

DEVELOPMENT OF A QUANTITATIVE DETECTION CARD FOR HEART-TYPE FATTY ACID-BINDING PROTEIN BASED ON BACKGROUND FLUORESCENCE QUENCHING IMMUNE CHROMATOGRAPHY

KARTE ZA KVANTITATIVNU DETEKCIJU SRČANOG TIPA MASNE KISELINE-VEZUJUĆEG PROTEINA ZASNOVANOG NA FLUORESCENTNOJ IMUNOHROMATOGRAFIJI

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Summary

Background: To establish a fast and simple quantitative method for detection of heart-type fatty acid-binding protein (H-FABP) in serum based on a background fluorescence quenching immunochromatographic assay.

Methods: A detection card based on the double-antibody sandwich double-antibody method with background fluorescence quenching was developed for quantitative measurement of H-FABP in serum. The optimal concentrations of control for coating the test and control lines were determined as well as the concentrations of gold-labeled antibodies used in preparing the detection system. The detection method for H-FABP in serum was established and validated using real-world clinical samples.

Results: The optimal concentrations of labeling antibody and coating antibody were 5.0 µg/mL and 1.0 mg/mL, respectively. The test card had a sensitivity of 1.15 ng/mL over a linear concentration range of 0–100 ng/mL. Based on three batches prepared for testing the card, the relative standard deviation (RSD) within batches was less than 15% without a significant difference ($P=0.942$). The detection method was tested against common interfering substances in serum, such as bilirubin, triglyceride and serum anticoagulants ethylenediamine tetraacetic acid (EDTA), heparin,

Kratik sadržaj

Uvod: Cilj ovog rada je bio da se izradi brz i jednostavan kvantitativni metod za detekciju srčanog vezujućeg proteina za masne kiseline (H-FABP) u serumu na osnovu fluorescentnog imunohromatografskog određivanja.

Metode: Detekciona karta se zasniva na duploj antitela sendvič duploj-antitela metodi zasnovanoj na fluorescenciji i razvijenoj za kvantitativno merenje H-FABP u serumu. Određene su optimalne koncentracije za presvlačenje linija testa i kontrola kao i koncentracije antitela označenih zlatom pri pripremi samog sistema. Uspostavljen je detekcioni metod za H-FABP u serumu i validiran primenom priznatih svetskih kliničkih metoda.

Rezultati: Optimalne koncentracije označenih antitela i presvučenih antitela bile su 5,0 µg/mL odnosno 1,0 mg/mL. Test karta imala je osetljivost 1,15 ng/mL u opsegu linearne koncentracije od 0–100 ng/mL. Na osnovu pripremljene tri grupe uzoraka za ispitivanje karte dobijena je relativna standardna devijacija (SD) između grupa manja od 15% bez značajnih razlika ($P = 0,942$). Metod detekcije ispitivan je naspram uobičajenih interferujućih supstanci u serumu, kao što su bilirubin, trigliceridi i serumski antikoagulansi etilendiamin tetrasirćetna kiselina (EDTA), heparin i natrijum citrat i pri tom nije nađena bilo kakva interferencija. Metoda je

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List of abbreviations: FABPs, Fatty acid-binding proteins; H-FABP, Heart-type fatty acid-binding protein; AMI, Acute myocardial infarction; ACS, Acute coronary syndrome; CK-MB, Creatine kinase MB; LTIA, Latex turbidimetric immunoassay.

and sodium citrate, and no significant cross-reaction was detected. The test method was further validated with 50 clinical serum samples, and the test results were comparable with standard reference detection methods with good correlation ($R=0.95$).

Conclusion: Our study presents a new method with strong specificity and sensitivity for the detection of H-FABP in serum, which could promote H-FABP detection in a broad range of applications.

Keywords: heart-type fatty acid binding protein, background fluorescence quenching, immunochromatography

Introduction

Fatty acid-binding proteins (FABPs) are small cytoplasmic proteins that are abundantly expressed in tissues with active fatty acid metabolism. Among the nine FABP families that have been identified, heart-type fatty acid-binding protein (H-FABP) has been the most widely studied. Abundantly expressed in cardiomyocytes, H-FABP is a low molecular weight, 14.5-kDa protein composed of 132 amino acids and constituting 5–15% of cytoplasmic protein in the human heart (1). With a reference concentration of 2.0 ng/mL, H-FABP levels usually remain low in plasma, but H-FABP is released rapidly upon the onset of acute myocardial infarction (AMI) (2). This increase in H-FABP can be detected as early as 30 min following the onset of AMI, and the H-FABP concentration typically peaks within 6–8 hours after symptom onset. H-FABP as a biomarker offers high sensitivity across the full spectrum of acute coronary syndrome (ACS) (3–5). The combination of high sensitivity and early specificity of H-FABP makes it a promising biomarker for early diagnosis and treatment of AMI and ACS that could be superior to the commonly applied Troponin I and Troponin T (initial elevation in blood occurs within 3–6 hours and time-to-peak is 10–48 hours) and creatine kinase MB (CK-MB; initial elevation 3–8 hours and time-to-peak 9–24 hours) (6–9).

