OXFORD

https://doi.org/10.1093/ijnp/pyac035 Advance Access Publication: June 23, 2022 Regular Research Article

REGULAR RESEARCH ARTICLE

Positive Allosteric Modulation of α5-GABA_A Receptors Reverses Stress-Induced Alterations in Dopamine System Function and Prepulse Inhibition of Startle

Alexandra M. McCoy[®], Thomas D. Prevot,[®] Md Yenus Mian, James M. Cook, Alan Frazer, Etienne L. Sibille, Flavia R. Carreno, Daniel J. Lodge

Department of Pharmacology and Center for Biomedical Neuroscience, UT Health San Antonio, San Antonio, Texas, USA (Mrs McCoy, Dr Frazer, Dr Carreno, and Dr Lodge); South Texas Veterans Health Care System, Audie L. Murphy Division, San Antonio, Texas, USA (Mrs McCoy, Dr Frazer, Dr Carreno, and Dr Lodge); Campbell Family Mental Health Research Institute of CAMH, Toronto, ON, Canada (Drs Prevot and Sibille); Department of Psychiatry, University of Toronto, Toronto, ON, Canada (Drs Prevot and Sibille); Department of Biochemistry, University of Wisconsin-Milwaukee, Milwaukee, Wisconsin, USA (Dr Mian); Department of Pharmacology and Toxicology, University of Toronto, Toronto, ON, Canada (Dr Sibille).

Correspondence: Alexandra M. McCoy, BS, 7703 Floyd Curl Drive, MC 7764, San Antonio, TX, 78229, USA (mccoya1@livemail.uthscsa.edu).

Abstract

Background: Up to 64% of patients diagnosed with posttraumatic stress disorder (PTSD) experience psychosis, likely attributable to aberrant dopamine neuron activity. We have previously demonstrated that positive allosteric modulators of α 5-GABA_ARs can selectively decrease hippocampal activity and reverse psychosis-like physiological and behavioral alterations in a rodent model used to study schizophrenia; however, whether this approach translates to a PTSD model remains to be elucidated. **Methods:** We utilized a 2-day inescapable foot shock (IS) procedure to induce stress-related pathophysiology in male Sprague-Dawley rats. We evaluated the effects of intra-ventral hippocampus (vHipp) administration GL-II-73, an α 5-GABA_AR, or viral overexpression of the α 5 subunit, using in vivo electrophysiology and behavioral measures in control and IS-treated rats. **Results:** IS significantly increased ventral tegmental area dopamine neuron population activity, or the number of dopamine neurons firing spontaneously (n = 6; P = .016), consistent with observation in multiple rodent models used to study psychosis. IS also induced deficits in sensorimotor gating, as measured by reduced prepulse inhibition of startle (n=12; P=.039). Interestingly, intra-vHipp administration of GL-II-73 completely reversed IS-induced increases in dopamine neuron population activity (n=6; P=.024) and deficits in prepulse inhibition (n=8; P=.025), whereas viral overexpression of the α 5

subunit in the vHipp was not effective.

Conclusions: Our results demonstrate that pharmacological intervention augmenting α 5-GABA_AR function, but not α 5 overexpression in itself, can reverse stress-induced deficits related to PTSD in a rodent model, providing a potential site of therapeutic intervention to treat comorbid psychosis in PTSD.

Keywords: PTSD, psychosis, α5-GABA, receptor, ventral hippocampus, positive allosteric modulators

Received: March 4, 2022; Revised: May 24, 2022; Accepted: June 21, 2022

© The Author(s) 2022. Published by Oxford University Press on behalf of CINP.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (https:// creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

Significance Statement

Patients with posttraumatic stress disorder (PTSD) often experience psychosis (hallucinations and delusions); however, treatments remain inadequate. This study examined a novel therapeutic approach in a rodent model of stress-related pathophysiology associated with PTSD. Specifically, decreasing ventral hippocampal activity with a novel α 5-GABA_A receptor-positive allosteric modulator (GL-II-73) reversed electrophysiological (abnormal dopamine neuron activity) and behavioral (deficits in sensorimotor gating) alterations associated with psychosis. These results suggest that the α 5-GABA_A receptors in the ventral hippocampus may be a novel site of intervention for the treatment of psychosis in PTSD.

Introduction

Approximately 8% of the United States general population and 20% of US combat veterans are diagnosed with posttraumatic stress disorder (PTSD), a mental health condition that develops after exposure to a traumatic event (Kilpatrick et al., 2003, 2013). Core symptoms of PTSD can include hyperarousal, avoidance of trauma-related stimuli, cognitive deficits, and sleep dysfunction (Braakman et al., 2009; Battle, 2013), along with the relatively high (approximately 64% patients) comorbid presentation of psychosis symptoms, including hallucinations and delusions (Battle, 2013; Hardy and Mueser, 2017; Compean and Hamner, 2019). Undesirable side effects and limited efficacy of antipsychotic medications often lead to discontinuation of treatment (Lieberman et al., 2005). Therefore, it is critical to understand the pathophysiology of psychosis associated with PTSD to develop more effective therapies.

Imaging studies have provided evidence that dopamine signaling is positively correlated with psychosis, contributing to the long-standing dopamine hypothesis of psychosis (Laruelle and Abi-Dargham, 1999; Howes et al., 2009). Accordingly, antipsychotics (D2-like receptor antagonists) reduce symptoms of psychosis in some patients; however, they are associated with severe side effects, such as metabolic disorders and dyskinesias (Lieberman et al., 2005). To avoid side effects associated with direct modulation of dopamine receptors, targeting upstream brain regions that modulate dopamine neuron activity may provide a more favorable therapeutic approach (Lodge and Grace, 2006b, 2007, 2011). One region of interest known to modulate dopamine neuron activity is the ventral hippocampus (vHipp). This area of the hippocampus is a stress-sensitive region that has been implicated in the pathology of both psychosis (Lodge and Grace, 2007, 2011) and PTSD (Joshi et al., 2020). In rodent studies, pathological activation of the hippocampus can drive aberrant dopamine system function via a multi-synaptic pathway and induce psychosis-like deficits (Lodge and Grace, 2006b, 2007; Perez and Lodge, 2018). Therefore, targeting aberrant hippocampal activity may represent a novel approach for the treatment of psychosis. Furthermore, targeting hippocampal activity may also reverse PTSD-related deficits attributable to pathology in other brain regions (e.g., the thalamus) (Perez and Lodge, 2018).

