

Research article

Open Access

## Adenomatous polyposis coli-mediated control of $\beta$ -catenin is essential for both chondrogenic and osteogenic differentiation of skeletal precursors

Razvan L Miclea<sup>†1</sup>, Marcel Karperien<sup>\*†2</sup>, Cathy AJ Bosch<sup>3</sup>, Geertje van der Horst<sup>4</sup>, Martin A van der Valk<sup>5</sup>, Tatsuya Kobayashi<sup>6</sup>, Henry M Kronenberg<sup>6</sup>, Georges Rawadi<sup>7</sup>, Pinar Akçakaya<sup>3</sup>, Clemens WGM Löwik<sup>8</sup>, Riccardo Fodde<sup>9</sup>, Jan Maarten Wit<sup>1</sup> and Els C Robanus-Maandag<sup>3</sup>

Address: <sup>1</sup>Department of Pediatrics, Leiden University Medical Centre, Leiden, the Netherlands, <sup>2</sup>Department of Tissue Regeneration, Institute for Biomedical Technology, University of Twente, Enschede, the Netherlands, <sup>3</sup>Department of Human Genetics, LUMC, Leiden, the Netherlands, <sup>4</sup>Department of Urology, Leiden University Medical Centre, Leiden, the Netherlands, <sup>5</sup>Department of Animal Pathology, The Netherlands Cancer Institute, Amsterdam, the Netherlands, <sup>6</sup>Department of Medicine, Endocrine Unit, Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts, USA, <sup>7</sup>Galapagos, Romainville, 93230, France, <sup>8</sup>Department of Endocrinology and Metabolic Diseases, Leiden University Medical Centre, Leiden, the Netherlands and <sup>9</sup>Department of Pathology, Josephine Nefkens Institute, Erasmus Medical Centre, Rotterdam, the Netherlands

Email: Razvan L Miclea - r.l.miclea@lumc.nl; Marcel Karperien\* - h.b.j.karperien@tnw.utwente.nl; Cathy AJ Bosch - c.a.j.bosch@lumc.nl; Geertje van der Horst - g.van\_der\_horst@lumc.nl; Martin A van der Valk - ma.vd.valk@nki.nl; Tatsuya Kobayashi - kobayash@HELIX.MGH.HARVARD.EDU; Henry M Kronenberg - hkronenberg@partners.org; Georges Rawadi - georges.rawadi@glpg.com; Pinar Akçakaya - pinarakakaya@gmail.com; Clemens WGM Löwik - c.w.g.m.lowik@lumc.nl; Riccardo Fodde - r.fodde@erasmusmc.nl; Jan Maarten Wit - j.m.wit@lumc.nl; Els C Robanus-Maandag - e.c.robanus@lumc.nl

\* Corresponding author †Equal contributors

Published: 8 April 2009

Received: 4 December 2008

BMC Developmental Biology 2009, 9:26 doi:10.1186/1471-213X-9-26

Accepted: 8 April 2009

This article is available from: <http://www.biomedcentral.com/1471-213X/9/26>

© 2009 Miclea et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** During skeletogenesis, protein levels of  $\beta$ -catenin in the canonical Wnt signaling pathway determine lineage commitment of skeletal precursor cells to osteoblasts and chondrocytes. Adenomatous polyposis coli (Apc) is a key controller of  $\beta$ -catenin turnover by down-regulating intracellular levels of  $\beta$ -catenin.

**Results:** To investigate whether Apc is involved in lineage commitment of skeletal precursor cells, we generated conditional knockout mice lacking functional Apc in *Col2a1*-expressing cells. In contrast to other models in which an oncogenic variant of  $\beta$ -catenin was used, our approach resulted in the accumulation of wild type  $\beta$ -catenin protein due to functional loss of Apc. Conditional homozygous Apc mutant mice died perinatally showing greatly impaired skeletogenesis. All endochondral bones were misshaped and lacked structural integrity. Lack of functional Apc resulted in a pleiotropic skeletal cell phenotype. The majority of the precursor cells lacking Apc failed to differentiate into chondrocytes or osteoblasts. However, skeletal precursor cells in the proximal ribs were able to escape the noxious effect of functional loss of Apc resulting in formation of highly active osteoblasts. Inactivation of Apc in chondrocytes was associated with dedifferentiation of these cells.

**Conclusion:** Our data indicate that a tight Apc-mediated control of  $\beta$ -catenin levels is essential for differentiation of skeletal precursors as well as for the maintenance of a chondrocytic phenotype in a spatio-temporal regulated manner.

## Background

During vertebrate embryogenesis, the axial and appendicular skeleton develop through endochondral bone formation. In this process, mesenchymal cells aggregate to form a chondrocytic template that prefigures the shape of the future bone. At the periphery of this cartilaginous mold, osteoblasts differentiate to form the bone collar. The cartilaginous mold is eventually replaced by bone in a step-wise program. Besides chondrocytes and osteoblasts, the skeleton also contains osteoclasts, which are of haematopoietic origin and play pivotal roles in both cartilage and bone resorption and remodelling [1-3].

Every step in the proliferation, differentiation, maturation, apoptosis, and resorption of both chondrocytes and osteoblasts is characterized by a specific transcriptional guideline [4]. Sox9, a high-mobility-group transcription factor, and Runx2, a Runt domain transcription factor, are both expressed in bi-potential skeletal precursor cells differentiating into either chondrocytes or osteoblasts [5-7]. Sox9 and Runx2 play leading roles in lineage commitment of these precursors: upregulation of Sox9 leads to chondrogenic differentiation [8], while activation of Runx2 is required for their osteogenic commitment [9].

Recently, based on mouse models, the canonical Wnt/ $\beta$ -catenin signaling pathway was found to act upstream of Sox9 and Runx2. In this pathway, in the absence of a Wnt signal, cytosolic  $\beta$ -catenin is degraded by the ubiquitination/proteasome system upon its phosphorylation at specific Ser-Thr residues by a destruction complex consisting of scaffold proteins such as Axin1, Axin2 (also known as Conductin) and the adenomatous polyposis coli (APC) tumor suppressor, and two kinases, namely glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) and casein-kinase 1 $\alpha$  (CK1 $\alpha$ ). Binding of Wnt to a complex composed of the transmembrane frizzled receptor and low-density lipoprotein receptor-related protein 5 or 6 (LRP5 or 6) co-receptor results in inactivation of the destruction complex and accumulation of cytoplasmic  $\beta$ -catenin. Upon its nuclear translocation,  $\beta$ -catenin acts as transcriptional co-activator in complex with transcription factors of the TCF/LEF family, leading to transcriptional activation of Wnt target genes [10]. In wild type mouse embryos, high levels of  $\beta$ -catenin and activation of canonical Wnt signaling have been found in osteoblastic precursors in developing skull and limb bones [11]. Accumulating evidence suggests that increased levels of canonical Wnt/ $\beta$ -catenin signaling inhibit Sox9 expression and activity, and stimulate Runx2 expression, leading to decreased chondrocyte differentiation and increased osteoblast differentiation, respectively [12-15]. Similar results have been found in transgenic mice with *Wnt14* overexpression in *Collagen 2a1* (*Col2a1*)-expressing cells [11].

It has been also demonstrated that  $\beta$ -catenin is required at an early stage to repress chondrocytic differentiation [15]. Upon conditional inactivation of  $\beta$ -catenin in the limb and head mesenchyme before or during early mesenchymal condensations, *Prx1*-expressing and *Dermo1*-expressing skeletal precursors, respectively, differentiate into chondrocytes instead of osteoblasts [11,15]. Finally, results on both constitutively active and inactivated  $\beta$ -catenin in *Osterix* (*Osx*)-, *Collagen 1a1* (*Col1a1*)- or *Osteocalcin* (*Osc*)-expressing osteoblasts suggest that Wnt/ $\beta$ -catenin signaling coordinates bone formation by controlling the differentiation and activity of both osteoblasts and osteoclasts in a sequential, stage-specific manner [16,17].

Little is known about the mechanisms regulating  $\beta$ -catenin activity in skeletal precursors. Through its wide range of specific motifs and domains, APC is involved in multiple cellular processes such as signal transduction, cytoskeletal organization, apoptosis, cell adhesion and motility, cell fate determination, and chromosomal stability [18]. However, biochemical and genetic evidence has been provided showing that APC's main tumor suppressing activity resides in its ability to bind to  $\beta$ -catenin and induce its degradation, thereby acting as a strong negative regulator of the canonical Wnt pathway [19-21].

Familial adenomatous polyposis (FAP) patients heterozygous for an APC mutation frequently develop osteomas and dental anomalies [22]. Heterozygous *Apc*<sup>1638N</sup> mutant mice occasionally develop osteomas (R. Fodde, personal communication). Homozygosity for the severely truncated *Apc*<sup>Min</sup> and for the more hypomorphic *Apc*<sup>1638N</sup> allele in the mouse results in a failure of primitive ectoderm development shortly after implantation, leading to lethality prior to gastrulation [23,24]. Mutant *Apc* disturbs the differentiation capacity of mouse embryonic stem (ES) cells in a quantitative and qualitative fashion depending on the dose of  $\beta$ -catenin signaling. Aberrant differentiation capacity of ES cells ranges from a strong differentiation blockade in case of two severely truncated *Apc*<sup>Min</sup> alleles, to more specific neuroectodermal, dorsal mesodermal, and endodermal defects (e.g., no differentiation in bone or cartilage) in case of two hypomorphic *Apc*<sup>1638N</sup> alleles [25,26]. Osteoblast-specific loss of *Apc* in the mouse leads to early onset of dramatically increased bone deposition and to lethality early in life [17]. However, *Apc* has not yet been linked with a role in the differentiation of skeletal precursor cells.

Here, we report that skeletal precursors of the axial and appendicular skeleton, when exposed to an uncontrolled rise of the  $\beta$ -catenin level due to conditional inactivation of *Apc*, lose their differentiation capacity to both the chon-

drogenic and osteogenic lineage. Moreover, conditional *Apc* mutant ribs show enhanced osteoblast activity, while the mutant nasal septum displays chondrocyte dedifferentiation. These results provide the first genetic evidence that *Apc* plays a crucial role throughout mouse skeletogenesis by regulating the differentiation of skeletal progenitor cells and maintenance of chondrocytes.