Despite establishment of its clinical value a decade ago, available methods for measuring H-FABP concentrations are limited and include radioimmunoassay (RIA) (10), latex turbidimetric immunoassay (LTIA) (11), enzyme-linked immunosorbent assay (ELISA) (12), and colloidal gold immunochromatography (13). Critical drawbacks of these methods have been identified. Specifically, the radioimmunoassay method is limited by its expense and the hazards of preparing and handling radioactive antigen. ELISAs are prone to the introduction of accidental errors during complicated procedures, and the wide application of LTIA is limited by the high cost of testing equipment. A one-step immunochromatography technique (13) for the detection of H-FABP in whole blood samples has been reported with a sensitivity of 6.2 ng/mL, which barely reaches the cut-off value for the diagnosis of AMI. In the present study, we developed and validated a background fluorescence quenching-

zatim validovana na 50 kliničkih serumskih uzoraka, pri čemu su rezultati bili uporedivi sa standardnim detekcionim metodama i dobrom korelacijom ($R = 0,95$).

Zaključak: Naša izučavanja predstavljaju novu metodu sa snažnom specifičnošću i osetljivošću za detekciju H-FABP u serumu, pa se može preporučiti za širu upotrebu.

Ključne reči: srčani tip masne kiseline-vezujući protein, fluorescencija, imunohemotografija

immunochromatographic technique for quantitative detection of H-FABP based on colloidal gold immunochromatography (14). Fluorescence quenching includes static quenching, dynamic quenching, resonance energy transfer and internal filtration effect (IFE). The fluorescent film is fixed on the rubber sheet and will not contact with the solution. The IFE requires that the absorption spectra of the quencher is effectively overlapped with the excitation or emission spectra of the fluorescence film, and that colloidal gold is used as quencher. A uniform fluorescence film was added to the solid phase film of colloidal gold immunochromatography as background fluorescence. The fluorescence intensity ratio was analyzed by detecting the background fluorescence and the test line. The fluorescence intensity ratio was quantitatively determined by detecting the background fluorescence and the test line of the strip. Strong anti-background interference characteristics can eliminate system errors.

Materials and Methods

Materials

H-FABP Standard (RDR-9, 100 μ g) was purchased from Medix Biochemica. Anti-human H-FABP antibody (FABP3 2302 SPTN-5, 5.1 mg/mL) and anti-human H-FABP antibody (FABP3 2304 SPTN-5, 5.0 mg/mL) were obtained from Medix Biochemica. Goat anti-mouse polyclonal antibody (1.0 mg/mL) was purchased from Shanghai Ronghui Biotechnology Co. Bovine serum albumin (lot No. 4529010350) was purchased from Genview. The following reagents were all obtained from Sinopharm Group Chemical Reagent Company: gold chloride (lot No. 20161118), sodium carbonate (lot No. 20140902), sodium bicarbonate (lot No. 20140904), disodium hydrogen phosphate (lot No. 20140902), sodium dihydrogen phosphate (lot No. 20140710), Tween-20 (lot No. 20170916), sodium chloride (lot No. 20140818), sucrose (lot No. 20140922), D-trehalose (dehydrated, lot No. Y18J718031), sodium citrate (lot No. 20161209), ethylene diamine tetraacetic acid (lot No. 20140829), and heparin sodium (lot No. 20170426). Bilirubin (lot No. 20170623) was purchased from Shanghai Rongsheng Bio-phar-

maceutical Co., and whole blood control samples (lot No. 11130581) were purchased from Sichuan Xincheng Biotechnology LLC. Serum matrix (lot No. 0010316) was purchased from Biological Industries. Clinical serum samples were kindly provided by Shanghai Xuhui District Central Hospital.