In both psychosis patients and animal models, hippocampal hyperactivity is suggested to be attributed to loss of gamma aminobutyric acid (GABA) interneuron function (Zhang and Reynolds, 2002; Lodge et al., 2009; Konradi et al., 2011; Perez and Lodge, 2013; Perez et al., 2014, 2019; Heckers and Konradi, 2015). A loss of interneurons may also contribute to decreased hippocampal volumes, one of the most consistent neurological changes observed in PTSD patients (Bremner et al., 1995, 1997, 2003; Stein et al., 1997; Gilbertson et al., 2002; Villarreal et al., 2002). Further, hippocampal volume is negatively correlated with the severity of core PTSD symptoms as well as dissociation severity (Stein et al., 1997; Gilbertson et al., 2002; Villarreal et al., 2002; Bremner et al., 2003). A large postmortem study reported significant downregulation of interneuron-related transcripts in the brains of PTSD participants (Girgenti et al., 2021). Similarly, stress vulnerability has been associated with decreased levels of GABA-related transcripts (Banasr et al., 2017; Sun et al., 2018), whereas increased expression of specific GABA receptors in the hippocampus appears to confer stress resiliency (Ardi et al., 2016; Sun et al., 2018). Additionally, there is substantial evidence to suggest that stress-related deficits in mood and cognition may be attributable to dysfunction of specific interneuron subtypes. Interneurons expressing the neuropeptide somatostatin appear to be vulnerable in stress-related disorders (Lin and Sibille, 2013; Prévot and Sibille, 2020; Tomoda et al., 2022), with reductions in mRNA and protein being noted in postmortem brains of depressed patients (Tripp et al., 2011; Guilloux et al., 2012; Seney et al., 2015). Likewise, silencing somatostatin interneurons induces depressive- and anxiety-like behaviors that can be reversed by administration of a GABA_A receptor-positive allosteric modulator (PAM) (Fee et al., 2021). Based on this evidence, we posit that hippocampal hyperactivity in psychosis is related to deficits in GABAergic function and thus represents a potential site for therapeutic intervention. Indeed, we have repeatedly demonstrated that decreasing activity in the vHipp can reverse psychosis-like alterations in dopamine neuron activity and behavior in a rodent model used to study schizophrenia-like pathophysiology (Perez et al., 2013; Donegan et al., 2017, 2019; Perez and Lodge, 2018); however, whether this approach extends to psychosis in models of stress-induced pathology remains to be established.

Hippocampal activity can be modulated by targeting GABA_A receptors that contain the α5 subunit (α5-GABA_ARs). GABA_ARs typically consist of 2 alpha (α 1-6), 2 beta (β 1-3), and 1 gamma (γ 1-3) or delta (δ) subunits, although 19 genes for GABA_ARs subunits have been identified (Simon et al., 2004; Jacob, 2019). The subunit composition of GABA_ARs confers distinct functional properties as well as unique patterns of expression throughout the brain. Of interest, α 5-GABA Rs are enriched in the hippocampus, where they make up nearly 25% of all hippocampal GABA Rs (Sur et al., 1999; Olsen and Sieghart, 2009). Further, within the hippocampus, there is higher α 5 expression in ventral vs dorsal subregions, making it an ideal target to manipulate activity in the vHipp (Fritschy and Mohler, 1995; Sur et al., 1999). Deficits in α 5 expression have been noted in animal models of psychosis (Kiemes et al., 2021), and knockdown of α 5-GABA Rs in otherwise healthy animals has been shown to induce psychosis-like deficits in sensorimotor gating, as measured by prepulse inhibition (PPI) (Hauser et al., 2005). Furthermore, overexpression of the α 5 subunit in the vHipp (Donegan et al., 2019) or administration of an α 5-GABA₄R PAM (Gill et al., 2011; Perez et al., 2022) can reverse dopamine system dysfunction and behavioral deficits associated with psychosis in a model used to study schizophrenia. These studies provide preclinical evidence that targeting α5-GABA_ARs may represent an effective treatment for psychosis. Although aberrant dopamine neuron activity and deficits in PPI are also observed in animal models used to study

PTSD (Elam et al., 2021), the underlying pathology likely differs and whether the efficacy of therapeutic approaches is consistent across diagnosis is not currently known. Nonetheless, we posit that augmenting hippocampal interneuron function by targeting α5-GABA Rs will correct this deficit. In this study, we investigated if pharmacological increase in function or genetic augmentation of vHipp a5-GABA R expression could reverse inescapable foot shock (IS)- induced alterations in dopamine system function and deficits in sensorimotor gating and whether IS altered vHipp α 5-GABA, R expression. We found that intra-vHipp administration of GL-II-73, a novel α5-GABA R PAM, reversed the electrophysiological and behavioral effects of IS, whereas vHipp overexpression of α 5-GABA_ARs was ineffective. IS exposure had no effect on the expression of α 5-GABA_ARs in the vHipp. The results of this study provide evidence that pharmacological targeting of a5-GABA, Rs may represent a novel therapeutic strategy for the treatment of PTSD and comorbid psychosis.

MATERIALS AND METHODS

All experiments were performed in accordance with the guidelines outlined in the USPH Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and the Use Committees of UT Health San Antonio and US Department of Veterans Affairs.

Animals

Adult, male Sprague Dawley rats (350–400 g; Envigo, Indianapolis, IN, USA) were used for all experiments. Rats were group housed to avoid social isolation stress and were kept on a 12-hour-light/-dark cycle. Food and water were provided ad libitum.

Inescapable Foot Shock Stress

Adult rats were randomly assigned to control (no shock) or shock groups who received 2 consecutive days of inescapable foot shock stress. The 2-day IS paradigm consists of rats placed in a 30.5 × 25.4 × 30.5 cm³ square conditioning chamber with a stainless-steel grid shock floor (Coulbourn Instruments, Whitehall, PA, USA). One session of IS consisted of 60×15 -second, 0.8-mA foot shocks with an average inter-trial interval of 30 seconds with a 25% deviation (±7.5 seconds) and lasts approximately 40 minutes. Control rats were handled daily but not exposed to conditioning chambers. Rats were further assigned to specific treatment groups (n=5–6 electrophysiology, n=8–12 PPI). Electrophysiology and behavioral experiments were conducted 24–48 hours after the last day of IS.

Materials

The proprietary compound GL-II-73 was supplied by the Centre for Addiction and Mental Health, Campbell Family Mental Health Research Institute (Toronto, ON, Canada). GL-II-73 or vehicle (85% H₂0, 14% propylene glycol, 1% Tween80) was administered directly into the vHipp (100 ng/uL; 0.75 µL; AP –5.3, ML±5.0, DV –6.0 mm from bregma) 20 minutes prior to electrophysiology or behavior. This dose and timing was selected based on previous characterization (Prevot et al., 2019) as well as our own data (Perez et al., 2022). Although systemic administration would be a more therapeutically relevant approach, we chose intra-vHipp administration because rats seem to rapidly metabolize this compound (Prevot et al., unpublished observations). Importantly, this effect appears to be exclusive to rats, because GL-II-73 can be administered systemically to mice and achieves

sufficient brain concentrations to produce robust behavioral effects (Prevot et al., 2019). Chloral hydrate (C8383), propylene glycol (P4347), and Tween80 (P1754) were obtained from Sigma-Aldrich (St. Louis, MO, USA).

Intra-Hippocampal Viral Administration

To overexpress the α 5 subunit of the GABA₄ receptor under the control of the CAMKII promoter, lentiviral vectors were used as previously described (Donegan et al., 2019). In brief, a single cohort of rats was anesthetized using Fluriso (2-5% isoflurane, USP with oxygen flow at 1 L/min) and placed on a stereotaxic apparatus. Bilateral cannula lowered into the vHipp (AP -5.3, ML±5.0, DV –6.0 mm from bregma) were used to inject the α 5 lentiviral vector expressing enhanced green fluorescent protein (EGFP) as a reporter (pLV-CaMKII-rGabr α 5-IRES-EGFP; 2.86 × 10⁹ TU/mL) or the control vector that expressed the reporter (pLV-EGFP:T2A:puro-EF1A > mCherry; 1.26×10^9 TU/mL) into each hemisphere (0.75 μ L; VectorBuilder, Chicago, IL, USA) using an injector that extended 1 mm past the guide cannula. Coordinates were selected based on an atlas (Paxinos and Watson, 1988). Animals were allowed to recover for >6 weeks before behavioral and electrophysiological experiments to allow maximal gene expression.

