Leptomeningeal contrast enhancement and blood-CSF barrier dysfunction in aseptic meningitis

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ABSTRACT

Objective: To investigate the blood-CSF barrier (BCSFB) dysfunction in aseptic meningitis.

Methods: In our case series of 14 patients with acute aseptic meningitis, we compared MRI characteristics with CSF findings.

Results: Contrast enhancement in the sulcal space in a leptomeningeal pattern was visualized in 7 patients with BCSFB dysfunction categorized as moderate to severe as evidenced by the CSF/serum albumin ratio (Q_{alb}) but was not present in those with mild or no barrier disturbance (p=0.001). The Q_{alb} as a marker for the leakiness of the BCSFB and, more indirectly, of the blood-brain barrier (BBB) was positively correlated with the incidence of leptomeningeal contrast enhancement seen on postcontrast fluid-attenuated inversion recovery (FLAIR) MRI (p=0.003). Patients with a more pronounced brain barrier dysfunction recovered more slowly and stayed longer in the hospital.

Conclusions: The severity of meningeal BBB disturbance can be estimated on postcontrast FLAIR MRI, which may be of diagnostic value in patients with aseptic meningitis. *Neurol Neuroimmunol Neuroinflamm* 2015;2:e164; doi: 10.1212/NXI.000000000000164

GLOSSARY

 $\textbf{BBB} = \text{blood-brain barrier}; \textbf{BCSFB} = \text{blood-CSF barrier}; \textbf{CMV} = \text{cytomegalovirus}; \textbf{FLAIR} = \text{fluid-attenuated inversion recovery}; \textbf{HSV} = \text{herpes simplex virus}; \textbf{Q}_{\textbf{alb}} = \text{CSF/serum albumin ratio}; \textbf{Q}_{\textbf{lgG}} = \text{CSF/serum IgG ratio}; \textbf{VZV} = \text{varicella-zoster virus}.$

The term "aseptic meningitis" encompasses nonbacterial inflammation syndromes of the meninges predominantly caused by viral infections. All viral pathogens share a common step in their pathophysiology when traversing brain barriers followed by penetration into the CNS. In their function to maintain CNS homeostasis, blood-brain barrier (BBB) endothelial cells also restrict the entry of both pathogens and immune cells. Meningeal CNS microvessels on the surface of the brain also establish a barrier with somewhat different morphology in order to separate the CSF-drained leptomeningeal space from the blood. In viral meningitis, disruption of the brain barriers occurs mostly via indirect effects of the virus on the immune system, including induction of chemokines and cytokines and leukocyte homing, among others. Disintegration of the brain barriers, in turn, can lead to efficient viral clearance by enhanced infiltration of immune cells into the CNS but may also cause a fatal clinical course with propagation to encephalitic involvement and entry of deleterious serum molecules into the brain parenchyma.

The diagnosis of meningitis is based on the combination of typical clinical symptoms and positive CSF findings. Whereas an increase in CSF leukocyte count reflects the inflammatory reaction with transmigration of immune cells into the CSF, the CSF/serum albumin ratio (Q_{alb}) indicates a leakage of blood-borne proteins due to blood-CSF barrier (BCSFB) dysfunction and,

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more indirectly, BBB dysfunction. MRI is commonly used to evaluate complications such as encephalitic involvement, hydrocephalus, or cerebral vasculitis with subsequent ischemia. However, recent work has highlighted the role of postcontrast fluid-attenuated inversion recovery (FLAIR) imaging in the detection of leptomeningeal pathologies, including meningitis or meningeal carcinomatosis.⁴ Mainly due to the suppression of CSF signal, postcontrast FLAIR images show a higher sensitivity in delineation of meningeal pathologies.⁵

In our study, we sought to compare CSF findings with postcontrast FLAIR findings in patients with aseptic meningitis.

METHODS In our case series, we analyzed 14 patients from our MRI and clinical database admitted between 2011 and 2014 with CSF-confirmed diagnosis of aseptic meningitis.