Solution preparation

The Base1 solution was prepared by dissolving 8.48 g Tris Base, 200 g sucrose, 50 g trehalose, and 10 g bovine serum albumin in 1000 mL pure water. The Base2 solution was prepared by dissolving 2.42 g Tris Base, 1 g PEG20000, 2 g sucrose, 20 g bovine serum albumin, and 3 mL Tween-20 in 1000 mL pure water. The pH was adjusted to 8.0 with 6 mol/L hydrochloric acid. Base1 solution and Base2 solution were mixed in a ratio of 3: 7.

Serial dilutions of H-FABP solution were prepared from 100 µg/mL H-FABP standard solution to obtain 1.0, 2.0, 5.0, 10.0, 20.0, 50.0, and 100.0 ng/mL standard solutions.

Gold colloid-conjugated antibodies

Antibodies were conjugated with gold colloid via the following method. Briefly, auric chloride (AuCl_3) was weighed and dissolved in deionized water to form 0.01% auric chloride solution. Then 100 mL solution was heated to boiling, and 2 mL trisodium citrate solution (1%) was added. The mixture was stirred vigorously until the solution turning claret, followed by heating for another 10 min. Then the solution was cooled down to room temperature. The colloidal gold

solution was adjusted to pH 8.0 with potassium carbonate solution (0.1%). First, 1 mL of gold colloid solution was added to each test tube, and then 3 µL, 5 µL, and 7 µL H-FABP antibody solution was added to respective tubes, followed by addition of 20 µL phosphate-buffered saline (PBS) solution containing 10% bovine serum albumin. The mixture was centrifuged at 9000 rpm and 4 °C for 15 min. The supernatant was aspirated and discarded, and the pellet was suspended in matrix solution added for a total of 200 µL. Finally, 10 µL of the solution was taken and dried in a vacuum for 3 h. The optical density of colloid-conjugated antibody was measured at 520 nm using an ultraviolet spectrophotometer (L6S, Shanghai Instrument Electric Analysis Instrument Company).

Immobilization of antibody onto nitrocellulose membrane

Anti-human H-FABP monoclonal antibody was immobilized along a straight line at a distance of 1.0 cm from the edge of a nitrocellulose membrane (NC). This line served as a detection line for binding of H-FABP/gold colloid-conjugated antibody complexes (Figure 1). The membrane was dried overnight at room temperature.

Assembly of test system

The test system is composed of two parts: the test cup and test panel. A schematic diagram of the plasma panel test is shown in Figure 1. The test cup is where binding occurs between the plasma sample and gold colloid-conjugated antibody complex. The

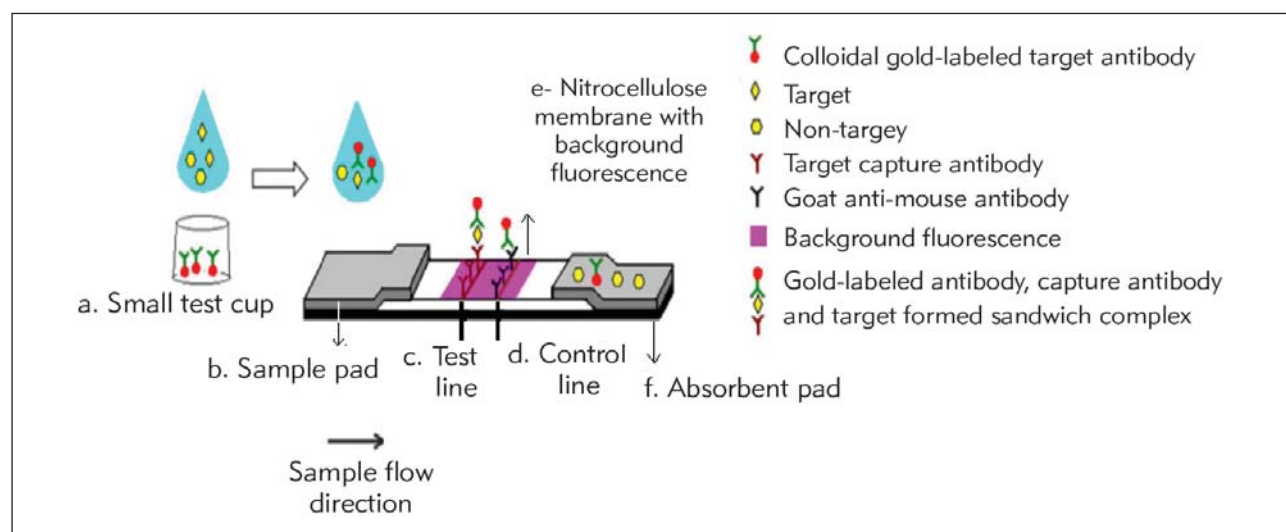


Figure 1 Schematic diagram of plasma panel test for H-FABP.

a. Small cup with bottom coated with gold colloid-conjugated antibodies; b. sample pad to which the sample plasma is added; c. test line (T-line), coated with capture antibody; d. control line (C-line); e. nitrocellulose membrane, on which background fluorescence for T-line and C-line is read; and f. absorbent pad that absorbs excess sample solution.