Verification of Electrode and Cannula Placement

To verify electrode and cannula placement, rats were killed via chloral hydrate overdose and rapidly decapitated at completion of all experiments. Brains were extracted, fixed for at least 24 hours (4% phosphate buffered formaldehyde), and cryoprotected (10% w/v sucrose in phosphate-buffered saline) until saturated. Brains were coronally sectioned (20–25 µm) using a cryostat (Leica, Buffalo Grove, IL, USA). Sections containing electrode or cannula tracks were mounted onto gelatin-coated slides, stained with neutral red (0.1%) and thionin acetate (0.01%), and cover-slipped with DPX Mountant for histochemical confirmation within the VTA or vHipp (Fig 1A, B). For overexpression studies, images were captured on an upright fluorescent microscope (Zeiss, AxioLab. A1; Oberkochen, Germany), and stable transgene expression was confirmed in a subset of animals by green fluorescent protein (GFP)+ fluorescence in vHipp pyramidal cells (Fig 1C).

In Vivo Extracellular Dopamine Recordings

Adult rats were anesthetized with 8% chloral hydrate (400 mg/kg, i.p.) and placed in a stereotaxic apparatus for the duration of the experiment (no longer than 3 hours). A maximum of 2 rats were recorded each day. Extracellular glass microelectrodes (impedance approximately 6–10 M Ω) were lowered into the VTA (from bregma: AP -5.3, ML ±0.6, and DV -6.5 to -9.0 mm) using a hydraulic micro-positioner (Model 640, Kopf Instruments, Tujunga, CA, USA). Spontaneously active dopamine neurons were identified using open filter settings (low-frequency cutoff: 30 Hz; highfrequency cutoff: 30 kHz) according to previously established electrophysiological criteria (Ungless and Grace, 2012). Three parameters of dopamine activity were measured and analyzed: the number of dopamine neurons firing spontaneously per track (population activity) (Lodge and Grace, 2011), firing rate, and proportion of action potentials occurring in bursts (defined as the incidence of spikes with <80 ms between them; termination of the burst is defined as >180 ms between spikes). Whereas population activity can be altered via the hippocampus (Lodge and Grace, 2007, 2011; Perez and Lodge, 2018; Donegan et al., 2019), changes in burst firing and firing rate are primarily mediated by other VTA afferents (Floresco et al., 2003; Lodge and Grace,



В



С



Figure 1. Verification of electrode and cannula placement and stable transgene expression. Representative brain slice with electrode placement in the ventral tegmental area (VTA) (A, left) and cannula placement for drug administration in the ventral hippocampus (vHipp) (B, left) indicated by the black arrow, with corresponding schematics of the brain sections (-5.40 mm posterior to bregma) with a box indicating the area in which tracks were found (right). Representative image containing a schematic of a brain slice (left -5.40 mm from bregma) with a box indicating the area in the vHipp near the tip of the cannula track where green fluorescent protein (GFP)-expressing pyramidal neurons were found (C, right, -5.40 mm from bregma). Scale bar denotes 50 μm.

2006a; Grace and Gomes, 2019). Analysis of dopamine neuron activity was performed using LabChart software (ADInstruments, Sydney, Australia).

Prepulse Inhibition of Startle

Adults rats were placed into a sound-attenuated chamber (SD Instruments) and allowed to acclimate to 65-dB background noise for 5 minutes. Rats were then exposed to 10 startle-only trials (40 ms, 120 dB, 15-second average inter-trial intervals). Next, rats were exposed to 24 trials where a pre-pulse (20 ms at 69, 73, and 81 dB) was presented 100 ms before the startle pulse. Each prepulse + startle pulse trial was presented 6 times in a pseudo-random order (15-second average inter-trial interval). The startle response was measured from 10 to 80 milliseconds after the onset of the startle pulse and recorded using SR-LAB Analysis Software (SD Instruments, San Diego, CA, USA); the entire test lasted approximately 15 minutes. Two cohorts of rats were used: 1 for drug experiments and 1 for virus experiments. All comparisons were made within cohorts (i.e., control groups were not combined).

Western Blot

Twenty-four hours after the last day of IS, brains (n=9/group)were extracted and punches containing the vHipp were sonicated in lysis buffer and the proteins in the total lysate were separated in an sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) gel followed by blotting onto a 0.2-µm nitrocellulose membrane. Membranes were incubated with an antibody against α 5-GABA₄Rs (UC Davis/NIH NeuroMab Facility, Davis, CA, USA 75-401; 1:1000 overnight at 4°C) and washed with Tris-buffered saline with 0.1% Tween 20 (TBST) prior to incubation with the secondary antibody (goat anti-mouse Sigma-Aldrich A9917; 1:30,000) for 1 hour at room temperature. Membranes were washed with TBST and incubated with a chemiluminescence kit (Pierce, Rockford, IL, USA) followed by exposure to X-ray film for detection. Antibodies were stripped using a commercially available stripping buffer (Thermo Scientific, Waltham, MA, USA), washed, blocked, and re-probed with an antibody against horseradish peroxidase (HRP)-conjugated β -actin (1:200000 in 1% bovine serum albumin (BSA) in TBST, Sigma-Aldrich A3854). Densitometry analyses of immunoreactive bands for α 5-GABA₄Rs or β -actin (loading control) were performed using the NIH Image J software from the scanned films. Densitometric arbitrary units obtained for α 5-GABA, Rs immunoreactive bands were normalized to β -actin and expressed as percent from no shock control.

Statistical Analysis

In all figures, data are shown as mean±SEM, and n values representing either the number of rats or neurons are indicated in the figure legend. In all experiments, data were analyzed using SigmaPlot (Systat Software Inc., Chicago, IL, USA) either with a 2-way ANOVA (electrophysiology) or 3-way ANOVA (PPI) or t test (western blot). When significant main effects or interactions were detected, Holm–Sidak post-hoc tests were used. All tests were 2-tailed, and significance was determined at P<.05. Graphs were plotted with Prism software (GraphPad Software Inc., San Diego, CA, USA).

RESULTS

Intra-vHipp Administration of GL-II-73 Reversed Shock-Induced Increases in VTA Dopamine Neuron Population Activity

In rats, changes in mesolimbic dopamine activity can be directly examined using in vivo extracellular electrophysiology to quantify the number of spontaneously active dopamine neurons in the VTA (population activity). Following exposure to 2 days of IS, a significant increase in dopamine neuron activity was observed. Specifically, a main effect of shock and drug were observed (Fig 2A; 2-way ANOVA; factors: shock × drug; $F_{(1,19)}$ shock =7.024; P=.016; $F_{(1,19) drug}$ =7.258; P=.014; $F_{(1,19) interaction}$ =4.149; P=.056). Shocked animals treated with vehicle displayed a significantly higher number of spontaneously active dopaminergic cells per track compared with nonshocked, vehicle-treated animals (Holm–Sidak; t=3.238, P=.026, control/vehicle: 0.97±0.07 cells per track; shock/vehicle: 1.68±0.30 cells per track). This shock-induced increase in dopamine neuron activity was completely reversed by the intra-hippocampal administration of GL-II-73 (100 ng/ μ L; 0.75 μ L) 20 minutes prior to electrophysiology (Holm–Sidak; t=3.268, P=.024, 0.96±0.09 cells per track). There was no significant effect of GL-II-73 on population activity in nonshocked rats (Holm–Sidak; t=0.476, P=.998, 0.87±0.10 cells per track). Two additional parameters of dopamine cell activity were examined: firing rate and percentage bursting. As expected, no significant differences were observed in average firing



