¹. Different superscript letters with the same columns are significantly different ($p < 0.05$ by Duncan's multiple range test).

The hysteresis loop area is an indicator to value the structural breakdown during shearing [52]. Compared with the control, the hysteresis loop area of RS gels containing guar gum and locust bean gum were reduced by 37.7% and 24.2%, respectively, indicating remarkable shear resistance, which has been attributed to the enhancement of the three-dimensional structure of the starch [53]. A similar trend was observed in the dynamic viscoelastic results (Figure 2), which, in turn, reflected the retrogradation extent of RS gels (Table 6).

3.4. Thermodynamic Properties

Various information such as the starch–hydrocolloid interaction, the effects of water, and related properties could be provided by using DSC analysis. The thermodynamic properties of RS gels in the presence or absence of gums and their corresponding retrograded gels during refrigerated storage (4 °C for 3, 5, and 12 days) are listed in Table 5.

Table 5. The retrogradation thermodynamic parameters of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG) *.

Samples	First Run	Second Run (3 d at 4 °C)		Third Run (5 d at 4 °C)		Fourth Run (12 d at 4 °C)	
	ΔH_1	ΔH_2	$\Delta H_2/\Delta H_1$	ΔH_3	$\Delta H_3/\Delta H_1$	ΔH_4	$\Delta H_4/\Delta H_1$
RS alone	2.66	0.30	0.11	1.48	0.56	2.32	0.87
RS/LBG	2.76	0.40	0.14	1.38	0.50	1.43	0.52
RS/GG	2.74	0.33	0.12	1.11	0.41	1.47	0.54

* ΔH_1 , gelatinization enthalpy; ΔH_2 , retrogradation enthalpy; $\Delta H_2/\Delta H_1$, retrogradation ratio.

Starch–galactomannan interactions affected the retrogradation of samples. The gelatinization enthalpies (ΔH_1) of the starch system, when added with gums, were slightly increased at first, which may be related to the higher viscosity. In the third and last run, the retrogradation enthalpies (ΔH_2) and retrogradation ratios ($\Delta H_2/\Delta H_1$) of retrograded RS gels decreased significantly in the presence of both guar gum and locust bean gum, indicating that gums slowed the retrogradation rate of RS gels during refrigerated storage. Starch retrogradation is generally considered a liquid state event, which requires orientational mobility of the polymer chains in the amylopectin molecule [54]. The presence of guar gum and locust bean gum decreased the orientational mobility of the starch gels and, thus, decreased the retrogradation rate. Moreover, the addition of hydrocolloids could hinder the formation of spongy structures in the starch system, where the gum combined with the starch separated from the starch granules.

The reduction in free water content would inhibit the rearrangement of starch chains, thereby effectively suppressing retrogradation [43]. Interestingly, it seems guar gum suppressed, more than locust bean gum, the short-term retrogradation of RS samples stored

for the first 5 days, which was attributed to lower water availability due to the stronger hydration capacity of guar gum [55]. Instead, locust bean gum exhibited the potential to suppress long-term retrogradation, which may be related to its viscous properties, as mentioned above.

The melting enthalpy of recrystallized starch was lower than that of gelatinization, which is consistent with the easier melting properties of recrystallized starch than that of native starch granules [56]. It also found that guar gum and locust bean gum increased or decreased the melting temperatures of recrystallized starch differently during storage time (please see Supplementary Materials Table S1). Similar research [57] has been reported in gum–tapioca starch systems. The results indicated that the addition of guar gum and locust bean gum could modify the thermal properties of RS dependent on the gum type and storage time.

3.5. Determination of Texture Properties

The texture properties of RS gels in the presence or absence of gums during refrigerated storage are presented in Table 6. The hardness of starch gel is always an indicator of the degree of starch retrogradation [58]. The higher hardness values of the starch gel during the initial 3 days indicated its rapid retrogradation. This observation is similar to the retrogradation of amaranth starch in that retrogradation is accelerated at refrigerated temperatures [59]. As a result, a more compact and ordered structure is formed by the amylose and amylopectin in the gelatinized starch, thereby increasing the gel hardness of the RS [60].