## Results

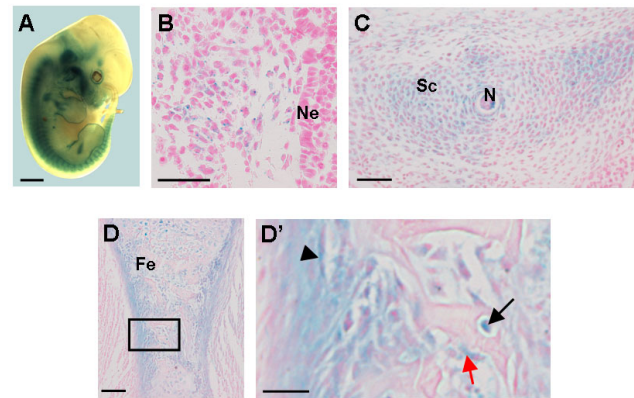
### Conditional *Apc*<sup>15lox</sup> mice and transgenic *Col2a1-Cre* mice

Recently, we (ECR-M and RF) generated a novel mouse model carrying a conditional *Apc*<sup>15lox</sup> allele where exon 15, encoding the majority of the coding region of *Apc*, and the polyadenylation signal, is flanked by loxP sites. Mice heterozygous and homozygous for the conditional *Apc*<sup>15lox</sup> allele did not show any major abnormalities or susceptibility to tumors. *Apc*<sup>A15/+</sup> mice, heterozygous for the *Apc*<sup>A15</sup> mutant allele obtained by germline Cre-mediated deletion of exon 15, developed multiple intestinal tumors at an early age similar to *Apc*<sup>Min/+</sup> animals. These results indicate that Cre-mediated recombination of the *Apc*<sup>15lox</sup> allele leads to inactivation of the *Apc* protein and to the constitutive activation of Wnt/ $\beta$ -catenin signaling (Robanus-Maandag et al., in preparation).

Next, we investigated the temporal and spatial expression pattern of *Cre* in transgenic *Col2a1-Cre* mice [27] using *LacZ* reporter mice ("Rosaflox") [28]. *Col2a1-Cre;Rosaflox* embryos expressed *Cre* specifically at all sites of endochondral bone formation (Fig. 1A). In accordance with previous studies suggesting that *Col2a1* is already expressed at E9.5 in the sclerotome of the somites [29], we detected *Cre* activity (based on positive *LacZ* staining) in mesenchymal condensations forming the sclerotome at E9.5 (Fig. 1B). At E12.5, *LacZ*-positive cells were identified in cartilage primordia later forming the vertebrae, long bones, sternum and cranial bones (Fig. 1C; data not shown). As reported in other *Col2a1-Cre* mouse lines [11,30], we found *LacZ* staining in the perichondrium at E14.5 (data not shown), and in the periosteum and primary spongiosa of long bones at E16.5, sites where osteoblasts normally differentiate (Fig. 1D, D'). The early onset (E9.5) of the *LacZ* expression in the sclerotome as well as its presence at later developmental stages (E14.5 and E16.5) in cells of the osteogenic lineage prompted us to conclude that the *Col2a1-Cre*-mediated recombination occurred in skeletal precursors characterized by both a chondrogenic and osteogenic differentiation potential.

### Heterozygous *Apc*<sup>15lox/+</sup> mice do not show any skeletal defect upon *Col2a1*-driven *Cre* expression

*Apc*<sup>15lox/15lox</sup> mice were bred with *Col2a1-Cre* mice to generate conditional heterozygous *Col2a1-Cre;Apc*<sup>15lox/+</sup> mice. Microscopical analysis performed on *Col2a1-Cre;Apc*<sup>15lox/+</sup> and control *Apc*<sup>15lox/+</sup> embryos at various

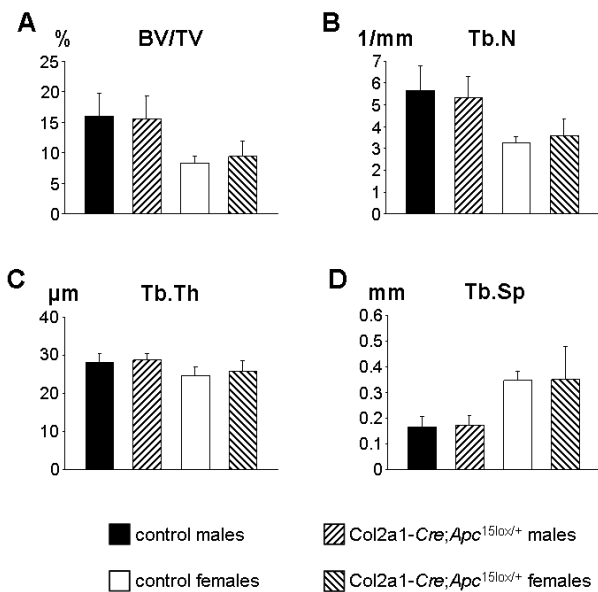


**Figure 1**

***Col2a1-Cre;Rosaflox* mice express *Cre* at sites of endochondral bone formation.** (A-D) *LacZ* expression in *Col2a1-Cre;Rosaflox* embryos following *Cre* recombination, detected by whole-mount X-Gal staining. (A) Macroscopic picture of E12.5 *Col2a1-Cre;Rosaflox* embryo. (B) Transversal section of E9.5 embryo showing  $\beta$ -galactosidase-positive sclerotomal cells adjacent to the neural tube. (C) Transversal section of E12.5 embryo showing *LacZ* expression in vertebrae primordia. (D) Sagittal section of E16.5 embryo showing *LacZ* expression in the femur. The boxed region in D is magnified in D' showing *LacZ* expression in the periosteum (arrow head), osteoblasts (red arrow) and osteocytes (black arrow). Ne, neuroepithelium; Sc, sclerotome; N, notochord; Fe, femur. Scale bars: 1 mm in A; 50  $\mu$ m in B, D'; 100  $\mu$ m in C, D.

developmental stages (E12.5, E14.5, E16.5) displayed a normal spatio-temporal expression of all chondrogenic and osteogenic markers investigated (data not shown).

To study postnatal growth and bone acquisition, 18 *Col2a1-Cre;Apc*<sup>15lox/+</sup> mice (7 males, 11 females) and 11 *Apc*<sup>15lox/+</sup> mice (7 males, 4 females) were monitored for 12 weeks after birth. Mice of both genotypes were healthy, similar in appearance, size, body length/weight ratio and growth rate (data not shown). We next assessed bone architecture in these animals by micro-computed tomography ( $\mu$ CT) of the distal femora. No difference was detected between *Col2a1-Cre;Apc*<sup>15lox/+</sup> mice and gender-matched *Apc*<sup>15lox/+</sup> control littermates with respect to bone mineral density, trabecular bone volume fraction, trabecular number, trabecular thickness, and trabecular separation (Fig. 2A-D; data not shown). We further wanted to study whether conditional heterozygous *Apc* inactivation would lead to skeletal anomalies later in life. For this purpose, 10 *Col2a1-Cre;Apc*<sup>15lox/+</sup> mice (5 males and 5 females) and 5 *Apc*<sup>15lox/+</sup> male mice were followed for 24 months. At the end of this period, animals were sacrificed and tissues were analyzed microscopically using hematoxylin/eosine-stained sections. No important abnormalities



**Figure 2**  
**Skeletal development occurs normally in Col2a1-Cre;Apc<sup>15lox/+</sup> mice.** (A-D)  $\mu$ CT analysis of the distal diaphysis of the femur did not reveal significant differences between 12-week-old Col2a1-Cre;Apc<sup>15lox/+</sup> mice and control littermates in any of the parameters investigated: (A) trabecular bone volume [BV/TV (%)], (B) number of trabeculae [Tb.N (1/mm)], (C) trabecular thickness [Tb.Th ( $\mu$ m)], and (D) trabecular separation [Tb.Sp (mm)]. All data represent mean values  $\pm$  s.d

could be distinguished in the skull, ribs, vertebral column and long bones. We evenly detected in both groups signs of cartilage degradation, fibrosis, and osteochondritis, pathological findings which most likely were all age-related (data not shown). Altogether, we considered conditional heterozygous *Apc* mutant embryos as controls for the next experiments.

#### **Homozygous Col2a1-Cre;Apc<sup>15lox/15lox</sup> mice die perinatally due to severe defects in skeletogenesis**

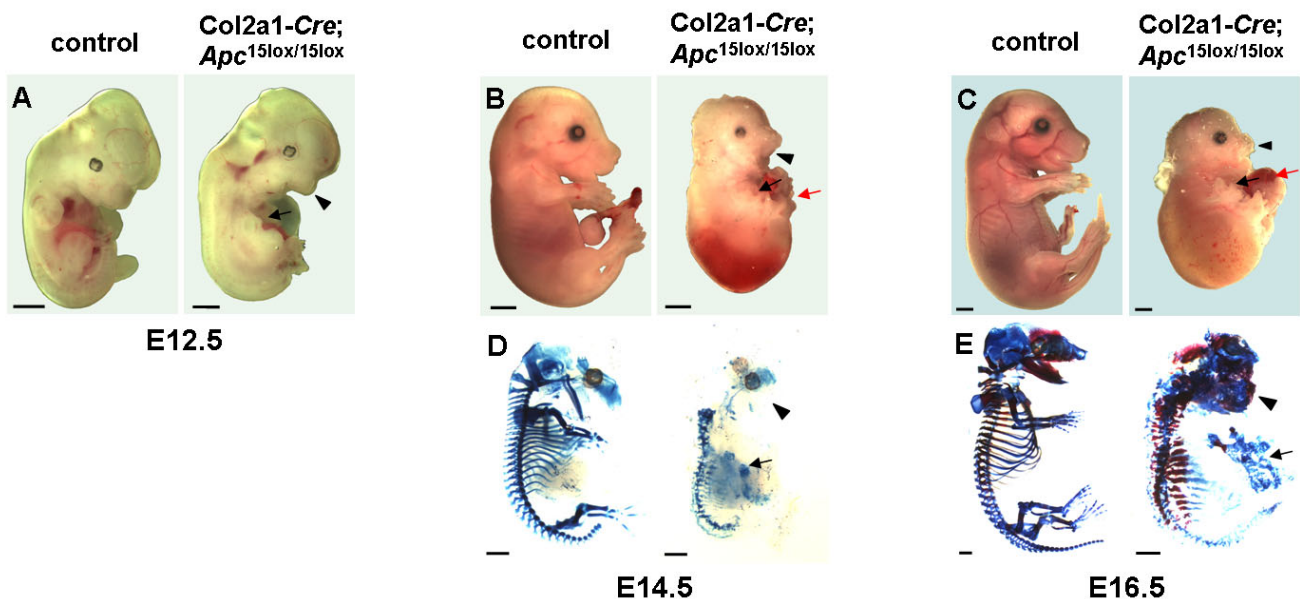
Col2a1-Cre;Apc<sup>15lox/+</sup> mice were crossed with Apc<sup>15lox/15lox</sup> mice to generate conditional homozygous Col2a1-Cre;Apc<sup>15lox/15lox</sup> mice (1:4). None of these mice were found at one month of age among 77 liveborn offspring. Of 27 dead pups found within the first month after delivery, only 5 pups on the day after delivery were Col2a1-Cre;Apc<sup>15lox/15lox</sup>. To further investigate the Col2a1-Cre;Apc<sup>15lox/15lox</sup> phenotype, embryonic litters at various developmental stages were isolated. Eight of 31 embryos isolated between E16.5 and E19.5 were Col2a1-Cre;Apc<sup>15lox/15lox</sup> (26%). We concluded that conditional homozygosity for this *Apc* mutant allele was perinatally lethal. At E12.5, Col2a1-Cre;Apc<sup>15lox/15lox</sup> embryos, although normal in size, displayed poor mandible and