Clinical data including age, sex, presenting symptoms and clinical course, duration of hospitalization, medication, and neurologic status at discharge were collected in all patients. Routine CSF analyses included leukocyte count, CSF protein concentration, Q_{alb} , CSF/serum IgG ratio (Q_{IgG}), CSF culture, CSF serology (herpes simplex virus [HSV] IgG/IgM, varicella-zoster virus

0.1000 | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 80% | 60% | 40% | 20% | 60% | 40% | 20% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40% | 40%

Increasing albumin quotients reflect increasing blood-brain barrier dysfunction (x-axis). The cutoff value for brain barrier dysfunction depends on the patient's age and is therefore not marked. On the y-axis, the blue line separates blood-borne IgG (below the line) and intrathecal IgG synthesis (above the line). In our patient cohort, the CSF/serum albumin ratio (Q_{alb}) was significantly higher in patients with leptomeningeal enhancement on postcontrast fluid-attenuated inversion recovery (FLAIR) imaging (red) than in those without leptomeningeal enhancement (green). $Q_{lgG} = CSF/serum \, lgG \, ratio$.

[VZV] IgG/IgM, measles IgG/IgM, mumps IgG/IgM, Borreliaburgdorferi IgG/IgM, syphilis, HIV), and PCR analysis for neurotropic viruses (HSV 1/2, VZV, CMV, Epstein-Barr virus, enterovirus, human herpesvirus 6). Brain barrier dysfunction and intrathecal synthesis of IgG were determined based on the method of Reiber. The resulting data are presented in Reibergrams (figure 1), discriminating the Q_{IgG} from the IgG/serum albumin ratio with hyperbolic reference range. The upper limit of the reference range for an intact brain barrier function is age-dependent and was calculated for each patient according to the formula $Q_{alb} < (4 + age/15) \times 10^3$. Parain barrier dysfunction was classified as mild $(Q_{alb} < 10 \times 10^3)$, moderate $(Q_{alb} 10-20 \times 10^3)$, or severe $(Q_{alb} > 20 \times 10^3)$.

Clinical data and CSF studies are given in the table.

MRI studies were performed on a 3T MRI system (MAGNETOM Skyra; Siemens Healthcare, Erlangen, Germany) using a standardized protocol including T2-weighted FLAIR echo planar diffusion-weighted imaging including apparent diffusion coefficient calculations and subsequent contrast-enhanced T1-weighted and FLAIR sequences. The MRI data were analyzed with a structured reporting scheme independently by 2 reviewers unaware of clinical detail and experienced with diagnostic and research studies using postcontrast FLAIR MRI.

Statistical analysis was performed using SPSS version 22.0.0.0 (IBM, Armonk, NY). Group comparisons between patients with and without leptomeningeal postcontrast FLAIR hyperintensity regarding the severity of brain barrier dysfunction and CSF protein concentration, $Q_{\rm alb}$, and $Q_{\rm IgG}$ were calculated by Mann-Whitney U test and 2-tailed Student t test, as appropriate. A p value < 0.05 was considered to be statistically significant.

Standard protocol approvals, registrations, and patient consents. This study was approved by the local institutional review board (Medizinische Ethikkommission II der Medizinischen Fakultät Mannheim, University of Heidelberg) and performed in accordance with the ethical standards of the 1964 Declaration of Helsinki and its later amendments. Patient consent was not required by our institutional review board due to the retrospective nature of the study and the lack of patient interaction.

RESULTS During the analyzed period, 15 patients with a diagnosis of aseptic meningitis were examined with a standardized 3T MRI protocol including post-contrast FLAIR sequences and were potentially eligible for the study. One patient had to be excluded due to incomplete CSF workup (Q_{alb} missing).