Table 6. TPA texture parameters of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG) during cold storage *.

Sample	Days	Hardness	Springiness	Adhesiveness	Cohesiveness	Gumminess	Chewiness
RS alone	3	1134.16 ± 58.91 ^a	0.37 ± 0.02 ^a	77.64 ± 4.16 ^b	0.24 ± 0.02 ^a	269.40 ± 38.87 ^a	99.06 ± 20.13 ^a
	5	907.52 ± 6.93 ^a	0.26 ± 0.01 ^a	17.63 ± 3.21 ^a	0.1 ± 0.01 ^a	94.42 ± 5.81 ^a	24.80 ± 2.97 ^a
RS/LBG	3	931.82 ± 14.62 ^b	0.36 ± 0.05 ^a	123.20 ± 14.03 ^a	0.25 ± 0.02 ^a	233.56 ± 24.78 ^a	84.93 ± 20.51 ^a
	5	861.36 ± 71.83 ^a	0.32 ± 0.05 ^a	11.90 ± 2.06 ^a	0.10 ± 0.01 ^a	84.02 ± 1.88 ^a	26.73 ± 3.90 ^a
RS/GG	3	861.02 ± 66.74 ^b	0.32 ± 0.01 ^a	54.32 ± 5.98 ^c	0.24 ± 0.04 ^a	202.81 ± 19.53 ^b	64.49 ± 4.68 ^a
	5	762.69 ± 0.54 ^b	0.28 ± 0.04 ^a	18.66 ± 4.97 ^a	0.10 ± 0.02 ^a	79.81 ± 13.04 ^a	21.81 ± 0.23 ^a

* Values represent the mean ± SD of triplicate tests. Columns with different superscripts are significantly different during different samples ($p < 0.05$ by Duncan's multiple range test).

Both guar gum and locust bean gum exhibited excellent potential in the texture modification of RS products during cold storage time. The numerical reduction in hardness was observed with the addition of gums after 3 days as compared with the control. This observation could be interpreted as the possible interaction between gums and the RS granules. The presence of galactomannans tends to combine with RS molecules, thereby inhibiting the leaching of amylose and hindering the retrogradation of starch-related systems [61]. The arrangement of leached amylose is also retarded by the combination, which further suppresses the recrystallization of starch molecules. Generally, the addition of hydrocolloids would improve water retention and inhibit the rearrangement of amylose of starch, which helps to maintain better texture characteristics during refrigerated storage [62].

To specify, the guar gum-containing system exhibited better textural properties during cold storage time, as shown in Table 6. It has also been reported that hydrogen bonding plays a critical role in the gelatinization and retrogradation of starch [11]. A large amount of hydroxyl groups in guar gum, rather than locust bean gum, tends to transform the nucleus of starch recrystallization from amylose to an amylose–gum mixture. As a result, the arrangement of starch molecules is retarded, leading to a lower retrogradation rate of starch [63]. In general, the hardness of RS alone gel would be increased during cold storage owing to starch retrogradation. Meanwhile, the addition of the gums to RS gels helped to

slow down the changes in textural characteristics during refrigerated storage in the order of guar gum > locust bean gum.

3.6. Freeze–Thaw Stability

The syneresis value of freeze–thaw RS is often determined as an indicator to evaluate its ability to maintain desirable physical properties during the freezing and thawing process [64,65]. The freeze–thaw stability of RS in the presence or absence of locust bean gum or guar gum is shown in Figure 4. For the RS gels alone, a high syneresis value (37.2%) was observed after the first freeze–thaw cycle. With an increase in freeze–thaw cycles, the syneresis values of gels increased consequently. During repeated freeze–thaw cycles, the starch molecules will recombine, coagulate, and even form a spongy structure, and water will precipitate from the starch body, resulting in dehydration and condensation [24,66].