limb outgrowth compared to control littermates (Fig. 3A). At E14.5 and E16.5, Col2a1-Cre;Apc<sup>15lox/15lox</sup> embryos were much smaller in comparison to controls, displayed craniofacial abnormalities, short trunk, and an incomplete closure of both thoracic and abdominal cavities (Fig. 3B, C). Gross analysis further indicated a severe truncation of both upper and lower limbs. Already at E14.5, but more significantly at E16.5, Col2a1-Cre;Apc<sup>15lox/15lox</sup> embryos presented large skin blisters especially in the dorso-lumbar region (Fig. 3C).

Skeletal preparations of mouse embryos stained with Alcian blue (chondrocyte matrix) and Alizarin red (mineralized matrix) of embryos at E14.5 revealed a clear difference in size between Col2a1-Cre;Apc<sup>15lox/15lox</sup> mutants and control littermates (Fig. 3D). All mutant structures were severely misshaped and fragmented. Mutants failed to develop a cartilaginous mold of both the mandibles and the occipital bone. The axial skeleton contained patchy and irregular cartilaginous structures that did not organize in vertebrae. All 13 rib pairs could be individually distinguished, however, due to their inadequate orientation, size, and shape and due to lack of a sternum, no thoracic basket was formed (Fig. 4A, B). Distorted cartilage rudiments were found where forelimbs should normally arise (Fig. 4E, F), while no signs of bone formation were found in hindlimb rudiments. Furthermore, no cartilaginous primordia of pelvic bones were observed. Similar observations were made in Col2a1-Cre;Apc<sup>15lox/15lox</sup> embryos at E16.5 (Fig. 3E). At this developmental stage however, distinct areas of mineralization were observed in most parts of the mutant skeleton. The mutant hind skull showed mineralized regions, whereas the control occipital and temporal bone primordia stained only with Alcian blue (Fig. 4I, J). Mutant proximal ribs in these Col2a1-Cre;Apc<sup>15lox/15lox</sup> embryos were much thicker and shorter in comparison to those in control embryos, and stained intensively with Alizarin red (Fig. 4C, D). In the mutant forelimb, a hypoplastic scapula could be identified, whereas more distal components were agenetic and replaced by an irregular cartilaginous structure (Fig. 4G, H).

#### **Loss of functional Apc inhibits differentiation of skeletal precursors**

Lack of functional *Apc* results in accumulation of cytoplasmic  $\beta$ -catenin, which subsequently translocates into the nucleus. This process can be well detected by immunohistochemistry (IHC). To investigate endochondral bone formation in Col2a1-Cre;Apc<sup>15lox/15lox</sup> embryos, we analyzed vertebra formation at E12.5 and E14.5, and humerus development at E16.5 using IHC for  $\beta$ -catenin in combination with Alcian blue staining, and *in situ* hybridization (ISH) for several chondrocyte- and osteoblast-specific genes. Strongly elevated levels of  $\beta$ -catenin were seen at all sites of endochondral ossification in the Col2a1-

**Figure 3**

**Skeletogenesis is severely impaired in *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos.** (A-E) Greatly impaired skeletal development and growth arrest of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos. Gross appearance (A-C) and Alcian blue and Alizarin red staining (D-E) of skeletal preparations of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos and control littermates at indicated developmental stages. Conditional *Apc* mutants showed lack of mandible outgrowth (arrowheads), poor limb development (black arrows), and an open thoracic and abdominal cavity (red arrows). Scale bars: 1 mm.

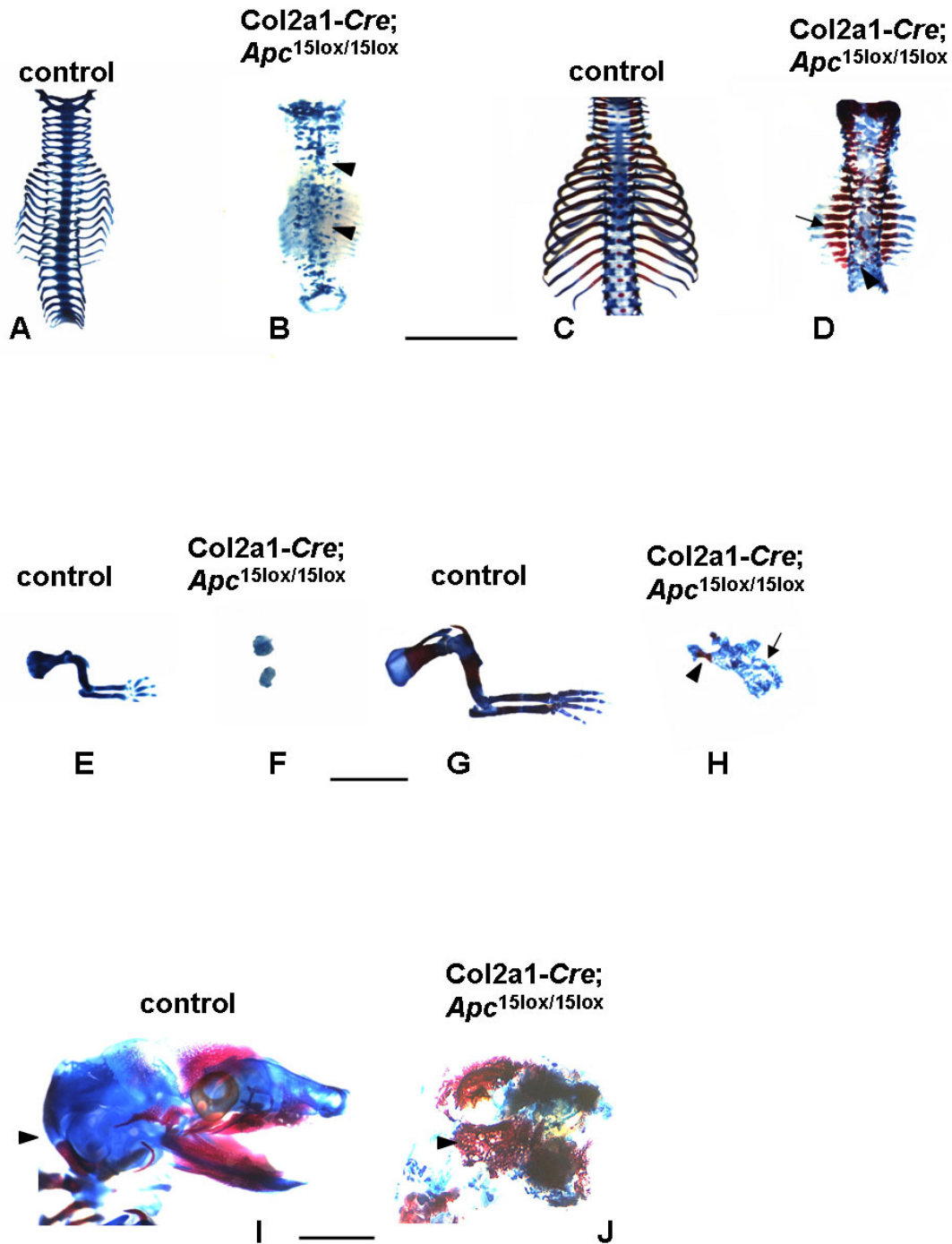
*Cre;Apc<sup>15lox/15lox</sup>* embryos at E12.5, E14.5 and E16.5, indicating efficient *Col2a1-Cre*-mediated *Apc* inactivation.

At E12.5, transversal sections of control vertebral primordia showed normal mesenchymal cell condensation and subsequent chondrogenic differentiation (Fig. 5A-C). Chondrocytes stained negatively for  $\beta$ -catenin, started to deposit an Alcian blue-stained matrix, and expressed the nascent chondrocyte markers *Sox9* and *Col2a1*. In marked contrast, mutant sclerotomal cells failed to condense into skeletal primordia. They showed strong nuclear  $\beta$ -catenin staining and displayed a mesenchymal-like spindle shape morphology. These cells expressed neither *Sox9*, nor *Col2a1*, implying that conditional loss of functional *Apc* in skeletal precursors inhibited mesenchymal cell condensation and chondrogenic differentiation. Next, we investigated whether these cells had switched their commitment to the osteogenic lineage due to the accumulation of  $\beta$ -catenin. Surprisingly, they did not express the early osteoblast markers *Runx2* and *Col1a1*, suggesting that  $\beta$ -catenin accumulation due to *Apc* inactivation impaired osteogenic differentiation of skeletal precursors as well (Fig. 5D, E).

