

# Heart failure with preserved ejection fraction in humans and mice: embracing clinical complexity in mouse models

Coenraad Withaar <sup>1</sup>, Carolyn S.P. Lam <sup>1,2</sup>, Gabriele G. Schiattarella <sup>3,4,5,6,7</sup>, Rudolf A. de Boer <sup>1\*†</sup>, and Laura M.G. Meems <sup>1†</sup>

<sup>1</sup>Department of Cardiology, University of Groningen, University Medical Center Groningen, Hanzeplein 1, 9713 GZ, Groningen, the Netherlands; <sup>2</sup>National Heart Centre, Singapore and Duke-National University of Singapore; <sup>3</sup>Translational Approaches in Heart Failure and Cardiometabolic Disease, Max Delbrück Center for Molecular Medicine in the Helmholtz Association (MDC), Berlin, Germany; <sup>4</sup>Department of Cardiology, Center for Cardiovascular Research (CCR), Charité - Universitätsmedizin Berlin, Berlin, Germany; <sup>5</sup>DZHK (German Centre for Cardiovascular Research), Partner Site Berlin, Berlin, Germany; <sup>6</sup>Division of Cardiology, Department of Advanced Biomedical Sciences, Federico II University, Naples, Italy; and <sup>7</sup>Department of Internal Medicine (Cardiology), University of Texas Southwestern Medical Center, Dallas, TX, USA

Received 16 February 2021; revised 15 April 2021; editorial decision 25 May 2021; accepted 2 June 2021; online publish-ahead-of-print 20 August 2021

Heart failure (HF) with preserved ejection fraction (HFpEF) is a multifactorial disease accounting for a large and increasing proportion of all clinical HF presentations. As a clinical syndrome, HFpEF is characterized by typical signs and symptoms of HF, a distinct cardiac phenotype and raised natriuretic peptides. Non-cardiac comorbidities frequently co-exist and contribute to the pathophysiology of HFpEF. To date, no therapy has proven to improve outcomes in HFpEF, with drug development hampered, at least partly, by lack of consensus on appropriate standards for pre-clinical HFpEF models. Recently, two clinical algorithms (HFA-PEFF and H<sub>2</sub>FPEF scores) have been developed to improve and standardize the diagnosis of HFpEF. In this review, we evaluate the translational utility of HFpEF mouse models in the context of these HFpEF scores. We systematically recorded evidence of symptoms and signs of HF or clinical HFpEF features and included several cardiac and extra-cardiac parameters as well as age and sex for each HFpEF mouse model. We found that most of the pre-clinical HFpEF models do not meet the HFpEF clinical criteria, although some multifactorial models resemble human HFpEF to a reasonable extent. We therefore conclude that to optimize the translational value of mouse models to human HFpEF, a novel approach for the development of pre-clinical HFpEF models is needed, taking into account the complex HFpEF pathophysiology in humans.

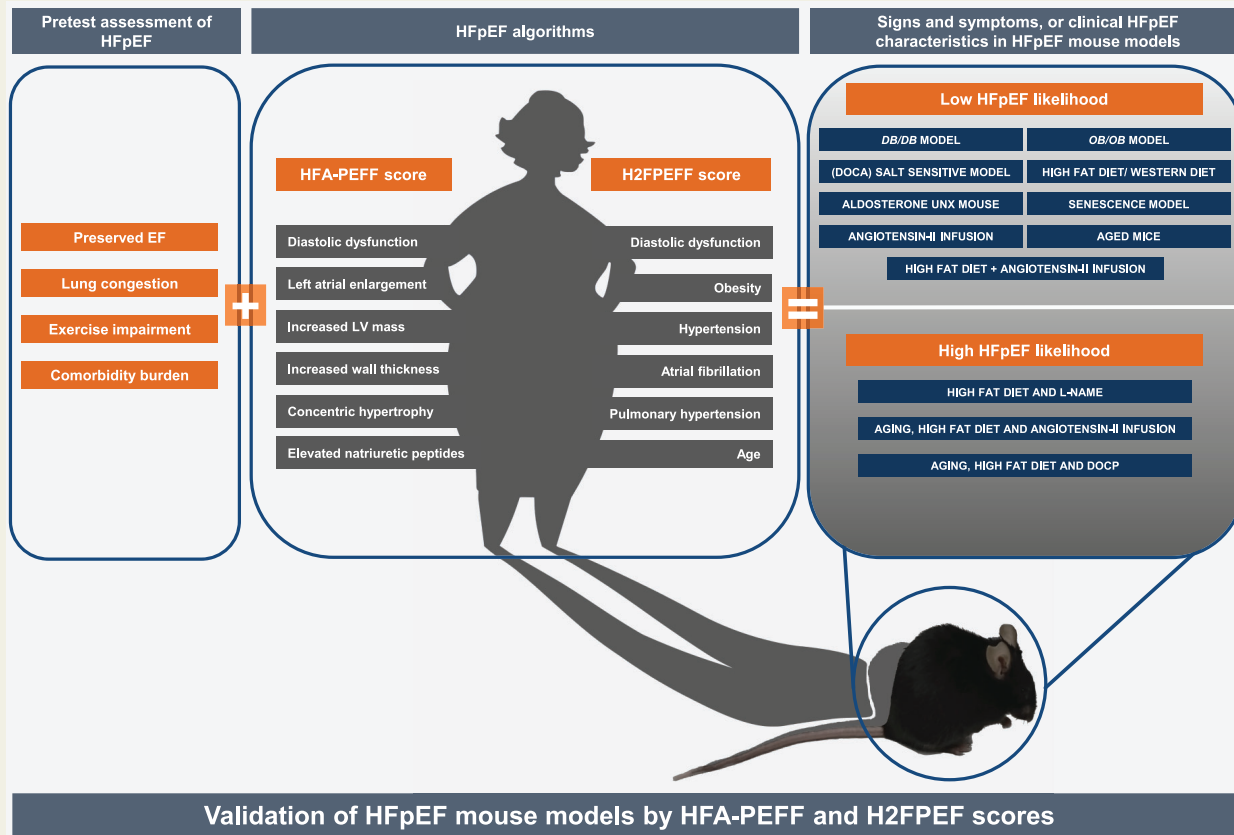
\* Corresponding author. Tel: +31 50 3612355; Email: [r.a.de.boer@umcg.nl](mailto:r.a.de.boer@umcg.nl)

† These authors contributed equally to this work.

© The Author(s) 2021. Published by Oxford University Press on behalf of the European Society of Cardiology.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact [journals.permissions@oup.com](mailto:journals.permissions@oup.com)

## Graphical Abstract



An in-depth review of existing pre-clinical HFpEF mouse models with validation of their translational value using the HFA-PEFF and H2FPEF scores.

## Keywords

HFpEF • Mouse • Human • Translational • H2FPEF • HFA-PEFF

## HFpEF: a heterogeneous disease with multiple disease mechanisms

Heart failure (HF) with preserved ejection fraction (HFpEF) is a complex clinical syndrome that is characterized by both extra-cardiac and cardiac features.<sup>1–3</sup> Prevalence is still rising<sup>4–8</sup> and survival of patients with HFpEF is poor, with a 5-year survival rate after first hospitalization of 35–40%.<sup>9,10</sup> So far no treatment has been proven successful in reducing morbidity and mortality rates in HFpEF, potentially due to the large pathophysiological heterogeneity and diversity in HFpEF phenotypes.<sup>11</sup> Recent studies have identified HFpEF as a systemic disease that is associated with, or may be triggered by a wide range of clinical risk factors and comorbidities such as aging, female sex, hypertension,<sup>12,13</sup> pulmonary congestion, metabolic syndrome, obesity,<sup>7,12,14–16</sup> type 2 diabetes mellitus (T2DM), hyperlipidaemia, renal disease, atrial fibrillation (AF), and skeletal muscle weakness.<sup>11</sup> These risk factors and comorbidities give rise to intertwining disease

mechanisms in the pathophysiology of HFpEF.<sup>17,18</sup> Due to the wide range of comorbidities and clinical presentations, potential underlying aetiology of HFpEF is diverse; HFpEF can result from various structural abnormalities of the myocardium, or may result from abnormal loading conditions, e.g. as seen in hypertension, valvular diseases, volume overload, or rhythm disorders.<sup>19</sup>

Although HFpEF patients thus represent a heterogeneous group with a broad extent of extra-cardiac features, the cardiac phenotype has less interpatient variability and includes (concentric) left ventricular (LV) hypertrophy,<sup>20</sup> LV diastolic dysfunction,<sup>21</sup> cardiac stiffening, atrial dilatation, fibrosis,<sup>22</sup> (systemic) inflammation, microvascular endothelial dysfunction,<sup>23,24</sup> and elevated natriuretic peptides.<sup>19,25,26</sup>

The definition of HFpEF as a clinical syndrome, based on typical symptoms and signs, presents challenges due to non-specificity of cardinal symptoms such as breathlessness and effort intolerance. Recently, two diagnostic HFpEF algorithms, the HFA-PEFF<sup>20</sup> and

Step 1		Pretest assessment of signs and symptoms and clinical features of HF 1) Shortness of breath 2) Comorbidity burden 3) Exercise tolerance		
Step 2	<b>HFA-PEFF score</b>	Points	<b>H<sub>2</sub>FPEF score</b>	Points
	<b>Functional aspects</b> -Diastolic function E/e' (≥ 15) GLS (<16%)	2 1	Obesity (BMI >30 kg/m <sup>2</sup> )	2
	<b>Morphological aspects</b> -Left atrial enlargement -Left ventricular mass -Wall thickness -Concentric hypertrophy	2	Hypertension	1
	<b>Natriuretic peptides</b>	2	Atrial fibrillation	3
			Pulmonary hypertension (PASP >35 mmHg)	1
			Age (>60 years)	1
			Diastolic function (E/e' > 9)	1
	<b>≥ 5 points: HFpEF</b>		<b>≥ 6 points: HFpEF</b>	

**Figure 1** Diagnostic HFpEF scoring algorithms used to score HFpEF animal models. Both algorithms first include a pretest assessment to evaluate signs and symptoms and clinical features of HFpEF that include congestion, increased comorbidity burden and reduced exercise tolerance. The second step of the HFA-PEFF<sup>19</sup> score assesses three domains that include functional aspects [echocardiographic diastolic function ( $E/e'$  and GLS)], morphological aspects (left atrial enlargement, LV mass and wall thickness and concentric hypertrophy) as well as levels of circulating natriuretic peptides.<sup>2,28</sup> The H<sub>2</sub>FPEF<sup>27</sup> score combines clinical and echocardiographic patient characteristics: obesity, hypertension, AF, pulmonary hypertension, age >60 years and diastolic function ( $E/e'$ ). A higher score represents a higher likelihood of having HFpEF (HFA-PEFF ≥5 points; H<sub>2</sub>FPEF >6 points), while a lower score is used to rule out HFpEF. For patients with an intermediate score, both algorithms recommend additional testing to refine the diagnosis by exercise echocardiography or invasive measurements of cardiac filling pressures in a non-resting state.<sup>19,27</sup> AF, atrial fibrillation; GLS, global longitudinal strain; HF, heart failure; LV, left ventricle; PASP, pulmonary artery systolic pressure.

H<sub>2</sub>FPEF<sup>27</sup> scores, were developed to standardize and improve the accuracy of HFpEF diagnosis. Both of these scores (Figure 1) use a stepwise diagnostic approach to score and evaluate probability of HFpEF presence. The H<sub>2</sub>FPEF score uses functional echocardiographic data and places emphasis on the presence of comorbidities (e.g. hypertension, obesity) and the effect of age, while not including natriuretic peptide levels. The HFA-PEFF algorithm also assesses pretest probability based on clinical features (including age and comorbidities) and similarly includes a score but based on both functional and structural echocardiographic data, including morphological aspects of the left atrium and LV, as well as levels of natriuretic peptides, such as N-terminal pro brain natriuretic peptide (NT-proBNP).

Both HFpEF scores have recently been validated in various patient cohorts<sup>29–33</sup> and communities studies<sup>34</sup> and it was concluded that both HFpEF scores categorized patients well, especially in those patients with intermediate and high scores. These scores, however, are not without controversy, with criticisms ranging from over-simplification of the diagnostic challenges to over-complicating the diagnostic process by requiring expensive tests or the scores

largely disagree.<sup>35,36</sup> In addition, misclassification has been reported, especially in those patients with low HFpEF scores, potentially due to the fact that both scores use resting parameters in a phenotype in which physiological abnormalities augment during exercise.<sup>33,37</sup> Nevertheless, both scores have been shown to have prognostic utility in human patients,<sup>38,39</sup> suggesting that they capture key pathophysiological components that determine outcomes in HFpEF.

Of note, the combined considerations of the phenotypic complexity of HFpEF, the interplay of cardiac and non-cardiac comorbidities, and the role that these comorbidities play in the pathophysiology of HFpEF have not been adequately taken into account in the evaluation of pre-clinical models of HFpEF. While the HFA-PEFF and H<sub>2</sub>FPEF algorithms have been developed to standardize and improve HFpEF diagnosis in patients, these scores may represent a novel approach to improve putative applicability of HFpEF mouse models. Therefore, this review aims to evaluate the translational aspects of currently available pre-clinical mouse models of HFpEF in the context of the HFA-PEFF and H<sub>2</sub>FPEF scores and proposes a novel approach to the assessment and development of future pre-clinical HFpEF models.

## HFpEF in mice: where do we stand?

Over the last decades, development of HFpEF specific treatments has been disappointing. Standard, successful, HF with reduced ejection fraction (HFrEF) treatment options, such as angiotensin-converting enzyme inhibitors (ACEi), angiotensin receptor 1 blockers and mineralocorticoid receptor antagonists (MRA) did not convincingly reduce mortality and morbidity rates in HFpEF patients.<sup>40–42</sup> Trials with other types of drugs, such as nitric oxide donors and cyclic guanosine monophosphate (cGMP) stimulating therapies failed to improve clinical status,<sup>43–47</sup> or were neutral for the primary endpoint (angiotensin receptor–neprilysin inhibitor, PARAGON-HF trial<sup>48,49</sup>). To date, no HFpEF specific treatment options exist and there is an unmet need to improve morbidity and mortality rate in these patients.

Drug development typically progresses in stages, from pre-clinical to clinical. Valuable HFpEF animal models presenting clinical HFpEF phenotypes are crucial for the successful design of new therapies. This has been neglected so far, which has led to the failure of many clinical studies. Sildenafil, for example, successfully reduced LV hypertrophy and cardiac remodelling in mice that suffered from angiotensin II (ANGII)-induced or transverse aortic constriction (TAC) induced HF.<sup>50,51</sup> Clinical studies of sildenafil in HFpEF patients, however, did not observe these beneficial effects on clinical or hemodynamic parameters.<sup>45</sup> Studies with ACEi in myocardial infarction models (MI),<sup>52,53</sup> successfully reduced hypertrophy and fibrosis with a concomitant improvement of cardiac function. However, studies in patients with HFpEF have yielded inconsistent results.<sup>40</sup> This was also the case for the MRA spironolactone: in pre-clinical studies in diet induced<sup>51,54</sup> and myocardial infarction (MI)<sup>55,56</sup> models this drug improved systolic and diastolic cardiac function. A subsequent large randomized controlled trial on the other hand, remained neutral and did not meet its endpoint.<sup>41</sup> The unsuccessful bench-to-bedside translation may, at least partly, be explained by the fact that pre-clinical animals models not fully recapitulate the clinical HFpEF phenotype and TAC or MI models cannot be considered as HFpEF model.