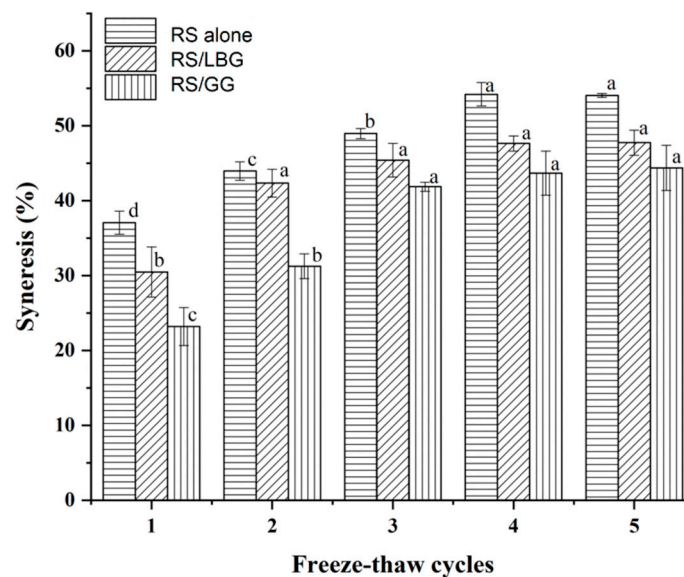


Figure 4. Freeze–thaw stability of RS alone, RS containing locust bean gum (RS/LBG), and RS containing guar gum (RS/GG). Different letters indicate significant differences among group ($p < 0.05$ by Duncan’s multiple range test).

The evidence is demonstrated by the lower syneresis value of the RS/gums gels than of the control, showing better freeze–thaw stability in the gels containing gums during freeze–thaw cycles. It could be interpreted by the thickening properties of guar gum and locust bean gum, which would reduce the ice crystal size [43]. The presence of hydrocolloids in starch gel could also bind to water molecules, which reduces the syneresis degree during the freeze–thaw cycles [67].

Compared with locust bean gum, guar gum exhibit better stability during all the freeze–thaw cycles. This is consistent with previous research in which guar gum showed pronounced freeze–thaw stability and a more remarkable synergistic combination with sweet potato starch [43] and corn starch [15]. Although locust bean gum showed similar gelatinization viscosity (Table 1), it was not as effective as guar gum. This is because of their different physical structures [68], especially the different ratios of galactose branching to the mannose backbone [69]. For the locust bean gum-containing system, more bulky phase water would be generated during the repeated FT cycles owing to the ease in the chain association [43].

4. Conclusions

In the present study, the effects of guar gum and locust bean gum on the gelatinization properties, rheological properties, and freeze–thaw stability of RS gels were investigated. RVA results showed that both guar gum and locust bean gum had inhibitory effects on the

retrogradation of RS, with a lower SBV value and higher PV. Higher viscoelastic behavior and less thixotropic behavior were reviewed in the presence of galactomannans in the rheological measurements, which related to the enhanced weak gel structure and higher shear resistance. The thermal properties of RS gels could be modified with the addition of galactomannans, which were dependent on the gum type and storage time. Moreover, the textural properties and freeze–thaw stability of the RS gel were significantly improved by the addition of galactomannans. Particularly, the guar gum-containing system exhibits a more significant effect rather than that of locust bean gum, which could be attributed to the different mannose to galactose ratios. From these findings, guar gum could be a better alternative for gluten-free products, as it confers a higher retrogradation inhibition effect, freeze–thaw stability, and fewer texture changes in gels. Future attempts with more detailed parameters optimization are strongly required to unveil the possible interaction mechanism between hydrocolloids and starch and, finally, pinpoint their effects on human digestibility and the sensory properties of food products.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/foods11162508/s1>, Figure S1: The creep-recovery curves of RS alone, RS containing locust bean gum (RS/LBG) and RS containing guar gum (RS/GG); Table S1: Gelatinization temperature and enthalpy and retrogradation ratio for RS alone, RS containing locust bean gum (RS/LBG) and RS containing guar gum (RS/GG) measured by the differential scanning calorimeter (DSC).

Author Contributions: Conceptualization, X.X.; data curation, S.Y.; formal analysis, X.Z. and X.X.; funding acquisition, S.F.; methodology, X.Z. and S.Y.; project administration, S.F.; resources, X.X.; software, X.X.; supervision, S.F. and X.X.; validation, S.F.; writing—original draft, S.Y. and S.F. All authors have read and agreed to the published version of the manuscript.