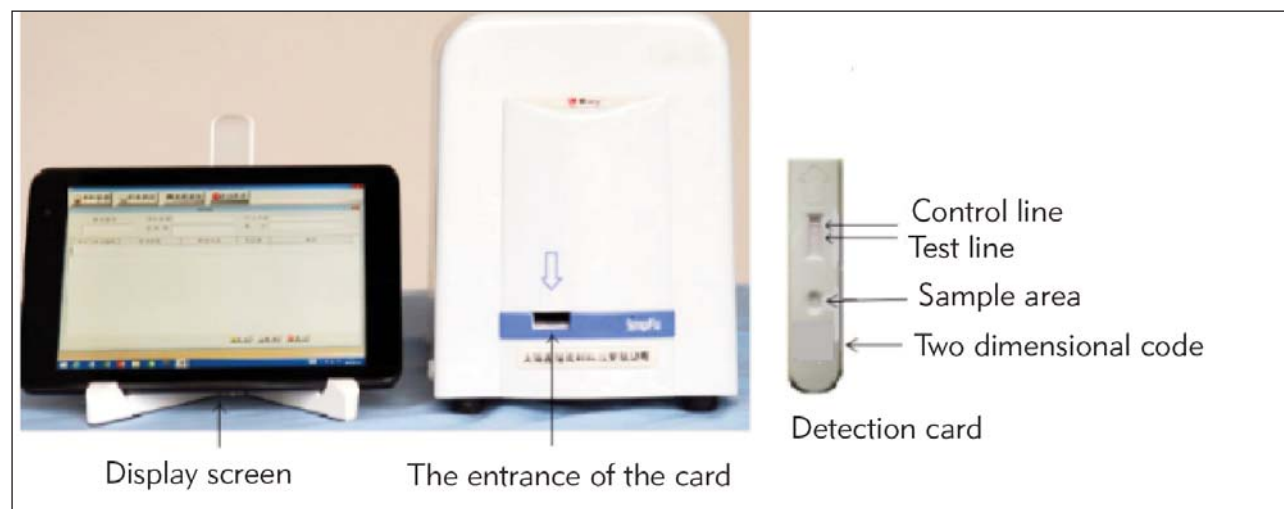


Figure 2 Background fluorescence quenching immunochromatographic analyzer and test card.

combined sample is then added to the test strip in the test card, which has a sample pad, nitrocellulose membrane bearing the test line (T-line) and control line (C-line), and an absorbent pad. The sample pad is used for dropping the sample solution. T-line is coated with capture antibody and C-line is coated with sheep anti-mouse antibody and used to detect the effectiveness of the test strip. Nitrocellulose membrane is used to carry T/C line. The absorbent pad is used to absorb excess sample solution (Figure 1). The sample pad containing gold colloid-conjugated antibody was cut into 21 mm-long and 3.4 mm-wide strips. The absorbent pad was cut into 19 mm-long and 3.4 mm-wide strips. The H-FABP antibodies were immobilized on the nitrocellulose membrane, which was then cut into 65-mm-long and 3.4-mm-wide strips. A sample pad strip was attached to one side of a nitrocellulose membrane strip. A absorbent pad strip was attached to other side of a nitrocellulose membrane strip. Next, a cellulose membrane for absorbing excess plasma and H-FABP/gold colloid-conjugated antibody complexes was attached to the other side of the nitrocellulose membrane strip. A background fluorescence quenching immunochromatographic analyzer (Shanghai Simp Bio-Science Co., Ltd.), as shown in Figure 2, was used for immunochromatographic detection. The analyzer was used for detection of respective analytes conjugated with gold colloid and antibodies on the test card.

