## Expression of NG2 proteoglycan in the degenerated intervertebral disc in dachshunds

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ABSTRACT. The pathogenesis of intervertebral disc (IVD) degeneration is not fully understood. The biomolecular signaling pathways involved in the IVD degeneration require further investigation. The aim of this study was to investigate the expression of NG2 proteoglycan in the degenerated IVD. IVD samples were obtained from 16 Dachshunds that were confirmed to have IVD herniation and subsequently underwent hemilaminectomy. The samples were subjected to histological and immunohistochemical (IHC) examinations. IHC revealed positive results for the expression of NG2 proteoglycan in all examined samples. The results showed the expression of NG2 proteoglycan by the degenerated IVDs.

KEY WORDS: dachshund, immunohistochemistry, intervertebral disc (IVD) degeneration, NG2 proteoglycan

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The detailed pathogenesis of intervertebral disc (IVD) degeneration and its underlying mechanism in dogs is not fully known [5]. Although the process of IVD degeneration has been described histopathologically [3, 10], the biomolecular signaling pathways involved in the IVD degeneration require further investigation [7, 21, 22].

The mechanism of degeneration may be associated with a variety of changes that involve the cells and extracellular matrix (ECM) of all parts of the IVD [5, 9]. The normal IVD is composed of cells embedded in the ECM [5, 6, 10]. The ECM is mainly formed from collagen and proteoglycan, which assemble into multiple matrix compartments [6, 10]. Chondroitin sulfate proteoglycans (CSPGs), such as neurocan, aggrecan, versican, brevican, phosphacan and NG2 (nerve/glial antigen 2), are expressed throughout the developing and adult central nervous system (CNS) in rat and human [13, 20, 28].

NG2 (or CSPG4), is a transmembrane CSPG that participates in both extracellular and intracellular events in various tissues [1, 8, 16, 19, 26]. Moreover, NG2 binds to type VI collagen, which is the main constituent of the pericellular matrix in the IVD annulus fibrosis (AF) and nucleus pulposus (NP) and articular cartilage [1, 18]. The changes in the pericellular microenvironment contribute to the pathoetiology of connective tissue diseases, such as osteoarthritis and IVD degeneration [1]. The expression of NG2 proteoglycan has been observed in cells of the human IVD with a high level of advanced degeneration [1]. There is no report on the expression of NG2 in IVD cells in dogs.

The purpose of this study was to investigate the expression of NG2 proteoglycan in the degenerated IVD of dachshund dogs using immunohistochemistry.

Herniated disc samples were obtained from 16 clientowned dachshunds that underwent to surgical treatment for IVD herniation. The animals were subjected to routine physical, clinical and neurological examinations, and blood samples were collected for CBC (complete blood count) and serum biochemical analysis. Radiography, myelography and computed tomography (CT) or magnetic resonance imaging (MRI) were performed. All the animals were subjected to decompression surgery, and the extruded disc samples were removed and preserved for histological and immunohistochemical examinations.

All histological samples were immediately fixed in 10% neutral buffered formalin and subsequently embedded in paraffin wax. Serial sections (4  $\mu$ m) were cut and fixed onto silanized glass slides for routine hematoxylin and eosin (H&E) staining and were examined using light microscopy. The sections were assessed by two independent observers. Histopathological grading was performed according to the modified Bergknut histological grading system [15].

Immunohistochemistry (IHC) was conducted on formalin-fixed, paraffin-embedded sections. Antigen retrieval

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Fig. 1. Correlation between the NG2 proteoglycan expression level and age of dogs (A), clinical grade (B) and histological grade (C). NG2 expression was not correlated with age of dogs, clinical grades and histological grades (r=-0.04, P=0.887; r=0.111, P=0.68 and r=0.178, P=0.51, respectively, confidence interval (CI)=95%). P>0.05 was considered to be nonsignificant.

and tissue section processing were carried out as standard procedures. To determine the NG2 proteoglycan expression, a rabbit polyclonal anti-human CSPG4 antibody (diluted 1:100, Sigma-Aldrich, St. Louis, MO, U.S.A.) was used. In negative control sections (*canine* lung tissue), the primary antibody was omitted or replaced with its respective serum. Samples of *canine* melanoma expressing the antigen were used as a positive control [17]. All sections were stained in the same session.

The adoptive scoring system was that described by Itoh *et al.* [11]; cell counts were performed by 2 independent observers. More than 100 cells in each sample were examined to evaluate the proportion of cells that showed a positive reaction for NG2 proteoglycan.