Funding: This study was funded by the Natural Science Foundation of Zhejiang Province (LQ22C200010) and Zhejiang Shuren University Basic Scientific Research Special Funds (2022XZ008).

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare that they have no conflict of interest.

References

1. Tarique, J.; Sapuan, S.M.; Khalina, A.; Ilyas, R.A.; Zainudin, E.S. Thermal, flammability, and antimicrobial properties of arrowroot (*Maranta arundinacea*) fiber reinforced arrowroot starch biopolymer composites for food packaging applications. *Int. J. Biol. Macromol.* **2022**, *213*, 1–10. [[CrossRef](#)] [[PubMed](#)]
2. Gatade, A.A.; Sahoo, A.K. Effect of additives and steaming on quality of air dried noodles. *J. Food Sci. Technol.* **2015**, *52*, 8395–8402. [[CrossRef](#)] [[PubMed](#)]
3. Benkadri, S.; Salvador, A.; Sanz, T.; Nasreddine Zidoune, M. Optimization of xanthan and locust bean gum in a gluten-free infant biscuit based on rice-chickpea flour using response surface methodology. *Foods* **2020**, *10*, 12. [[CrossRef](#)]
4. Anil, M.; Durmus, Y.; Tarakci, Z. Effects of different concentrations of guar, xanthan and locust bean gums on physicochemical quality and rheological properties of corn flour tarhana. *Nutr. Food Sci.* **2020**, *51*, 137–150. [[CrossRef](#)]
5. Pongjaruvat, W.; Methacanon, P.; Seetapan, N.; Fuongfuchat, A.; Gamonpilas, C. Influence of pregelatinised tapioca starch and transglutaminase on dough rheology and quality of gluten-free jasmine rice breads. *Food Hydrocoll.* **2014**, *36*, 143–150. [[CrossRef](#)]
6. Kirchmayer, D.M.; Steinhoff, B.; Warren, H.; Clark, R.; In het Panhuis, M. Enhanced gelation properties of purified gellan gum. *Carbohydr. Res.* **2014**, *388*, 125–129. [[CrossRef](#)] [[PubMed](#)]
7. Fang, S.; Zhao, X.; Liu, Y.; Liang, X.; Yang, Y. Fabricating multilayer emulsions by using OSA starch and chitosan suitable for spray drying: Application in the encapsulation of β -carotene. *Food Hydrocoll.* **2019**, *93*, 102–110. [[CrossRef](#)]
8. Hu, Q.H.; Wu, Y.L.; Zhong, L.; Ma, N.; Zhao, L.Y.; Ma, G.X.; Cheng, N.H.; Nakata, P.A.; Xu, J. In vitro digestion and cellular antioxidant activity of β -carotene-loaded emulsion stabilized by soy protein isolate-Pleurotus eryngii polysaccharide conjugates. *Food Hydrocoll.* **2021**, *112*, 106340. [[CrossRef](#)]
9. Sun, J.; Zuo, X.B.; Fang, S.; Xu, H.N.; Chen, J.; Meng, Y.C.; Chen, T. Effects of cellulose derivative hydrocolloids on pasting, viscoelastic, and morphological characteristics of rice starch gel. *J. Texture Stud.* **2017**, *48*, 241–248. [[CrossRef](#)]
10. Clemens, R.A.; Pressman, P. Food gums: An overview. *Nutr. Today* **2017**, *52*, 41–43. [[CrossRef](#)]
11. Li, J.X.; Yadav, M.P.; Li, J.L. Effect of different hydrocolloids on gluten proteins, starch and dough microstructure. *J. Cereal Sci.* **2019**, *87*, 85–90. [[CrossRef](#)]