Figure 2. Direct intra-ventral hippocampus (vHipp) administration of the α 5-selective positive allosteric modulator (PAM), GL-II-73, completely restored dopamine system function in shocked rats. In vivo extracellular electrophysiology was used to measure dopamine cell activity in the ventral tegmental area. (A) Inescapable shock exposure significantly increased the number of spontaneously active cells/track (population activity), which was reversed by intra-ventral hippocampus injection of GL-II-73 (100 ng/µL; 0.75 µL). (B) Firing rate was unaffected by shock or drug treatment; however, (C) there was a small effect of GL-II-73 on burst firing. Approximately 10-second example traces from control (D) or shocked (E) rats (bottom) and individual action potentials (top). *P<.05, significance from respective nonshocked control; *P<.05, significance from shock/vehicle group, n=5–6/group.

rate (Fig 2B; 2-way ANOVA; factors: shock \times drug; $F_{(1,91) drug}$ =0.075, P = .785; $F_{(1, 91) \text{ shock}} = 1.417$, P = .237; $F_{(1, 91) \text{ interaction}} = 1.320$, $\bar{P} = .254$). There was a main effect of GL-II-73 on percentage bursting (Fig 2C; 2-way ANOVA; factors: shock × drug; $F_{(1, 87) drug}$ = 4.000, P = .049; $F_{(1, 87) \text{ shock}} = 1.027$, P = .314; $F_{(1, 87) \text{ interaction}} < 0.003$, P = .958); however, post-hoc analysis revealed no significant differences between individual groups (shock/vehicle: 23.36±3.75%; shock/GL-II-73: 33.99±6.09%; control/vehicle: 28.61±6.24%; control/GL-II-73: 39.81±5.86%). The lack of robust effects of GL-II-73 on burst firing and firing rate are expected because, unlike VTA population activity, which can be altered via the hippocampus (Lodge and Grace, 2007, 2011; Perez and Lodge, 2018; Donegan et al., 2019), changes in burst firing and firing rate are primarily mediated by other VTA afferents, such as the laterodorsal tegmental nucleus and pedunculopontine tegmental nucleus (Floresco et al., 2003; Lodge and Grace, 2006a; Grace and Gomes, 2019) (Fig. 2D,E).

GL-II-73 Reversed Shock-Induced Deficits in PPI

Consistent with previous reports (Elam et al., 2021), IS induced a deficit in PPI (Fig. 3; 3-way mixed ANOVA; factors: shock × drug × pre-pulse intensity; $F_{(1, 33) \text{ shock}}$ =1.431; P=.240; $F_{(1, 33) \text{ drug}}$ =1.159; P=.2894; $F_{(2, 66) \text{ interaction}}$ =5.543; P=.025). Interestingly, this shock-induced deficit in sensorimotor gating was completely reversed at 73 dB by the intra-hippocampal administration of GL-II-73 (Holm–Sidak; t=3.546, P<.001). There was no significant effect of GL-II-73 on PPI in nonshocked animals (Holm–Sidak; t=1.240, P=.218).

Overexpression of $\alpha 5$ in the vHipp Did Not Alter Dopamine Population Activity

We confirmed stable transgene expression in the vHipp by the presence of GFP-labeled cells in the pyramidal cell layer of the vHipp, similar to previous characterizations of this approach (Donegan et al., 2019) (Fig. 1C). Consistent with our previous findings (Elam et al., 2021) and data from vehicle-treated rats (Fig. 2A), inescapable shock significantly increased VTA dopamine neuron population activity, but there was no effect of α 5-subunit overexpression (Fig. 4A; 2-way ANOVA; factors: shock × virus; $F_{(1,23) \text{ shock}=} 22.33$; P<0.0001; $F_{(1,23) \text{ virus}} = 2.092$; P=.162; $F_{(1,23) \text{ interaction}} = 0.4849$; P=.485). Post hoc tests revealed a significant difference between control/nonshocked and control/shock groups (Holm–Sidak; t=3.018, P=.024; control/nonshocked: 0.98 ± 0.05 cells per track; control/shock: 1.56 ± 0.19 cells per track) and between α 5/nonshocked and α 5/shock groups (Holm– Sidak: t=3.641, P=.007; α 5/nonshocked: 1.08 ± 0.11 cells per



Figure 3. Direct intra-ventral hippocampus (vHipp) administration of GL-II-73 reversed shock-induced deficits in prepulse inhibition of startle. Two days of inescapable shock significantly decreased prepulse inhibition in rats that received vehicle, but not rats that received GL-II-73. *P<0.05, difference between no shock/vehicle and shock/vehicle; #P<0.05, difference between shock/vehicle and shock/GL-II-73, n=8-12/group.

track; α 5/shock: 1.87±0.20 cells per track). There was no effect of α 5-subunit overexpression on firing rate (Fig. 4B; 2-way ANOVA; factors: stress × virus; $F_{(1,203) \text{ shock}} = 1.776$; P=.185; $F_{(1,203) \text{ virus}} = 1.445$; P=.231; $F_{(1,203) \text{ interaction}} = 0.011$; P=.918; control/GFP: 3.98±0.32 Hz; stress/GFP: 3.65±0.21 Hz; control/ α 5: 3.68±0.32 Hz; stress/a5: 3.29±0.23 Hz). There was a main effect of shock on percentage bursting (Fig. 4C; 2-way ANOVA; factors: stress×virus; $F_{(1,202)}$; shock = 5.388; P=.021; $F_{(1,202) \text{ virus}} = 0.982$; P=.323; $F_{(1,202) \text{ interaction}} = 0.049$; P=.824), though post-hoc analysis revealed no significant differences between groups (control/GFP: 30.82±3.38%; shock/GFP: 23.61 ± 2.93%; control/ α 5: 35.00±4.68%; shock/ α 5: 26.25±3.00%). These results demonstrate that viral overexpression of α 5-GABA_AR is insufficient to reverse aberrant dopamine neuron activity following shock exposure.

α5 Overexpression Did Not Alter PPI

Consistent with our earlier results (Fig. 3), a main effect of shock was observed and there was a trend towards a main effect of the virus (Fig. 5; 3-way mixed ANOVA; factors: shock × virus × PPI; $F_{(1,32)}$ shock =4.653; P=.039; $F_{(1,32)}$ virus =3.369; P=.070; $F_{(1,32)}$ interaction =0.021; P=.887), replicating our laboratory's previous finding that IS induces deficits in PPI (Elam et al., 2021). However, posthoc analysis revealed no relevant statistical differences between individual groups.

Inescapable Shock Did Not Alter α 5-GABA_AR Expression Within the vHipp

Other rodent models used to study psychosis display deficits in hippocampal α 5-GABA_AR expression (Kiemes et al., 2021). To determine whether inescapable shock altered total levels of α 5-GABA_ARs in the vHipp, we performed western-blot analysis in control and shocked rats and demonstrated that IS did not significantly alter α 5-GABA_ARs levels in the vHipp (Fig. 6: t test; t=0.469, P=.645).