At E14.5, chondrocytes in the control vertebrae did not stain positively for  $\beta$ -catenin, displayed an intensely Alcian blue-stained matrix and expressed both early (*Sox9*,

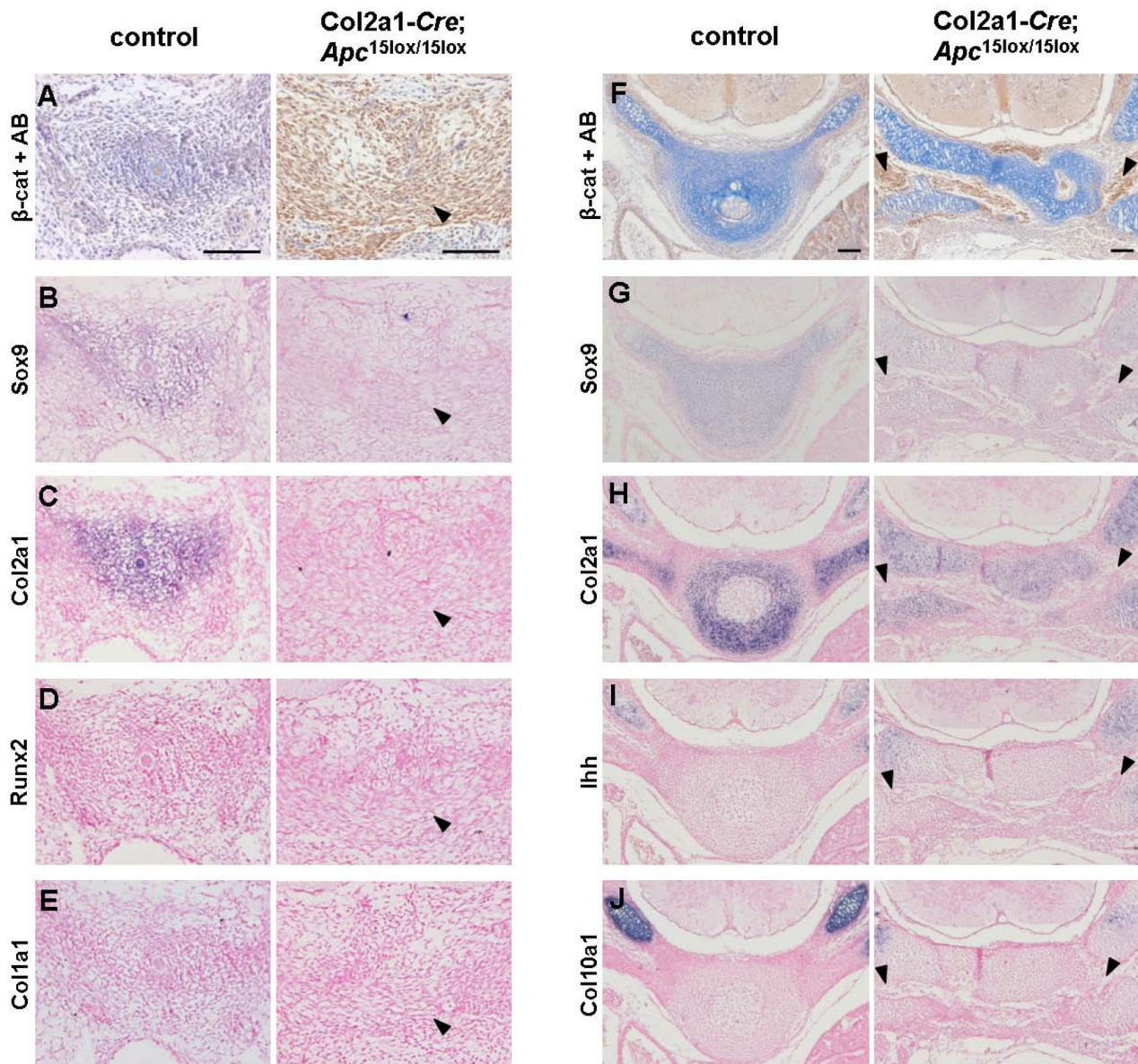
*Col2a1*) and mature chondrocyte markers, like *Indian hedgehog (Ihh)* and *Collagen 10a1 (Col10a1)*, indicating a normal progression of endochondral ossification (Fig. 5F-J). Although somite formation was present, mutant vertebrae were heavily crumbled and failed to organize in a cartilaginous anlage. Occasionally Alcian blue-positive clusters of chondrocytes were seen, which lacked detectable  $\beta$ -catenin immunostaining and were positive for chondrogenic marker expression. These cells were probably derived from non-recombined cells due to mosaicism of *Cre* expression. Surrounding these cartilage islands, mesenchymal-like spindle-shaped cells were observed. Comparable to the defects observed at E12.5, these cells expressed high levels of nuclear  $\beta$ -catenin due to *Apc* inactivation and lacked not only an Alcian blue-positive matrix but also expression of both chondrogenic and osteogenic markers (Fig. 5F-J; data not shown).

At E16.5, chondrocytes of control proximal humeri did not express detectable  $\beta$ -catenin protein levels and were surrounded by a proteoglycan-rich matrix, which stained positively with both Alcian blue and Toluidine blue (Fig. 6A, B). They were organized in growth plates with a characteristic spatial expression pattern of the chondrogenic markers *Sox9*, *Col2a1*, *Ihh*, and *Col10a1* (Fig. 6C-F). Young osteoblasts in the perichondrium, periosteum and



**Figure 4**

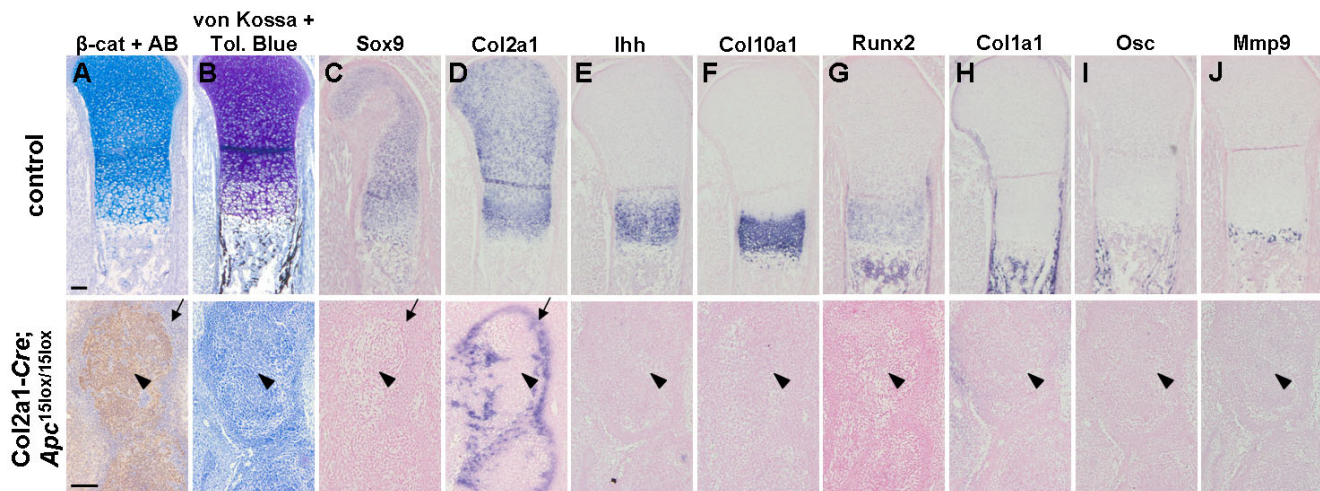
**Details of skeletal preparations.** (A-D) Vertebral column of control and mutant littermates at E14.5 (A, B) and E16.5 (C, D). Mutant vertebrae lacked structural integrity (arrowheads). At E16.5, mineralization was enhanced in the proximal part of the mutant rib (arrow). (E-H) Forelimb of control and mutant littermates at E14.5 (E, F) and E16.5 (G, H). At E16.5, only the scapula was identified (arrowhead), while more distal parts were represented by patchy cartilage aggregations (arrow). (I, J) Skull of control and mutant littermates at E16.5. The mutant displayed mineral deposition in the back skull corresponding to the cartilaginous structure in the control (arrowheads). Scale bars: 3 mm in A-H; 1 mm in I, J.



**Figure 5**  
**Abnormal axial skeleton formation of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos already detectable at E12.5.** (A) Immunostaining for  $\beta$ -catenin combined with Alcian blue (AB) staining, and (B-E) gene expression analysis by in situ hybridization with indicated probes on consecutive transversal sections of the sclerotome of a *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryo and control littermate at E12.5. (F-J) Similar analysis of vertebrae primordia at E14.5.  $\beta$ -Catenin-positive spindle-shaped cells lacked expression of all indicated chondrogenic and osteogenic markers (arrowheads). Scale bars: 100  $\mu$ m.

primary spongiosa were surrounded by a mineralized osteoid as detected by von Kossa staining (Fig. 6B) and expressed *Runx2* and *Col1a1* (Fig. 6G, H). Mature osteoblasts expressed *Osc* (Fig. 6I), while osteoclasts expressed Matrix metalloproteinase 9 (*Mmp9*) (Fig. 6J). In contrast, mutant humeri were completely misshaped and contained nuclear  $\beta$ -catenin-positive cells that were organized in clusters, showing a mesenchymal-like shape (Fig. 6A). Similar to our observations at E12.5 and E14.5, these cells

expressed neither chondrogenic, nor osteogenic markers (Fig. 6C-I). In addition, no *Mmp9* expression could be detected (Fig. 6J), suggesting that differentiation of bone-resorbing cells was impaired as well. These  $\beta$ -catenin-positive cell clusters were surrounded by chondrocytes expressing *Sox9* and *Col2a* and lacked positive staining for  $\beta$ -catenin. These cells most likely have not undergone a recombination event as observed at E14.5 (Fig. 6A, C, D).

**Figure 6**

**No chondrogenic and osteogenic differentiation in the developing humerus due to lack of functional *Apc*.** (A-B) Immunostaining for  $\beta$ -catenin combined with Alcian blue (AB) staining (A), combined von Kossa-Toluidine blue staining (B), and (C-J) gene expression analysis by in situ hybridization with indicated probes for (C-F) chondrocytes, (G-I) osteoblasts and (J) osteoclasts on consecutive transversal sections of the developing humerus of a *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryo and control littermate at E16.5.  $\beta$ -Catenin-positive spindle-shaped cells organized in clusters and failed to express chondrogenic and osteogenic markers (arrowheads).  $\beta$ -Catenin-negative cells at the periphery of these clusters expressed early chondrogenic markers only (arrows), probably due to lack of Cre-mediated loss of functional *Apc*. Scale bars: 100  $\mu$ m.

