Shruti Gupta⁴ David E. Leaf⁴

¹Division of Hematology, Massachusetts General Hospital, Harvard Medical School, Boston, MA, ²Department of Internal Medicine, Hackensack Meridian School of Medicine at Seton Hall, Nutley, NJ, ³Department of Internal Medicine, Heart and Vascular Hospital, Hackensack Meridian Health Hackensack University Medical Center, Hackensack, NJ and ⁴Division of Renal Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, USA. E-mail: rkarp-leaf@partners.org

[†]These authors contributed equally to this work.

Keywords: blood groups, ABO, COVID-19

First published online 30 July 2020 doi: 10.1111/bjh.16984

References

- Anstee DJ. The relationship between blood groups and disease. Blood. 2010;115:4635–43.
- Cooling L. Blood groups in infection and host susceptibility. *Clin Microbiol Rev.* 2015;28:801–70.

- Zhao JYY, Huang H, Li D, Gu D, Lu X, Zhang Z, et al. Relationship between the ABO blood group and the COVID-19 susceptibility. *MedRxiv*. 2020 [Epub ahead of print]. DOI: https://doi.org/10.1101/2020.03.11. 20031096
- Ellinghaus D, Degenhardt F, Bujanda L, Buti, M, Albillos, A, Invernizzi, P, et al. Genomewide Association Study of severe Covid-19 with respiratory failure. N Engl J Med. 2020 [Online ahead of print]. DOI: https://doi.org/ 10.1056/NEJMoa2020283
- Cheng Y, Cheng G, Chui CH, Lau FY, Chan PK, Ng MH, et al. ABO blood group and susceptibility to severe acute respiratory syndrome. *JAMA*. 2005;293:1450–1.
- Guillon P, Clement M, Sebille V, Rivain JG, Chou CF, Ruvoën-Clouet N, et al. Inhibition of the interaction between the SARS-CoV spike protein and its cellular receptor by anti-histo-blood group antibodies. *Glycobiology*. 2008;18:1085–93.
- Garratty G, Glynn SA, McEntire R; Retrovirus Epidemiology Donor Study. ABO and Rh(D) phenotype frequencies of different racial/ethnic groups in the United States. *Transfusion*. 2004;44:703–6.
- O'Sullivan JM, Ward S, Fogarty H, O'Donnell JS. More on 'association between ABO blood groups and risk of SARS-CoV-2 pneumonia'. Br J Haematol. 2020;190:27–8.
- Reilly JP, Meyer NJ, Shashaty MGS, Feng R, Lanken PN, Gallop R, et al. ABO blood type A is associated with increased risk of ARDS in whites following both major trauma and severe sepsis. *Chest.* 2014;145:753–61.
- Lee SJ, Grobe JE, Tiro JA. Assessing race and ethnicity data quality across cancer registries and EMRs in two hospitals. J Am Med Inform Assoc. 2016;23:627–34.

Concerns about how to use established minimal residual disease monitoring in the treatment of *NPM1*-mutant acute myeloid leukaemia (AML) following reduced intensity chemotherapy protocols for AML given as a result of the COVID-19 pandemic

In view of the coronavirus disease 2019 (COVID-19) pandemic and the predicted risk of severe infection in immunocompromised patients, chemotherapy protocols for patients with acute myeloid leukaemia (AML) have been modified in some patients to newer, less myelosuppressive regimens than standard induction chemotherapy. However, the modifications to treatment have occurred at such a considerable pace, due to the urgency of the pandemic, that optimal time points for measuring minimal residual disease (MRD) to assess disease response and monitor for relapse have not yet been established for the new regimens. Thus, decisions about duration of therapy and appropriate time points to intensify therapy prove very challenging.

The combination of the B-cell lymphoma 2 (BCL-2) inhibitor venetoclax and the hypomethylating agent azacitidine (Ven-Aza) has recently been introduced as a treatment option for patients with AML during the COVID-19

e208

pandemic, instead of the standard more intensive chemotherapy regimen of daunorubicin and cytarabine. It has been approved by the National Institute for Health and Care Excellence¹ and was introduced in our institution on the 19 March 2020. The use of this combination of drugs in AML is based on evidence that it produces high rates of rapid and durable responses for patients who were not eligible for intensive chemotherapy.² In particular, AML with nucleophosmin-1 (NPM1) mutations is shown to be particularly responsive to this combination of treatment.^{3,4} Moreover, Ven-Aza can be used to treat persistent or rising NPM1 MRD levels after intensive induction chemotherapy.⁵ This combination of drugs is also well tolerated^{3,6} and has a lower rate of death than that expected with induction chemotherapy,⁷ although to date there has not been a randomised trial to compare Ven-Aza directly with standard induction chemotherapy.