Discussion

The dopamine hypothesis of psychosis suggests that symptoms of psychosis are attributable to a pathological increase in mesolimbic dopamine transmission. Increasing evidence from our laboratory and others supports the hypothesis that dysregulation of the dopamine system observed in animal models of psychosis is driven by aberrant hippocampal output. Specifically, hippocampal hyperactivity can drive increases in VTA dopamine neuron population activity (Lodge and Grace, 2007, 2011; Perez and Lodge, 2013; Perez et al., 2013; Donegan et al., 2019). Therefore, we examined if reducing hippocampal activity in a rodent model of stress-related psychopathology used to study PTSD could normalize dopamine neuron activity as well as behavioral correlates of psychosis. In the current study, we utilized a 2-day IS procedure, which produces physiological and behavioral changes consistent with those of PTSD (i.e., hyperarousal, anxiety-like behavior, and sleep alterations) (Van Dijken et al., 1992; Pynoos et al., 1996; Pawlyk et al., 2005) to study psychosis-like alterations in physiology and behavior. Consistent with previous work from our laboratory and others, we found that foot shock exposure produced robust changes in dopamine neuron population activity and deficits in PPI (Valenti et al., 2011; Gomes and Grace, 2017; Gomes et al., 2020; Elam et al., 2021). Further, intra-vHipp administration of GL-II-73, a selective α5-GABA, R PAM, reversed the effects of IS on dopamine neuron population activity and PPI, suggesting that α5-GABA Rs



Figure 4. Ventral hippocampus (vHipp) overexpression of the α 5 subunit of the GABA_AR failed to restore dopamine system function in stressed rats. (A) Shock exposure significantly increased ventral tegmental area population activity in both control rats and rats with overexpression of the α 5 subunit. (B) Firing rate was unaffected by shock or α 5 overexpression. (C) A small decrease in burst firing was observed in shocked animals, but there was no effect of virus. Approximately 10-second example traces from control (D) or overexpression (E) rats (bottom) and individual action potentials (top). *P<.05; **P<.01 denotes significance from nonshocked control, n=6–8/ group.

may be a novel target for treating PTSD and comorbid psychosis. Although administration of GL-II-73 did produce a statistically significant change in the burst firing activity of VTA dopamine neurons, we do not believe this to be physiologically relevant because our previous work demonstrates that multiple α 5-GABA_AR PAMs (Perez et al., 2022), or other hippocampal manipulations (Perez et al., 2013; Perez and Lodge, 2013, 2018), do not consistently alter burst firing. Indeed, previous studies have established that burst firing is largely independent of hippocampal

activity and mediated by other regions, such as the laterodorsal tegmental nucleus and pedunculopontine tegmental nucleus (Floresco et al., 2003; Lodge and Grace, 2006a; Grace and Gomes, 2019).

The hippocampus, specifically the vHipp, has received considerable attention as a target for treating symptoms of psychosis. Previous studies have demonstrated that the vHipp can regulate dopamine activity in the VTA through a multisynaptic pathway, including the nucleus accumbens and ventral pallidum



Figure 5. Ventral hippocampus (vHipp) overexpression of the α 5 subunit of the GABA_AR failed to reverse deficits in prepulse inhibition of startle. Two days of inescapable shock significantly decreased prepulse inhibition in both control and overexpression rats. There was no effect of α 5 overexpression in control or shocked animals. *P<.05, main effect of shock, n=8–10/group.



Figure 6. Inescapable shock did not change the α 5-GABA_AR levels in the ventral hippocampus (vHipp) . Representative image of western blot (WB) of vHipp α 5-GABA_ARs (top) and actin (bottom) immunoreactive bands (A). Quantification (B) revealed no differences between no shock (NS) and inescapable shock (IS) groups, P=.64, n=9/group.

(Lodge and Grace, 2006b, 2007, 2011; Perez and Lodge, 2018). This indirect approach to modulating dopamine neuron activity via the vHipp may provide a more favorable side effect profile compared with currently available antipsychotic drugs, which directly target dopamine D2-like receptors and are associated with severe side effects (Lieberman et al., 2005). Furthermore, it differentially modulates activity in the VTA and substantia nigra, potentially limiting extrapyramidal effects (Bortz and Grace, 2018). In this study, we utilized pharmacological and overexpression approaches to examine whether increasing GABAergic tone to vHipp pyramidal cells can normalize stress-induced, psychosislike alterations to physiology and behavior. We chose intravHipp administration of GL-II-73, a novel PAM selective for α5-GABA_ARs (Prevot et al., 2019), rather than systemic administration for 2 reasons. First, although α 5-GABA_ARs are enriched in the hippocampus, they are not exclusively expressed in the hippocampus (Fritschy and Mohler, 1995; Sur et al., 1999), and this method allows for the effects of GL-II-73 to be attributed to α 5-GABA_AR in the vHipp specifically. The second reason is that we have discovered, when given to rats systemically, this compound is rapidly metabolized, resulting in insufficient brain concentrations to produce measurable psychoactive effects (Prevot et al., unpublished observations). Importantly, this effect appears to be exclusive to rats, because GL-II-73 can be administered systemically to mice and achieves sufficient brain concentrations to produce robust behavioral effects (Prevot et al., 2019). Here, we demonstrated that intra-vHipp administration of GL-II-73 can reverse aberrant dopamine neuron activity and deficits in PPI observed after IS. These results are consistent

with previous work demonstrating that another α 5-GABA_A PAM, SH-053-2'F-R-CH3, can reverse aberrant dopamine neuron activity in a rodent model used to study schizophrenia (Gill et al., 2011). Of note, both positive and negative α 5-GABA_AR modulators are being developed for clinical applications, including as antidepressants, anxiolytics, and cognitive enhancers (Atack et al., 2006; Collinson et al., 2006; Carreno et al., 2017; Prevot et al., 2019, 2020). This is particularly exciting for the treatment of PTSD because comorbid depression and anxiety as well as cognitive deficits are commonly present in this disorder (Brady et al., 2000; Quinones et al., 2020). In combination with its robust effects on dopamine neuron activity and PPI, it appears that GL-II-73 may be beneficial for treating multiple comorbidities associated with PTSD as well as other psychiatric disorders (Prevot et al., 2019, 2020; Perez et al., 2022).