In this review we discuss and score several pre-clinical HFpEF models using the HFA-PEFF and H<sub>2</sub>FPEF scores. We found that several major discrepancies exist between pre-clinical HF models and clinical HFpEF. Pre-clinical HFpEF models do not always recognize the importance of signs and symptoms of HFpEF, or clinical HFpEF characteristics (*graphical abstract*). Several so-called HFpEF models would have obtained high scores according to the HFA-PEFF and H<sub>2</sub>FPEF risk scores (*Figure 2*) due to functional or morphological features, while signs of lung congestion or exercise impairment were absent and levels of natriuretic peptides low (*Table 1*). Thus, a model without pulmonary congestion may relate to hypertensive heart disease in humans rather than clinical HFpEF (for example *db/db* or *ob/ob* models). The currently developed HFA-PEFF and H<sub>2</sub>FPEF scores both emphasize typical symptoms and signs of HF, or clinical HFpEF characteristics as key for the diagnosis of HFpEF. Although the assessment of signs and symptoms or diagnostic HF criteria may be more challenging in animals than in humans, it is not impossible. Pulmonary congestion can be demonstrated by increased lung weight, and

reduced exercise tolerance can be measured via voluntary or forced exercise testing. Reduced exercise tolerance is one of the hallmarks in human HFpEF and should ideally be part of phenotyping HFpEF animal models.

Importantly, the demonstration of LV diastolic dysfunction has been the cornerstone of validation of a HFpEF animal model; however, the presence of diastolic dysfunction alone is neither synonymous nor sufficient for a diagnosis of HFpEF. Indeed, diastolic dysfunction, as occurs with aging, can exist without the presence of symptomatic HF. Nonetheless, aging is a potent risk factor for HFpEF.<sup>7,57,58</sup> Aging itself is associated with ventricular-vascular stiffening and fibrosis, key mechanisms in the pathogenesis of HFpEF.<sup>59,60</sup> The aging process also exacerbates chronic systemic inflammation, dysregulation of energy supply<sup>61–63</sup> and increased cardiomyocyte stiffness and increased hypertrophy that may all result in HFpEF specific diastolic dysfunction and cardiac remodelling.<sup>64,65</sup> We realize that aging itself can have major practical limitations (>20 months to produce the phenotype); however, because it is such an important factor, we encourage researchers to include it.

Another major difference between animal and human HFpEF can be found in disease complexity and disease heterogeneity. In humans, HFpEF is considered a multifactorial and heterogeneous disease with a plethora of clinical manifestations.<sup>11</sup> For many years, pre-clinical HFpEF models have relied upon a single perturbation. The development of several recent multifactorial models has shown that it is feasible to develop a HFpEF-like phenotype in mice by using multiple perturbations, and these models may represent a new era of multifactorial pre-clinical HFpEF models.

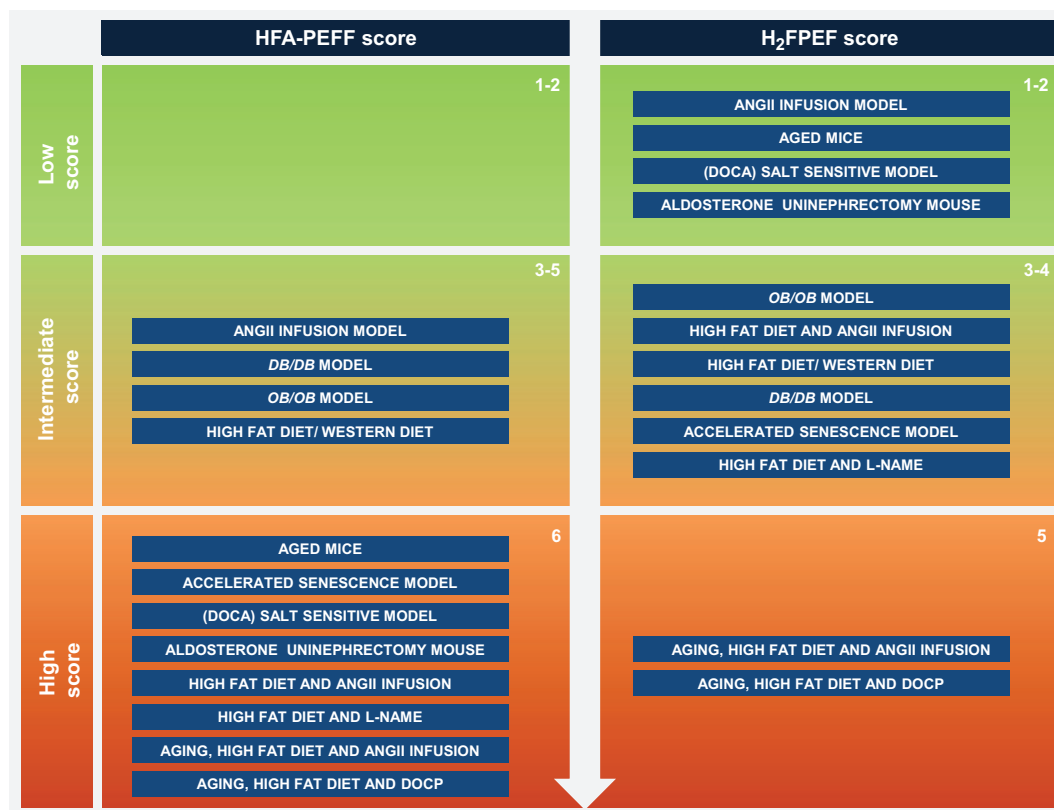
## HFpEF in mice: fundamental checklist

We do not believe that ‘one-size-fits-all’ pre-clinical HFpEF model exists. Several animal models of HFpEF have been developed that only focused on a limited aspect of this multifactorial syndrome. This strategy has been proven unsuccessful and the recent development of combinatory models is very promising.<sup>66–68</sup> Although recent multifactorial HFpEF models have been proven valuable, and may improve bench-to-bed translation, these models also focus on specific HFpEF phenotypes and do not recapitulate the entire heterogeneity of the clinical HFpEF syndrome. In addition, technical challenges remain in developing mouse models. AF, for example, has not been included in any of the pre-clinical HFpEF models so far.

We therefore suggest that all pre-clinical HFpEF studies should include a mouse model that fulfils (a majority of) the following requirements in order to perform a reliable and accurate pre-clinical HFpEF study. This has been schematically presented in *Figure 3*.

## Pretest assessment of signs and symptoms and clinical HFpEF features

First of all, ejection fraction should be preserved. Assessment of symptoms such as shortness of breath, fatigue, oedema, tachycardia, and exercise impairment in animals may be less straightforward than in humans, but various parameters are available to provide a global



**Figure 2** HFA-PEFF and H<sub>2</sub>FPEF scores obtained by HF models. All HF models have been scored for cardiac and extra-cardiac domains of HFA-PEFF and H<sub>2</sub>FPEF scores. Based upon these scores, mouse HF models are differentiated into more or less likely to fulfil the criteria of the HFA-PEFF or H<sub>2</sub>FPEF score. If we solely record the scores, several of so-called HFpEF models would have obtained high scores due to functional or morphological features, while signs of lung congestion or exercise impairment were absent and levels of natriuretic peptides low. ANGII, angiotensin II; DOCA, deoxycorticosterone acetate; DOCP, desoxycorticosterone pivalate; *db/db*, leptin receptor-deficient model; HFpEF, heart failure with preserved ejection fraction; L-NAME, *N*( $\omega$ )-nitro-L-arginine methyl ester; *ob/ob*, leptin-deficient model.

impression if signs and symptoms and clinical HFpEF features are present:

- *Increased natriuretic peptide levels.* Natriuretic peptide levels should be measured in plasma or LV tissue. Elevated natriuretic peptide levels play an important part in the HFA-PEFF score and also provide a global impression if HFpEF is likely to be present in animals.
- *Impaired exercise performance.* Impaired exercise capacity caused by skeletal muscle weakness, fatigue, or cardiovascular to muscle mismatch should be measured by voluntary or forced exercise. This is a typical feature of HFpEF, and analysis of exercise capacity, including assessment of skeletal muscle function, will provide essential information regarding HFpEF severity.<sup>69–71</sup>
- *Lung congestion.* Analysis of lung weight and pulmonary vasculature will be helpful to determine increased diastolic filling pressures and presence of diastolic dysfunction.

In case surrogate measurements of signs and symptoms and clinical HFpEF features (increased natriuretic peptides, preserved ejection fraction and increased comorbidity burden) are not present, the pre-clinical model does not meet the HFpEF criteria as suggested by the

two scores and should therefore not be regarded as a pre-clinical HFpEF model.

### A distinct cardiac phenotype with preserved systolic lv function with concentric hypertrophy and diastolic dysfunction

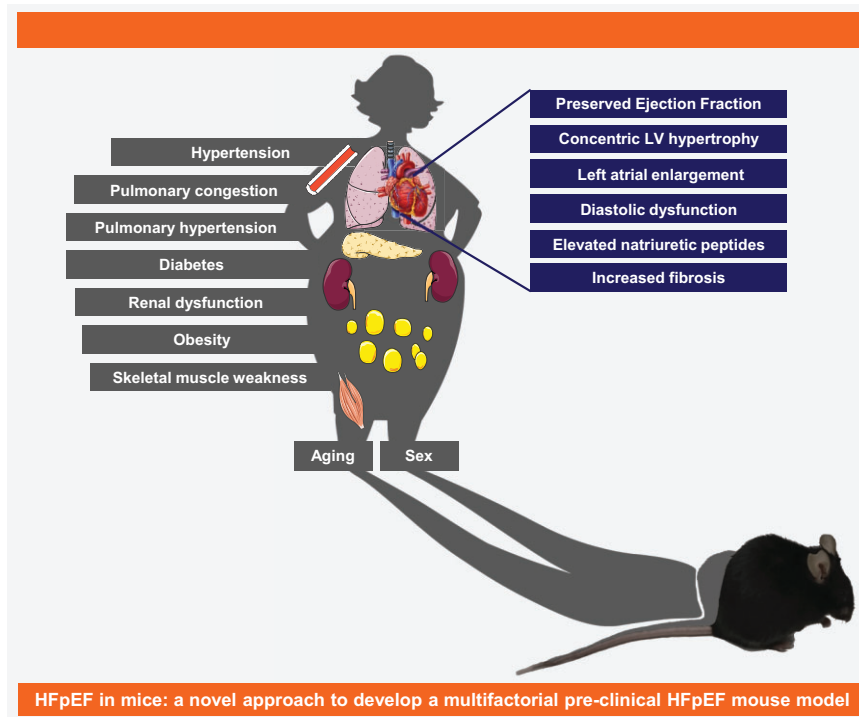
- *Assessment of systolic cardiac function.* Systolic cardiac function should be assessed by transthoracic echocardiography and should include measurement of LV dimensions to assess concentric hypertrophy and LV systolic function. Post-mortem analysis (weighing and staining) of the total heart and LV should take place to assess amount of cardiac hypertrophy and fibrosis.
- *Assessment of diastolic function.* Diastolic function should be determined by morphological criteria (atrial enlargement) or functional parameters. In mice, evaluation of diastolic function is complex and the E/A and *E/e'* ratio is difficult to assess and highly variable.<sup>72</sup> Global longitudinal strain (GLS) and reverse peak longitudinal strain rate (RPLSR) are easily obtained, highly reproducible, and

**Table 1 Validation of HFpEF mouse models by HFA-PEFF and H<sub>2</sub>FPEF scores**

Model	Pretest assessment of signs and symptoms, clinical HFpEF features and biological factors (age and sex)										HFA-PEFF score			H <sub>2</sub> FPEF score										
	Preserved EF	Sex	Age (months)	Lung congestion	Impaired Exercise capacity	Hyper-tension	Obesity	T2DM	Renal Dysfunction	Functional aspects	Diastolic dysfunction	Left atrial enlargement	Left ventricular mass	Increased wall thickness	Concentric hypertrophy	Increased natriuretic peptides	Total Obesity points	Hyper-tension	Atrial Fibrillation	Pulmonary hypertension	Age	Diastolic dysfunction	Total points	
<b>Low HFpEF likelihood</b>																								
Aldosterone uninephrectomy mouse	Yes	M	3	Yes	Yes	Yes	Yes	Yes	N/A	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	No	Yes	N/A	No	Yes	2	
High fat diet/Western diet	Yes	M/F	3–16	Yes	Yes	Yes	Yes	No	Yes	Yes	N/A	Yes	Yes	Yes	No	No	4	Yes	No	N/A	No	Yes	4	
Aged mice (24–30 months)	Yes	M	24–30	Yes	Yes	No	No	No	Yes	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	No	No	N/A	Yes	Yes	2	
Angiotensin-II infusion models	Yes	M/F	3	Yes	Yes	Yes	No	No	Yes	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	No	Yes	yes	No	No	2	
Accelerated senescence model (SAMP)	Yes	F	3–12	No	Yes	Yes	No	No	N/A	Yes	Yes	Yes	Yes	Yes	Yes	Yes	6	No	Yes	N/A	Yes	Yes	4	
Leptin receptor-deficient model (db/db)	Yes	M/F	3	No	Yes	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No	4	Yes	Yes	N/A	No	Yes	4	
Leptin-deficient model (ob/ob)	Yes	M/F	3	No	Yes	No	Yes	Yes	No	Yes	No	Yes	Yes	Yes	Yes	No	4	Yes	Yes	Yes	No	Yes	3	
(DOCA) salt-sensitive model	Yes	M	3	No	No	Yes	No	No	Yes	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	No	No	No	No	Yes	1	
High fat diet and angiotensin II	Yes	M	3	No	No	Yes	Yes	N/A	Yes	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	Yes	Yes	N/A	No	Yes	3	
<b>High HFpEF likelihood</b>																								
High fat diet and L-NAME	Yes	M/F	3	Yes	Yes	Yes	Yes	Yes	N/A	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	Yes	Yes	N/A	No	Yes	4	
Aging, high fat diet and angiotensin II	Yes	F	22	Yes	Yes	Yes	Yes	N/A	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	6	Yes	Yes	N/A	Yes	Yes	5	
Aging, high fat and DOCP	Yes	M/F	18	Yes	Yes	Yes	Yes	N/A	Yes	Yes	N/A	Yes	Yes	Yes	Yes	Yes	6	Yes	Yes	N/A	Yes	Yes	5	

HF models are scored for signs and symptoms or clinical HFpEF features, included age, sex, as well as cardiac and extra-cardiac domains of HFA-PEFF and H<sub>2</sub>FPEF scores. Based upon these scores, mouse HF models were differentiated into more or less likely to fulfil the criteria of the human HFpEF situation, with higher scores representing pre-clinical HF models that most resembled clinical HFpEF. Models that presented full signs and symptoms and clinical HFpEF features are shown in the high HFpEF likelihood box.

db/db, leptin receptor-deficient model; DOCA, deoxycorticosterone acetate; DOCP, desoxycorticosterone pivalate; EF, Ejection fraction; L-NAME, N(ω)-nitro-L-arginine methyl ester; ob/ob, leptin-deficient model; T2DM, type 2 diabetes mellitus.