12. Charoenrein, S.; Tatirat, O.; Rengsutthi, K.; Thongngam, M. Effect of konjac glucomannan on syneresis, textural properties and the microstructure of frozen rice starch gels. *Carbohydr. Polym.* **2011**, *83*, 291–296. [[CrossRef](#)]
13. Busch, V.; Rozycki, V.; Buera, M.P. Galactomannans from different prosopis species: Extraction, characterization, and applications. In *Prosopis as a Heat Tolerant Nitrogen Fixing Desert Food Legume*; Academic Press: Cambridge, MA, USA, 2022; pp. 305–317.
14. Das, A.; Kundu, S.; Gupta, M.; Mukherjee, A. Guar gum propionate-kojic acid films for *Escherichia coli* biofilm disruption and simultaneous inhibition of planktonic growth. *Int. J. Biol. Macromol.* **2022**, *211*, 57–73. [[CrossRef](#)] [[PubMed](#)]
15. Sudhakar, V.; Singhal, R.S.; Kulkarni, P.R. Starch-galactomannan interactions: Functionality and rheological aspects. *Food Chem.* **1996**, *55*, 259–264. [[CrossRef](#)]
16. Huang, C.C. Physicochemical, pasting and thermal properties of tuber starches as modified by guar gum and locust bean gum. *Int. J. Food Sci. Technol.* **2009**, *44*, 50–57. [[CrossRef](#)]
17. Kim, W.W.; Yoo, B. Rheological and thermal effects of galactomannan addition to acorn starch paste. *LWT Food Sci. Technol.* **2011**, *44*, 759–764. [[CrossRef](#)]
18. Bahnassey, Y.A.; Breene, W.M. Rapid visco-analyzer (RVA) pasting profiles of wheat, corn, waxy corn, tapioca and amaranth starches (*A. hypochondriacus* and *A. cruentus*) in the presence of konjac flour, gellan, guar, xanthan and locust bean gums. *Starch-Stärke* **1994**, *46*, 134–141. [[CrossRef](#)]
19. Prajapati, V.D.; Jani, G.K.; Moradiya, N.G.; Randeria, N.P.; Nagar, B.J.; Naikwadi, N.N.; Variya, B.C. Galactomannan: A versatile biodegradable seed polysaccharide. *Int. J. Biol. Macromol.* **2013**, *60*, 83–92. [[CrossRef](#)]
20. Borries-Medrano, E.V.; Jaime-Fonseca, M.R.; Aguilar-Méndez, M. Tapioca starch-galactomannan systems: Comparative studies of rheological and textural properties. *Int. J. Biol. Macromol.* **2018**, *122*, 1173–1183. [[CrossRef](#)]
21. Richardson, P.H.; Willmer, J.; Foster, T.J. Dilute solution properties of guar and locust bean gum in sucrose solutions. *Food Hydrocoll.* **1998**, *12*, 339–348. [[CrossRef](#)]
22. Salehi, F. Effect of common and new gums on the quality, physical, and textural properties of bakery products: A review. *J. Texture Stud.* **2020**, *51*, 361–370. [[CrossRef](#)] [[PubMed](#)]
23. Tarique, J.; Zainudin, E.S.; Sapuan, S.M.; Ilyas, R.A.; Khalina, A. Physical, mechanical, and morphological performances of arrowroot (*Maranta arundinacea*) fiber reinforced arrowroot starch biopolymer composites. *Polymers* **2022**, *14*, 388. [[CrossRef](#)] [[PubMed](#)]
24. Qiu, S.; Yadav, M.P.; Liu, Y.; Chen, H.; Tatsumi, E.; Yin, L. Effects of corn fiber gum with different molecular weights on the gelatinization behaviors of corn and wheat starch. *Food Hydrocoll.* **2016**, *53*, 180–186. [[CrossRef](#)]
25. Nazrin, A.; Sapuan, S.M.; Zuhri, M.Y.M.; Tawakkal, I.S.M.A.; Ilyas, R.A. Flammability and physical stability of sugar palm crystalline nanocellulose reinforced thermoplastic sugar palm starch/poly(lactic acid) blend bionanocomposites. *Nanotechnol. Rev.* **2021**, *11*, 86–95. [[CrossRef](#)]