Assay procedure for the test panel

An 80- μ L aliquot of plasma sample is applied to the sample cup coated with colloidal gold/antibody complex. After binding of H-FABP in the sample with gold colloid-conjugated antibody complex, a 60- μ L aliquot is dropped into the sample well of the test card. The sample moves along the pad through capillary migration and combines to the NC generating

background fluorescence. The H-FABP/colloidal gold-labeled antibody complex binds with capture antibody immobilized along the T-line to form a double antibody sandwich complex, causing background fluorescence quenching and development of a visible T-line. The C-line serves as the quality control line. The goat anti-mouse antibody combines with the immobilized antibody and develops the C-line. The intensity of background fluorescence is set as F_0 , and the fluorescence intensity of the T-line as F_1 . Then the fluorescence quenching degree (F_0/F_1) is related to the H-FABP concentration in the plasma sample.

Statistical analysis

Statistical Package for the Social Sciences (SPSS, version 17.0, Chicago, IL) was used for all analyses. Data are presented as means \pm standard deviation (SD). Statistical significance was set at $P < 0.05$.

Results

Detection of capture antibody at the T-line

Test cards on which H-FABP antibodies were immobilized from solutions at varying concentrations (0.5, 0.75, 1.0, or 1.5 mg/mL) were prepared to determine the optimal capture concentration. For each respective capture antibody concentration, tests were conducted with varying H-FABP/gold colloid-conjugated antibody complex solutions (0, 1, 5, 10, 25, 50, and 100 ng/mL). For measurement, 60 μ L of complex solution was added to the test card and the F_0/F_1 values were measured after 15 min of chromatography at room temperature. As shown in Figure 3, when the sample cup was coated using 5 μ L gold colloid-conjugated antibody complex and 1 mg/mL capture antibody solution was used for immobilization