**Figure 3** HFpEF in mice: a novel approach to develop a multifactorial pre-clinical HFpEF mouse model. The following clinical HFpEF features are essential to develop a reliable and accurate pre-clinical HFpEF model: (1) pulmonary congestion and elevated natriuretic peptides; (2) a distinct cardiac phenotype with preserved systolic LV function with concentric hypertrophy, fibrosis, atrial enlargement and diastolic dysfunction; (3) extra-cardiac comorbidities such as hypertension, obesity, T2DM and renal dysfunction and skeletal muscle weakness; and (4) incorporate and evaluate the effect of sex and aging. LV, left ventricle. Parts of the figure were drawn by using pictures from Smart Servier Medical Art (<http://smart.servier.com>), licensed under a Creative Commons Attribution 3.0 Generic License (<https://creativecommons.org/licenses/by/3.0/>).

have therefore to be integrated as indices of diastolic dysfunction in mice.<sup>73,74</sup> Post-mortem analysis (weighing) of atria should take place to evaluate atrial enlargement.

- **Assessment of cardiac hemodynamics.** Although considered as gold standard for diagnosis of HFpEF, invasive hemodynamic measurements are performed to a limited scale in humans due to a lack of expertise, availability, risks, and costs. A distinct advantage in animal models is that this gold standard assessment can be done more easily and more frequently but requires experience to be reliable. Invasive hemodynamic measurements provide information on intracardiac volumes, filling pressures, contractile and relaxation forces and derive measures such as tau,  $dP/dT$  of the LV. Although measurements of systolic pulmonary artery pressure and pulmonary capillary wedge pressure yield additional information about diastolic function and pulmonary hypertension, measuring right-sided invasive hemodynamics presents more of a challenge in pre-clinical models and may not be required if gold standard left-sided invasive hemodynamics are already evaluated.

### Extra-cardiac comorbidities such as hypertension, obesity, type 2 diabetes mellitus, and renal dysfunction

Assessment of extra-cardiac features of HFpEF should take place in all pre-clinical HFpEF models. This assessment should include

evaluation of several comorbidities that are closely related to the development of HFpEF.

- **Hypertension.** Assessment of hypertension can be performed in several ways, including invasive hemodynamic measurements at sacrifice or by using tail-cuff measurements or continuous registrations throughout the study period.
- **Renal function.** Plasma should be obtained to determine kidney function. Post-mortem analysis of kidneys should take place (weighing + staining).
- **Obesity.** Mice should be repeatedly weighed during the experiment. Body mass composition should be determined throughout the experiment and prior to sacrifice.
- **T2DM.** Fasting plasma glucose levels or glycated hemoglobin should be obtained throughout the experiment. Glucose tolerance can be evaluated by oral glucose tolerance test and insulin sensitivity can be tested by insulin tolerance test.
- **Skeletal muscle weakness.** Post-mortem analysis of skeletal muscle should take place to evaluate reduced mass, and address impaired skeletal oxidative metabolism and abnormal skeletal muscle composition.

AF is a well-known comorbidity for HFpEF and represents an important part of the H<sub>2</sub>FPEF score (three points if AF is present). Unfortunately, induction of AF in mice is challenging and so far none

of the experimental AF models resemble typical clinical HFpEF characteristics.<sup>75–77</sup> We therefore excluded AF from this section.

## Effect of sex and aging

Epidemiological evidence suggests that HFpEF is highly represented in older women.<sup>78</sup> The effect of aging and sex should therefore be taken into account when developing a pre-clinical model.

- **Aging.** The life span of a rodent is shorter than humans, and mice are already considered 'old' after 18 months and 'very old' when >24 months.<sup>79</sup> Aging may represent an important contributing factor to the development of HFpEF and should therefore be considered when studying HFpEF.<sup>57,80</sup>
- **Female sex.** Sex-specific differences are known to exist in humans and mice<sup>4,12,81–86</sup> and for various interventions, young female mice have been shown to be less susceptible to develop a cardiac phenotype as compared to young males.<sup>87,88</sup> Hormonal differences or hormonal changes (such as menopause) are thought to play an important role in the increased cardiovascular risk profile of older females.<sup>21,89</sup> Interestingly, the development of LV hypertrophy may also occur in a sex-specific manner: females more often display concentric remodeling<sup>89</sup> while males develop eccentric LV remodeling.<sup>90</sup> Since the meaning of these differences are not fully understood yet we strongly advise to develop pre-clinical HFpEF models that take into account the effect sex may have. At the very least, investigators may consider including females rather than performing exclusively male experiments as is often the case.

## Validation and translation of the H<sub>2</sub>FPEF and HFA-PEFF scores in animal models

For most experimental HFpEF models, mice are preferred small animals since they are easy to handle, quick to breed, allow genetic experiments, and are known to produce reliable and highly reproducible outcomes. Larger animal models of HFpEF, such as rat,<sup>91–108</sup> dogs<sup>109,110</sup> and pigs,<sup>111–117</sup> also exist (summarized in [Supplementary material online, Table S1](#)); nevertheless, ethical issues, difficulty in introducing high throughput genetic and molecular studies, cost, and duration of study limit large animal models. We included mice models that were widely used in HF research, and are presented as 'HFpEF' models, or were used to evaluate several HFpEF treatment options in the pre-clinical phase, often without translational success.

All models were scored for pre-clinical sign and symptoms or clinical HFpEF features (including age and sex), as well as cardiac and extra-cardiac domains of HFA-PEFF and H<sub>2</sub>FPEF scores ([Table 1](#)). Based upon these scores, mouse HF models have been differentiated into more or less likely to fulfil the criteria of the HFA-PEFF or H<sub>2</sub>FPEF score, schematically presented in [Figure 2](#). In the [Graphical abstract](#), we presented the models in less or high likelihood for HFpEF, including whether models with higher scores also present pre-clinical signs and symptoms or clinical HFpEF characteristics.

## Angiotensin-II infusion models

Chronic stimulation of the ANGII type 1 receptor with ANGII infusion by osmotic mini-pumps is a well-known and reliable model

to induce HF with cardiac hypertrophy and increased remodelling. Remodeling takes place with<sup>118–122</sup> or without<sup>123</sup> hypertension, depending on the dosage of ANGII. The ANGII effects seems to be strain specific: treatment with ANGII in Balb/c<sup>124</sup> mice typically results in lung congestion and LV dilatation, whereas treatment with ANGII in C57BL6 mice results in lung congestion, as well as exercise intolerance, concentric remodelling with fibrosis, and increased levels of natriuretic peptides.<sup>120,122</sup> ANGII treated mice develop diastolic dysfunction that includes worsening LV isovolumetric relaxation time, increased LV end-diastolic pressure and increased *E/e'*.<sup>50,120–124</sup> In mice, exogenous ANGII administration does not interfere with kidney function,<sup>123</sup> but may induce skeletal muscle alterations.<sup>125</sup> ANGII models, and especially the ANGII induced hypertension models, resemble cardiac features of human HFpEF to a large extent. Effects of age and obesity, however, are neglected in this model resulting in the following scores:

- Pretest assessment of signs and symptoms and clinical HFpEF features: lung congestion, hypertension and reduced exercise capacity.
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy, increased natriuretic peptide levels);
- Total H<sub>2</sub>FPEF score: 2 (hypertension and increased filling pressures).

## Leptin receptor-deficient model (*db/db*)

Genetically modified *db/db* mice have a point mutation in the gene encoding for the leptin receptor that leads to malfunctioning of this receptor.<sup>126</sup> These mice are typically used for cardiometabolic research, especially for studies in the field of non-insulin dependent T2DM. Young *db/db* mice develop obesity, hyperglycaemia and severe dyslipidemia without hypertension.<sup>127</sup> The onset of symptoms in mice is severe and early in life, and therefore not directly translatable to the human situation in which progression of obesity and T2DM is a slower and chronic process. *db/db* mice have been from different strains, different ages and different sex<sup>128</sup> and results from studies performed in these mice are therefore not always comparable.

In general, *db/db* mice develop diastolic dysfunction including atrial enlargement, concentric hypertrophy, and fibrosis at older ages.<sup>129,130</sup> LV ejection fraction remains preserved, with decreased GLS rates after 16 weeks. Hypertension may be present, with<sup>131,132</sup> or without<sup>133,134</sup> ANGII infusion. Development of cardiac hypertrophy may already be present at early age (8–9 weeks<sup>133,135</sup>) or develops at a later point in time (up to 16 weeks<sup>136,137</sup>). Most *db/db* mice develop concentric hypertrophy, although eccentric hypertrophy has been observed as well.<sup>138</sup> Signs of congestion are usually not present in these mice, and natriuretic peptide levels are not elevated.<sup>139,140</sup>

- Pretest assessment of signs and symptoms and clinical HFpEF features: increased comorbidity burden (obesity and diabetes) and reduced exercise capacity.
- Total HFA-PEFF score: 4 (diastolic dysfunction, LV hypertrophy);
- Total H<sub>2</sub>FPEF score: 4 (obesity, hypertension, diastolic dysfunction).



## Leptin-deficient model (*ob/ob*)

The *ob/ob* is a leptin-deficient mouse that spontaneously develops obesity (within 4 weeks) and T2DM secondary to hyperglycaemia and hyperinsulinemia.<sup>141,142</sup> The mice develop concentric hypertrophy with diastolic dysfunction possible due to lipid accumulation.<sup>143</sup> The ejection fraction is preserved without congestion or exercise impairment and natriuretic peptide levels are unchanged or reduced.<sup>144–146</sup> The observed maladaptive cardiac alterations appear to be related to the loss of leptin mediated signaling and are reversed by recombinant leptin treatment.<sup>129,147</sup> However, obese HFpEF patients with leptin deficiency are rarely observed, so the *ob/ob* mice do not mimic the human HFpEF phenotype.<sup>148</sup>

- Pretest of signs and symptoms and clinical HFpEF features: increased comorbidity burden (obesity and diabetes) and reduced exercise capacity.
- Total HFA-PEFF score: 4 (diastolic dysfunction, LV hypertrophy);
- Total H<sub>2</sub>FPEF score: 3 (obesity, diastolic dysfunction).

## High fat diet/western diet

Obesity is an important comorbidity in patients with HFpEF and has been suggested to play an important role in (development of) HFpEF.<sup>149,150</sup> In pre-clinical models, unhealthy food consumption is mimicked by a high fat diet (HFD) (>60% fat of daily caloric intake) or by a Western diet (36% fat and 36% sucrose of daily intake). Both of these diets are able to induce an unfavourable cardiometabolic phenotype with obesity and glucose intolerance in young male and female animals<sup>138,151–155</sup> albeit in a strain-specific manner.<sup>156–159</sup> In older animals, the HFD appears to result in more profound cardiometabolic changes including hyperglycaemia and insulin resistance and more profound inflammation.<sup>160,161</sup> There may also be sex-specific effect as female mice tend to gain more weight than age-matched male littermates.<sup>81,138,154–164</sup>

Besides an unfavourable cardiometabolic phenotype, these models result in concentric LV hypertrophy with preserved ejection fraction, and mild to moderate diastolic dysfunction.<sup>154,163,164</sup> Furthermore, pulmonary hypertension has been described as well as increased levels of cardiac fibrosis.<sup>164,165</sup> Pulmonary congestion is absent and levels of natriuretic peptides are usually not elevated.<sup>166</sup> Renal dysfunction may occur after long term diet (>20 weeks) in young mice or at earlier point in time in aged mice.<sup>161,167,168</sup> Mice fed on an HFD or Western diet typically show reduced exercise capacity, most likely related to their obese state as skeletal muscle weakness is not observed in these mice.<sup>71,157,169</sup>

- Pretest assessment of signs and symptoms and clinical HFpEF features: increased comorbidity burden (obesity and pre-diabetes) and reduced exercise capacity.
- Total HFA-PEFF score: 4 (diastolic dysfunction, LV hypertrophy).
- Total H<sub>2</sub>FPEF score: 4 (obesity, pulmonary hypertension, diastolic dysfunction)

## Aged mice (24–30 months)

Similar to humans, natural aging in mice (with or without dietary intervention) is a main driver of development of a maladaptive cardiac HFpEF phenotype.<sup>170</sup> At an age of 24–30 months, mice recapitulate many hallmarks of human HFpEF pathophysiology, including diastolic

dysfunction, concentric hypertrophy with fibrosis and reduced exercise capacity.<sup>171,172</sup> These mice furthermore have lung congestion and increased natriuretic peptide levels. Hypertension or T2DM, however, have not been described.

- Pretest assessment of signs and symptoms and clinical HFpEF features: lung congestion, increased natriuretic peptide levels, reduced exercise capacity, but no comorbidity burden.
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy);
- Total H<sub>2</sub>FPEF score: 2 (age, diastolic dysfunction).

## Accelerated senescence model (SAMP)

Senescence accelerated prone (SAMP) mice belong to a strain of mice that were generated by selective inbreeding of AKR/J mice.<sup>173</sup> These mice show accelerated senescence and age-related pathological phenotypes, similar to aging disorders seen in humans. In addition, they start displaying features of aging at younger age (10 months) than normal mice (8 months).<sup>174</sup> Deleterious mutations in the DNA repair genes are to be involved in their genetic vulnerability for enhanced aging, and specific gene analyses show involvement of oxidative and stress response pathways.<sup>175</sup> SAMP mice develop age-related diastolic dysfunction with atrial enlargement and adverse cardiac remodelling including LV hypertrophy and fibrosis.<sup>102,159,176</sup> Levels of natriuretic peptides are elevated in these mice.<sup>159</sup> When fed a Western diet, SAMP mice also develop hypertension and lung congestion, albeit without obesity or T2DM.<sup>159</sup> It has not been described if female or male SAMPs age differently.

- Pretest assessment of signs and symptoms and clinical HFpEF features: increased natriuretic peptide levels, lung congestion and reduced exercise capacity.
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy, elevated natriuretic peptides).
- Total H<sub>2</sub>FPEF score: 4 (hypertension, effect of aging, increased filling pressures).

## Progress in pre-clinical HF models: development of multifactorial models

The abovementioned models are mostly unifactorial disease models that use one perturbation to induce HF. More recently, progress has been made in the development of pre-clinical HFpEF models and this has led to multifactorial models that use two or more perturbations to mimic the human HFpEF phenotype. In the following section, we will again use the HFA-PEFF and H<sub>2</sub>FPEF score to describe and validate a traditional multifactorial model as well as newer multifactorial HFpEF models.

## Deoxycorticosterone acetate salt-sensitive model

The deoxycorticosterone acetate salt-sensitive model was already developed in 1969 to study hypertension in young mice and rats.<sup>177</sup> This model relies upon a combination of multiple perturbations including administration of deoxycorticosterone acetate, increased salt intake (addition of 1% NaCl to drinking water) and

uninephrectomy. This typically results in cardiac hypertrophy with fibrosis, increased levels of natriuretic peptides, while blood pressure remains unchanged or only mildly increased.<sup>178,179</sup> LV function remains preserved while moderate diastolic dysfunction can be observed.<sup>180</sup> Nevertheless, these mice do not display lung congestion.<sup>181</sup> Again, the effect of age and sex has not been described in this model.