26. Culetu, A.; Duta, D.E.; Papageorgiou, M.; Varzakas, T. The role of hydrocolloids in gluten-free bread and pasta; rheology, characteristics, staling and glycemic index. *Foods* **2021**, *10*, 3121. [[CrossRef](#)]
27. Syafiq, R.M.O.; Sapuan, S.M.; Zuhri, M.Y.M.; Othman, S.H.; Ilyas, R.A. Effect of plasticizers on the properties of sugar palm nanocellulose/cinnamon essential oil reinforced starch bionanocomposite films. *Nanotechnol. Rev.* **2022**, *11*, 423–437. [[CrossRef](#)]
28. Hamdani, A.M.; Wani, I.A. Guar and Locust bean gum: Composition, total phenolic content, antioxidant and antinutritional characterisation. *Bioact. Carbohydr. Diet. Fibre* **2017**, *11*, 53–59. [[CrossRef](#)]
29. Schorsch, C.; Garnier, C.; Doublier, J.L. Viscoelastic properties of xanthangalactomannan mixtures: Comparison of guar gum with locust bean gum. *Carbohydr. Polym.* **1997**, *34*, 165–175. [[CrossRef](#)]
30. Maier, H.; Anderson, M.; Karl, C.; Magnuson, K.; Whistler, R.L. Guar, locust bean, Tara, and fenugreek gums: Industrial gums. In *Industrial Gums*; Academic Press: Cambridge, MA, USA, 1993; pp. 181–226.
31. Ge, H.; Wu, Y.; Woshnak, L.L.; Mitmesser, S.H. Effects of hydrocolloids, acids and nutrients on gelatin network in gummies. *Food Hydrocoll.* **2021**, *113*, 106549. [[CrossRef](#)]
32. Xu, X.; Fang, S.; Li, Y.; Zhang, F.; Shao, Z.; Zeng, Y.; Chen, J.; Meng, Y. Effects of low acyl and high acyl gellan gum on the thermal stability of purple sweet potato anthocyanins in the presence of ascorbic acid. *Food Hydrocoll.* **2018**, *86*, 116–123. [[CrossRef](#)]
33. Fang, S.; Wang, J.; Xu, X.J.; Zuo, X.B. Influence of Low Acyl and High Acyl Gellan Gums on Pasting and Rheological Properties of Rice Starch Gel. *Food Biophys.* **2018**, *13*, 116–123. [[CrossRef](#)]
34. Kim, C.; Yoo, B. Rheological properties of rice starch-xanthan gum mixtures. *J. Food Eng.* **2006**, *75*, 120–128. [[CrossRef](#)]
35. Meng, Y.C.; Sun, M.H.; Fang, S.; Chen, J.; Li, Y.H. Effect of sucrose fatty acid esters on pasting, rheological properties and freeze-thaw stability of rice flour. *Food Hydrocoll.* **2014**, *40*, 64–70. [[CrossRef](#)]
36. Wang, B.; Wang, L.J.; Li, D.; Özkan, N.; Li, S.J.; Mao, Z.H. Rheological properties of waxy maize starch and xanthan gum mixtures in the presence of sucrose. *Carbohydr. Polym.* **2009**, *77*, 472–481. [[CrossRef](#)]
37. Kamaruddin, Z.H.; Jumaidin, R.; Ilyas, R.A.; Selamat, M.Z.; Alamjuri, R.H.; Yusof, F.A.M. Biocomposite of cassava starch-cymbopogan citratus fibre: Mechanical, thermal and biodegradation properties. *Polymers* **2022**, *14*, 514. [[CrossRef](#)] [[PubMed](#)]
38. Zhang, D.; Lin, Z.; Lei, W.; Zhong, G. Synergistic effects of acetylated distarch adipate and sesbania gum on gelatinization and retrogradation of wheat starch. *Int. J. Biol. Macromol.* **2020**, *156*, 171–179. [[CrossRef](#)]
39. Zhai, Y.H.; Xing, J.L.; Luo, X.H.; Zhang, H.; Yang, K.; Shao, X.F.; Li, Y.N. Effects of pectin on the physicochemical properties and freeze-thaw stability of waxy rice starch. *Foods* **2021**, *10*, 2419. [[CrossRef](#)]