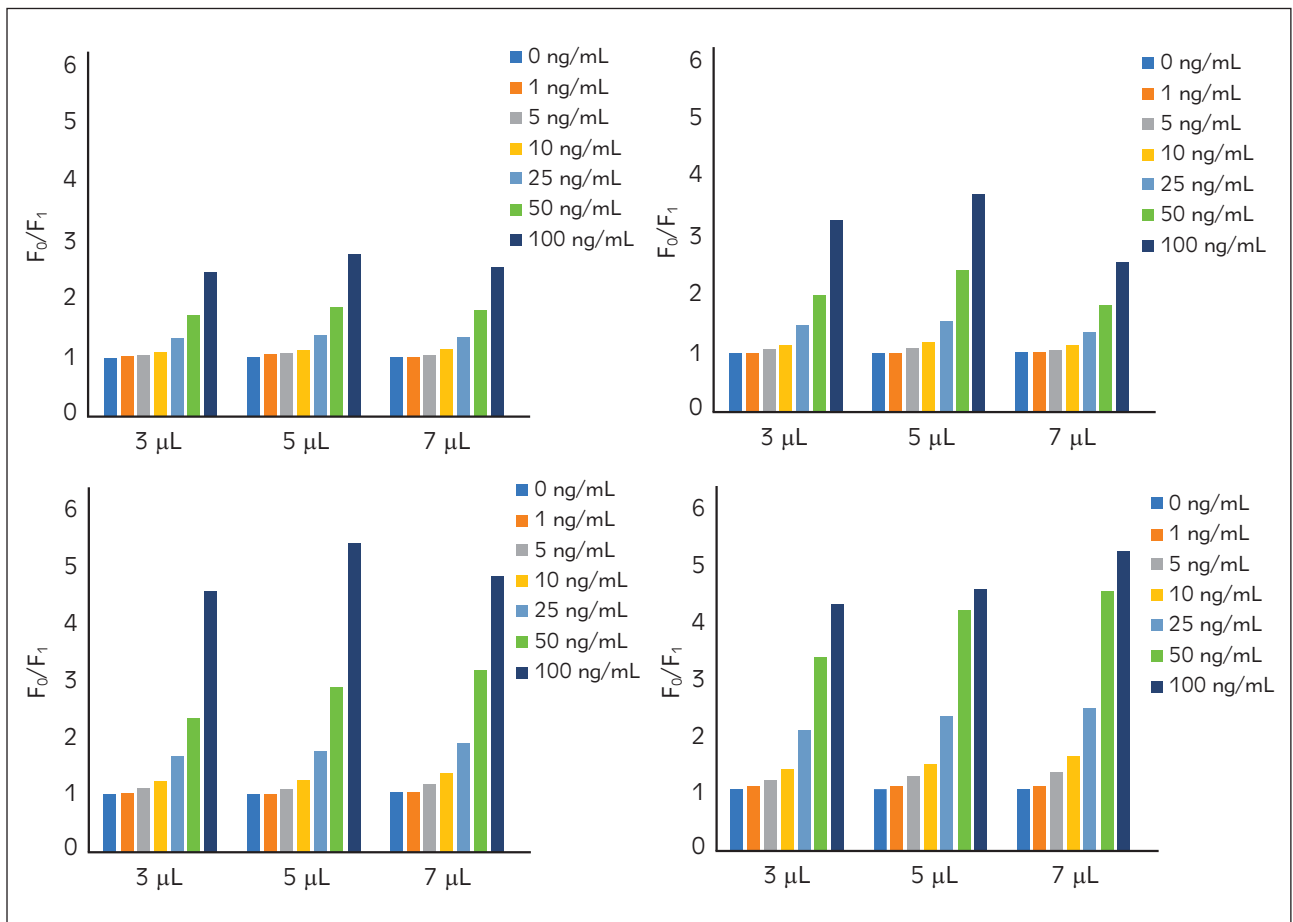


Figure 3 Background fluorescence quenching with varying amounts of immobilized capture antibody on the T-line and a series of H-FABP/gold colloid-conjugated antibody complex concentrations.

H-FABP antibodies were immobilized on test cards from solutions of varying concentrations (0.5, 0.75, 1.0, or 1.5 mg/mL) for testing to determine the optimal capture concentration. Briefly, serial H-FABP/gold colloid-conjugated antibody complex solutions were prepared by mixing 3, 5, or 7 mL of 1.0 mg/mL H-FABP solution with gold colloid-conjugated antibody solution prepared as described in the »Materials and Methods«. The H-FABP capture solution was diluted in PBS to 0.5, 0.75, 1.0, or 1.5 mg/mL. The T-line on the nitrocellulose membrane was marked at density of 0.9 mL/cm. Goat anti-mouse H-FABP antibody (0.2 mg/mL) was immobilized on the C-line at a density of 0.9 mL/cm. After application of the T-line and C-line, the nitrocellulose membrane was dried at room temperature for 8 hours. Finally, 80- μL solutions of different H-FABP/gold colloid-conjugated antibody complex solutions were applied to test cards on which the T-line was prepared by immobilizing varying amounts of capture antibody: 0.5 mg/mL (a), 0.75 mg/mL (b), 1.0 mg/mL (c), and 1.5 mg/mL (d).

on the test card at the T-line, the most significant increasing trend of the F_0/F_1 ratio was observed.

Validation of test method

The test card was validated with serially diluted solutions of H-FABP (1.0, 2.0, 5.0, 10.0, 20.0, 50.0, and 100.0 ng/mL). With the concentration of test solution along the x-axis and F_0/F_1 along the y-axis, the regression equation was fitted to be: $Y=0.0001+0.0311x+1.0007$, with a correlation coefficient = 0.9997, which indicated that the test method provides good linear correlation over an H-FABP concentration range of 0–100.0 ng/mL (Table I). Blank solution was taken as the negative control for

measurement of the minimum detection limit, which was found to be 1.15 ng/mL. Measurement of 5.0, 25.0, and 50.0 ng/mL H-FABP standard solutions showed that the RSDs for each concentration were 10.8%, 6.8%, and 9.2%, respectively, indicating the high repeatability of this assay (Table II).

Variation between batches was measured further using three batches of test cups coated with gold colloid-conjugated antibody complex and test cards with immobilized H-FABP capture antibody. The three batches were validated using H-FABP standard solutions (5.0, 25.0, and 50.0 ng/mL), and the calculated RSDs for each concentration are listed in Table III. The results for the three groups were employed for variance analysis, which yielded a $P=0.942$.

Table I Validation with H-FABP standard solutions: H-FABP concentration in solution vs. F_0/F_1 .

H-FABP concentration (ng/mL)	0	1	5	10	25	50	100
F_0/F_1	1.036±0.01	1.050±0.02	1.142±0.01	1.303±0.06	1.808±0.04	2.951±0.25	5.467±0.95

n=3. Correlation coefficient = 0.9997

Table II Repeatability of the test results.

H-FABP concentration (ng/mL)	F_0/F_1	Actual concentration (ng/mL)	RSD (%)
5.0±0.5	1.1659±0.02	6.03±0.65	10.8
25.0±2.5	1.6861±0.06	21.41±1.46	6.8
50.0±5.0	2.6232±0.13	43.17±2.87	9.2

n=10. No significant difference was detected according to the RSDs for the different concentrations.