- Pretest assessment of signs and symptoms and clinical HFpEF features: increased natriuretic peptide levels, and reduced exercise capacity.
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy, increased levels of natriuretic peptides);
- Total H<sub>2</sub>FPEF score: 1 (diastolic dysfunction).

### Aldosterone uninephrectomy mouse

Impaired renal function is frequently observed in patients with HFpEF. Renal dysfunction may be attributed to fluid overload, blood pressure elevation, and thus congestion.<sup>182</sup> In C57BL6 or FB/N background, the combination of uninephrectomy and aldosterone infusion results in the development of hypertension, lung congestion, and reduced exercise capacity without obesity or T2DM.<sup>183,184</sup> Preserved LV ejection fraction is observed with concentric remodeling, mild-to-moderate diastolic dysfunction, and increased levels of natriuretic peptides.<sup>185–188</sup> The effect of female sex or aging is unknown and obesity or T2DM is not observed.

- Pretest assessment of signs and symptoms and clinical HFpEF features: lung congestion, increased natriuretic peptide levels and reduced exercise capacity.
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy, increased levels of natriuretic peptides);
- Total H<sub>2</sub>FPEF score: 2 (hypertension, increased filling pressures).

### Combinatory model of high fat diet and L-NAME

Schiattarella *et al.*<sup>169</sup> were the first to present a two-hit pre-clinical mouse model that resembles human HFpEF. In short, C57BL/6N wild-type mice were subjected to a combination of HFD and hypertension that was induced by L-NAME (constitutive nitric oxide synthase inhibitor). They observed that mice that were subjected to both stress factors developed a typical HFpEF phenotype, including lung congestion and reduced exercise tolerance and increased natriuretic peptides. On the contrary, mice that were only exposed to one stressor did not develop this phenotype.<sup>169</sup> More recently, sex-dependent effects have also been shown: young female mice were more resilient for development of HFpEF, as the combination of high-fat and L-NAME resulted in a more attenuated cardiac phenotype as compared to young male mice.<sup>189</sup> The effect of aging was not studied.

- Pretest assessment of signs and symptoms and clinical HFpEF features: increased natriuretic peptides, lung congestion, reduced exercise capacity, and increased comorbidity burden (hypertension, obesity and pre-diabetes).
- Total HFA-PEFF score: 6 points (increased natriuretic peptides, diastolic dysfunction, concentric LV hypertrophy).

- Total H<sub>2</sub>FPEF score: 4 (obesity, hypertension, increased filling pressures).

### Combinatory model of high fat diet and ANGII infusion

The combination of HFD and ANGII infusion induces hypertension, obesity and T2DM in young male mice.<sup>151,190,191</sup> This intervention also results in preserved LV function with diastolic dysfunction, concentric hypertrophy with fibrosis and increased natriuretic peptides. However, signs and symptoms or clinical features of HFpEF, if any, appear to be very mild since lung congestion in young animals is absent and effect on exercise capacity is unknown.<sup>151,190–192</sup>

- Pretest assessment of signs and symptoms and clinical HFpEF features: increased natriuretic peptide levels, increased comorbidity burden (hypertension and pre-diabetes);
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy, elevated levels of natriuretic peptides);
- Total H<sub>2</sub>FPEF score: 4 (obesity, hypertension, increased filling pressures).

### Combinatory model of aging, high fat diet, and ANGII infusion

We have recently developed a multifactorial mouse model that combines aging (18–22 months) with HFD and ANGII infusion.<sup>193</sup> In these older female C57BL6/J mice, a HFpEF-like phenotype is present including concentric LV hypertrophy and LV fibrosis, diastolic dysfunction, lung congestion, increased natriuretic peptide levels, and elevated blood pressures. The effect of sex has not been studied yet.

- Pretest assessment of signs and symptoms and clinical HFpEF features: lung congestion, increased natriuretic peptide levels, reduced exercise capacity, and increased comorbidity burden (hypertension, obesity and pre-diabetes);
- Total HFA-PEFF score: 6 (diastolic dysfunction, concentric LV hypertrophy, elevated natriuretic peptide levels);
- Total H<sub>2</sub>FPEF score: 5 (obesity, hypertension, elderly, increased filling pressures).

### Combinatory model of aging, high fat diet and desoxycorticosterone pivalate

A very recent study by Deng *et al.*<sup>194</sup> used a combinatory model of 16 months of ageing, long-term HFD (13 months) and 3 months of desoxycorticosterone pivalate challenge in mice to induce a HFpEF-like phenotype. Their model resulted in many typical HFpEF features, including lung congestion, hypertension and impaired exercise tolerance. They also showed diastolic dysfunction, LV hypertrophy, fibrosis and increased levels of natriuretic peptides. Both sexes were included but not further studied.

- Pretest assessment of signs and symptoms and clinical HFpEF features: lung congestion, increased natriuretic peptide levels, reduced exercise capacity and increased comorbidity burden (hypertension, obesity and pre-diabetes);
- Total HFA-PEFF score: 6 (diastolic dysfunction, LV hypertrophy, elevated natriuretic peptide levels);
- Total H<sub>2</sub>FPEF score: 5 (obesity, hypertension, elderly, increased filling pressures).

## Conclusion

HFpEF remains a major public health problem worldwide with still increasing prevalence and incidence. So far, HFpEF treatment mostly focuses on symptom reduction since HFpEF-specific drugs do not exist. Despite numerous efforts to develop HFpEF-specific drugs, bench-to-bedside translation has not been successful, and this may, at least partly, be due to the lack of pre-clinical HFpEF models that adequately recapitulate the complexities of the human condition.

HFpEF is a multifactorial disease in which comorbidities contribute to the pathophysiology of the clinical syndrome. While this complicates the development of preclinical models, progress in the field will be aided by consensus on key elements that a HFpEF animal model should manifest. The recent development of two clinical HFpEF scores has led to a novel clinical standard for defining the key clinical features of HFpEF. This state-of-the-art review is the first to apply clinical scores to HFpEF mouse models to improve putative applicability and translational value of pre-clinical HFpEF research. It proposes a novel approach to follow when performing a pre-clinical HFpEF study to optimize bench-to-bed translation and provide a checklist for small HFpEF animal models. Although this checklist may not capture all human HFpEF variables, it will help to provide better and more relevant small animal HFpEF models with better putative application and translational value. So far, most of the pre-clinical models do not fully meet these criteria (presented in [Graphical abstract](#)). Of course, pathophysiology of the mouse heart cannot be translated to humans 1 on 1, and translation of pre-clinical findings to human conditions should always be done cautiously. Of note, clinical studies should be challenged as well to account for diverse HFpEF physiology to optimize bench-to-bed translation.

This review furthermore describes some multifactorial models that resemble human HFpEF to a large extent, and suggests that these small animal models remain attractive models for future HFpEF research. Based on this review, we advocate that future HFpEF pre-clinical studies that test potential new therapeutic agents should consider use of multiple HFpEF animal models so that their effects can be tested on multiple HFpEF phenotypes. Following this approach we believe that pre-clinical HFpEF models will be able to help fill major gaps in HFpEF pathophysiology and will eventually facilitate development of novel HFpEF therapeutics.

## Supplementary material

Supplementary material is available at *European Heart Journal* online.

## Funding

C.W. and C.S.L. are supported by the University of Groningen (Rosaling Franklin fellowship). G.G.S. is supported by the German Center for Cardiovascular Research (DZHK) Junior Research Group Excellence Grant. C.S.L. is supported by a Clinician Scientist Award from the National Medical Research Council of Singapore. This work was supported by grants from the Netherlands Heart Foundation (CVON SHE-PREDICTS-HF, grant 2017-21; CVON RED-CVD, grant 2017-11; CVON PREDICT2, grant 2018-30; and CVON DOUBLE DOSE, grant 2020B005), by a grant from the leDucq Foundation (Cure PhosphoLambaN induced Cardiomyopathy (Cure-PLaN)), and by a grant from the European Research Council (ERC CoG 818715, SECRETE-HF).

**Conflict of interest:** C.W. reports grants from NovoNordisk and AstraZeneca, outside the submitted work. In addition, G.G.S. has a patent PCT/US17/37019 pending. C.S.P.L. reports grants from Boston Scientific, Bayer, Roche Diagnostics, AstraZeneca, Medtronic, and Vifor Pharma, personal fees from Abbott Diagnostics, Amgen, Applied Therapeutics, AstraZeneca, Bayer, Biofourmis, Boehringer Ingelheim, Boston Scientific, Corvia Medical, Cytokinetics, Darma Inc., Us2.ai, JanaCare, Janssen Research & Development LLC, Medtronic, Menarini Group, Merck, MyoKardia, Novartis, Novo Nordisk, Radcliffe Group Ltd, Roche Diagnostics, Sanofi, Stealth BioTherapeutics, The Corpus, Vifor Pharma, and WebMD Global LLC, outside the submitted work. In addition, C.S.P.L. has a patent PCT/SG2016/050217 pending and a patent 16/216,929 issued. R.A.d.B. reports grants from Abbott, grants from AstraZeneca, grants from Boehringer Ingelheim, grants from Cardior Pharmaceuticals GmbH, grants from Ionis Pharmaceuticals, Inc., grants from Novo Nordisk, grants from Roche, personal fees from Abbott, personal fees from AstraZeneca, personal fees from Bayer, personal fees from Novartis, and personal fees from Roche, outside the submitted work. L.M.G.M. reports grants from Mandema Stipendium (UMCG), outside the submitted work.

## References

- Borlaug BA. Evaluation and management of heart failure with preserved ejection fraction. *Nat Rev Cardiol* 2020;**17**:559–573.
- Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JGF, Coats AJS, Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GMC, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P; ESC Scientific Document Group. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: the Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC). Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur Heart J* 2016;**37**:2129–2200.
- Dunlay SM, Roger VL, Redfield MM. Epidemiology of heart failure with preserved ejection fraction. *Nat Rev Cardiol* 2017;**14**:591–602.
- Benjamin EJ, Virani SS, Callaway CW, Chamberlain AM, Chang AR, Cheng S, Chiuve SE, Cushman M, Delling FN, Deo R, De FS, Ferguson JF, Fornage M, Gillespie C, Isasi CR, Jiménez MC, Jordan LC, Judd SE, Lackland D, Lichtman JH, Lisabeth L, Liu S, Longenecker CT, Lutsey PL, MacKey JS, Matchar DB, Matsushita K, Mussolino ME, Nasir K, O'Flaherty M, Palaniappan LP, Pandey A, Pandey DK, Reeves MJ, Ritchey MD, Rodriguez CJ, Roth GA, Rosamond WD, Sampson UKA, Satou GM, Shah SH, Spartano NL, Tirschwell DL, Tsao CW, Voeks JH, Willey JZ, Wilkins JT, Wu JH, Alger HM, Wong SS, Muntner P. Heart disease and stroke statistics – 2018 update: a report from the American Heart Association. *Circulation* 2018;**137**:E67–E492.
- Gori M, Lam CSP, Gupta DK, Santos ABS, Cheng S, Shah AM, Claggett B, Zile MR, Kraigher-Krainer E, Pieske B, Voors AA, Packer M, Bransford T, Lefkowitz M, McMurray JJV, Solomon SD; for the PARAMOUNT Investigators. Sex-specific cardiovascular structure and function in heart failure with preserved ejection fraction. *Eur J Heart Fail* 2014;**16**:535–542.
- Boonman-de Winter LJM, Rutten FH, Cramer MJM, Landman MJ, Liem AH, Rutten GEHM, Hoes AW. High prevalence of previously unknown heart failure and left ventricular dysfunction in patients with type 2 diabetes. *Diabetologia* 2012;**55**:2154–2162.
- Ho JE, Enserro D, Brouwers FP, Kizer JR, Shah SJ, Psaty BM, Bartz TM, Santhanakrishnan R, Lee DS, Chan C, Liu K, Blaha MJ, Hillege HL, van der Harst P, van Gilst WH, Kop WJ, Gansevoort RT, Vasan RS, Gardin JM, Levy D, Gottdiener JS, de Boer RA, Larson MG. Predicting heart failure with preserved and reduced ejection fraction. *Circ Heart Fail* 2016;**9**:e003116.
- Packer M, Kitzman DW. Obesity-related heart failure with a preserved ejection fraction. *JACC Heart Fail* 2018;**6**:633–639.
- Oktay AA, Rich JD, Shah SJ. The emerging epidemic of heart failure with preserved ejection fraction. *Curr Heart Fail Rep* 2013;**10**:401–410.
- Tribouilloy C, Rusinaru D, Mahjoub H, Soulière V, Lévy F, Peltier M, Slama M, Massy Z. Prognosis of heart failure with preserved ejection fraction: a 5 year prospective population-based study. *Eur Heart J* 2008;**29**:339–347.
- Shah SJ, Kitzman DW, Borlaug BA, van Heerebeek L, Zile MR, Kass DA, Paulus WJ. Phenotype-specific treatment of heart failure with preserved ejection fraction. *Circulation* 2016;**134**:73–90.