40. Sasaki, T.; Yasui, T.; Matsuki, J. Influence of non-starch polysaccharides isolated from wheat flour on the gelatinization and gelation of wheat starches. *Food Hydrocoll.* **2000**, *14*, 295–303. [[CrossRef](#)]
41. Brennan, C.S.; Suter, M.; Luethi, T.; Matia-Merino, L.; Qvortrup, J. The relationship between wheat flour and starch pasting properties and starch hydrolysis: Effect of non-starch polysaccharides in a starch gel system. *Starch Strke* **2008**, *60*, 23–33. [[CrossRef](#)]
42. Torres, M.D.; Moreira, R.; Chenlo, F.; Morel, M.H. Effect of water and guar gum content on thermal properties of chestnut flour and its starch. *Food Hydrocoll.* **2013**, *33*, 192–198. [[CrossRef](#)]
43. Lee, M.H.; Baek, M.H.; Cha, D.S.; Park, H.J.; Lim, S.T. Freeze–thaw stabilization of sweet potato starch gel by polysaccharide gums. *Food Hydrocoll.* **2002**, *16*, 345–352. [[CrossRef](#)]
44. Ptaszek, P.; Grzesik, M. Viscoelastic properties of maize starch and guar gum gels. *J. Food Eng.* **2007**, *82*, 227–237. [[CrossRef](#)]
45. Alloncle, M.; Doublier, J.L. Viscoelastic properties of maize starch/hydrocolloid pastes and gels. *Food Hydrocoll.* **1991**, *5*, 455–467. [[CrossRef](#)]
46. Gao, J.; Mao, Y.; Xiang, C.; Cao, M.; Ren, G.; Wang, K.; Ma, X.; Wu, D.; Xie, H. Preparation of β -lactoglobulin/gum arabic complex nanoparticles for encapsulation and controlled release of EGCG in simulated gastrointestinal digestion model. *Food Chem.* **2021**, *354*, 129516. [[CrossRef](#)] [[PubMed](#)]
47. Wu, X.; Hu, Q.; Liang, X.; Fang, S. Fabrication of colloidal stable gliadin-casein nanoparticles for the encapsulation of natamycin: Molecular interactions and antifungal application on cherry tomato. *Food Chem.* **2022**, *391*, 133288. [[CrossRef](#)] [[PubMed](#)]
48. Gao, J.; Liu, C.; Shi, J.; Ni, F.; Shen, Q.; Xie, H.; Wang, K.; Lei, Q.; Fang, W.; Ren, G. The regulation of sodium alginate on the stability of ovalbumin-pectin complexes for vd3 encapsulation and in vitro simulated gastrointestinal digestion study. *Food Res. Int.* **2021**, *140*, 110011. [[CrossRef](#)] [[PubMed](#)]
49. Ptaszek, A.; Berski, W.; Ptaszek, P.; Witczak, T.; Repelewicz, U.; Grzesik, M. Viscoelastic properties of waxy maize starch and selected non-starch hydrocolloids gels. *Carbohydr. Polym.* **2009**, *76*, 567–577. [[CrossRef](#)]
50. Zhang, F.; Fu, S.L.; Jin, T. Rheological properties of maize starch pastes. *Transactions of the CSAE* **2008**, *24*, 294–297.
51. Hussain, S.; Mohamed, A.A.; Alamri, M.S.; Ibraheem, M.A.; Qasem, A.A.A.; Shahzad, S.A.; Ababtain, I.A. Use of gum cordia (*cordia myxa*) as a natural starch modifier; effect on pasting, thermal, textural, and rheological properties of corn starch. *Foods* **2020**, *9*, 909. [[CrossRef](#)]
52. Banchathanakij, R.; Supphantharika, M. Effect of different β -glucans on the gelatinisation and retrogradation of rice starch. *Food Chem.* **2009**, *114*, 5–14. [[CrossRef](#)]
53. Achayuthakan, P.; Supphantharika, M. Pasting and rheological properties of waxy corn starch as affected by guar gum and xanthan gum. *Carbohydr. Polym.* **2008**, *71*, 9–17. [[CrossRef](#)]