We previously reported that overexpression of the α 5 subunit in the vHipp can correct aberrant dopamine neuron activity and psychosis-like behaviors in a rodent model of schizophrenia (Donegan et al., 2019). As a proof of concept, we tested whether α 5 overexpression in the vHipp could reverse shock-induced, psychosis-like alterations in a model of PTSD. Interestingly, over expression of α 5 in the vHipp did not affect dopamine neuron population activity and although there was a trend towards an effect of α 5 overexpression on PPI, this was not statistically significant (P=.07). This may be due to low transfection; however, we do not believe this to be the case because we observed robust GFP expression in the vHipp. Alternatively, the additional a5 subunits may not have successfully incorporated into functional receptors. Indeed, oligomerization of GABA, Rs is highly inefficient; only approximately 25% of translated subunits incorporate into receptors (Jacob et al., 2008). Even if overexpression produced functional receptors, it has been shown that α 5-GABA_AR localization (extrasynaptic vs synaptic) is highly dynamic (Loebrich et al., 2006; Hausrat et al., 2015; Davenport et al., 2021), and overexpression may shunt α5-GABA_AR receptors, the majority of which are found extrasynaptically, into the synapse. This relocation is relevant to the work done here because we previously published data that suggest that targeting extrasynaptic, but not synaptic, GABA_ARs in the vHipp can restore aberrant dopamine system function (Donegan et al., 2019; Perez et al., 2022). Although we did not measure inhibitory currents in the vHipp in this study, we previously observed an increase in tonic inhibition following transfection with the same vector and protocol utilized here, suggesting that this overexpression approach yields functional, extrasynaptic α5-GABA_ARs (Donegan et al., 2019). Thus, the lack of effect of α 5 overexpression here is likely due to differences between models used and the divergent pathology underlying developmental and stress-based insults. PTSD is a complex disorder, and numerous brain regions are implicated in its pathology, including, but not limited to, the vHipp (Semple et al., 2000; Shin et al., 2006). Indeed, the role of the paraventricular nucleus of the thalamus (PVT) in PTSD and psychosis is an active area of study (Elam et al., 2021). However, we have also demonstrated that afferents from the vHipp and PVT converge on single neurons within the NAc to regulate dopamine population activity in the VTA (Perez and Lodge, 2018), and regardless of which region is activated, decreasing activity in either the vHipp or PVT can restore dopamine system function. Thus, although the vHipp may not be the primary site of pathology, it may still be a promising site of pharmacologic intervention. A more parsimonious explanation for these results is that receptor overexpression does not address the underlying deficits observed in PTSD. In the methylazoxymethanol acetate (MAM) model used to study psychosis, rats have been shown to have

reduced number of interneurons (Lodge et al., 2009) as well as decreased α 5-GABA_AR density in the hippocampus (Kiemes et al., 2021), which may account for why α 5 overexpression successfully treated dopamine system dysfunction in MAM rats (Donegan et al., 2019). However, aberrant hippocampal function in PTSD may not be due to decreased GABARs (Gilbertson et al., 2002; Villarreal et al., 2002; Bremner et al., 2003; Girgenti et al., 2021). Indeed, we demonstrated that there is no deficit in vHipp α 5-GABA_AR expression following IS exposure (Fig. 6). Thus, it is not surprising that increasing the number of α 5-GABA Rs was ineffective in this model, although it does raise the question as to why GL-II-73 was effective. Although there may not be a deficit in receptor expression, a low-GABA state that is present in multiple psychiatric conditions (Prévot and Sibille, 2020) may also be present in the IS model, and enhancing response to GABA with a α 5-GABA, R may address this pathology if, for example, concentrations of GABA are lower.

In summary, we have demonstrated that GL-II-73, a novel α 5-GABA_AR PAM, can reverse stress-induced, psychosis-like alterations in a rodent model used to study PTSD. Although α 5 overexpression did not successfully reverse deficits associated with our shock model, the results with GL-II-73 demonstrate that pharmacological targeting of α 5-GABA_ARs is a useful strategy for treating psychosis. This study adds to the growing literature implicating α 5-GABA_ARs as a common therapeutic target for the treatment of psychosis in multiple animal models and provides evidence that GL-II-73 may be a novel compound that can treat psychosis in addition to a range of psychiatric disorders.

Acknowledgments

The authors thank Angela Boley, PhD, for assistance with data acquisition. Some figures were created with BioRender.com. This work was supported by Merit Awards from the United States Department of Veterans Affairs, Biomedical Laboratory Research (BX004693 and BX004646 to D.J.L.) and Development Service and National Institutes of Health grants (R01-MH090067 to D.J.L. and T32-NS082145 to A.M.M.)

Interest Statement

E.S., T.D.P., and J.C. are co-inventors or listed on US patent applications that cover GABAergic ligands, including GL-II-73, and their use in brain disorders. E.S. is co-founder of DAMONA Pharmaceuticals, a biopharmaceutical company dedicated to treat cognitive deficits in brain disorders.

References

- Ardi Z, Albrecht A, Richter-Levin A, Saha R, Richter-Levin G (2016) Behavioral profiling as a translational approach in an animal model of posttraumatic stress disorder. Neurobiol Dis 88:139– 147.
- Atack JR, Bayley PJ, Seabrook GR, Wafford KA, McKernan RM, Dawson GR (2006) L-655,708 enhances cognition in rats but is not proconvulsant at a dose selective for alpha5-containing GABAA receptors. Neuropharmacology 51:1023–1029.
- Banasr M, Lepack A, Fee C, Duric V, Maldonado-Aviles J, DiLeone R, Sibille E, Duman RS, Sanacora G (2017). Characterization of GABAergic marker expression in the chronic unpredictable stress model of depression. Chronic Stress (Thousand Oaks, Calif.) 1:2470547017720459. doi:10.1177/2470547017720459

- Battle DE (2013) Diagnostic and statistical manual of mental disorders (DSM). CoDAS 25:191–192.
- Bortz DM, Grace AA (2018) Medial septum differentially regulates dopamine neuron activity in the rat ventral tegmental area and substantia nigra via distinct pathways. Neuropsychopharm. 43:2093–2100.
- Braakman MH, Kortmann FAM, van den Brink W (2009). Validity of "post-traumatic stress disorder with secondary psychotic features": a review of the evidence. Acta Psychiatr Scand 119:15–24.
- Brady KT, Killeen TK, Brewerton T, Lucerini S (2000) Comorbidity of psychiatric disorders and posttraumatic stress disorder. J Clin Psychiatry 61:22–32.
- Bremner JD, Randall P, Scott TM, Bronen RA, Seibyl JP, Southwick SM, Delaney RC, McCarthy G, Charney DS, Innis RB (1995) MRI-based measurement of hippocampal volume in patients with combat-related posttraumatic stress disorder. Am J Psychiatry 152:973–981.
- Bremner JD, Randall P, Vermetten E, Staib L, Bronen RA, Mazure C, Capelli S, McCarthy G, Innis RB, Charney DS (1997) Magnetic resonance imaging-based measurement of hippocampal volume in posttraumatic stress disorder related to childhood physical and sexual abuse—a preliminary report. Biol Psychiatry 41:23–32.
- Bremner JD, Vythilingam M, Vermetten E, Southwick SM, McGlashan T, Nazeer A, Khan S, Vaccarino LV, Soufer R, Garg PK, Ng CK, Staib LH, Duncan JS, Charney DS (2003) MRI and PET study of deficits in hippocampal structure and function in women with childhood sexual abuse and posttraumatic stress disorder. Am J Psychiatry 160:924–932.
- Carreno FR, Collins GT, Frazer A, Lodge DJ (2017) Selective pharmacological augmentation of hippocampal activity produces a sustained antidepressant-like response without abuse-related or psychotomimetic effects. Intl J Neuropsychopharmacol 20:504–509.
- Collinson N, Atack JR, Laughton P, Dawson GR, Stephens DN (2006) An inverse agonist selective for alpha5 subunitcontaining GABAA receptors improves encoding and recall but not consolidation in the Morris water maze. Psychopharmacology 188:619–628.
- Compean E, Hamner M (2019) Posttraumatic stress disorder with secondary psychotic features (PTSD-SP): diagnostic and treatment challenges. Prog Neuropsychopharmacol Biol 88:265–275.
- Davenport CM, Rajappa R, Katchan L, Taylor CR, Tsai M-C, Smith CM, de Jong JW, Arnold DB, Lammel S, Kramer RH (2021). Relocation of an extrasynaptic GABAA receptor to inhibitory synapses freezes excitatory synaptic strength and preserves memory. Neuron 109(1):123–134.e4.
- Donegan JJ, Tyson JA, Branch SY, Beckstead MJ, Anderson SA, Lodge DJ (2017) Stem cell-derived interneuron transplants as a treatment for schizophrenia: Preclinical validation in a rodent model. Mol Psychiatry 22:1492–1501.
- Donegan JJ, Boley AM, Yamaguchi J, Toney GM, Lodge DJ (2019) Modulation of extrasynaptic GABAA alpha 5 receptors in the ventral hippocampus normalizes physiological and behavioral deficits in a circuit specific manner. Nat Commun 10:2819.
- Elam HB, Perez SM, Donegan JJ, Lodge DJ (2021) Orexin receptor antagonists reverse aberrant dopamine neuron activity and related behaviors in a rodent model of stress-induced psychosis. Transl Psychiatry 11:114.
- Fee C, Prevot TD, Misquitta K, Knutson DE, Li G, Mondal P, Cook JM, Banasr M, Sibille E (2021) Behavioral deficits