12. Goyal P, Paul T, Almarzooq ZI, Peterson JC, Krishnan U, Swaminathan RV, Feldman DN, Wells MT, Karas MG, Sobol I, Maurer MS, Horn EM, Kim LK. Sex- and race-related differences in characteristics and outcomes of hospitalizations for heart failure with preserved ejection fraction. *J Am Heart Assoc* 2017;**6**:e003330.
13. Eaton CB, Pettinger M, Rossouw J, Martin LW, Foraker R, Quddus A, Liu S, Wampler NS, Hank Wu W-C, Manson JE, Margolis K, Johnson KC, Allison M, Corbie-Smith G, Rosamond W, Breathett K, Klein L. Risk factors for incident hospitalized heart failure with preserved versus reduced ejection fraction in a multiracial cohort of postmenopausal women. *Circ Heart Fail* 2016;**9**:e002883.
14. Savji N, Meijers WC, Bartz TM, Bhambhani V, Cushman M, Naylor M, Kizer JR, Sarma A, Blaha MJ, Gansevoort RT, Gardin JM, Hillege HL, Ji F, Kop WJ, Lau ES, Lee DS, Sadreyev R, van Gilst WH, Wang TJ, Zanni MV, Vasani RS, Allen NB, Psaty BM, van der Harst P, Levy D, Larson M, Shah SJ, de Boer RA, Gottdiener JS, Ho JE. The association of obesity and cardiometabolic traits with incident HFpEF and HFrEF. *JACC Heart Fail* 2018;**6**:701–709.
15. De Boer RA, Naylor M, DeFilippi CR, Enserro D, Bhambhani V, Kizer JR, Blaha MJ, Brouwers FP, Cushman M, Lima JAC, Bahrami H, Van Der Harst P, Wang TJ, Gansevoort RT, Fox CS, Gaggin HK, Kop WJ, Liu K, Vasani RS, Psaty BM, Lee DS, Hillege HL, Bartz TM, Benjamin EJ, Chan C, Allison M, Gardin JM, Januzzi JL, Shah SJ, Levy D, Herrington DM, Larson MG, Van Gilst WH, Gottdiener JS, Bertoni AG, Ho JE. Association of cardiovascular biomarkers with incident heart failure with preserved and reduced ejection fraction. *JAMA Cardiol* 2018;**3**:215–224.
16. Kanter R, Caballero B. Global gender disparities in obesity: a review. *Adv Nutr* 2012;**3**:491–498.
17. Maréchaux S, Six-Carpentier MM, Bouabdallaoui N, Montaigne D, Bauchart JJ, Mouquet F, Auffray JL, Le Tourneau T, Asseman P, Lejemtel TH, Ennezat PV. Prognostic importance of comorbidities in heart failure with preserved left ventricular ejection fraction. *Heart Vessels* 2011;**26**:313–320.
18. Mentz RJ, Kelly JP, Von Lueder TG, Voors AA, Lam CSP, Cowie MR, Kjeldsen S, Jankowska EA, Atar D, Butler J, Fiuzat M, Zannad F, Pitt B, Connor, OCM. Noncardiac comorbidities in heart failure with reduced versus preserved ejection fraction. *J Am Coll Cardiol* 2014;**64**:2281–2293.
19. Pieske B, Tschöpe C, De Boer RA, Fraser AG, Anker SD, Donal E, Edelmann F, Fu M, Guazzi M, Lam CSP, Lancellotti P, Melenovsky V, Morris DA, Nagel E, Pieske-Kraigher E, Ponikowski P, Solomon SD, Vasani RS, Rutten FH, Voors AA, Ruschitzka F, Paulus WJ, Seferovic P, Filipatos G. How to diagnose heart failure with preserved ejection fraction: the HFA-PEFF diagnostic algorithm: a consensus recommendation from the Heart Failure Association (HFA) of the European Society of Cardiology (ESC). *Eur Heart J* 2019;**40**:3297–3317.
20. Tadic M, Cuspidi C, Plein S, Belyavskiy E, Heinzel F, Galderisi M. Sex and heart failure with preserved ejection fraction: from pathophysiology to clinical studies. *J Clin Med* 2019;**8**:792.
21. De Simone G, Devereux RB, Chinali M, Roman MJ, Barac A, Panza JA, Lee ET, Howard BV. Sex differences in obesity-related changes in left ventricular morphology: the Strong Heart Study. *J Hypertens* 2011;**29**:1431–1438.
22. Hahn VS, Yanek LR, Vaishnav J, Ying W, Vaidya D, Lee YZJ, Riley SJ, Subramanya V, Brown EE, Hopkins CD, Ononogbu S, Perzel Mandell K, Halushka MK, Steenbergen C, Rosenberg AZ, Tedford RJ, Judge DP, Shah SJ, Russell SD, Kass DA, Sharma K. Endomyocardial biopsy characterization of heart failure with preserved ejection fraction and prevalence of cardiac amyloidosis. *JACC Heart Fail* 2020;**8**:712–724.
23. Paulus WJ, Tschöpe C. A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J Am Coll Cardiol* 2013;**62**:263–271.
24. Shah SJ, Lam CSP, Svedlund S, Saraste A, Hage C, Tan RS, Beussink-Nelson L, Faxén UL, Fermer ML, Broberg MA, Gan LM, Lund LH. Prevalence and correlates of coronary microvascular dysfunction in heart failure with preserved ejection fraction: PROMIS-HFpEF. *Eur Heart J* 2018;**39**:3439–3450.
25. Chirinos JA, Orlenko A, Zhao L, Basso MD, Cvijic ME, Li Z, Spires TE, Yarde M, Wang Z, Seiffert DA, Prenner S, Zamani P, Bhattacharya P, Kumar A, Margulies KB, Car BD, Gordon DA, Moore JH, Cappola TP. Multiple plasma biomarkers for risk stratification in patients with heart failure and preserved ejection fraction. *J Am Coll Cardiol* 2020;**75**:1281–1295.
26. Pfeffer MA, Shah AM, Borlaug BA. Heart failure with preserved ejection fraction in perspective. *Circ Res* 2019;**124**:1598–1617.
27. Reddy YNV, Carter RE, Obokata M, Redfield MM, Borlaug BA. A simple, evidence-based approach to help guide diagnosis of heart failure with preserved ejection fraction. *Circulation* 2018;**138**:861–870.
28. Komajda M, Lam CSP. Heart failure with preserved ejection fraction: a clinical dilemma. *Eur Heart J* 2014;**35**:1022–1032.
29. Faxen UL, Venkateshvaran A, Shah SJ, Lam CSP, Svedlund S, Saraste A, Beussink-Nelson L, Lagerstrom Fermer M, Gan L-M, Hage C, Lund LH. Generalizability of HFA-PEFF and H<sub>2</sub>FPEF Diagnostic Algorithms and associations with heart failure indices and proteomic biomarkers: insights from PROMIS-HFpEF. *J Card Fail* 2021;**5**:1071-9164(21)00077-4 doi: 10.1016/j.cardfail.2021.02.005 [Epub ahead of print].
30. Barandiarán Aizpurua A, Sanders-van WS, Brunner-La Rocca H, Henkens M, Heymans S, Beussink-Nelson L, Shah SJ, Empel VPM. Validation of the HFA-PEFF score for the diagnosis of heart failure with preserved ejection fraction. *Eur J Heart Fail* 2020;**22**:413–421.
31. Iwakura K, Onishi T, Okada M, Inoue K, Koyama Y, Okamura A, Yamada T, Yasumura Y, Tamaki S, Hayashi T, Yano M, Fujii K, Hikoso S, Sakata Y. Validation of the HFA-PEFF- and H<sub>2</sub>FPEF score in Japanese patients with heart failure with preserved ejection fraction [abstract]. *Eur Heart J* 2020;**41**:998.
32. Myhre PL, Vaduganathan M, Claggett BL, Lam CSP, Desai AS, Anand IS, Sweitzer NK, Fang JC, O'Meara E, Shah SJ, Shah AM, Lewis EF, Rouleau J, Pitt B, Solomon SD. Application of the H<sub>2</sub>FPEF score to a global clinical trial of patients with heart failure with preserved ejection fraction: the TOPCAT trial. *Eur J Heart Fail* 2019;**21**:1288–1291.
33. Churchill TV, Li SX, Curreri L, Zern EK, Lau ES, Liu EE, Farrell R, Shoenike MW, Sbarbaro J, Malhotra R, Naylor M, Tschöpe C, de Boer RA, Lewis GD, Ho JE. Evaluation of 2 existing diagnostic scores for heart failure with preserved ejection fraction against a comprehensively phenotyped cohort. *Circulation* 2021;**143**:289–291.
34. Selvaraj S, Myhre PL, Vaduganathan M, Claggett BL, Matsushita K, Kitzman DW, Borlaug BA, Shah AM, Solomon SD. Application of diagnostic algorithms for heart failure with preserved ejection fraction to the community. *JACC Heart Fail* 2020;**8**:640–653.
35. Sanders-van Wijk S, Barandiarán Aizpurua A, Brunner-La Rocca H-P, Henkens MTHM, Weerts J, Knackstedt C, Uszko-Lencer N, Heymans S, van Empel V. The HFA-PEFF and H<sub>2</sub>FPEF scores largely disagree in classifying patients with suspected heart failure with preserved ejection fraction. *Eur J Heart Fail* 2020; doi: 10.1002/ehfj.2019 [Epub ahead of print].
36. Coats AJS. Validating the HFA-PEFF score – or how to define a disease? *Eur J Heart Fail* 2020;**22**:428–431.
37. Ho JE, Redfield MM, Lewis GD, Paulus WJ, Lam CSP. Deliberating the diagnostic dilemma of heart failure with preserved ejection fraction. *Circulation* 2020;**142**:1770–1780.
38. Parcha V, Kalra R, Malla G, Patel N, Sanders-van Wijk S, Shah SJ, Arora G, Arora P. Diagnostic and prognostic implications of heart failure with preserved ejection fraction scoring systems. *J Card Fail* 2020;**26**:S38.
39. Sueta D, Yamamoto E, Nishihara T, Tokitsu T, Fujisue K, Oike F, Takae M, Usuku H, Takashio S, Arima Y, Suzuki S, Nakamura T, Ito M, Kanazawa H, Sakamoto K, Kaikita K, Tsujita K. H<sub>2</sub>FPEF score as a prognostic value in HFpEF patients. *Am J Hypertens* 2019;**32**:1082–1090.
40. Khan MS, Fonarow GC, Khan H, Greene SJ, Anker SD, Gheorghiadu M, Butler J. Renin-angiotensin blockade in heart failure with preserved ejection fraction: a systematic review and meta-analysis. *ESC Heart Fail* 2017;**4**:402–408.
41. Pitt B, Pfeffer MA, Assmann SF, Boineau R, Anand IS, Claggett B, Clausell N, Desai AS, Diaz R, Fleg JL, Gordeev I, Harty B, Heitner JF, Kenwood CT, Lewis EF, O'Meara E, Probstfield JL, Shaburishvili T, Shah SJ, Solomon SD, Sweitzer NK, Yang S, McKinlay SM; TOPCAT Investigators. Spironolactone for heart failure with preserved ejection fraction. *N Engl J Med* 2014;**370**:1383–1392.
42. Beldhuis IE, Myhre PL, Claggett B, Damman K, Fang JC, Lewis EF, O'Meara E, Pitt B, Shah SJ, Voors AA, Pfeffer MA, Solomon SD, Desai AS. Efficacy and safety of spironolactone in patients with HFpEF and chronic kidney disease. *JACC Heart Fail* 2019;**7**:25–32.
43. Pieske B, Maggioni AP, Lam CSP, Pieske-Kraigher E, Filipatos G, Butler J, Ponikowski P, Shah SJ, Solomon SD, Scalise A-V, Mueller K, Roessig L, Gheorghiadu M. Vericiguat in patients with worsening chronic heart failure and preserved ejection fraction: results of the SOLuble guanylate Cyclase stimulator in heart failure (SOLVD) study. *Eur Heart J* 2017;**38**:1119–1127.
44. Redfield MM, Chen HH, Borlaug BA, Semigran MJ, Lee KL, Lewis G, LeWinter MM, Rouleau JL, Bull DA, Mann DL, Deswal A, Stevenson LW, Givertz EO, Ofili MM, O'Connor CM, Felker GM, Goldsmith SR, Bart BA, McNulty SE, Ibarra JC, Lin G, Oh JK, Patel MR, Kim RJ, Tracy RP, Velazquez EJ, Anstrom KJ, Hernandez AF, Mascette AM, Braunwald E. Effect of phosphodiesterase-5 inhibition on exercise capacity and clinical status in heart failure with preserved ejection fraction: a randomized clinical trial. *JAMA* 2013;**309**:1268–1277.
45. Hoendermis ES, Liu LCY, Hummel YM, Van Der Meer P, De Boer RA, Berger RMF, Van Veldhuisen DJ, Voors AA. Effects of sildenafil on invasive haemodynamics and exercise capacity in heart failure patients with preserved ejection fraction and pulmonary hypertension: a randomized controlled trial. *Eur Heart J* 2015;**36**:2565–2573.
46. Borlaug BA, Anstrom KJ, Lewis GD, Shah SJ, Levine JA, Koepff GA, Givertz MM, Felker GM, Lewinter MM, Mann DL, Margulies KB, Smith AL, Tang WHW, Whellan DJ, Chen HH, Davila-Roman VG, McNulty S, Desvigne-Nickens P, Hernandez AF, Braunwald E, Redfield MM. Effect of inorganic nitrite vs placebo