Fourteen patients (8 men, age range 18–58 years, all immunocompetent) were included in our study. All patients presented with moderate to severe new-onset headache; 8/14 had fever at presentation. In the clinical course, 2 patients developed cranial nerve palsy (abducens nerve in 1, facial and vestibulocochlear nerve in 1). One patient had new-onset epileptic seizures (table).

On the initial MRI, no parenchymal signal alterations were detected in precontrast sequences including native FLAIR imaging in any of the patients. On post-contrast FLAIR images, 7/14 patients showed leptomeningeal and/or sulcal contrast enhancement. In 4/7, a distinct leptomeningeal enhancement was present along each cerebral lobe of both hemispheres

Table Clinical data and CSF analyses of the included patients							
Age, y, sex	Etiology	Postcontrast FLAIR	CSF WBC count, n/μL	CSF protein, mg/L	Q_{alb}	Q_{lgG}	Brain barrier dysfunction
27, F	Viral, CMV	+	188	1,646.6	32.57	17.63	Severe
18, M	ND	+	436	1,526.9	26.28	13.22	Severe
58, F	ND	+	97	665.8	20.88	11.95	Severe
43, M	ND	+	27	586.0	13	6.5	Moderate
54, M	Viral, VZV	+	336	938.4	16.66	10.49	Moderate
31, M	Viral, HSV2	+	491	879.6	15.2	7.32	Moderate
19, F	ND	+	120	1,031.6	15.1	8.6	Moderate
52, M	ND	_	98	1,059.7	16.4	8.19	Moderate
20, F	ND	_	170	425.4	6.9	3.71	Mild
37, F	Viral, enterovirus	_	66	541.1	9.1	4.07	Mild
39, M	ND	_	48	468.6	7.49	3.9	Mild
18, M	Viral, enterovirus	_	426	396.0	6.58	3.42	Mild
26, F	Viral, enterovirus	_	366	493.6	8.78	4.41	Mild
42, M	Viral, enterovirus	_	21	409.2	6.36	3.06	None

Abbreviations: CMV = cytomegalovirus; FLAIR = fluid-attenuated inversion recovery; HSV = herpes simplex virus; ND = not detectable; Q_{alb} = CSF/serum albumin ratio; Q_{lgG} = CSF/serum IgG ratio; VZV = varicella-zoster virus; WBC = white blood cell. Brain barrier dysfunction was classified as mild (Q_{alb} <10 × 10³), moderate (Q_{alb} 10-20 × 10³) or severe (Q_{alb} >20 × 10³).

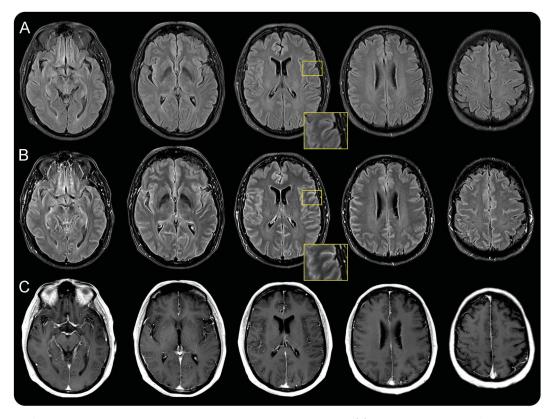
and the cerebellum; in 3/7, contrast enhancement was more subtle and confined to the supratentorial leptomeninges. An example of ubiquitous contrast enhancement of the meninges in a patient with meningitis caused by VZV is given in figure 2. On contrast-enhanced T1-weighted images, no parenchymal or meningeal signal abnormalities were noted.