54. Slade, L.; Levine, H. Recent advances in starch retrogradation. In *Industrial polysaccharides-The impact of biotechnology and advanced methodologies*; Stivala, S.S., Crescenzi, V., Dea, I.C.M., Eds.; Gordon and Breach Science Publishers: New York, NY, USA, 1987; pp. 387–430.
55. Lee, S.; Yoo, B. Effect of sucrose addition on rheological and thermal properties of rice starch-gum mixtures. *Int. J. Food Eng.* **2014**, *10*, 849–856. [[CrossRef](#)]
56. Babić, J.; Šubarić, D.; Ackar, D.; Kovačević, D.; Piližota, V.; Kopjar, M. Preparation and characterization of acetylated tapioca starches. *Dtsch. Lebensm. -Rundsch.* **2007**, *103*, 580–585.
57. Šubarić, D.; Babić, J.; Ačkar, Đ.; Piližota, V.; Kopjar, M.; Ljubas, I.; Ivanovska, S. Effect of galactomannan hydrocolloids on gelatinization and retrogradation of tapioca and corn starch. *Croatian J. Food Sci. Technol.* **2011**, *3*, 26–31.
58. Luo, Y.M.; Niu, L.Y.; Li, D.M.; Xiao, J.H. Synergistic effects of plant protein hydrolysates and xanthan gum on the short- and long-term retrogradation of rice starch. *Int. J. Biol. Macromol.* **2019**, *144*, 967–977. [[CrossRef](#)]
59. Baker, L.A.; Rayas-Duarte, P. Retrogradation of amaranth starch at different storage temperatures and the effects of salt and sugars. *Cereal Chem.* **1998**, *75*, 308–314. [[CrossRef](#)]
60. Wu, Y.; Chen, Z.X.; Li, X.X.; Mei, L. Effect of tea polyphenols on the retrogradation of rice starch. *Food Res. Int.* **2009**, *42*, 221–225. [[CrossRef](#)]
61. Chen, L.; Ren, F.; Zhang, Z.P.; Tong, Q.Y.; Rashed, M.M.A. Effect of pullulan on the short-term and long-term retrogradation of rice starch. *Carbohydr. Polym.* **2015**, *115*, 415–421. [[CrossRef](#)]
62. Chen, L.; Tian, Y.Q.; Tong, Q.Y.; Zhang, Z.P.; Jin, Z.Y. Effect of pullulan on the water distribution, microstructure and textural properties of rice starch gels during cold storage. *Food Chem.* **2016**, *214*, 702–709. [[CrossRef](#)]
63. Tian, Y.Q.; Xu, X.M.; Li, Y.; Jin, Z.Y.; Chen, H.Q.; Wang, H. Effect of β -cyclodextrin on the long-term retrogradation of rice starch. *Eur. Food Res. Technol.* **2009**, *228*, 743–748. [[CrossRef](#)]
64. Yuan, R.C.; Thompson, D.B. Freeze–thaw stability of three waxy maize starch pastes measured by centrifugation and calorimetry. *Cereal Chem.* **1998**, *75*, 571–573. [[CrossRef](#)]
65. Wang, Y.J.; Jane, J. Correlation between glass transition temperature and starch retrogradation in the presence of sugars and maltodextrins. *Cereal Chem.* **1994**, *71*, 527–531.
66. Teng, L.Y.; Chin, N.L.; Yusof, Y.A. Rheological and textural studies of fresh and freeze–thawed native sago starch-sugar gels. II. Comparisons with other starch sources and reheating effects. *Food Hydrocoll.* **2013**, *31*, 156–165. [[CrossRef](#)]
67. Arunyanart, T.; Charoenrein, S. (Eds.) Effects of sucrose, xylitol and konjac glucomannan on stability of frozen rice starch gels. In *Proceedings of the 34th Congress on Science and Technology of Thailand, Bangkok, Thailand, 31 October–2 November 2008*.

-
68. Sprenger, M. New stabilising systems using galactomannans. *Dairy Ind. Int.* **1990**, *55*, 19–20.
 69. Choi, H.M.; Yoo, B. Rheology of Mixed Systems of Sweet Potato Starch and Galactomannans. *Starch Stärke.* **2008**, *60*, 263–269. [[CrossRef](#)]