Statistical analysis was performed using Pearson linear regression to determine the association between age, clinical grades and degenerative grades with the level of NG2 proteoglycan expression. The linear correlation coefficient (r) and 95% confidence interval (CI) were recorded. Values of P<0.05 were considered significant.

A total of 16 male client-owned dachshunds were included in this study. Their ages ranged from 3 to 12 years (mean 7.9 years). A preoperative neurological examination was performed for all dogs, and it showed that the overall median clinical grade was IV. The clinical neurological grades were III (1/16, 6.25%), IV (8/16, 50%) and V (7/16, 43.75%). There was no correlation between the age of animals (r=-0.04, CI=95% and P=0.887) (Fig. 1A), clinical grades (r=0.111, CI=95% and P=0.68) (Fig. 1B) and histological grades (r=0.178, CI=95% and P=0.51) (Fig. 1C) and the level of NG2 expression by the degenerated IVD samples.

The surgically obtained IVD samples consisted of the AF

and NP. These was marked chondrocyte proliferation in the AF (up to the outer layers of the AF) in all cases, while the proliferation of chondrocytes in the NP comprised small clones (2–7 cells) in 6/16 dogs (38%), medium sized clones (8–15 cells) in 4/16 dogs (25%), huge clones (>15 cells) in 5/16 dogs (31%) and formation of scar tissue in 1/16 dogs (6%). Notochordal cells were present (1–50%) in 5/16 dogs (31%) and completely absent in 11/16 dogs (69%). Representative histopathological examples are shown in Fig. 2.

The expression of NG2 using IHC was observed in all herniated disc tissues. The brown color in immunohistochemical staining was restricted to the cytoplasm and cell membrane of the chondrocyte, and the nucleus was stained blue with hematoxylin. Representative immunohistochemical examples are shown in Fig. 3.

The results of this study demonstrated, for the first time, that canine intervertebral disc cells express NG2 proteoglycan and that it can be detected by immunohistochemical examination.

The occurrence of IVD degeneration and the transformation of the NP from the gelatinous to dried form at the thoracolumbar discs during early life in chondrodystrophic dogs may have caused the lack of correlation between age of animals (3–12 years) and the level of NG2 expression in this study. Furthermore, the expression of NG2 proteoglycan might be restricted to dogs of a certain age, but it might be expressed by cells in fetal, normal and degenerative adult IVDs as reported in human osteoarthritic articular cartilage [10, 18, 24].

The clinical grades due to spinal cord dysfunction depend upon the location, size, severity and rate of development of the lesion that occurs as a result of the compressive force of



Fig. 2. Representative histological images showing chondrocyte-like cells proliferation in the nucleus pulposus (NP) of degenerative IVDs (H&E staining). (A) Moderate sized clones (8–15 cells) in the case of grade IV IVD degeneration. (B) Huge clones (more than 15 cells) in the case of grade V degeneration. Bar: 200 μm (A), 100 μm (B).



Fig. 3. Immunohistochemical staining of canine degenerative IVD samples with an anti CSPG4 antibody showing positively labeled NG2-expressing chondrocyte-like cells (brown-colored cells). (A) grade IV, (B) grade V degenerated IVD samples. Bar: 200 μm (A), 100 μm (B).

extruded disc material on the spinal cord parenchyma and/ or the nerve roots [23]. So, as shown in the results of this study, the clinical grades are not correlated with the degree of degeneration of IVDs and level of NG2 expression. In addition, there is a difference between IVD degeneration and herniation. The former can give rise to a number of diseases, one of them IVD herniation [4], which may occur in normal (non-degenerated) discs due to severe trauma [10].

Although a previous study reported a positive relationship between the degenerative grade of IVDs and the level of NG2 expression [1], the results of the present study did not confirm this relationship. This may be due to the degree of degeneration of almost examined samples which was either grade IV or V, while the other grades were not represented in this study. Moreover, the small number of animals included in this study is considered one of its limitations.

The expression of NG2 was restricted only to the cytoplasm and cell membrane of the chondrocytes. This is in agreement with previous studies that described the structure of NG2 as a large extracellular domain (2224 amino acids), a single small transmembrane domain (25 amino acids) and a short cytoplasmic domain (76 amino acids) [25, 27].

Although the loss of notochordal cells from the NP is as-

sociated with the aging process and disc degeneration, the results of this study showed the presence of notochordal cells in samples from 5/16 dogs, which ranged in age from 3-12 years. The persistence of notochordal cells in chondrodystrophic dogs was reported in previous studies [6, 10, 12]. The normal NP in chondrodystrophic dogs has 2 types of notochordal cells. Large cells represent about 13% of total NP cells, and small cells account for the rest. The large cells are able to synthesize 1.5 times proteoglycan than the small cells [6]. The degenerative process of the IVD is characterized by a decrease in proteoglycan level and increase in collagen fibers as a result of the replacement of the notochordal cells with chondrocyte-like cells. So, the large notochordal cells may have a more important role in maintaining the integrity of IVD than smaller cells [2, 6], some of which may persist in the NP of the IVD of chondrodystrophic breeds.