induced by somatostatin-positive GABA neuron silencing are rescued by alpha 5 GABA-A receptor potentiation. Intl J Neuropsychopharmacol. 24:505–518.

- Floresco SB, West AR, Ash B, Moore H, Grace AA (2003) Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic dopamine transmission. Nat Neurosci 6:968–973.
- Fritschy JM, Mohler H (1995) GABAA-receptor heterogeneity in the adult rat brain: differential regional and cellular distribution of seven major subunits. J Comp Neurol 359:154–194.
- Gilbertson MW, Shenton ME, Ciszewski A, Kasai K, Lasko NB, Orr SP, Pitman RK (2002) Smaller hippocampal volume predicts pathologic vulnerability to psychological trauma. Nat Neurosci 5:1242–1247.
- Gill KM, Lodge DJ, Cook JM, Aras S, Grace AA (2011) A novel α5GABAAR-positive allosteric modulator reverses hyperactivation of the dopamine system in the MAM model of schizophrenia. Neuropsychopharmacology 36:1903–1911.
- Girgenti MJ, Wang J, Ji D, Cruz DA, Stein MB, Gelernter J, Young KA, Huber BR, Williamson DE, Friedman MJ, Krystal JH, Zhao H, Duman RS (2021) Transcriptomic organization of the human brain in post-traumatic stress disorder. Nat Neurosci 24:24– 33.
- Gomes FV, Grace AA (2017) Prefrontal cortex dysfunction increases susceptibility to schizophrenia-like changes induced by adolescent stress exposure. Schizophr Bull 43:592–600.
- Gomes FV, Zhu X, Grace AA (2020) The pathophysiological impact of stress on the dopamine system is dependent on the state of the critical period of vulnerability. Mol Psychiatry 25:3278–3291.
- Grace AA, Gomes FV (2019) The circuitry of dopamine system regulation and its disruption in schizophrenia: insights into treatment and prevention. Schizophr Bull 45:148–157.
- Guilloux J-P, Douillard-Guilloux G, Kota R, Wang X, Gardier AM, Martinowich K, Tseng GC, Lewis DA, Sibille E (2012) Molecular evidence for BDNF- and GABA-related dysfunctions in the amygdala of female subjects with major depression. Mol Psychiatry 17:1130–1142.
- Hardy KV, Mueser KT (2017) Editorial: trauma, psychosis and posttraumatic stress disorder. Front Psychiatry 8:220.
- Hauser J, Rudolph U, Keist R, Möhler H, Feldon J, Yee BK (2005) Hippocampal α 5 subunit-containing GABAA receptors modulate the expression of prepulse inhibition. Mol Psychiatry 10:201–207.
- Hausrat TJ, Muhia M, Gerrow K, Thomas P, Hirdes W, Tsukita S, Heisler FF, Herich L, Dubroqua S, Breiden P, Feldon J, Schwarz JR, Yee BK, Smart TG, Triller A, Kneussel M (2015) Radixin regulates synaptic GABAA receptor density and is essential for reversal learning and short-term memory. Nat Commun 6:6872. doi:https://doi.org/10.1038/ ncomms7872.
- Heckers S, Konradi C (2015) GABAergic mechanisms of hippocampal hyperactivity in schizophrenia. Schizophr Res 167:4–11.
- Howes OD, Montgomery AJ, Asselin M-C, Murray RM, Valli I, Tabraham P, Bramon-Bosch E, Valmaggia L, Johns L, Broome M, McGuire PK, Grasby PM (2009) Elevated striatal dopamine function linked to prodromal signs of schizophrenia. Arch Gen Psychiatry 66:13–20.
- Jacob TC (2019) Neurobiology and therapeutic potential of α5-GABA type A receptors. Front Mol Neurosci 12:179.
- Jacob TC, Moss SJ, Jurd R (2008) GABAA receptor trafficking and its role in the dynamic modulation of neuronal inhibition. Nat Rev Neurosci 9:331–343.

- Joshi SA, Duval ER, Kubat B, Liberzon I (2020) A review of hippocampal activation in post-traumatic stress disorder. Psychophysiology 57:e13357.
- Kiemes A, Gomes FV, Cash D, Uliana DL, Simmons C, Singh N, Vernon AC, Turkheimer F, Davies C, Stone JM, Grace AA, Modinos G (2021) GABAA and NMDA receptor density alterations and their behavioral correlates in the gestational methylazoxymethanol acetate model for schizophrenia. Neuropsychopharmacology 47:687–695. doi: 10.1038/s41386-021-01213-0.
- Kilpatrick DG, Ruggiero KJ, Acierno R, Saunders BE, Resnick HS, Best CL (2003) Violence and risk of PTSD, major depression, substance abuse/dependence, and comorbidity: results from the National Survey of Adolescents. J Consult Clin Psychol 71:692–700.
- Kilpatrick DG, Resnick HS, Milanak ME, Miller MW, Keyes KM, Friedman MJ (2013) National estimates of exposure to traumatic events and PTSD prevalence using DSM-IV and DSM-5 criteria. J Trauma Stress 26:537–547.
- Konradi C, Yang CK, Zimmerman EI, Lohmann KM, Gresch P, Pantazopoulos H, Berretta S, Heckers S (2011) Hippocampal interneurons are abnormal in schizophrenia. Schizophr Res 131:165–173.
- Laruelle M, Abi-Dargham A (1999) Dopamine as the wind of the psychotic fire: New evidence from brain imaging studies. J Psychopharmacol (Oxford, England) 13:358–371.
- Lieberman JA, Stroup TS, McEvoy JP, Swartz MS, Rosenheck RA, Perkins DO, Keefe RSE, Davis SM, Davis CE, Lebowitz BD, Severe J, Hsiao JK (2005) Effectiveness of antipsychotic drugs in patients with chronic schizophrenia. N Engl J Med 353:1209– 1223.
- Lin L-C, Sibille E (2013) Reduced brain somatostatin in mood disorders: a common pathophysiological substrate and drug target? Front Pharmacol 4:110.
- Lodge DJ, Behrens MM, Grace AA (2009) A loss of parvalbumincontaining interneurons is associated with diminished oscillatory activity in an animal model of schizophrenia. J Neurosci 29:2344–2354.
- Lodge DJ, Grace AA (2006a) The laterodorsal tegmentum is essential for burst firing of ventral tegmental area dopamine neurons. Proc Natl Acad Sci USA 103:5167–5172.
- Lodge DJ, Grace AA (2006b) The hippocampus modulates dopamine neuron responsivity by regulating the intensity of phasic neuron activation. Neuropsychopharmacology 31:1356–1361.
- Lodge DJ, Grace AA (2007) Aberrant hippocampal activity underlies the dopamine dysregulation in an animal model of schizophrenia. J Neurosci 27:11424–11430.
- Lodge DJ, Grace AA (2011) Hippocampal dysregulation of dopamine system function and the pathophysiology of schizophrenia. Trends in Pharmacol Sci. 32:507–513.
- Loebrich S, Bähring R, Katsuno T, Tsukita S, Kneussel M (2006) Activated radixin is essential for GABAA receptor α 5 subunit anchoring at the actin cytoskeleton. EMBO J 25:987–999.
- Olsen RW, Sieghart W (2009) GABAA receptors: subtypes provide diversity of function and pharmacology. Neuropharmacology 56:141–148.
- Pawlyk AC, Jha SK, Brennan FX, Morrison AR, Ross RJ (2005) A rodent model of sleep disturbances in posttraumatic stress disorder: the role of context after fear conditioning. Biol Psychiatry 57:268–277.
- Paxinos G, Watson C (1998) The rat brain in stereotaxic coordinates. San Diego. Academic Press.
- Perez SM, Boley A, Lodge DJ (2019) Region specific knockdown of Parvalbumin or Somatostatin produces neuronal and