© 2020 British Society for Haematology and John Wiley & Sons Ltd British Journal of Haematology, 2020, **190**, e181–e232

(A) Bone Marrow MRD



(B)

Fig 1. *NPM1* MRD levels in (A) bone marrow MRD and (B) peripheral blood MRD. Shows the *NPM1* transcripts by RT-qPCR in relation to *ABL1* transcripts at each date that the sample was taken. (A) Shows transcript levels in the bone marrow and (B) shows transcript levels in the peripheral blood. *NPM1* transcript levels are depicted as triangles and the sensitivity of the assay is depicted as squares. The sensitivity of the assay shows the lower limit of detection of transcript levels, and so *NPM1* levels below the sensitivity level of the assay are considered *NPM1* MRD negative. MRD levels are shown on the logarithmic scale on the y-axis. The x-axis (not drawn to scale) shows the dates of the samples and the timing of diagnosis and chemotherapy cycles below the dates. RT-qPCR, reverse transcriptase quantitative polymerase chain reaction; MRD, minimal residual disease.

However, reducing the intensity of chemotherapy now comes with new challenges, as it has to be applied rapidly to well-established protocols of MRD monitoring. This is illustrated in the case below of a 40-year-old male patient with *NPM1*-mutant AML. The *NPM1* transcript level, as an MRD marker, is well established, particularly its level in peripheral blood, which has prognostic significance.^{8,9}

This patient presented to our institution on the 13 February 2020 with AML. His marrow showed 30% blasts which were $CD34^{-}CD33^{+}CD13^{+}HLADR^{+}CD117^{+}CD38^{+}MPO^{+}$ by flow cytometry and that had a normal karyotype. Molecular typing showed that the marrow was positive for the *NPM1* Type A mutation, and negative for the fms related receptor tyrosine kinase 3 (*FLT3*) internal tandem duplication (ITD) and the *FLT3* D835/I836 variant.

He was initially treated with daunorubicin, cytarabine and gemtuzumab ozogamicin (Mylotarg) chemotherapy, which was complicated by a difficult course with haemophagocytic lymphohistiocytosis (HLH) and possible COVID-19 infection, although several nucleic acid tests for COVID-19 were negative. He was admitted to intensive care and treated with the interleukin 1 receptor antagonist anakinra, as described previously.¹⁰ He made a good recovery and was in complete morphological remission following regeneration of his blood counts. MRD using *NPM1* transcript levels was measured

© 2020 British Society for Haematology and John Wiley & Sons Ltd British Journal of Haematology, 2020, **190**, e181–e232 using the reverse transcription quantitative polymerase chain reaction assay (RT-qPCR) comparing it to the reference ABL proto-oncogene 1, non-receptor tyrosine kinase (*ABL1*) transcript levels as described by the UK National Cancer Research Institute AML Working Group.⁸ Following induction chemotherapy *NPM1* transcript levels in the bone marrow were positive at 3×10^{-4} (sensitivity level of assay at 2.67×10^{-7}).

Peripheral Blood MRD

As a result of the COVID-19 pandemic, and the fact that this patient had a very serious complication during the intensive induction chemotherapy, this patient proceeded to cycle 2 with Ven-Aza combination therapy. This was uneventful and he was in complete morphological remission after cycle 2 with *NPM1* mutation levels in the bone marrow at 5.98×10^{-5} (sensitivity level of 8.67×10^{-7}). The *NPM1* mutation levels in the peripheral blood at this point were negative (mutation level of 1.12×10^{-8} , with sensitivity level of assay at 2.8×10^{-6}). As *NPM1* MRD levels at this time point and from this source of sample (peripheral blood after cycle 2) is established as having prognostic impact for patients with *NPM1* mutant AML,⁸ this patient then proceeded to have cycle 3 chemotherapy with the same drug combination of Ven-Aza.

The third course of chemotherapy was also uneventful. However, at the end of this course, although the peripheral blood *NPM1* MRD level remains negative $(2 \cdot 13 \times 10^{-7} \text{ with} \text{ sensitivity level of assay of } 7 \cdot 36 \times 10^{-7}$), the bone marrow *NPM1* MRD level is still positive and higher than after cycle 2 $(1 \cdot 65 \times 10^{-4} \text{ with assay sensitivity level of } 2 \cdot 01 \times 10^{-6})$ (Fig 1).

It is of concern that the bone marrow NPM1 MRD level is still positive and increasing, and this presents a significant challenge for the next therapeutic decision. This is because it is crucial to note that the time points of assessment of NPM1 MRD and the prognostic impact of each of these assessments as applied for this patient, were based on outcomes of patients having intensive standard chemotherapy during the AML17 trial (International Standard Randomised Controlled Trial Number ISRCTN55675535).8 There is no evidence yet to extrapolate these decision time points to patients being treated with the reduced intensity protocols with Ven-Aza. However, in view of the persistent and rising NPM1 MRD level in the bone marrow, and following multidisciplinary meeting review, we have decided to treat this patient with intensive chemotherapy using the fludarabine-idarubicin (FLA-IDA) protocol followed by allogeneic haematopoietic transplantation.