#### **Increased osteoblastogenesis in proximal ribs of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos**

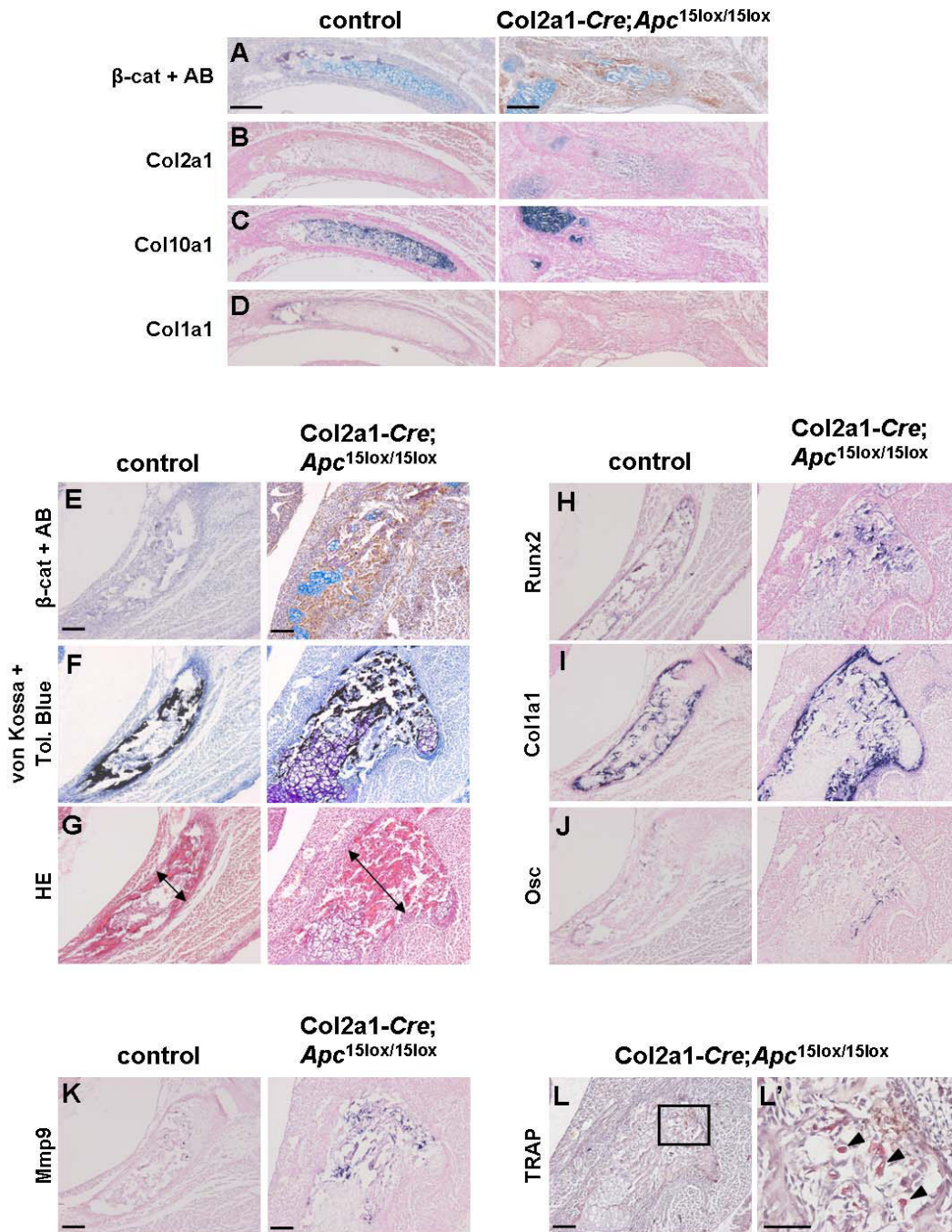
Despite the inhibitory effect of *Apc* inactivation on differentiation of skeletal precursors in long bones and vertebrae, proximal ribs of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos at E16.5 showed clearly enhanced mineralization upon skeletal staining (Fig. 4D). Therefore, we analyzed the development of these skeletal structures in more detail. The ribs develop through endochondral ossification from the paired lateral sclerotomic areas [31]. Formation of the proximal rib depends on the notochord and the ventral neural tube, whereas development of the distal part depends on the surface ectoderm [32]. At E14.5, proximal ribs of control embryos were cartilaginous and contained mature chondrocytes that did not stain for  $\beta$ -catenin (Fig. 7A-C). Mutant proximal ribs were severely misshaped and contained  $\beta$ -catenin negatively stained cartilage islands, accounting for the positive Alcian blue staining observed upon skeletal preparation (Fig. 4B, 7A).  $\beta$ -Catenin-positive cells were negative for chondrogenic and osteogenic markers (Fig. 7A-D). At E16.5, the  $\beta$ -catenin-negative proximal ribs of control embryos consisted of cartilage and mineralized bone matrix as indicated by combined von Kossa-Toluidine blue staining (Fig. 7E, F; data not shown). They contained chondrocytes, osteoblasts, and osteoclasts as assessed by ISH (Fig. 7H-K; data not shown). In contrast, proximal ribs of mutant littermates stained strongly positive for  $\beta$ -catenin and were signif-

icantly thicker and shorter compared to those of control embryos (Fig. 4C, D and 7E-G). They consisted of a massive mineralized bone matrix and a poorly developed bone marrow cavity, although osteoclast differentiation and activity were normal as assessed by ISH for *Mmp9* and TRAP staining, respectively (Fig. 7K, L, L'). Interestingly,  $\beta$ -catenin-positive cells expressed all osteogenic markers analyzed (*Runx2*, *Col1a1*, and *Osc*), indicating that, unlike in the long bones and vertebrae, *Apc* inactivation in skeletal precursors of the proximal ribs did not impair osteoblastogenesis (Fig. 7H-J). Since *Ihh* is a critical regulator of osteoblastogenesis, we subsequently tested whether the increased ossification might be due to increased *Ihh* expression in the non-recombined neighbouring chondrocytes. The  $\beta$ -catenin-negative cells, however, matured normally expressing all chondrocyte markers investigated (*Sox9*, *Col2a1*, *Ihh* and *Col10a1*) at similar levels compared to control cartilage (data not shown). The abundant presence of a bone matrix combined with evidence of functional osteoclasts suggested that the  $\beta$ -catenin-positive osteoblasts were sclerotic.

#### **Chondrocyte dedifferentiation in the nasal septum of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos**

The nasal septum is a midline vertical plate of hyaline cartilage, which undergoes endochondral ossification in postnatal life [33]. Endochondral ossification of the caudal and dorsal borders of the septum, when combined





**Figure 7**  
**Conditional *Apc* inactivation enhances osteoblast formation and mineral deposition in the developing proximal rib.** (A-L) Immunostaining for  $\beta$ -catenin combined with Alcian blue (AB) staining (A, E), combined von Kossa-Toluidine blue staining (F), hematoxylin/eosin staining (G), gene expression analysis by in situ hybridization with indicated probes for chondrogenic (B, C), osteogenic (D, H-J) and osteoclastogenic differentiation (K) on consecutive transversal sections of the developing proximal rib of a *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryo and control littermate at E14.5 (A-D) and E16.5 (E-K). The double-headed arrows in G indicate the thickness of the rib. (L) Tartrate-resistant acid phosphatase (TRAP) staining of the developing proximal rib of a *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryo at E16.5. The boxed region in L is magnified in L' showing multinucleated osteoclasts (arrowheads) staining positive for TRAP. Scale bars: 100  $\mu$ m in A-L; 50  $\mu$ m in L'.

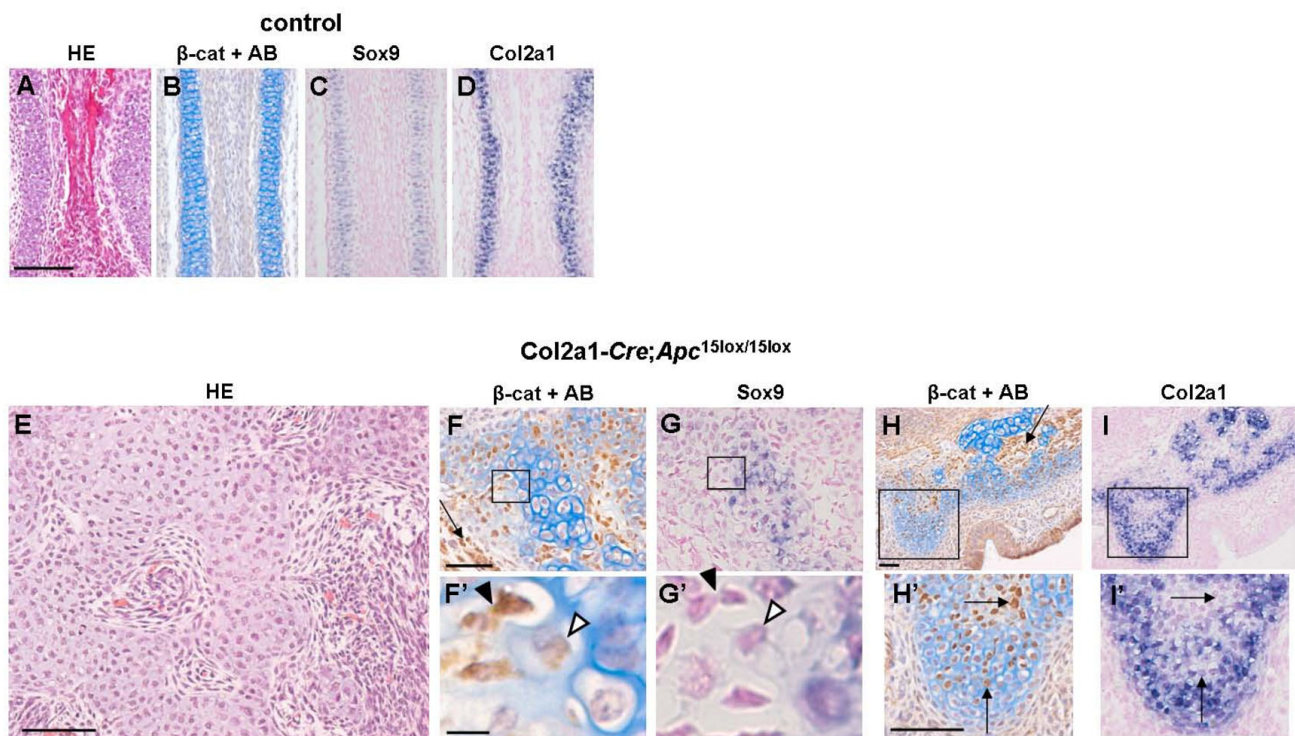
with interstitial expansion, has the effect of displacing the facial skeleton away from the neurocranium and thus enlarging the skull [34]. At E16.5, chondrocytes forming the nasal septum of control mice did not stain for  $\beta$ -catenin, were surrounded by an Alcian blue-positive matrix, and expressed *Sox9* and *Col2a1* (Fig. 8A–D). In the mutant nasal cartilage we distinguished crumbled chondrogenic islands surrounded by  $\beta$ -catenin-positive cells with an undifferentiated mesenchymal-like phenotype (Fig. 8F–H). The chondrogenic islands consisted of round cells embedded in chondrons surrounded by extracellular matrix (ECM) (Fig. 8E). Interestingly, molecular analysis of these chondrogenic islands revealed the presence of two cell populations:  $\beta$ -catenin-negative and  $\beta$ -catenin-positive cells. The former expressed chondrogenic markers like *Sox9* and *Col2a1*, and their ECM stained positive with Alcian blue, whereas the latter did not express any chondrogenic or osteogenic markers, while their ECM stained significantly less with Alcian blue (Fig. 8F–I; data not shown).