behavioral deficits consistent with those observed in schizophrenia. Transl Psychiatry 9:1–13.

- Perez SM, Carreno FR, Frazer A, Lodge DJ (2014) Vagal nerve stimulation reverses aberrant dopamine system function in the methylazoxymethanol acetate rodent model of schizophrenia. J Neurosci 34:9261–9267.
- Perez SM, Lodge DJ (2013) Hippocampal interneuron transplants reverse aberrant dopamine system function and behavior in a rodent model of schizophrenia. Mol Psychiatry 18:1193–1198.
- Perez SM, Lodge DJ (2018) Convergent inputs from the hippocampus and thalamus to the nucleus accumbens regulate dopamine neuron activity. J Neurosci 38:10607–10618.
- Perez SM, McCoy AM, Prevot TD, Mian Y, Carreno FR, Frazer A, Cook JM, Sibille E, Lodge DJ (2022) Hippocampal α5-GABAA receptors modulate dopamine neuron activity in the rat ventral tegmental area. Biol Psychiatry GOS S2667174322000039. doi: 10.1016/j.bpsgos.2021.12.010 In press. Available online at: https://www.sciencedirect.com/science/article/pii/ S2667174322000039.
- Perez SM, Shah A, Asher A, Lodge DJ (2013) Hippocampal deep brain stimulation reverses physiological and behavioural deficits in a rodent model of schizophrenia. Int J Neuropsychopharmacol 16:1331–1339.
- Prevot TD, Li G, Vidojevic A, Misquitta KA, Fee C, Santrac A, Knutson DE, Stephen MR, Kodali R, Zahn NM, Arnold LA, Scholze P, Fisher JL, Marković BD, Banasr M, Cook JM, Savic M, Sibille E (2019) Novel benzodiazepine-like ligands with various anxiolytic, antidepressant, or pro-cognitive profiles. Mol Neuropsychiatry 5:84–97.
- Prévot T, Sibille E (2020) Altered GABA-mediated information processing and cognitive dysfunctions in depression and other brain disorders. Mol Psychiatry. doi: 10.1038/s41380-020-0727-3
- Prevot TD, Sumitomo A, Tomoda T, Knutson DE, Li G, Mondal P, Banasr M, Cook JM, Sibille E (2020) Reversal of age-related neuronal atrophy by α5-GABAA receptor positive allosteric modulation. Cereb (New York, N.Y.: 1991). doi: 10.1093/cercor/ bhaa310
- Pynoos RS, Ritzmann RF, Steinberg AM, Goenjian A, Prisecaru I (1996) A behavioral animal model of posttraumatic stress disorder featuring repeated exposure to situational reminders. Biol Psychiatry 39:129–134.
- Quinones MM, Gallegos AM, Lin FV, Heffner K (2020) Dysregulation of inflammation, neurobiology, and cognitive function in PTSD: an integrative review. Cogn Affect Behav Neurosci 20:455–480.
- Semple WE, Goyer PF, McCormick R, Donovan B, Muzic RF, Rugle L, McCutcheon K, Lewis C, Liebling D, Kowaliw S, Vapenik K,

Semple MA, Flener CR, Schulz SC (2000) Higher brain blood flow at amygdala and lower frontal cortex blood flow in PTSD patients with comorbid cocaine and alcohol abuse compared with normals. Psychiatry 63:65–74.

- Seney ML, Tripp A, McCune S, Lewis DA, Sibille E (2015) Laminar and cellular analyses of reduced somatostatin gene expression in the subgenual anterior cingulate cortex in major depression. Neurobiol Dis 73:213–219.
- Shin LM, Rauch SL, Pitman RK (2006) Amygdala, medial prefrontal cortex, and hippocampal function in PTSD. Ann N Y Acad Sci 1071:67–79.
- Simon J, Wakimoto H, Fujita N, Lalande M, Barnard EA (2004) Analysis of the set of GABAA receptor genes in the human genome*. J Biol Chem 279:41422–41435.
- Stein MB, Koverola C, Hanna C, Torchia MG, McClarty B (1997) Hippocampal volume in women victimized by childhood sexual abuse. Psychol Med 27:951–959.
- Sun X, Song Z, Si Y, Wang J-H (2018) MicroRNA and mRNA profiles in ventral tegmental area relevant to stress-induced depression and resilience. Prog Neuropsychopharmacol Biol Psychiatry 86:150–165.
- Sur C, Fresu L, Howell O, McKernan RM, Atack JR (1999) Autoradiographic localization of alpha5 subunit-containing GABAA receptors in rat brain. Brain Res 822:265–270.
- Tomoda T, Sumitomo A, Newton D, Sibille E (2022) Molecular origin of somatostatin-positive neuron vulnerability. Mol Psychiatry. doi: 10.1038/s41380-022-01463-4
- Tripp A, Kota RS, Lewis DA, Sibille E (2011) Reduced somatostatin in subgenual anterior cingulate cortex in major depression. Neurobiol Dis 42:116–124.
- Ungless MA, Grace AA (2012) Are you or aren't you? Challenges associated with physiologically identifying dopamine neurons. Trends Neurosci 35:422–430.
- Valenti O, Lodge DJ, Grace AA (2011) Aversive stimuli alter ventral tegmental area dopamine neuron activity via a common action in the ventral hippocampus. J Neurosci 31:4280–4289.
- Van Dijken HH, Van der Heyden JA, Mos J, Tilders FJ (1992) Inescapable footshocks induce progressive and long-lasting behavioural changes in male rats. Physiol Behav 51:787–794.
- Villarreal G, Hamilton DA, Petropoulos H, Driscoll I, Rowland LM, Griego JA, Kodituwakku PW, Hart BL, Escalona R, Brooks WM (2002) Reduced hippocampal volume and total white matter volume in posttraumatic stress disorder. Biol Psychiatry 52:119–125.
- Zhang ZJ, Reynolds GP (2002) A selective decrease in the relative density of parvalbumin-immunoreactive neurons in the hippocampus in schizophrenia. Schizophr Res 55:1–10.