Table III Test results for different sample batches.

Batch No.	H-FABP concentration (ng/mL)	F_0/F_1	Actual concentration (ng/mL)	RSD (%)
1	5.0±0.5	1.3074±0.02	4.77±0.39	8.3
	25.0±2.5	2.6489±0.05	22.17±0.58	2.6
	50.0±5.0	4.8288±0.18	45.99±2.02	4.4
2	5.0±0.5	1.3520±0.02	5.56±0.32	5.7
	25.0±2.5	2.6163±0.10	21.80±1.14	5.3
	50.0±5.0	4.5676±0.28	43.11±3.09	7.2
3	5.0±0.5	1.3249±0.04	5.07±0.66	13.0
	25.0±2.5	2.6143±0.02	21.78±0.26	1.2
	50.0±5.0	4.4087±0.30	41.37±3.30	7.9

n=10. No significant difference was detected according to the RSDs for different concentrations.

Table IV Recovery rates in test samples.

F_0/F_1	Added amount (m/ng)	Measured amount (m/ng)	Recovery rate (%)	RSD (%)
1.2309±0.03	8.06	8.49±0.96	105.5	11.5
1.3969±0.02	14.35	13.80±0.64	96.2	4.8
2.009±0.06	28.76	29.94±1.53	104.1	5.2

n=3. No significant difference was detected according to the RSDs for the tested concentrations.

Table V Potential interference of co-present substances in H-FABP solutions.

Interfering substance	H-FABP concentration (ng/mL)	F_0/F_1	Actual H-FABP concentration (ng/mL)	Recovery rate (%)
Bilirubin	5.0±0.5	1.2222	5.01	95.3
	25.0±2.5	1.9630	24.34	97.3
	50.0±5.0	3.0209	49.48	101.3
Hemoglobin	5.0±0.5	1.1620	3.22	61.3
	25.0±2.5	1.7103	18.04	72.1
	50.0±5.0	2.5881	39.36	80.6
Triglyceride	5.0±0.5	1.2235	5.05	96.1
	25.0±2.5	2.1023	27.73	110.8
	50.0±5.0	3.3403	56.85	116.4
Heparin sodium	5.0±0.5	1.2689	6.33	109.5
	25.0±2.5	1.9630	31.86	117.3
	50.0±5.0	4.0844	83.85	111.2
EDTA	5.0±0.5	1.2594	6.07	115.6
	25.0±2.5	2.0646	26.82	107.2
	50.0±5.0	3.0637	50.47	103.3
Sodium citrate	5.0±0.5	1.1998	4.35	82.8
	25.0±2.5	1.9165	23.18	92.6
	50.0±5.0	3.0923	51.13	104.7

n=3. Recovery rates for substances other than hemoglobin ranged from 80%–120%, indicating no significant effect on H-FABP measurement.

The recovery rate was also validated for the test system. Briefly, 20- μ L, 30- μ L, and 50- μ L aliquots of H-FABP solution (100 ng/mL) were added to 5.0 ng/mL, 10.0 ng/mL, and 25.0 ng/mL standard solutions to prepare test solutions of high, medium, and low concentrations, respectively. The F_0/F_1 ratio was measured for each concentration, and the results fell within the range of 85%–115%, indicating that the detection method was accurate and reliable (Table IV).

Interference by other substances present in serum

Possible interference by various substances was investigated using plasma control samples containing H-FABP at concentrations of 5.0, 25.0, and 50.0 ng/mL as well as bilirubin (2 mg/mL), hemoglobin (10 mg/mL), triglyceride (40 mg/mL), heparin sodium (100 mg/mL), EDTA (1.5 mg/mL), or sodium citrate (0.1 mg/mL). All recovery rates for test solutions fell within the range of 80%–120%, except those for plasma samples containing hemoglobin, suggesting that the test system is inappropriate to hemolysis-free blood samples (Table V).

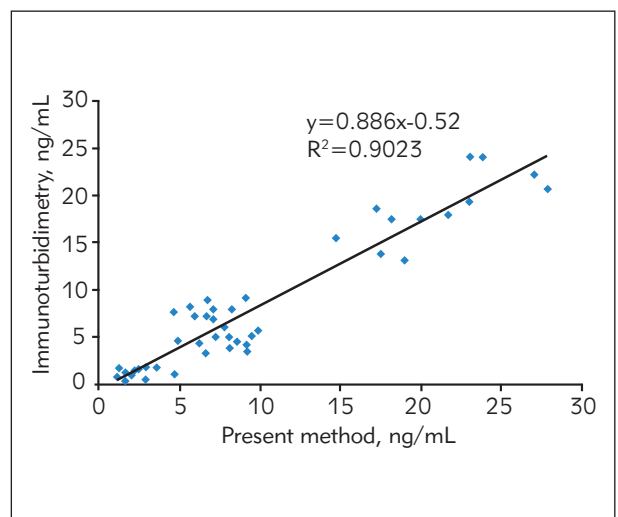


Figure 4 Linear correlation between test system and immunoturbidimetry results.