The presence of  $\beta$ -catenin-positive cells in the chondrogenic islands suggested that these cells, due to mosaicism of *Cre* expression, had initiated normal chondrocyte differentiation before undergoing *Apc* inactivation. Subsequently, the increased level of  $\beta$ -catenin triggered the loss of expression of the early chondrogenic markers and initiated degradation of the ECM. These observations were indicative of dedifferentiated chondrocytes. Similar observations were made in cartilaginous rudiments at other sites of endochondral bone formation, but the effect was most pronounced in the nasal septum (data not shown).

## Discussion

### Conditional homozygous loss of functional *Apc* severely disrupts mouse skeletogenesis via stabilized $\beta$ -catenin

According to most of the transgenic mouse studies reported, levels of  $\beta$ -catenin, the effector of the canonical Wnt ligands, need to be downregulated in skeletal precursor cells to enable chondrogenic differentiation, whereas elevated  $\beta$ -catenin levels promote differentiation into



**Figure 8**

**Dedifferentiation in the nasal septum of *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos at E16.5.** (A, E) Hematoxylin/eosin staining, (B, F, H) immunostaining for  $\beta$ -catenin combined with Alcian blue (AB) staining, and (C–D, G, I) gene expression analysis by in situ hybridization with indicated probes for chondrogenic differentiation on consecutive transversal sections of the developing nasal septum of a *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryo and control littermate at E16.5. (F', G', H', I') High magnification pictures of the boxed regions in F, G, H, and I, respectively. Mesenchymal-like  $\beta$ -catenin-positive cells (arrow in F, H) were present between crumbled cartilage islands. Within these cartilage islands, although displaying chondrocytic morphology and an Alcian blue stained matrix, most of the  $\beta$ -catenin-positive cells did not express *Sox9* (arrowheads in G') or *Col2a1* (arrows in I'). Scale bars: 100  $\mu$ m in A, E, F, H; 5  $\mu$ m in F', H'.

osteoblasts [11,12,15,35,36]. This theory is partly based on observations in heterozygous gain-of-function models in which Cre-mediated recombination results in the expression of oncogenic  $\beta$ -catenin. The cellular mechanisms controlling the biological effects of oncogenic  $\beta$ -catenin in the presence of wild type  $\beta$ -catenin are largely unknown. In addition, there are no reports on the role of Apc in regulation of skeletal precursor differentiation via control of  $\beta$ -catenin in the mouse. Here, we have focused on this important role of the multifunctional protein Apc, binding to and downregulating  $\beta$ -catenin. We have selectively inactivated one or both alleles of *Apc* in murine *Col2a1*-expressing cells. Our data indicate that the *Col2a1* promoter is suitable for this study, since Cre-mediated recombination starts very early (E9.5) in skeletal precursor cells that have not yet committed to the chondrogenic or the osteogenic lineage, consistent with previous findings in other *Col2a1-Cre* lines [11,30].

Conditional heterozygous inactivation of *Apc* does not result in a detectable level of its target  $\beta$ -catenin as determined by IHC. Moreover, heterozygous *Col2a1-Cre*-mediated *Apc* inactivation does not interfere with embryonic skeletal development, postnatal growth or bone acquisition up to 24 months of age, as determined by histological and  $\mu$ CT analysis. Our data imply that the level of Apc protein produced by a single functional *Apc* allele is sufficient to mediate appropriate  $\beta$ -catenin degradation. This is in agreement with normal body weight, size, and growth of young *Apc*<sup>Min/+</sup> mice [37].

In marked contrast, conditional inactivation of *Apc* results in a strongly elevated level of (wild type)  $\beta$ -catenin in skeletal precursors, leading to greatly impaired embryogenesis and perinatal lethality. The significantly reduced size and the vast range of skeletal malformations in these embryos is most likely due to the specific *Col2a1-Cre* activity in skeletal primordia at a very early embryonic stage starting at E9.5 resulting in massive  $\beta$ -catenin accumulation in the developing endochondral skeleton. Probably several factors, like the open rib cage and the severe malformation, from E14.5 on have led to the perinatal lethality. The loss of the multiple  $\beta$ -catenin-independent functions of the Apc protein might have contributed to the gravity and complexity of the skeletal phenotype observed in *Col2a1-Cre;Apc*<sup>15lox/15lox</sup> mice as well [18]. Moreover, since *Col2a1* expression is not completely restricted to skeletal tissues during mouse embryogenesis [29], we can not exclude that the severity of the phenotype might have been partly due to loss of functional *Apc* in other *Col2a1-Cre*-expressing cell types.

#### **Apc is crucial for both chondrogenic and osteogenic differentiation of skeletal precursors**

Wnt/ $\beta$ -catenin signaling represents a mechanism in mesenchymal precursor cells for selecting between chondro-

cytic and osteoblastic fates. This key regulating role in lineage commitment has been attributed to  $\beta$ -catenin. Indeed, conditional gain-of-function mutation of  *$\beta$ -catenin* leads to decreased chondrocyte differentiation in *Prx1*-expressing and *Col2a1*-expressing cells [12,15]. However, corresponding increased osteoblast differentiation has not been observed in these models, instead, a decreased osteoblast marker expression has been seen in case of *Prx1*-expressing cells, suggesting that activation of  $\beta$ -catenin negatively affects skeletogenesis [12,15]. In addition, conditional loss-of-function mutation of  *$\beta$ -catenin* in *Prx1*-expressing cells leads to increased expression of not only chondrocyte but also early osteoblast markers [15]. These data strongly suggest that  $\beta$ -catenin negatively regulates the differentiation of mesenchymal cells into a common skeletal precursor [38].

We report here that in the vast majority of endochondral skeletal elements, precursor cells lacking functional Apc express strong nuclear  $\beta$ -catenin staining and fail to differentiate into both chondrogenic and osteogenic lineages. These data are in line with the inability of mouse embryonic stem cells carrying specific bi-allelic *Apc* mutations to differentiate into bone and cartilage [25]. Our data are also consistent with those based on conditional stabilization of  $\beta$ -catenin in mesenchymal skeletal precursors which had an undifferentiated appearance [15]. This consistency strongly suggests that, notwithstanding the multiple functions of Apc, its  $\beta$ -catenin-controlling role is the most important during skeletogenesis. We conclude that Apc plays a crucial role in differentiation of skeletal precursors in vertebrae and long bones: it enables the differentiation into both skeletal lineages by decreasing the level of  $\beta$ -catenin.

#### **Loss of functional Apc in skeletal precursors of the proximal rib stimulates osteogenesis**

Although in the vast majority of the endochondral skeleton both chondrogenic and osteogenic differentiation is inhibited due to loss of functional *Apc* in skeletal precursors, we find a different phenotype in the proximal ribs. Notwithstanding the cartilaginous structure at E14.5, proximal ribs of *Col2a1-Cre;Apc*<sup>15lox/15lox</sup> mutants at E16.5 show abundant bone matrix deposited by osteoblasts, invariably expressing high levels of nuclear  $\beta$ -catenin. Since osteoblasts do not express *Col2a1*, these cells are most likely derived from *Col2a1*-expressing skeletal precursors lacking functional Apc. This implies that, in contrast to other skeletal elements, skeletal precursors of the proximal ribs are able to escape from the noxious effects of strongly elevated  $\beta$ -catenin levels on differentiation of precursor cells by an as yet unknown mechanism. Since *Ihh* expression is normal in the non-recombined neighbouring chondrocytes, we speculate that *Ihh* may be a prime target for inducing osteoblastogenesis in the recom-

bined precursor cells counteracting the noxious effect of  $\beta$ -catenin.

Despite the evidence of functional osteoclasts, the intensely ossified proximal ribs show a strongly diminished bone marrow cavity, rendering it likely that the increased bone formation is due to osteosclerosis. These observations are in agreement with other data, showing that enhanced canonical Wnt signaling can increase bone mass through stimulation of osteoblast activity rather than inhibition of osteoclast formation and activity [39-41]. Such an osteopetrotic phenotype has only been seen in mice with conditional loss of functional *Apc* or constitutively active  $\beta$ -catenin in already differentiated osteoblasts, resulting in dramatically increased bone deposition [17,42].