CSF analyses were performed a mean of 2.6 days (±2.1 days, range 0-8 days) prior to MRI and showed lymphocytic pleocytosis (mean 206 cells/µL, range 21-491) indicating aseptic meningitis in all patients. There was no evidence of noninfectious aseptic meningitis (postvaccinal, drugs, systemic inflammatory disease, neoplastic disorder) in any of the patients. A specific pathogen was detected in 7 patients. Of these, the 3 patients with leptomeningeal enhancement were infected by Herpesviridae (VZV, HSV2, CMV), whereas the 4 patients without leptomeningeal enhancement were positive for enterovirus RNA. In relation to Q_{IgG}, none of the patients had intrathecal synthesis of IgG antibodies. In 13/14 patients, Q_{alb} accounted for a dysfunction of the brain barriers that was classified as severe in 3/13, moderate in 5/13, and mild in 5/13. Patients with leptomeningeal enhancement on postcontrast FLAIR imaging showed significantly higher values of brain barrier dysfunction (p = 0.001). Leptomeningeal enhancement on postcontrast FLAIR imaging was present in all but 1 patient with moderate or severe brain barrier dysfunction, whereas none of the patients with mild or no disturbance of the brain barriers showed contrast enhancement of the leptomeninges. The presence of postcontrast leptomeningeal enhancement was associated with higher Q_{alb} (p=0.003) and Q_{IgG} (p=0.002) as well as a higher CSF protein concentration (p=0.016). Leukocyte count did not differ between patients with or without postcontrast leptomeningeal enhancement (p=0.449). No post–lumbar puncture complications occurred.

Clinical outcome was favorable, with resolution of symptoms within 2 weeks in all but 1 patient. This patient showed marked leptomeningeal enhancement and moderate brain barrier disturbance on CSF analysis and had several complications, including sixth cranial nerve palsy, undulating leukocyte count >100/µL over 4 weeks, and increase of intracranial pressure requiring continuous lumbar CSF drainage for 7 days. In patients without leptomeningeal contrast enhancement, symptoms regressed more quickly than in those showing postcontrast FLAIR signal enhancement. As a result, the average length of hospital stay was longer in patients with leptomeningeal contrast enhancement (mean 19 days vs 7 days in those without enhancing leptomeninges).

DISCUSSION Postcontrast FLAIR MRI sequences have been shown to be valuable in the detection and evaluation of different leptomeningeal diseases, including subarachnoid hemorrhage, meningeosis, and infectious meningitis. The nulling of the CSF signal due to the inversion time facilitates an optimized delineation of hyperintense pathology adjacent

Figure 2 Exemplary MRI of a patient with contrast enhancement on postcontrast FLAIR



MRI of a 54-year-old patient with viral meningitis caused by varicella-zoster virus. (A) Unenhanced precontrast fluid-attenuated inversion recovery (FLAIR) images. On postcontrast FLAIR images (B), sulcal contrast enhancement and a fine hyperintense lining can be seen, probably involving the leptomeninges in the absence of obvious white or gray matter lesions on both FLAIR and postcontrast T1-weighted MRI (C). The magnified inserts highlight sulcal enhancement.

to the CSF in the sulcal and leptomeningeal space while the T1 shortening in FLAIR sequences is responsible for the highly sensitive delineation of pathologic contrast enhancement following application of gadolinium.¹⁰ In a large cohort of infants with bacterial meningitis, contrast enhancement of the leptomeninges was the most common pathologic finding on MRI.11 From a pathophysiologic point of view, leakage from damaged pial vessels leading to focal extravasation of contrast agent into the CSF adjacent to the brain is the likely cause for leptomeningeal enhancement on FLAIR images. 12 Contrast enhancement of the leptomeninges on postcontrast FLAIR sequences may thus be a surrogate for dysfunction of the BCSFB and, more indirectly, a marker of BBB dysfunction. In contrast to patients with ischemic stroke¹³ or cerebral amyloid angiopathy,¹⁴ in whom localized brain barrier breakdown can be observed, in patients with meningitis a more generalized pattern of contrast enhancement was seen, involving both hemispheres and also the infratentorial space in some patients.