- on exercise capacity among patients with heart failure with preserved ejection fraction: the INDIE-HFpEF Randomized Clinical Trial. *JAMA* 2018;**320**:1764–1773.
47. Komajda M, Isnard R, Cohen-Solal A, Metra M, Pieske B, Ponikowski P, Voors AA, Dominjon F, Henon-Goburdhun C, Pannaux M, Böhm M; on behalf of the prServed left ventricular ejection fraction chronic heart Failure with ivabradine study (EDIFY) Investigators. Effect of ivabradine in patients with heart failure with preserved ejection fraction: the EDIFY randomized placebo-controlled trial. *Eur J Heart Fail* 2017;**19**:1495–1503.
  48. Wachter R, Shah SJ, Cowie MR, Szecsódy P, Shi V, Ibram G, Zhao Z, Gong J, Klebs S, Pieske B. Angiotensin receptor neprilysin inhibition versus individualized RAAS blockade: design and rationale of the PARALLAX trial. *ESC Heart Fail* 2020;**7**:856–864.
  49. Solomon SD, McMurray JVV, Anand IS, Ge J, Lam CSP, Maggioni AP, Martinez F, Packer M, Pfeffer MA, Pieske B, Redfield MM, Rouleau JL, van Veldhuisen DJ, Zannad F, Zile MR, Desai AS, Claggett B, Jhund PS, Boytsov SA, Comin-Colet J, Cleland J, Düngen H-D, Gonçalvesova E, Katova T, Kerr Saraiva JF, Lelonek M, Merkely B, Senni M, Shah SJ, Zhou J, Rizkala AR, Gong J, Shi VC, Lefkowitz MP; PARAGON-HF Investigators and Committees. Angiotensin–neprilysin inhibition in heart failure with preserved ejection fraction. *N Engl J Med* 2019;**381**:1609–1620.
  50. Westermann D, Becher PM, Lindner D, Savvatis K, Xia Y, Fröhlich M, Hoffmann S, Schultheiss H-PP, Tschöpe C. Selective PDE5A inhibition with sildenafil rescues left ventricular dysfunction, inflammatory immune response and cardiac remodeling in angiotensin II-induced heart failure in vivo. *Basic Res Cardiol* 2012;**107**:308.
  51. Takimoto E, Champion HC, Li M, Belardi D, Ren S, Rodriguez ER, Bedja D, Gabrielson KL, Wang Y, Kass DA. Chronic inhibition of cyclic GMP phosphodiesterase 5A prevents and reverses cardiac hypertrophy. *Nat Med* 2005;**11**:214–222.
  52. Pfeffer MA, Pfeffer JM, Steinberg C, Finn P. Survival after an experimental myocardial infarction: beneficial effects of long-term therapy with captopril. *Circulation* 1985;**72**:406–412.
  53. Pfeffer JM, Pfeffer MA, Braunwald E. Influence of chronic captopril therapy on the infarcted left ventricle of the rat. *Circ Res* 1985;**57**:84–95.
  54. Bostick B, Habibi J, DeMarco VG, Jia G, Demeier T L, Lambert MD, Aroor AR, Nistala R, Bender SB, Garro M, Hayden MR, Ma L, Manrique C, Sowers JR. Mineralocorticoid receptor blockade prevents western diet-induced diastolic dysfunction in female mice. *Am J Physiol – Hear Circ Physiol* 2015;**308**:H1126–H1135.
  55. Wang D, Liu YH, Yang XP, Rhaleb NE, Xu J, Peterson E, Rudolph AE, Carretero OA. Role of a selective aldosterone blocker in mice with chronic heart failure. *J Card Fail* 2004;**10**:67–73.
  56. Fraccarollo D, Galuppo P, Sieweke JT, Napp LC, Grobbeck P, Bauersachs J. Efficacy of mineralocorticoid receptor antagonism in the acute myocardial infarction phase: eplerenone versus spironolactone. *ESC Heart Fail* 2015;**2**:150–158.
  57. Kenchaiah S, Vasan RS. Heart failure in women – insights from the Framingham Heart Study. *Cardiovasc Drugs Ther* 2015;**29**:377–390.
  58. Brouwers FP, de Boer RA, van der Harst P, Voors AA, Gansevoort RT, Bakker SJ, Hillege HL, van Veldhuisen DJ, van Gilst WH. Incidence and epidemiology of new onset heart failure with preserved vs. reduced ejection fraction in a community-based cohort: 11-year follow-up of PREVENT. *Eur Heart J* 2013;**34**:1424–1431.
  59. Lam CSP, Roger VL, Rodeheffer RJ, Bursi F, Borlaug BA, Ommen SR, Kass DA, Redfield MM. Cardiac structure and ventricular-vascular function in persons with heart failure and preserved ejection fraction from Olmsted County, Minnesota. *Circulation* 2007;**115**:1982–1990.
  60. Redfield MM, Jacobsen SJ, Borlaug BA, Rodeheffer RJ, Kass DA. Age- and gender-related ventricular-vascular stiffening: a community-based study. *Circulation* 2005;**112**:2254–2262.
  61. Terman A, Kurz T, Navratil M, Arriaga EA, Brunk UT. Mitochondrial turnover and aging of long-lived postmitotic cells: the mitochondrial-lysosomal axis theory of aging. *Antioxidants Redox Signal* 2010;**12**:503–535.
  62. Dai D-F, Chen T, Wanagat J, Laflamme M, Marcinek DJ, Emond MJ, Ngo CP, Prolla TA, Rabinovitch PS. Age-dependent cardiomyopathy in mitochondrial mutator mice is attenuated by overexpression of catalase targeted to mitochondria. *Aging Cell* 2010;**9**:536–544.
  63. Schriener SE, Linford NJ, Martin GM, Treuting P, Ogburn CE, Emond M, Coskun PE, Ladiges W, Wolf N, Van Remmen H, Wallace DC, Rabinovitch PS. Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* 2005;**308**:1909–1911.
  64. Dai DF, Chen T, Szeto H, Nieves-Cintrón M, Kutyavin V, Santana LF, Rabinovitch PS. Mitochondrial targeted antioxidant peptide ameliorates hypertensive cardiomyopathy. *J Am Coll Cardiol* 2011;**58**:73–82.
  65. Dai DF, Santana LF, Vermulst M, Tomazela DM, Emond MJ, MacCoss MJ, Gollahon K, Martin GM, Loeb LA, Ladiges WC, Rabinovitch PS. Overexpression of catalase targeted to mitochondria attenuates murine cardiac aging. *Circulation* 2009;**119**:2789–2797.
  66. Conceição G, Heinonen I, Lourenço AP, Duncker DJ, Falcão-Pires I. Animal models of heart failure with preserved ejection fraction. *Neth Heart J* 2016;**24**:275–286.
  67. Valero-Muñoz M, Backman W, Sam F. Murine models of heart failure with preserved ejection fraction: a “Fishing Expedition”. *JACC Basic Transl Sci* 2017;**2**:770–789.
  68. Roh J, Houstis N, Rosenzweig A. Why Don't We Have Proven Treatments for HFpEF? *Circ Res* 2017;**120**:1243–1245.
  69. Brunjes DL, Kennel PJ, Christian Schulze P. Exercise capacity, physical activity, and morbidity. *Heart Fail Rev* 2017;**22**:133–139.
  70. Baltgalvis KA, White K, Li W, Claypool MD, Lang W, Alcantara R, Singh BK, Frieri AM, McLaughlin J, Hansen D, McCaughey K, Nguyen H, Smith IJ, Godinez G, Shaw SJ, Goff D, Singh R, Markovtsov V, Sun T-Q, Jenkins Y, Uy G, Li Y, Pan A, Gururaja T, Lau D, Park G, Hitoshi Y, Payan DG, Kinsella TM. Exercise performance and peripheral vascular insufficiency improve with AMPK activation in high-fat diet-fed mice. *Am J Physiol Heart Circ Physiol* 2014;**306**:H1128–H1145.
  71. Kitzman DW, Nicklas B, Kraus WE, Lyles MF, Eggebeen J, Morgan TM, Haykowsky M. Skeletal muscle abnormalities and exercise intolerance in older patients with heart failure and preserved ejection fraction. *Am J Physiol Heart Circ Physiol* 2014;**306**:H1364–70.
  72. Schnelle M, Catibog N, Zhang M, Nabeebaccus AA, Anderson G, Richards DA, Sawyer G, Zhang X, Toischer K, Hasenfuss G, Monaghan MJ, Shah AM. Echocardiographic evaluation of diastolic function in mouse models of heart disease. *J Mol Cell Cardiol* 2018;**114**:20–28.
  73. Ferferieva V, Van den Bergh A, Claus P, Jasaityte R, La Gerche A, Rademakers F, Herijgers P, D'hooge J. Assessment of strain and strain rate by two-dimensional speckle tracking in mice: comparison with tissue Doppler echocardiography and conductance catheter measurements. *Eur Heart J Cardiovasc Imaging* 2013;**14**:765–773.
  74. Zaccagna S, Paldino A, Falcão-Pires I, Daskalopoulos EP, Dal Ferro M, Vodret S, Lesizza P, Cannatà A, Miranda-Silva D, Lourenço AP, Pinamonti B, Sinagra G, Weinberger F, Eschenhagen T, Carrier L, Kehat I, Tocchetti CG, Russo M, Ghigo A, Cimino J, Hirsch E, Dawson D, Ciccarelli M, Olivetti M, Linke WA, Cuijpers I, Heymans S, Hamdani N, de Boer M, Duncker DJ, Kuster D, van der Velden J, Beauloye C, Bertrand L, Mayr M, Giacca M, Leuschner F, Backs J, Thum T. Towards standardization of echocardiography for the evaluation of left ventricular function in adult rodents: a position paper of the ESC Working Group on Myocardial Function. *Cardiovasc Res* 2021;**117**:43–59.
  75. Nishida K, Michael G, Dobrev D, Nattel S. Animal models for atrial fibrillation: clinical insights and scientific opportunities. *Eurpace* 2010;**12**:160–172.
  76. Zhang F, Hartnett S, Sample A, Schnack S, Li Y. High fat diet induced alterations of atrial electrical activities in mice. *Am J Cardiovasc Dis* 2016;**6**:1–9.
  77. Nakada Y, Canseco DC, Thet S, Abdulsalam S, Asaithamby A, Santos CX, Shah AM, Zhang H, Faber JE, Kinter MT, Szveda LI, Xing C, Hu Z, Deberardinis RJ, Schiattarella G, Hill JA, Oz O, Lu Z, Zhang CC, Kimura W, Sadek HA. Hypoxia induces heart regeneration in adult mice. *Nature* 2017;**541**:222–227.
  78. Scantlebury DC, Borlaug BA. Why are women more likely than men to develop heart failure with preserved ejection fraction? *Curr Opin Cardiol* 2011;**26**:562–568.
  79. Flurkey K, Currer JM, Harrison DE. Mpmouse models in aging research. In: J Fox, S Barthold, M Davison, C Newcomer, F Quimby, A Smith, eds. *The Mouse in Biomedical Research*. 2nd ed. New York, NY: Academic Press; 2007. p637–672.
  80. Loffredo FS, Nikolova AP, Pancoast JR, Lee RT. Heart failure with preserved ejection fraction: molecular pathways of the aging myocardium. *Circ Res* 2014;**115**:97–107.
  81. Salinero AE, Anderson BM, Zuloaga KL. Sex differences in the metabolic effects of diet-induced obesity vary by age of onset. *Int J Obes* 2018;**42**:1088–1091.
  82. Mitchell SJ, Matute JM, Scheibye-Knudsen M, Fang E, Aon M, González-Reyes JA, Cortassa S, Kaushik S, Patel B, Wahl D, Ali A, Calvo-Rubio M, María I, Guiterrez V, Ward TM, Palacios HH, Cai H, David W, Hine C, Broeskamp F, Habering L, Sinclair DA, Cohen P, Egan JM, Mitchell JR, Baur JA, Allison DB, Anson RM, Villalba JM, Cuervo AM, Pearson KJ, Ingram DK, Bernier M. Effects of sex, strain, and energy intake on hallmarks of aging in mice. *Cell Metab* 2017;**23**:1093–1112.
  83. Piro M, Della Bona R, Abbate A, Biasucci LM, Crea F. Sex-related differences in myocardial remodeling. *J Am Coll Cardiol* 2010;**55**:1057–1065.
  84. Krumholz HM, Larson M, Levy D. Sex differences in cardiac adaptation to isolated systolic hypertension. *Am J Cardiol* 1993;**72**:310–313.
  85. Douglas PS, Katz SE, Weinberg EO, Chen MH, Bishop SP, Lorell BH. Hypertrophic remodeling: gender differences in the early response to left ventricular pressure overload. *J Am Coll Cardiol* 1998;**32**:1118–1125.
  86. Harada E, Mizuno Y, Kugimiya F, Shono M, Maeda H, Yano N, Yasue H. Sex differences in heart failure with preserved ejection fraction reflected by B-type natriuretic peptide level. *Am J Med Sci* 2018;**356**:335–343.