#### **Functional *Apc* is required to maintain the chondrocyte phenotype**

We have found clear evidence for the occurrence of chondrocyte dedifferentiation due to  $\beta$ -catenin accumulation in the nasal septum. Morphologically characterized chondrocytes, which were nuclear  $\beta$ -catenin-positive, lacked expression of typical chondrocyte markers. Furthermore, they were imbedded in an ECM containing significantly less proteoglycans.

Given the noxious effect of increased  $\beta$ -catenin levels on chondrocyte formation (our data and [12,15]), these cells most likely have undergone Cre-mediated loss of functional *Apc* after completion of the initial stages of chondrocyte differentiation. Mouse models with an increased level of  $\beta$ -catenin in *Col2a1*-expressing cells show accelerated chondrocyte maturation [11,12]. We have found no indication for this phenomenon, implying that the high level of  $\beta$ -catenin due to loss of *Apc* does not result in chondrocyte maturation but in chondrocyte dedifferentiation. Our data suggest that accumulated  $\beta$ -catenin triggers this dedifferentiation program not only through inhibition of chondrogenic marker expression but also by enhancing the loss of ECM presumably through stimulation of matrix-degrading enzymes. It has been demonstrated that  $\beta$ -catenin increases expression and activity of a number of enzymes involved in matrix degradation [43-45].  $\beta$ -Catenin stabilization has been associated with dedifferentiation of articular chondrocytes in vitro upon serial monolayer culture, or treatment with retinoic acid or IL1 $\beta$  [46]. Dedifferentiated chondrocytes have also been observed at other sites of endochondral bone formation in the *Col2a1-Cre;Apc<sup>15lox/15lox</sup>* embryos, however, the presence of these cells was most pronounced in the nasal septum. Altogether, our data indicate that *Apc* is required to suppress  $\beta$ -catenin for maintenance of the chondrocytic phenotype.

#### **Conclusion**

We show here for the first time that *Apc*, by negatively controlling the levels of  $\beta$ -catenin, is a critical regulator of the differentiation of skeletal progenitor cells. Conditional inactivation of the mouse *Apc* gene results in a heterogeneous skeletal phenotype. Based on our results, we postulate that *Apc*-mediated control of the dosage of transcriptionally active  $\beta$ -catenin protein is directive for the differentiation program of skeletal precursor cells. In the vast majority of the skeletal precursors, loss of functional *Apc* leads to a strongly increased  $\beta$ -catenin level, resulting in the formation of an undifferentiated mesenchymal cell, which lacks differentiation potential for both osteogenic and chondrogenic lineages. When the inhibitory effect of a strongly increased  $\beta$ -catenin level in the skeletal precursors is reduced, highly active osteoblasts arise. Strong repression of  $\beta$ -catenin in these precursors is required for chondrogenesis. Support for our hypothesis on the importance of the dosage of *Apc* and  $\beta$ -catenin is provided by observations in *Col2a1-Wnt14* transgenic mice [11]. Higher levels of *Wnt14* expression resulting in a high level of  $\beta$ -catenin block differentiation of skeletal precursors into chondrocytes or osteoblasts, whereas lower levels of *Wnt14* expression result in enhanced ossification. We provide evidence that *Apc* plays a crucial role in modulating the  $\beta$ -catenin level during mouse skeletogenesis in a spatio-temporal regulated manner. In skeletal precursor cells, *Apc* is required for differentiation into both chondrocytes and osteoblasts. In addition, *Apc* is essential in chondrocytes to maintain their phenotype and enable their maturation.

#### **Methods**

##### **Transgenic mice**

All animal studies were approved by the ethical committee of the Leiden University Medical Centre and complied with national laws relating to the conduct of animal experiments. The *Apc<sup>15lox/+</sup>* mouse was generated by gene targeting in IB10 embryonic stem cells, using a 22.5 kb targeting vector containing loxP sites flanking the last exon of *Apc*, i.e. exon 15. LoxP sites were inserted in the *Bgl*III site of intron 14 and in the *Apa*I site approximately 350 bp downstream of the *Apc* polyadenylation signal. Exon 15 of the *Apc* gene encodes for codons 660 to 2842 of the *Apc* protein and harbours all the functional domains of *Apc* involved in  $\beta$ -catenin regulation as well as the C-terminal domains binding to microtubules, DLG, and EB1. Therefore, following Cre-mediated deletion of exon 15, functionality of the remaining protein will be fully impaired with respect to the main function of *Apc*, i.e.  $\beta$ -catenin regulating. Moreover, as deletion of exon 15 also removes the polyadenylation signal, no stable mRNA is produced and as a consequence no stable truncated protein will be generated. A full description of this new con-

ditional *Apc* mouse model is currently in preparation (Robanus-Maandag et al., in preparation). *Col2a1-Cre* mice [27] were mated with *Apc*<sup>15lox/15lox</sup> mice. Of the offspring, *Col2a1-Cre*<sup>+/-</sup>;*Apc*<sup>15lox/+</sup> mice were mated with *Apc*<sup>15lox/15lox</sup> mice to obtain *Col2a1-Cre*<sup>+/-</sup>;*Apc*<sup>15lox/15lox</sup> mice. *LacZ* reporter mice were obtained from Dr. Xiaohong Mao [28]. Routine mouse genotyping was performed on tail DNAs by PCR (Robanus-Maandag et al., in preparation).

### Skeletal analysis

Skeletons of mouse embryos were stained with Alcian blue and Alizarin red for cartilaginous and mineralized tissues, respectively, according to standard procedures [47]. For micro-computed tomography ( $\mu$ CT) analysis, femora were recovered from 12-week-old mice after death and processed as described [48].

### $\beta$ -galactosidase staining, Histology, Immunohistochemistry, In Situ Hybridization

Whole mount  $\beta$ -galactosidase staining was performed as described [49], from E16.5 on after removal of the skin. For histology, immunohistochemistry, and in situ hybridization, specimens were fixed in phosphate-buffered formalin, embedded in paraffin, and sectioned at 6  $\mu$ m. Hematoxylin/eosin, Nuclear red, Toluidine blue, and von Kossa stainings were performed according to standard procedures. For immunohistochemistry, sections were treated with 1% H<sub>2</sub>O<sub>2</sub> in 40% methanol/60% TBS for 30 minutes to reduce endogenous peroxidase activity. For antigen retrieval the sections were boiled in Tris-EDTA pH 9.0 for 20 minutes. Blocking was performed with 5% blocking buffer for 30 minutes at 37°C (Boehringer Ingelheim). Sections were incubated with the primary mouse monoclonal antibody against  $\beta$ -catenin (1:100; BD Transduction Laboratories) overnight at 4°C, followed by incubation with the second antibody biotin-conjugated rabbit anti-mouse IgG (1:300; Amersham Biosciences) for 45 minutes at 37°C. The biotinylated proteins were detected by incubation with horseradish peroxidase-conjugated streptavidin (1:200; Amersham Biosciences) for 30 minutes at 37°C and visualized with DAB (Sigma). After counterstaining with Alcian blue for 15 minutes and hematoxylin for 1 minute, sections were dehydrated and embedded in Histomount (BDH). For in situ hybridization, digoxigenin-labeled single-stranded RNA probes were prepared using a DIG RNA labeling kit (Boehringer) following the manufacturers' instructions. All probes are available upon request. In situ hybridization was carried out as described [15,50]. Images were taken with a DXM-1200 digital camera (Nikon).

### Authors' contributions

The studies were designed and initiated by MK as principal investigator, with the help of RF and ECR-M; *Col2a1-*

*Cre* mice were provided by TK and HMK; mutant mice were generated and genotyped by CAJB and ECR-M; embryo experimental work and analysis were performed by RLM;  $\mu$ CT analysis was performed by GR; data interpretation was carried out by RLM assisted by MK, GH, MAV, PA, CWGML, RF, JMW, and ECR-M; the manuscript was written by RLM with the assistance of all co-authors. All authors read and approved the final manuscript.

### Acknowledgements

We thank Christine Hartmann (IMP, Vienna, Austria) for the mouse *Runx2*, *Sox9*, *Osc*, and *Ihh* probes, Eero Vuorio (University of Turku, Finland) for the mouse *Colla1* and *Col2a1* probes, and Willy Hofstetter (University of Bern, Switzerland) for the mouse *Coll0a1* probe.

This work was financially supported by a short-term research fellowship from the European Society for Pediatric Endocrinology (RLM), an unrestricted educational grant from IPSEN FARMACEUTICA BV (RLM), a research grant from The Human Growth Foundation (MK), and a research grant from the Association for International Cancer Research (CAJB).

### References

1. Karsenty G: **Genetics of skeletogenesis.** *Dev Genet* 1998, **22**:301-313.
2. Karsenty G, Wagner EF: **Reaching a genetic and molecular understanding of skeletal development.** *Dev Cell* 2002, **2**:389-406.
3. Tuan RS: **Biology of developmental and regenerative skeletogenesis.** *Clin Orthop Relat Res* 2004:S105-S117.
4. Olsen BR, Reginato AM, Wang W: **Bone development.** *Annu Rev Cell Dev Biol* 2000, **16**:191-220.
5. Akiyama H, Chaboissier MC, Martin JF, Schedl A, de Crombrugge B: **The transcription factor Sox9 has essential roles in successive steps of the chondrocyte differentiation pathway and is required for expression of Sox5 and Sox6.** *Genes Dev* 2002, **16**:2813-2828.
6. Smith N, Dong Y, Lian JB, Pratap J, Kingsley PD, van Wijnen AJ, Stein JL, Schwarz EM, O'Keefe RJ, Stein GS, et al.: **Overlapping expression of Runx1(Cbfa2) and Runx2(Cbfa1) transcription factors supports cooperative induction of skeletal development.** *J Cell Physiol* 2005, **203**:133-143.
7. Zhou G, Zheng Q, Engin F, Munivez E, Chen Y, Sebald E, Krakow D, Lee B: **Dominance of SOX9 function over RUNX2 during skeletogenesis.** *Proc Natl Acad Sci USA* 2006, **103**:19004-19009.
8. Lefebvre V, Smits P: **Transcriptional control of chondrocyte fate and differentiation.** *Birth Defects Res C Embryo Today* 2005, **75**:200-212.
9. Kobayashi T, Kronenberg H: **Minireview: transcriptional regulation in development of bone.** *Endocrinology* 2005, **146**:1012-1017.
10. Clevers H: **Wnt/beta-catenin signaling in development and disease.** *Cell* 2006, **127**:469-480.
11. Day TF, Guo X, Garrett-Beal L, Yang Y: **Wnt/beta-catenin signaling in mesenchymal progenitors controls osteoblast and chondrocyte differentiation during vertebrate skeletogenesis.** *Dev Cell* 2005, **8**:739-750.
12. Akiyama H, Lyons JP, Mori-Akiyama Y, Yang X, Zhang R, Zhang Z, Deng JM, Taketo MM, Nakamura T, Behringer RR, et al.: **Interactions between Sox9 and beta-catenin control chondrocyte differentiation.** *Genes Dev* 2004, **18**:1072-1087.
13. Gaur T, Lengner CJ, Hovhannisyann H, Bhat RA, Bodine PV, Komm BS, Javed A, van Wijnen AJ, Stein JL, Stein GS, et al.: **Canonical WNT signaling promotes osteogenesis by directly stimulating Runx2 gene expression.** *J Biol Chem* 2005, **280**:33132-33140.
14. Dong YF, Soung DY, Schwarz EM, O'Keefe RJ, Drissi H: **Wnt induction of chondrocyte hypertrophy through the Runx2 transcription factor.** *J Cell Physiol* 2006, **208**:77-86.
15. Hill TP, Spater D, Taketo MM, Birchmeier W, Hartmann C: **Canonical Wnt/beta-catenin signaling prevents osteoblasts from differentiating into chondrocytes.** *Dev Cell* 2005, **8**:727-738.