We substantiate this hypothesis by correlating the CSF findings with the presence of leptomeningeal enhancement in patients with aseptic meningitis. Contrast enhancement of the meninges occurred only in patients with moderate to severe dysfunction of the brain barriers and not in those with mild or no brain barrier disturbance. Moreover, we demonstrate a positive correlation between the Qalb as a quantitative marker for brain barrier dysfunction and the occurrence of leptomeningeal enhancement. Partly in line with these findings, an association of log10transformed CSF protein and a leptomeningeal enhancement score has been reported in patients with infectious meningitis.15 However, log10-transformed CSF protein is only an approximate value for brain barrier disturbance, as it is not corrected for serum protein content. The positive correlation may thus be primarily based on the inclusion of patients with bacterial meningitis and marked CSF protein elevation.

Severe damage to the brain barriers, particularly as seen in bacterial meningitis, may predispose to complications and poor prognosis. Accordingly, a positive correlation between the extent of leptomeningeal enhancement on postcontrast FLAIR images and the occurrence of complications has been demonstrated in a study of 43 patients with meningitis of various etiologies. ¹⁵ However, complications affected only 3 patients with bacterial meningitis and 1 patient

with fungal meningitis, while the clinical course of patients with viral meningitis was benign overall.

Our study focused on patients with aseptic meningitis, characterized by mostly mild to moderate brain barrier dysfunction. Of interest, although only 1 patient in our cohort had a complicated clinical course, the presence of leptomeningeal enhancement still accounted for a delayed resolution of clinical symptoms and a longer hospital stay. On the other hand, a mild increase in brain barrier permeability in patients with viral meningitis may be a precondition for lymphocyte infiltration and subsequent effective viral clearance.16 Furthermore, the extent of brain barrier dysfunction may vary depending on the specific viral agent. Viral infections are characterized by individual patterns of chemokine secretion that promote vascular permeability.¹⁶ In our cohort, 3 patients with leptomeningeal enhancement were infected by Herpesviridae (VZV, HSV2, CMV), coated double-stranded DNA viruses, whereas 4 patients without leptomeningeal enhancement were positive for enteroviruses, uncoated double-stranded RNA viruses. Although data on HSV2 are lacking, HSV1 has been shown to affect the BBB mainly via increased MMP2 and MMP9 activity, leading to collagen type IV degradation.¹⁷ Instead, in vitro studies have shown that infection of choroid plexus cells with echovirus 30, a member of the enterovirus family, leads to enhanced secretion of CXCL1-3, chemoattractants for T lymphocytes. 18 This is motivation to study the potential association between viral agents and the occurrence of leptomeningeal enhancement in future cohorts with more patients.

Because contrast-enhanced MRI was performed after CSF analysis, it is theoretically possible that leptomeningeal enhancement was caused or enhanced by the lumbar puncture. However, meningeal enhancement seems to occur only in the case of post–lumbar puncture headache or intracranial hypotension, 19 which did not occur in any of our patients.

In patients with aseptic meningitis, leptomeningeal enhancement on postcontrast FLAIR images is closely related to the extent of leakage of bloodborne proteins into the CSF. Both leptomeningeal enhancement and $Q_{\rm alb}$ can thus serve as surrogate markers for brain barrier dysfunction.

AUTHOR CONTRIBUTIONS

Angelika Alonso: conceptualization of the study, data acquisition, analysis and interpretation of the data, drafting and revising the manuscript. Philipp Eisele: data acquisition, analysis and interpretation of the data, revising the manuscript. Anne D. Ebert: analysis and interpretation of the data, revising the manuscript, statistical analysis. Martin Griebe: acquisition, analysis, and interpretation of the data, revising the manuscript. Britta Engelhardt: analysis and interpretation of the data, revising the manuscript. Kristina Szabo: acquisition, analysis, and interpretation of the data, revising the manuscript. Michael G. Hennerici: analysis and interpretation of the data, revising the manuscript. Achim Gass: conceptualization of the study, data acquisition, analysis and interpretation of the data, drafting and revising the manuscript.

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DISCLOSURE

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