Fifty clinical plasma samples were tested with the test system and immunoturbidimetry, and the results were comparable ($t=1.985$ and $P>0.05$, paired t test). The linear regression equation was calculated as: $Y=0.886X-0.52$ (correlation coefficient $r=0.95$, $P<0.01$).

Comparison of plasma sample test with immunoturbidimetry

To examine the test system with real-world samples, we applied the panel test for 50 clinical serum samples and compared with the results obtained by immunoturbidimetry. The results in *Figure 4* indicate that measurements obtained with the two methods were comparable ($t=1.985$, $P>0.05$).

Discussion

A new method for measuring the H-FABP concentration in plasma samples was developed based on the principle of background fluorescence quenching immune chromatography and validated. The developed assay features a wide linear detection range, high repeatability, a low cut-off value, and low inter-batch variation. The whole process is simple and requires less time than conventional methods through the development of a visible T-line on the test pad within 15 min. The appearance of the C-line verifies the validity of test. The degree of background fluorescence quenching is measured by F_0/F_1 , which reflects the H-FABP concentration.

The clinical value of H-FABP has been long recognized (15). Its early release following myocardial ischemia and early detection within 30 min of onset make it a highly effective biomarker for the diagnosis and management of ACS. A gold immunochromatographic assay (GICA) has been developed and applied in diagnostic tests; however, it is a qualitative test in the majority of applications. In the present study, we established a novel, quantitative GICA for H-FABP based on fluorescence quenching and nitrocellulose membrane background signal. The test system consisting the test card and test cup was prepared, and the performance of the background fluorescence quenching immunochromatographic assay was assessed. High reproducibility was confirmed using plasma samples, and no interference for the detection of H-FABP was observed in samples containing bilirubin, ascorbic acid, human serum albumin, and human immunoglobulin.

The principle of reaction for the developed assay is quite simple. A plasma sample is first added to the test cup to bind with colloidal gold antibody complex. Then the treated solution is added to the test card. The presence of H-FABP is detected with the appearance of a red T-line on the nitrocellulose membrane. Unlike existing immunochromatographic assays that detect the coating or labeling fluorescence, colloidal gold, or capture antibodies, this background fluores-

cence quenching immunochromatographic assay takes advantage of background fluorescein signal on the NC membrane and collects the total fluorescence signal at the C-line, as quality control, and at the T-line. The method is sensitive enough to detect any fluctuation in fluorescence intensity compared with the baseline fluorescence to ensure accurate measurement. Therefore, the assay offers greater tolerance of potential changes in the test system. Moreover, greater sensitivity for the detection of H-FABP has been achieved with extension of the quantitation range to 0–100 ng/mL. In addition, with a minimum detection limit of 1.15 ng/mL. The new method has high sensitivity. Importantly, the new test system does not require the additional purchase of a special device, as existing assays do (15), and the detection period is reduced from 27 min to 15 min (16, 17). A one-step immunotest for H-FABP has been reported (18), but the effective detection limit was relatively higher (7 ng/mL) compared with the current system.

Limitations of the present study should be recognized. First, different types of clinical samples, such as whole blood, need to be tested with large sample sizes to further validate the clinical value of the developed assay. Second, we only validated the test system with immunoturbidimetry. However, given that other test measures, such as ELISA, have been widely applied in clinical practice, further comparison may be necessary to distinguish the advantages and disadvantages of the developed assay compared with other measures. Third, the background signal of the plasma sample tested in the study was heterogeneous, which may contribute to individual variation. Moreover, whether the assay can be used to accurately track the dynamic change of H-FABP levels observed in AMI patients remains to be studied for real-world cases.

In summary, background fluorescence quenching immunochromatography can detect the site of H-FABP fast, and the detection instrument is small and convenient, with low cost. It will be popularized in the community and primary medical institution.

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Conflict of interest statement

The authors stated that they have no conflicts of interest regarding the publication of this article.

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