16. Rodda SJ, McMahon AP: **Distinct roles for Hedgehog and canonical Wnt signaling in specification, differentiation and maintenance of osteoblast progenitors.** *Development* 2006, **133**:3231-3244.
17. Holmen SL, Zylstra CR, Mukherjee A, Sigler RE, Faugere MC, Bouxsein ML, Deng L, Clemens TL, Williams BO: **Essential role of beta-catenin in postnatal bone acquisition.** *J Biol Chem* 2005, **280**:21162-21168.
18. Fodde R: **The multiple functions of tumour suppressors: it's all in APC.** *Nat Cell Biol* 2003, **5**:190-192.
19. Korinek V, Barker N, Morin PJ, van Wichen D, de Weger R, Kinzler KW, Vogelstein B, Clevers H: **Constitutive transcriptional activation by a beta-catenin-Tcf complex in APC-/- colon carcinoma.** *Science* 1997, **275**:1784-1787.
20. Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B, Kinzler KW: **Activation of beta-catenin-Tcf signaling in colon cancer by mutations in beta-catenin or APC.** *Science* 1997, **275**:1787-1790.
21. Smits R, Houven van Oordt W van der, Luz A, Zurcher C, Jagmohan-Changur S, Breukel C, Khan PM, Fodde R: **Apc1638N: a mouse model for familial adenomatous polyposis-associated desmoid tumors and cutaneous cysts.** *Gastroenterology* 1998, **114**:275-283.
22. Bertario L, Russo A, Sala P, Varesco L, Giarola M, Mondini P, Pierotti M, Spinelli P, Radice P: **Multiple approach to the exploration of genotype-phenotype correlations in familial adenomatous polyposis.** *J Clin Oncol* 2003, **21**:1698-1707.
23. Fodde R, Edelmann W, Yang K, van Leeuwen C, Carlson C, Renault B, Breukel C, Alt E, Lipkin M, Khan PM: **A targeted chain-termination mutation in the mouse Apc gene results in multiple intestinal tumors.** *Proc Natl Acad Sci USA* 1994, **91**:8969-8973.
24. Moser AR, Shoemaker AR, Connelly CS, Clipson L, Gould KA, Luongo C, Dove WF, Siggers PH, Gardner RL: **Homozygosity for the Min allele of Apc results in disruption of mouse development prior to gastrulation.** *Dev Dyn* 1995, **203**:422-433.
25. Kielman MF, Rindapaa M, Gaspar C, van Poppel N, Breukel C, van Leeuwen S, Taketo MM, Roberts S, Smits R, Fodde R: **Apc modulates embryonic stem-cell differentiation by controlling the dosage of beta-catenin signaling.** *Nat Genet* 2002, **32**:594-605.
26. Fodde R, Smits R: **Cancer biology. A matter of dosage.** *Science* 2002, **298**:761-763.
27. Schipani E, Ryan HE, Didrickson S, Kobayashi T, Knight M, Johnson RS: **Hypoxia in cartilage: HIF-1alpha is essential for chondrocyte growth arrest and survival.** *Genes Dev* 2001, **15**:2865-2876.
28. Mao X, Fujiwara Y, Orkin SH: **Improved reporter strain for monitoring Cre recombinase-mediated DNA excisions in mice.** *Proc Natl Acad Sci USA* 1999, **96**:5037-5042.
29. Cheah KS, Lau ET, Au PK, Tam PP: **Expression of the mouse alpha 1 (II) collagen gene is not restricted to cartilage during development.** *Development* 1991, **111**:945-953.
30. Long F, Chung UI, Ohba S, McMahon J, Kronenberg HM, McMahon AP: **Ihh signaling is directly required for the osteoblast lineage in the endochondral skeleton.** *Development* 2004, **131**:1309-1318.
31. Christ B, Wiltling J: **From somites to vertebral column.** *Ann Anat* 1992, **174**:23-32.
32. Aoyama H, Mizutani-koseki S, Koseki H: **Three developmental compartments involved in rib formation.** *Int J Dev Biol* 2005, **49**:325-333.
33. Wealthall RJ, Herring SW: **Endochondral ossification of the mouse nasal septum.** *Anat Rec A Discov Mol Cell Evol Biol* 2006, **288**:1163-1172.
34. Roberts GJ, Lucas VS: **Growth of the nasal septum in the Snell strain of hypopituitary dwarf mouse.** *Eur J Orthod* 1994, **16**:138-148.
35. Hu H, Hilton MJ, Tu X, Yu K, Ornitz DM, Long F: **Sequential roles of Hedgehog and Wnt signaling in osteoblast development.** *Development* 2005, **132**:49-60.
36. Tamamura Y, Otani T, Kanatani N, Koyama E, Kitagaki J, Komori T, Yamada Y, Costantini F, Wakisaka S, Pacifici M, et al.: **Developmental regulation of Wnt/beta-catenin signals is required for growth plate assembly, cartilage integrity, and endochondral ossification.** *J Biol Chem* 2005, **280**:19185-19195.
37. Moser AR, Pitot HC, Dove WF: **A dominant mutation that predisposes to multiple intestinal neoplasia in the mouse.** *Science* 1990, **247**:322-324.
38. Hartmann C: **A Wnt canon orchestrating osteoblastogenesis.** *Trends Cell Biol* 2006, **16**:151-158.
39. Kato M, Patel MS, Levasseur R, Lobov I, Chang BH, Glass DA, Hartmann C, Li L, Hwang TH, Brayton CF, et al.: **Cbfa1-independent decrease in osteoblast proliferation, osteopenia, and persistent embryonic eye vascularization in mice deficient in Lrp5, a Wnt coreceptor.** *J Cell Biol* 2002, **157**:303-314.
40. Bodine PV, Zhao W, Kharode YP, Bex FJ, Lambert AJ, Goad MB, Gaur T, Stein GS, Lian JB, Komm BS: **The Wnt antagonist secreted frizzled-related protein-1 is a negative regulator of trabecular bone formation in adult mice.** *Mol Endocrinol* 2004, **18**:1222-1237.
41. Bennett CN, Longo KA, Wright WS, Suva LJ, Lane TF, Hankenson KD, Macdougald OA: **Regulation of osteoblastogenesis and bone mass by Wnt10b.** *Proc Natl Acad Sci USA* 2005, **102**:3324-3329.
42. Glass DA, Bialek P, Ahn JD, Starbuck M, Patel MS, Clevers H, Taketo MM, Long F, McMahon AP, Lang RA, et al.: **Canonical Wnt signaling in differentiated osteoblasts controls osteoclast differentiation.** *Dev Cell* 2005, **8**:751-764.
43. Yuasa T, Otani T, Koike T, Iwamoto M, Enomoto-Iwamoto M: **Wnt/beta-catenin signaling stimulates matrix catabolic genes and activity in articular chondrocytes: its possible role in joint degeneration.** *Lab Invest* 2008, **88**:264-74.
44. Enomoto-Iwamoto M, Otani T, Koike T, Iwamoto M: **Wnt/beta-Catenin Signaling in Chondrocyte Function and Cartilage Matrix Disruption.** *Current rheumatology reviews* 2006, **2**:31-38.
45. Zhu M, Tang D, Wu Q, Hao S, Chen M, Xie C, Rosier RN, O'Keefe RJ, Zuscik M, Chen D: **Activation of beta-catenin signaling in articular chondrocytes leads to osteoarthritis-like phenotype in adult beta-catenin conditional activation mice.** *J Bone Miner Res* 2009, **24**:12-21.
46. Ryu JH, Kim SJ, Kim SH, Oh CD, Hwang SG, Chun CH, Oh SH, Seong JK, Huh TL, Chun JS: **Regulation of the chondrocyte phenotype by beta-catenin.** *Development* 2002, **129**:5541-5550.
47. McLeod MJ: **Differential staining of cartilage and bone in whole mouse fetuses by alcian blue and alizarin red S.** *Teratology* 1980, **22**:299-301.
48. Clement-Lacroix P, Ai M, Morvan F, Roman-Roman S, Vayssiere B, Belleville C, Estrera K, Warman ML, Baron R, Rawadi G: **Lrp5-independent activation of Wnt signaling by lithium chloride increases bone formation and bone mass in mice.** *Proc Natl Acad Sci USA* 2005, **102**:17406-17411.
49. Dannenberg JH, Schuijff L, Dekker M, van der Valk M, te Riele H: **Tissue-specific tumor suppressor activity of retinoblastoma gene homologs p107 and p130.** *Genes Dev* 2004, **18**:2952-2962.
50. Eerden BC van der, van Til NP, Brinkmann AO, Lowik CW, Wit JM, Karperien M: **Gender differences in expression of androgen receptor in tibial growth plate and metaphyseal bone of the rat.** *Bone* 2002, **30**:891-896.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