Increase of the number of chondrocyte-like cells in the NP is in agreement with previous studies, which stated that the NP is replaced by chondrocyte-like cells embedded in a large amount of extracellular matrix at an early age in chondrodystrophic dogs in a process called chondrification or chondroid metaplasia of the NP [5, 6, 10, 14].

In conclusion, the results of this short study indicated that

NG2 proteoglycan is expressed by the degenerated IVD.

## REFERENCES

- Akeda, K., An, H. S., Pichika, R., Patel, K., Muehleman, C., Nakagawa, K., Uchida, A. and Masuda, K. 2007. The expression of NG2 proteoglycan in the human intervertebral disc. *Spine* 32: 306–314. [Medline] [CrossRef]
- Aguiar, D. J., Johnson, S. L. and Oegema, T. R. 1999. Notochordal cells interact with nucleus pulposus cells: regulation of proteoglycan synthesis. *Exp. Cell Res.* 246: 129–137. [Medline] [CrossRef]
- Bergknut, N., Meij, B. P., Hagman, R., de Nies, K., Rutges, J. P., Smolders, L. A., Creemers, L. B., Lagerstedt, A. S., Hazewinkel, H. A. and Grinwis, G. C. 2013. Intervertebral disc disease in dogs – Part 1: a new histological grading scheme for classification of intervertebral disc degeneration in dogs. *Vet. J.* 195: 156–163. [Medline] [CrossRef]
- Bergknut, N., Rutges, J. P., Kranenburg, H. J., Smolders, L. A., Hagman, R., Smidt, H. J., Lagerstedt, A. S., Voorhout, G., Hazewinkel, H. A. W., Grinwis, G. C., Creemers, L. B., Meij, B. P. and Dhert, W. J. 2012. The dog as an animal model for intervertebral disc degeneration? *Spine* 37: 351–358. [Medline] [CrossRef]
- Bergknut, N., Smolders, L. A., Grinwis, G. C., Hagman, R., Lagerstedt, A. S., Hazewinkel, H. A. W., Tryfonidou, M. A. and Meij, B. P. 2013. Intervertebral disc degeneration in the dog. Part 1: anatomy and physiology of the intervertebral disc and characteristics of intervertebral disc degeneration. *Vet. J.* 195: 282–291. [Medline] [CrossRef]
- Cappello, R., Bird, J. L., Pfeiffer, D., Bayliss, M. T. and Dudhia, J. 2006. Notochordal cell produce and assemble extracellular matrix in a distinct manner, which may be responsible for the maintenance of healthy nucleus pulposus. *Spine* **31**: 873–882. [Medline] [CrossRef]
- Erwin, W. M. 2010. The enigma that is the nucleus pulposus cell: the search goes on. *Arthritis Res. Ther.* 12: 118–119. [Medline] [CrossRef]
- Fidler, P. S., Schuette, K., Asher, R. A., Dobbertin, A., Thornton, S. R., Calle-Patino, Y., Muir, E., Levine, J. M., Geller, H. M., Rogers, J. H., Faissner, A. and Fawcett, J. W. 1999. Comparing astrocytic cell lines that are inhibitory or permissive for axon growth: the major axon-inhibitory proteoglycan is NG2. *J. Neurosci.* 19: 8778–8788. [Medline]
- Goupille, P., Jayson, M. I., Valat, J. P. and Freemont, A. J. 1998. Matrix metalloproteinases: the clue to intervertebral disc degeneration? *Spine* 23: 1612–1626. [Medline] [CrossRef]
- Hansen, H. J. 1952. A pathologic-anatomical study on disc degeneration in dog, with special reference to the so-called enchondrosis intervertebralis. *Acta Orthop. Scand. Suppl.* 11: 1–117. [Medline] [CrossRef]
- Itoh, H., Hara, Y., Tagawa, M., Kato, T., Ochi, H., Koga, D., Okawa, A. and Asou, Y. 2012. Evaluation of the association between runt-related transcription factor 2 expression and intervertebral disk aging in dogs. *Am. J. Vet. Res.* **73**: 1553–1559. [Medline] [CrossRef]
- Johnson, J. A., da Costa, R. C. and Allen, M. J. 2010. Micromorphometry and cellular characteristics of the canine cervical intervertebral discs. *J. Vet. Intern. Med.* 24: 1343–1349. [Medline] [CrossRef]
- 13. Jones, L. L., Margolis, R. U. and Tuszynski, M. H. 2003. The

chondroitin sulfate proteoglycans neurocan, brevican, phosphacan, and versican are differentially regulated following spinal cord injury. *Exp. Neurol.* **182**: 399–411. [Medline] [CrossRef]