87. Shioura KM, Geenen DL, Goldspink PH. Sex-related changes in cardiac function following myocardial infarction in mice. *Am J Physiol Integr Comp Physiol* 2008; **295**:R528–R534.
88. Pettersson US, Waldén TB, Carlsson P-O, Jansson L, Phillipson M. Female mice are protected against high-fat diet induced metabolic syndrome and increase the regulatory T cell population in adipose tissue. *PLoS One* 2012; **7**:e46057.
89. Kajstura J, Gurusamy N, Ogórek B, Goichberg P, Clavo-Rondon C, Hosoda T, D'Amario D, Bardelli S, Beltrami AP, Cesselli D, Bussani R, Del Monte F, Quaini F, Rota M, Beltrami CA, Buchholz BA, Leri A, Anversa P. Myocyte turnover in the aging human heart. *Circ Res* 2010; **107**:1374–1386.
90. Kuch B, Muscholl M, Luchner A, Döring A, Riegger GAJ, Schunkert H, Hense HW. Gender specific differences in left ventricular adaptation to obesity and hypertension. *J Hum Hypertens* 1998; **12**:685–691.
91. Nguyen ITN, Brandt MM, van de Wouw J, van Drie RWA, Wesseling M, Cramer MJ, de Jager SCA, Merkus D, Duncker DJ, Cheng C, Joles JA, Verhaar MC. Both male and female obese ZSF1 rats develop cardiac dysfunction in obesity-induced heart failure with preserved ejection fraction. *PLoS One* 2020; **15**:e0232399.
92. Lekawanvijit S, Kompa AR, Manabe M, Wang BH, Langham RG, Nishijima F, Kelly DJ, Krum H. Chronic kidney disease-induced cardiac fibrosis is ameliorated by reducing circulating levels of a non-dialysable uremic toxin, indoxyl sulfate. *PLoS One* 2012; **7**:e41281.
93. Suzuki H, Schaefer L, Ling H, Schaefer RM, Dämmrich J, Teschner M, Heidland A. Prevention of cardiac hypertrophy in experimental chronic renal failure by long-term ACE inhibitor administration: Potential role of lysosomal proteinases. *Am J Nephrol* 1995; **15**:129–136.
94. Kennedy DJ, Vetteh S, Periyasamy SM, Kanj M, Fedorova L, Khouri S, Kahaleh MB, Xie Z, Malhotra D, Kolodkin NI, Lakatta EG, Fedorova OV, Bagrov AY, Shapiro JL. Central role for the cardiotoxic steroid marinobufagenin in the pathogenesis of experimental uremic cardiomyopathy. *Hypertension* 2006; **47**:488–495.
95. Bongartz LG, Braam B, Verhaar MC, Cramer MJ, Goldschmeding R, Gaillard CA, Doevendans PA, Joles JA. Transient nitric oxide reduction induces permanent cardiac systolic dysfunction and worsens kidney damage in rats with chronic kidney disease. *Am J Physiol Integr Comp Physiol* 2010; **298**:R815–R823.
96. Hamdani N, Franssen C, Lourenço A, Falca O-Pires I, Fontoura D, Leite S, Plettig L, Lopez B, Ottenheim CA, Becher PM, Gonzalez A, Tscho Pe C, Diez J, Linke WA, Leite-Moreira AF, Paulus WJ. Myocardial titin hypophosphorylation importantly contributes to heart failure with preserved ejection fraction in a rat metabolic risk model. *Circ Heart Fail* 2013; **6**:1239–1249.
97. Van Dijk CGM, Oosterhuis NR, Xu YJ, Brandt M, Paulus WJ, Van Heerebeek L, Duncker DJ, Verhaar MC, Fontoura D, Lourenço AP, Leite-Moreira AF, Falcão-Pires I, Joles JA, Cheng C. Distinct endothelial cell responses in the heart and kidney microvasculature characterize the progression of heart failure with preserved ejection fraction in the obese ZSF1 rat with cardiorenal metabolic syndrome. *Circ Heart Fail* 2016; **9**:e002760.
98. Grobe JL, Mecca AP, Mao H, Katovich MJ. Chronic angiotensin-(1–7) prevents cardiac fibrosis in DOCA-salt model of hypertension. *Am J Physiol Circ Physiol* 2006; **290**:H2417–H2423.
99. Schauer A, Adams V, Augstein A, Jannasch A, Draskowski R, Kirchhoff V, Goto K, Mittag J, Galli R, Männel A, Barthel P, Linke A, Winzer EB. Sacubitril/valsartan improves diastolic function but not skeletal muscle function in a rat model of HFpEF. *Int J Mol Sci* 2021; **22**:3570.
100. Bode D, Rolim NPL, Guthof T, Hegemann N, Wakula P, Primessnig U, Berre AMO, Adams V, Wisløff U, Pieske BM, Heinzel FR, Hohendanner F. Effects of different exercise modalities on cardiac dysfunction in heart failure with preserved ejection fraction. *ESC Heart Fail* 2021; **8**:1806–1818.
101. Bode D, Semmler L, Wakula P, Hegemann N, Primessnig U, Beindorff N, Powell D, Dahmen R, Ruettgen H, Oeing C, Alogna A, Messroghli D, Pieske BM, Heinzel FR, Hohendanner F. Dual SGLT-1 and SGLT-2 inhibition improves left atrial dysfunction in HFpEF. *Cardiovasc Diabetol* 2021; **20**:7.
102. Penumatsa KC, Falcão-Pires I, Leite S, Leite-Moreira A, Bhedi CD, Nasirova S, Ma J, Sutliff RL, Fanburg BL. Increased transglutaminase 2 expression and activity in rodent models of obesity/metabolic syndrome and aging. *Front Physiol* 2020; **11**:560019.
103. Hohendanner F, Bode D, Primessnig U, Guthof T, Doerr R, Jeuthe S, Reimers S, Zhang K, Bach D, Wakula P, Pieske BM, Heinzel FR. Cellular mechanisms of metabolic syndrome-related atrial decompensation in a rat model of HFpEF. *J Mol Cell Cardiol* 2018; **115**:10–19.
104. Brenner DA, Apstein CS, Saupé KW. Exercise training attenuates age-associated diastolic dysfunction in rats. *Circulation* 2001; **104**:221–226.
105. Elkholy K, Morris L, Niewiadomska M, Houser J, Ramirez M, Tang M, Humphrey MB, Stavrakis S. Sex differences in the incidence and mode of death in rats with heart failure with preserved ejection fraction. *Exp Physiol* 2021; **106**:673–682.
106. Bustamante M, Garate-Carrillo A, R. Ito B, Garcia R, Carson N, Ceballos G, Ramirez-Sanchez I, Omens J, Villarreal F. Unmasking of oestrogen-dependent changes in left ventricular structure and function in aged female rats: a potential model for pre-heart failure with preserved ejection fraction. *J Physiol* 2019; **597**:1805–1817.
107. Curl CL, Danes VR, Bell JR, Raaijmakers AJA, Ip WTK, Chandramouli C, Harding TW, Porrello ER, Erickson JR, Charchar FJ, Kompa AR, Edgley AJ, Crossman DJ, Soeller C, Mellor KM, Kalman JM, Harrap SB, Delbridge LMD. Cardiomyocyte functional etiology in heart failure with preserved ejection fraction is distinctive—a new preclinical model. *J Am Heart Assoc* 2018; **7**:e007451.
108. Goto K, Schauer A, Augstein A, Methawasin M, Granzier H, Halle M, Craenenbroeck EMV, Rolim N, Gielen S, Pieske B, Winzer EB, Linke A, Adams V. Muscular changes in animal models of heart failure with preserved ejection fraction: what comes closest to the patient? *ESC Heart Fail* 2021; **8**:139–150.
109. Munagala VK, Hart CYT, Burnett JC, Meyer DM, Redfield MM. Ventricular structure and function in aged dogs with renal hypertension: a model of experimental diastolic heart failure. *Circulation* 2005; **111**:1128–1135.
110. Zakeri R, Moulay G, Chai Q, Ogut O, Hussain S, Takahama H, Lu T, Wang X-L, Linke WA, Lee H-C, Redfield MM. Left atrial remodeling and atrioventricular coupling in a canine model of early heart failure with preserved ejection fraction. *Circ Heart Fail* 2016; **9**:1128–1135.
111. Schwarzl M, Hamdani N, Seiler S, Alogna A, Manninger M, Reilly S, Zirngast B, Kirsch A, Steendijk P, Verderber J, Zweiker D, Eller P, Höfler G, Schauer S, Eller K, Maechler H, Pieske BM, Linke WA, Casadei B, Post H. A porcine model of hypertensive cardiomyopathy: implications for heart failure with preserved ejection fraction. *Am J Physiol Heart Circ Physiol* 2015; **309**:H1407–H1418.
112. Sharp TE, Scarborough AL, Li Z, Polhemus DJ, Hidalgo HA, Schumacher JD, Matsuura TR, Jenkins JS, Kelly DP, Goodchild TT, Lefler DJ. Novel Göttingen Miniswine model of heart failure with preserved ejection fraction integrating multiple comorbidities. *JACC Basic Transl Sci* 2021; **6**:154–170.
113. Zhang N, Feng B, Ma X, Sun K, Xu G, Zhou Y. Dapagliflozin improves left ventricular remodeling and aorta sympathetic tone in a pig model of heart failure with preserved ejection fraction. *Cardiovasc Diabetol* 2019; **18**:107.
114. Olver TD, Edwards JC, Jurrissen TJ, Veteto AB, Jones JL, Gao C, Rau C, Warren CM, Klutho PJ, Alex L, Ferreira-Nichols SC, Ivey JR, Thorne PK, McDonald KS, Krenz M, Baines CP, Solaro RJ, Wang Y, Ford DA, Domeier TL, Padilla J, Rector RS, Emtner CA. Western diet-fed, aortic-banded Ossabaw Swine: a preclinical model of cardio-metabolic heart failure. *JACC Basic Transl Sci* 2019; **4**:404–421.
115. Charles CJ, Lee P, Li RR, Yeung T, Ibrahim Mazlan SM, Tay ZW, Abdurrachim D, Teo XQ, Wang WH, de Kleijn DPV, Cozzone PJ, Lam CSP, Richards AM. A porcine model of heart failure with preserved ejection fraction: magnetic resonance imaging and metabolic energetics. *ESC Heart Fail* 2020; **7**:92–102.
116. Gyöngyösi M, Pavo N, Lukovic D, Zlabinger K, Spannauer A, Traxler D, Goliašč G, Mandić L, Bergler-Klein J, Gugerell A, Jakab A, Szankai Z, Toth L, Garamvölgyi R, Maurer G, Jaisser F, Zannad F, Thum T, Bátkai S, Winkler J. Porcine model of progressive cardiac hypertrophy and fibrosis with secondary postcapillary pulmonary hypertension. *J Transl Med* 2017; **15**:202.
117. Sorop O, Heinonen I, van Kranenburg M, van de Wouw J, de Beer VJ, Nguyen ITN, Octavia Y, van Duin RWB, Stam K, van Geuns R-J, Wielopolski PA, Krestin GP, van den Meiracker AH, Verjans R, van Bilsen M, Danser AHJ, Paulus WJ, Cheng C, Linke WA, Joles JA, Verhaar MC, van der Velden J, Merkus D, Duncker DJ. Multiple common comorbidities produce left ventricular diastolic dysfunction associated with coronary microvascular dysfunction, oxidative stress, and myocardial stiffening. *Cardiovasc Res* 2018; **114**:954–964.
118. Shen Y, Cheng F, Sharma M, Merkulova Y, Raithatha SA, Parkinson LG, Zhao H, Westendorf K, Bohunek L, Bozin T, Hsu I, Ang LS, Williams SJ, Bleackley RC, Eriksson JE, Seidman MA, McManus BM, Granville DJ. Granzyme B deficiency protects against angiotensin II-induced cardiac fibrosis. *Am J Pathol* 2016; **186**:87–100.
119. Ichihara S, Senbonmatsu T, Price E, Ichiki T, Gaffney FA, Inagami T. Angiotensin II type 2 receptor is essential for left ventricular hypertrophy and cardiac fibrosis in chronic angiotensin II-induced hypertension. *Circulation* 2001; **104**:346–351.
120. Glenn DJ, Cardema MC, Ni W, Zhang Y, Yeghiazarians Y, Grapov D, Fiehn O, Gardner DG. Cardiac steatosis potentiates angiotensin II effects in the heart. *Am J Physiol Heart Circ Physiol* 2015; **308**:H339–H350.
121. Murdoch CE, Chaubey S, Zeng L, Yu B, Ivetic A, Walker SJ, Vanhoutte D, Heymans S, Grieve DJ, Cave AC, Brewer AC, Zhang M, Shah AM. Endothelial NADPH oxidase-2 promotes interstitial cardiac fibrosis and diastolic dysfunction through proinflammatory effects and endothelial-mesenchymal transition. *J Am Coll Cardiol* 2014; **63**:2734–2741.
122. Becher PM, Lindner D, Miteva K, Savvatis K, Zietsch C, Schmack B, Van Linthout S, Westermann D, Schultheiss HP, Tschöpe C. Role of heart rate reduction in the prevention of experimental heart failure: comparison between if-channel blockade and  $\beta$ -receptor blockade. *Hypertension* 2012; **59**:949–957.

123. Regan JA, Mauro AG, Carbone S, Marchetti C, Gill R, Mezzaroma E, Raleigh JV, Salloum FN, Van Tassel BW, Abbate A, Toldo S, Mauro AG, Carbone S, Marchetti C, Gill R, Mezzaroma E, Valle Raleigh J, Salloum FN, Van Tassel BW, Abbate A, Toldo S. A mouse model of heart failure with preserved ejection fraction due to chronic infusion of a low suppressor dose of angiotensin II. *Am J Physiol Heart Circ Physiol* 2015;**309**:H771–8.
124. Peng H, Yang X-P, Carretero OA, Nakagawa P, D'Ambrosio M, Leung P, Xu J, Peterson EL, González GE, Harding P, Rhaleb N-E. Angiotensin II-induced dilated cardiomyopathy in Balb/c but not C57BL/6j mice. *Exp Physiol* 2011;**96**:756–764.
125. Kadoguchi T, Kinugawa S, Takada S, Fukushima A, Furihata T, Homma T, Masaki Y, Mizushima W, Nishikawa M, Takahashi M, Yokota T, Matsushima S, Okita K, Tsutsui H. Angiotensin II can directly induce mitochondrial dysfunction, decrease oxidative fibre number and induce atrophy in mouse hindlimb skeletal muscle. *Exp Physiol* 2015;**100**:312–322.
126. Chen H, Charlat O, Tartaglia LA, Woolf EA, Weng X, Ellis SJ, Lakey ND, Culpepper J, More KJ, Breitbart RE, Duyk GM, Tepper RI, Morgenstern JP. Evidence that the diabetes gene encodes the leptin receptor: identification of a mutation in the leptin receptor gene in db/db mice. *Cell* 1996;**84**:491–495.
127. Reil J-C, Hohl M, Reil G-H, Granzier HL, Kratz MT, Kazakov A, Fries P, Müller A, Lenski M, Custodis F, Gräber S, Fröhlig G, Steendijk P, Neuberger H-R, Böhm M. Heart rate reduction by If-inhibition improves vascular stiffness and left ventricular systolic and diastolic function in a mouse model of heart failure with preserved ejection fraction. *Eur Heart J* 2013;**34**:2839–2849.
128. Alex L, Russo I, Holoborodko V, Frangogiannis NG. Characterization of a mouse model of obesity-related fibrotic cardiomyopathy that recapitulates features of human heart failure with preserved ejection fraction. *Am J Physiol Heart Circ Physiol* 2018;**315**:H934–H949.
129. Barouch LA, Berkowitz DE, Harrison RW, O'Donnell CP, Hare JM. Disruption of leptin signaling contributes to cardiac hypertrophy independently of body weight in mice. *Circulation* 2003;**108**:754–759.
130. Van den Bergh A, Flameng W, Herijgers P. Type II diabetic mice exhibit contractile dysfunction but maintain cardiac output by favourable loading conditions. *Eur J Heart Fail* 2006;**8**:777–783.
131. Arow M, Waldman M, Yadin D, Nudelman V, Shainberg A, Abraham NG, Freimark D, Kornowski R, Aravot D, Hochhauser E, Arad M. Sodium-glucose cotransporter 2 inhibitor Dapagliflozin attenuates diabetic cardiomyopathy. *Cardiovasc Diabetol* 2020;**19**:7.
132. Van Bilsen M, Daniels A, Brouwers O, Janssen BJAA, Derks WJAA, Brouns AE, Munts C, Schalkwijk CG, Van Der Vusse GJ, Van Nieuwenhoven FA. Hypertension is a conditional factor for the development of cardiac hypertrophy in type 2 diabetic mice. *PLoS One* 2014;**9**:e85078.
133. Sukumaran V, Tsuchimochi H, Tatsumi E, Shirai M, Pearson JT. Azilsartan ameliorates diabetic cardiomyopathy in young db/db mice through the modulation of ACE-2/ANG 1–7/Mas receptor cascade. *Biochem Pharmacol* 2017;**144**:90–99.
134. Habibi J, Aroor AR, Sowers JR, Jia G, Hayden MR, Garro M, Barron B, Mayoux E, Rector RS, Whaley-Connell A, DeMarco VG. Sodium glucose transporter 2 (SGLT2) inhibition with empagliflozin improves cardiac diastolic function in a female rodent model of diabetes. *Cardiovasc Diabetol* 2017;**16**:9.
135. Yue P, Arai T, Terashima M, Sheikh AY, Cao F, Charo D, Hoyt G, Robbins RC, Ashley EA, Wu J, Yang PC, Tsao PS. Magnetic resonance imaging of progressive cardiomyopathic changes in the db/db mouse. *Am J Physiol Heart Circ Physiol* 2007;**292**:H2106–H2118.
136. Regitz-Zagrosek V, Brokat S, Tschöpe C. Role of gender in heart failure with normal left ventricular ejection fraction. *Prog Cardiovasc Dis* 2007;**49**:241–251.
137. Hutchinson KR, Lord CK, West TA, Stewart JA. Jr., Cardiac fibroblast-dependent extracellular matrix accumulation is associated with diastolic stiffness in type 2 diabetes. *PLoS One* 2013;**8**:e72080.
138. Bostick B, Habibi J, Ma L, Aroor A, Rehmer N, Hayden MR, Sowers JR. Dipeptidyl peptidase inhibition prevents diastolic dysfunction and reduces myocardial fibrosis in a mouse model of Western diet induced obesity. *Metabolism* 2014;**63**:1000–1011.
139. Gutkowska J, Broderick TL, Bogdan D, Wang D, Lavoie JM, Jankowski M. Downregulation of oxytocin and natriuretic peptides in diabetes: Possible implications in cardiomyopathy. *J Physiol* 2009;**587**:4725–4736.
140. Broderick TL, Jankowski M, Wang D, Danalache BA, Parrott CR, Gutkowska J. Downregulation of GATA4 and downstream structural and contractile genes in the db/db mouse heart. *ISRN Endocrinol* 2012;**2012**:1–12.
141. Lindström P. The physiology of obese-hyperglycemic mice [ob/ob mice]. *ScientificWorldJournal* 2007;**7**:666–685.
142. Ingalls AM, Dickie MM, Snell GD. Obese, a new mutation in the house mouse. *J Hered* 1950;**41**:317–318.
143. Christoffersen C, Bollano E, Lindegaard MLS, Bartels ED, Goetze JP, Andersen CB, Nielsen LB. Cardiac lipid accumulation associated with diastolic dysfunction in obese mice. *Endocrinology* 2003;**144**:3483–3490.
144. Broderick TL, Wang D, Jankowski M, Gutkowska J. Unexpected effects of voluntary exercise training on natriuretic peptide and receptor mRNA expression in the ob/ob mouse heart. *Regul Pept* 2014;**188**:52–59.
145. Christoffersen C, Bartels ED, Nielsen LB. Heart specific up-regulation of genes for B-type and C-type natriuretic peptide receptors in diabetic mice. *Eur J Clin Invest* 2006;**36**:69–75.
146. Manolescu DC, Jankowski M, Danalache BA, Wang D, Broderick TL, Chiasson JL, Gutkowska J. All-trans retinoic acid stimulates gene expression of the cardio-protective natriuretic peptide system and prevents fibrosis and apoptosis in cardiomyocytes of obese ob/ob mice. *Appl Physiol Nutr Metab* 2014;**39**:1127–1136.
147. Dong F, Zhang X, Yang X, Esberg LB, Yang H, Zhang Z, Culver B, Ren J. Impaired cardiac contractile function in ventricular myocytes from leptin-deficient ob/ob obese mice. *J Endocrinol* 2006;**188**:25–36.
148. Clément K. Genetics of human obesity. *C R Biol* 2006;**329**:608–622.
149. Sorimachi H, Obokata M, Takahashi N, Reddy YNV, Jain CC, Verbrugge FH, Koepf KE, Khosla S, Jensen MD, Borlaug BA. Pathophysiologic importance of visceral adipose tissue in women with heart failure and preserved ejection fraction. *Eur Heart J* 2021;**42**:1595–1605.
150. Withaar C, Meems LMG, De Boer RA. Fighting HFpEF in women: taking aim at belly fat. *Eur Heart J* 2021;**42**:1606–1608.
151. Piek A, Koonen DYYY, Schouten E-MM, Lindstedt EL, Michaëlsson E, de Boer RA, Silljé HHWW. Pharmacological myeloperoxidase (MPO) inhibition in an obese/hypertensive mouse model attenuates obesity and liver damage, but not cardiac remodeling. *Sci Rep* 2019;**9**:18765.
152. Ternacle J, Wan F, Sawaki D, Surenaud M, Pini M, Mercedes R, Ernande L, Audureau E, Dubois-Randé JL, Adnot S, Hue S, Czibik G, Derumeaux G. Short-term high-fat diet compromises myocardial function: a radial strain rate imaging study. *Eur Heart J Cardiovasc Imaging* 2017;**18**:1283–1291.
153. Bostick B, Aroor AR, Habibi J, Durante W, Ma L, DeMarco VG, Garro M, Hayden MR, Booth FW, Sowers JR. Daily exercise prevents diastolic dysfunction and oxidative stress in a female mouse model of western diet induced obesity by maintaining cardiac heme oxygenase-1 levels. *Metabolism* 2017;**66**:14–22.
154. Aroor AR, Habibi J, Kandikattu HK, Garro-Kacher M, Barron B, Chen D, Hayden MR, Whaley-Connell A, Bender SB, Klein T, Padilla J, Sowers JR, Chandrasekar B, DeMarco VG. Dipeptidyl peptidase-4 (DPP-4) inhibition with linagliptin reduces western diet-induced myocardial TRAF3IP2 expression, inflammation and fibrosis in female mice. *Cardiovasc Diabetol* 2017;**16**:61.
155. Luptak I, Sverdlov AL, Panagia M, Qin F, Pimentel DR, Croteau D, Siwik DA, Ingwall JS, Bachschmid MM, Balschi JA, Colucci WS. Decreased ATP production and myocardial contractile reserve in metabolic heart disease. *J Mol Cell Cardiol* 2018;**116**:106–114.
156. Agrawal V, Fortune N, Yu S, Fuentes J, Shi F, Nichols D, Gleaves L, Poovey E, Wang TJ, Brittain EL, Collins S, West JD, Hemmes AR. Natriuretic peptide receptor C contributes to disproportionate right ventricular hypertrophy in a rodent model of obesity-induced heart failure with preserved ejection fraction with pulmonary hypertension. *Pulm Circ* 2019;**9**:204589401987859.
157. Aboumsallem JP, Muthuramu I, Mishra M, De Geest B. Cholesterol-lowering gene therapy prevents heart failure with preserved ejection fraction in obese type 2 diabetic mice. *Int J Mol Sci* 2019;**20**:2222.
158. Carbone S, Mauro AG, Mezzaroma E, Kraskauskas D, Marchetti C, Buzzetti R, Van Tassel BW, Abbate A, Toldo S. A high-sugar and high-fat diet impairs cardiac systolic and diastolic function in mice. *Int J Cardiol* 2015;**198**:66–69.
159. Gevaert AB, Shakeri H, Leloup AJ, Van Hove CE, De Meyer GRV, Vrints CJ, Lemmens K, Van Craenenbroeck EM. Endothelial senescence contributes to heart failure with preserved ejection fraction in an aging mouse model. *Circ Heart Fail* 2017;**10**:e003806.
160. Zhong J, Gupta GK, Chiurchiù V, Ahnstedt H, Ahnstedt H, Reilly R-O, Ms S, As M, Bravo-Alegria J, Chauhan A, Aronowski J, Sp M, Ld M, Roy-O'reilly M, Spychala MS, Mobley AS, Bravo-Alegria J, Chauhan A, Aronowski J, Marrelli SP, McCullough LD. Sex differences in adipose tissue CD8+ T cells and regulatory T cells in middle-aged mice. *Front Immunol* 2018;**9**:659.
161. van der Heijden RA, Bijzet J, Meijers WC, Yakala GK, Kleemann R, Nguyen TQ, de Boer RA, Schalkwijk CG, Hazenberg BPC, Tietge UJF, Heeringa P. Obesity-induced chronic inflammation in high fat diet challenged C57BL/6j mice is associated with acceleration of age-dependent renal amyloidosis. *Sci Rep* 2015;**5**:16474.
162. Ingvorsen C, Karp NA, Lelliott CJ. The role of sex and body weight on the metabolic effects of high-fat diet in C57BL/6N mice. *Nutr Diabetes* 2017;**7**:e261.
163. Manrique C, DeMarco VG, Aroor AR, Mugerfeld I, Garro M, Habibi J, Hayden MR, Sowers JR. Obesity and insulin resistance induce early development of diastolic dysfunction in young female mice fed a western diet. *Endocrinology* 2013;**154**:3632–3642.
164. Meng Q, Lai YC, Kelly NJ, Bueno M, Baust JJ, Bachman TN, Goncharov D, Vanderpool RR, Radder JE, Hu J, Goncharova E, Morris AM, Mora AL, Shapiro SD, Gladwin MT. Development of a mouse model of metabolic syndrome,