Therefore, the impact of persistent bone marrow *NPM1* MRD levels after cycle 2 of Ven-Aza needs to be reassessed in this new treatment regimen and whether treatment escalation needs to occur at a different time point to that of standard intensive treatment regimens. This question highlights the urgent need to collect the data of response rates and MRD levels of patients with AML treated on reduced intensity protocols and this will need to be addressed in future collaborative studies and randomised control trials.

Acknowledgements

Nadine Farah, Richard Burt, Amr R. Ibrahim and Panagiotis D. Kottaridis wrote the manuscript. Robert Baker conducted the molecular *NPM1* MRD analysis. Panagiotis D. Kottaridis is the consultant in charge of the patient. Panagiotis D. Kottaridis, Nadine Farah, Richard Burt, Amr R. Ibrahim were all involved in the clinical care of this patient. All authors reviewed and proofread the manuscript.

Conflict of interest

The authors have no competing interests.

Nadine Farah^{1,2} Richard Burt^{1,2,3} Amr R. Ibrahim^{1,3} Robert Baker^{3,4} Panagiotis D. Kottaridis³ ¹HCA Healthcare at UCH, University College Hospitals NHS Foundation Trust, ²UCL Cancer Institute, UCL, ³Department of Haematology, University College Hospitals NHS Foundation Trust, and ⁴Molecular Pathology, Health Services Laboratories, The Halo Building, London, UK.

E-mail: n.farah@ucl.ac.uk

Keywords: AML, acute leukaemia, MRD, molecular haematology, molecular genetics

First published online 26 July 2020 doi: 10.1111/bjh.16985

References

- National Institute for Health and Care Excellence (NICE). Guideline NG161. Interim treatment change options during the covid19 pandemic endorsed by NHS England. https://www.nice.org.uk/guidance/ng161/ resources/interim-treatment-change-options-during-the-covid19-pande mic-endorsed-by-nhs-england-pdf-8715724381. Accessed July 2020.
- Pollyea DA, Pratz KW, Jonas BA, Letai A, Pullarkat VA, Wei A, et al. Venetoclax in combination with hypomethylating agents induces rapid, deep, and durable responses in patients with AML ineligible for intensive therapy. *Blood.* 2018;132(Suppl 1):285.
- DiNardo CD, Pratz K, Pullarkat V, Jonas BA, Arellano M, Becker PS, et al. Venetoclax combined with decitabine or azacitidine in treatmentnaive, elderly patients with acute myeloid leukemia. *Blood.* 2019;133:7– 17.
- Lachowiez CA, Loghavi S, Kadia TM, Daver N, Borthakur G, Pemmaraju N, et al. Outcomes of older patients with NPM1-mutated AML: current treatments and the promise of venetoclax-based regimens. *Blood Adv*. 2020;4:1311–20.
- Tiong IS, Dillon R, Ivey A, Teh TC, Vassili C, Donati VR, et al. Rapid elimination of NPM1 mutant measurable residual disease (MRD) using low intensity venetoclax-based combinations in acute myeloid leukemia (AML). *Blood.* 2019;**134**(Suppl 1):2648.
- DiNardo CD, Pratz KW, Letai A, Jonas BA, Wei AH, Thirman M, et al. Safety and preliminary efficacy of venetoclax with decitabine or azacitidine in elderly patients with previously untreated acute myeloid leukaemia: a non-randomised, open-label, phase 1b study. *Lancet Oncol.* 2018;19: 216–28.
- Winters AC, Gutman JA, Purev E, Nakic M, Tobin J, Chase S, et al. Realworld experience of venetoclax with azacitidine for untreated patients with acute myeloid leukemia. *Blood Adv.* 2019;3:2911–9.
- Ivey A, Hills RK, Simpson MA, Jovanovic JV, Gilkes A, Grech A, et al. Assessment of minimal residual disease in standard-risk AML. N Engl J Med. 2016;374:422–33.
- Balsat M, Renneville A, Thomas X, Botton SD, Caillot D, Marceau A, et al. Postinduction minimal residual disease predicts outcome and benefit from allogeneic stem cell transplantation in acute myeloid leukemia with NPM1 mutation: a study by the acute leukemia French Association Group. *J Clin Oncol.* 2017;35:185–93.
- Day JW, Fox TA, Halsey R, Carpenter B, Kottaridis PD. IL-1 blockade with anakinra in acute leukaemia patients with severe COVID-19 pneumonia appears safe and may result in clinical improvement. *Br J Haematol.* 2020 [Epub ahead of print]. https://doi.org/10.1111/bjh. 16873.