- Klauser, M., Forterre, F., Doherr, M., Zurbriggen, A., Spreng, D. and Forterre, S. 2012. Evaluation of apoptotic cell death in normal and chondrodystrophic canine intervertebral discs. *Veterinary Sci. Dev.* 2: 20–24. [CrossRef]
- Kranenburg, H. J., Grinwis, G. C., Bergknut, N., Gahrmann, N., Voorhout, G., Hazewinkel, H. A. and Meij, B. P. 2013. Intervertebral disc disease in dogs – Part 2: comparison of clinical, magnetic resonance imaging, and histological findings in 74 surgically treated dogs. *Vet. J.* 195: 164–171. [Medline] [CrossRef]
- Levine, J. M. 1994. Increased expression of the NG2 chondroitin-sulfate proteoglycan after brain injury. J. Neurosci. 14: 4716–4730. [Medline]
- Mayayo, S. L., Prestigio, S., Maniscalco, L., Rosa, G. L., Aricò, A., Maria, R. D., Cavallo, F., Ferrone, S., Buracco, P. and Iussich, S. 2011. Chondroitin sulfate proteoglycan-4: a biomarker and a potential immunotherapeutic target for canine malignant melanoma. *Vet. J.* **190**: e26–30. [Medline] [CrossRef]
- Midwood, K. S. and Salter, D. M. 2001. NG2/HMPG modulation of human articular chondrocyte adhesion to type VI collagen is lost in osteoarthritis. *J. Pathol.* 195: 631–635. [Medline] [CrossRef]
- Nishiyama, A., Dahlin, K. J., Prince, J. T., Johnstone, S. R. and Stallcup, W. B. 1991. The primary structure of NG2, a novel membrane-spanning proteoglycan. *J. Cell Biol.* **114**: 359–371. [Medline] [CrossRef]
- Pantazopoulos, H., Murray, E. A. and Berretta, S. 2008. Total number, distribution, and phenotype of cells expressing chondroitin sulfate proteoglycans in the normal human amygdala. *Brain Res.* 1207: 84–95. [Medline] [CrossRef]
- Risbud, M. V. and Shapiro, I. M. 2011. Notochordal cells in the adult intervertebral disc: new perspective on an old question. *Crit. Rev. Eukaryot. Gene Expr.* 21: 29–41. [Medline] [Cross-Ref]
- Risbud, M. V., Schaer, T. P. and Shapiro, I. M. 2010. Toward an understanding of the role of notochordal cells in the adult intervertebral disc: from discord to accord. *Dev. Dyn.* 239: 2141–2148. [Medline] [CrossRef]
- Sharp, N. and Wheeler, S. 2005. Small Animal Spinal Disorders: Diagnosis and Surgery, 2nd ed., Elsevier Mosby, Philadelphia.
- Smolders, L. A., Bergknut, N., Grinwis, G. C., Hagman, R., Lagerstedt, A. S., Hazewinkel, H. A., Tryfonidou, M. A. and Meij, B. P. 2013. Intervertebral disc degeneration in the dog. Part 2: chondrodystrophic and non-chondrodystrophic breeds. *Vet. J.* 195: 292–299. [Medline] [CrossRef]
- Stallcup, W. B. 1981. The NG2 antigen, a putative lineage marker: immunofluorescent localization in primary cultures of rat brain. *Dev. Biol.* 83: 154–165. [Medline] [CrossRef]
- Stallcup, W. B. 2002. The NG2 proteoglycan: past insights and future prospects. *J. Neurocytol.* 31: 423–435. [Medline] [Cross-Ref]
- Stallcup, W. B., Beasley, L. and Levine, J. M. 1983. Cell surface molecules that characterize different stages in the development of cerebellar interneurons. *Cold Spring Harb. Symp. Quant. Biol.* 48: 761–774. [Medline] [CrossRef]
- Yamaguchi, Y. 2000. Chondroitin sulfate proteoglycans in the nervous system. pp. 379–402. *In*: Proteoglycans: Structure, Biology, and Molecular Interactions, 1st ed. (Iozzo, R.V. ed.), Marcel Dekker, Inc., New York.