- pulmonary hypertension, and heart failure with preserved ejection fraction. *Am J Respir Cell Mol Biol* 2017;**56**:497–505.
165. Cannon MV, Silljé HHV, Sijbesma JVA, Khan MAF, Steffensen KR, van Gilst WH, de Boer RA. LXR $\alpha$  improves myocardial glucose tolerance and reduces cardiac hypertrophy in a mouse model of obesity-induced type 2 diabetes. *Diabetologia* 2016;**59**:634–643.
  166. Bartels ED, Nielsen JM, Bisgaard LS, Goetze JP, Nielsen LB. Decreased expression of natriuretic peptides associated with lipid accumulation in cardiac ventricle of obese mice. *Endocrinology* 2010;**151**:5218–5225.
  167. Bruder-Nascimento T, Ekeledo OJ, Anderson R, Le HB, Belin de Chantemele EJ. Long Term High Fat Diet Treatment: an Appropriate Approach to Study the Sex-Specificity of the Autonomic and Cardiovascular Responses to Obesity in Mice. *Front Physiol* 2017;**8**.
  168. Gai Z, Hiller C, Chin SH, Hofstetter L, Stieger B, Konrad D, Kullak-Ublick GA. Uninephrectomy augments the effects of high fat diet induced obesity on gene expression in mouse kidney. *Biochim Biophys Acta Mol Basis Dis* 2014;**1842**:1870–1878.
  169. Schiattarella GG, Altamirano F, Tong D, French KM, Villalobos E, Kim SY, Luo X, Jiang N, May HI, Wang ZV, Hill TM, Mammen PPA, Huang J, Lee DI, Hahn VS, Sharma K, Kass DA, Lavandro S, Gillette TG, Hill JA. Nitrosative stress drives heart failure with preserved ejection fraction. *Nature* 2019;**568**:351–356.
  170. Aurich AC, Niemann B, Pan R, Gruenler S, Issa H, Silber RE, Rohrbach S. Age-dependent effects of high fat-diet on murine left ventricles: role of palmitate. *Basic Res Cardiol* 2013;**108**:1–17.
  171. Roh JD, Houstis N, Yu A, Chang B, Yeri A, Li H, Hobson R, Lerchenmüller C, Vujic A, Chaudhari V, Damilano F, Platt C, Zlotoff D, Lee RT, Shah R, Jerosch-Herold M, Rosenzweig A. Exercise training reverses cardiac aging phenotypes associated with heart failure with preserved ejection fraction in male mice. *Aging Cell* 2020;**19**:e13159.
  172. Roh JD, Hobson R, Chaudhari V, Quintero P, Yeri A, Benson M, Xiao C, Zlotoff D, Bezzerides V, Houstis N, Platt C, Damilano F, Lindman BR, Elmariyah S, Biersmith M, Lee SJ, Seidman CE, Seidman JG, Gerszten RE, Lach-Trifileff E, Glass DJ, Rosenzweig A. Activin type II receptor signaling in cardiac aging and heart failure. *Sci Transl Med* 2019;**11**:8680.
  173. Takeda T, Hosokawa M, Higuchi K. Senescence-accelerated mouse (SAM): a novel murine model of senescence. *Exp Gerontol* 1997;**32**:105–109.
  174. Karuppagounder V, Arumugam S, Babu SS, Palaniyandi SS, Watanabe K, Cooke JP, Thandavarayan RA. The senescence accelerated mouse prone 8 (SAMP8): a novel murine model for cardiac aging. *Ageing Res Rev* 2017;**35**:291–296.
  175. Tanisawa K, Mikami E, Fuku N, Honda Y, Honda S, Ohsawa I, Ito M, Endo S, Ihara K, Ohno K, Kishimoto Y, Ishigami A, Maruyama N, Sawabe M, Iseki H, Okazaki Y, Hasegawa-Ishii S, Takei S, Shimada A, Hosokawa M, Mori M, Higuchi K, Takeda T, Higuchi M, Tanaka M. Exome sequencing of senescence-accelerated mice (SAM) reveals deleterious mutations in degenerative disease-causing genes. *BMC Genomics* 2013;**14**:248.
  176. Reed AL, Tanaka A, Sorescu D, Liu H, Jeong E-M, Sturdy M, Walp ER, Dudley SC, Sutliff RL. Diastolic dysfunction is associated with cardiac fibrosis in the senescence-accelerated mouse. *Am J Physiol Circ Physiol* 2011;**301**:H824–H831.
  177. Willard PW. A model for evaluation of thiazide-induced hypotension. *J Pharm Pharmacol* 1969;**21**:406–408.
  178. Lovelock JD, Monasky MM, Jeong EM, Lardin HA, Liu H, Patel BG, Taglieri DM, Gu L, Kumar P, Pokhrel N, Zeng D, Belardinelli L, Sorescu D, Solaro RJ, Dudley SC. Ranolazine improves cardiac diastolic dysfunction through modulation of myofilament calcium sensitivity. *Circ Res* 2012;**110**:841–850.
  179. Mohammed SF, Ohtani T, Korinek J, Lam CSP, Larsen K, Simari RD, Valencik ML, Burnett JC, Redfield MM. Mineralocorticoid accelerates transition to heart failure with preserved ejection fraction via “nongenomic effects”. *Circulation* 2010;**122**:370–378.
  180. Jeong EM, Monasky MM, Gu L, Taglieri DM, Patel BG, Liu H, Wang Q, Greener I, Dudley SC, Solaro RJ. Tetrahydrobiopterin improves diastolic dysfunction by reversing changes in myofilament properties. *J Mol Cell Cardiol* 2013;**56**:44–54.
  181. Bowen TS, Eisenkolb S, Drobner J, Fischer T, Werner S, Linke A, Mangner N, Schuler G, Adams V. High-intensity interval training prevents oxidant-mediated diaphragm muscle weakness in hypertensive mice. *FASEB J* 2017;**31**:60–71.
  182. ter Maaten JM, Damman K, Verhaar MC, Paulus WJ, Duncker DJ, Cheng C, van Heerebeek L, Hillege HL, Lam CSPP, Navis G, Voors AA. Connecting heart failure with preserved ejection fraction and renal dysfunction: the role of endothelial dysfunction and inflammation. *Eur J Heart Fail* 2016;**18**:588–598.
  183. Tanaka K, Valero-Muñoz M, Wilson RM, Essick EE, Fowler CT, Nakamura K, van den Hoff M, Ouchi N, Sam F. Follistatin-like 1 regulates hypertrophy in heart failure with preserved ejection fraction. *JACC Basic Transl Sci* 2016;**1**:207–221.
  184. Valero-Munoz M, Li S, Wilson RM, Hulsmans M, Aprahamian T, Fuster JJ, Nahrendorf M, Scherer PE, Sam F. Heart failure with preserved ejection fraction induces Beiging in adipose tissue. *Circ Heart Fail* 2016;**9**:e002724.
  185. Valero-Munoz M, Li S, Wilson RM, Boldbaatar B, Iglarz M, Sam F. Dual endothelin-A/endothelin-B receptor blockade and cardiac remodeling in heart failure with preserved ejection fraction. *Circ Heart Fail* 2016;**9**:e003381.
  186. Wilson RM, De Silva DS, Sato K, Izumiya Y, Sam F. Effects of fixed-dose isosorbide dinitrate/hydralazine on diastolic function and exercise capacity in hypertension-induced diastolic heart failure. *Hypertension* 2009;**54**:583–590.
  187. Garcia AG, Wilson RM, Heo J, Murthy NR, Baid S, Ouchi N, Sam F. Interferon- $\gamma$  ablation exacerbates myocardial hypertrophy in diastolic heart failure. *Am J Physiol Heart Circ Physiol* 2012;**303**:H587–H596.
  188. Tanaka K, Wilson RM, Essick EE, Duffen JL, Scherer PE, Ouchi N, Sam F. Effects of adiponectin on calcium-handling proteins in heart failure with preserved ejection fraction. *Circ Heart Fail* 2014;**7**:976–985.
  189. Tong D, Schiattarella GG, Jiang N, May HI, Lavandro S, Gillette TG, Hill JA. Female sex is protective in a preclinical model of heart failure with preserved ejection fraction. *Circulation* 2019;**140**:1769–1771.
  190. Reddy SS, Agarwal H, Barthwal MK. Cilostazol ameliorates heart failure with preserved ejection fraction and diastolic dysfunction in obese and non-obese hypertensive mice. *J Mol Cell Cardiol* 2018;**123**:46–57.
  191. Gaspari T, Brdar M, Lee HW, Spizzo I, Hu Y, Widdop RE, Simpson RW, Dear AE. Molecular and cellular mechanisms of glucagon-like peptide-1 receptor agonist-mediated attenuation of cardiac fibrosis. *Diabetes Vasc Dis Res* 2016;**13**:56–68.
  192. Du W, Piek A, Marloes Schouten E, van de Kolk CWA, Mueller C, Mebazaa A, Voors AA, de Boer RA, Silljé HHV. Plasma levels of heart failure biomarkers are primarily a reflection of extracardiac production. *Theranostics* 2018;**8**:4155–4169.
  193. Withaar C, Meems LMG, Markousis-Mavrogenis G, Boogerd CJ, Silljé HHV, Schouten EM, Dokter MM, Voors AA, Westenbrink BD, Lam CSP, de Boer RA. The effects of liraglutide and dapagliflozin on cardiac function and structure in a multi-hit mouse model of heart failure with preserved ejection fraction. *Cardiovasc Res* 2020;cvaa256. doi: 10.1093/cvr/cvaa256 [Epub ahead of print].
  194. Deng Y, Xie M, Li Q, Xu X, Ou W, Zhang Y, Xiao H, Yu H, Zheng Y, Liang Y, Jiang C, Chen G, Du D, Zheng W, Wang S, Gong M, Chen Y, Tian R, Li T. Targeting mitochondria-inflammation circuit by  $\beta$ -hydroxybutyrate mitigates HFpEF. *Circ Res* 2021;**128**:232–245.