

# The Complete Genome Sequence of the San Miguel Sea Lion Virus-8 Reveals that It Is Not a Member of the Vesicular Exanthema of Swine Virus/San Miguel Sea Lion Virus Species of the *Caliciviridae*

John D. Neill

Ruminant Diseases and Immunology Research Unit, National Animal Disease Center, Department of Agriculture, ARS, Ames, Iowa, USA

**The complete genome sequence of the San Miguel sea lion virus-8 (SMSV-8) was determined in this study. A comparison of this sequence to other calicivirus sequences in GenBank showed that this virus is genetically distinct from the vesicular exanthema of swine virus/San Miguel sea lion virus (VESV/SMSV) strains and belongs to a novel clade within the *Vesivirus* genus.**

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Address correspondence to john.neill@ars.usda.gov.

The *Caliciviridae* is composed of five recognized genera, *Norovirus*, *Sapovirus*, *Lagovirus*, *Vesivirus*, and *Nebovirus*. The *Vesivirus* genus contains viruses that primarily infect animals only, such as feline calicivirus, vesicular exanthema of swine virus (VESV), San Miguel sea lion virus (SMSV), and viruses that have not been assigned to a specific group. The first isolated SMSV type, SMSV-1, was reported in 1972 from vesicular lesions on the flipper of a California sea lion (1). The virus was shown to have biophysical properties in common with vesicular exanthema of swine viruses (2). The isolation of SMSV-8 was reported in 1981 from vesicular lesions on the flippers of northern fur seals and possessed the typical calicivirus morphology (3).

SMSV-8 was propagated on Vero cells and supernatant stocks stored at -80°C. To prepare for sequencing, viral RNA was purified following the treatment of the virus stock with a nuclease cocktail (4). The genomic RNA was sequenced using a random-primed single-primer method for the synthesis of barcoded double-stranded cDNA (5). Briefly, a single-tube reverse transcriptase followed by two second-strand cDNA synthesis reactions was performed using 20-mer primers of known sequence with 8 random nucleotides on the 3' ends. The cDNA was amplified using primers with the same 20-mer sequence. The amplified cDNA was prepared for sequencing using the Ion Torrent sequencing platform (Life Technologies, Inc., Grand Island, NY). The viral genome was assembled with SeqMan NGen software (DNASTar, Inc., Madison, WI) using *de novo* assembly of sequences and alignment to mink calicivirus strain MCV-DL/2007/CN (GenBank accession no. JX847605). The 3' terminal sequence was confirmed by 3' RACE.

The SMSV-8 library yielded 36,783 (34,978 virus) sequences. This number of sequences provided an average coverage over the genome of 489×. The genome length of SMSV-8 is 8,477 bases, excluding the poly(A) tail. The genomic RNA contains 3 open reading frames (ORFs), the first encoding the nonstructural polyprotein, the second encoding the capsid protein precursor (VP-1), and the third encoding a small basic protein (VP-2). A phylogenetic analysis of the genome se-

quence showed that the most closely related caliciviruses to SMSV-8 are members of the *Vesivirus* genus. The virus with the greatest genetic relatedness to SMSV-8 is mink calicivirus (MCV) (GenBank accession no. JX847605), at 71% identity. When the amino acid sequences of the three ORFs were compared to those of other vesiviruses, SMSV-8 was found to be distantly related. The amino acid sequences of ORF1, ORF2, and ORF3 have 73%, 63%, and 48% identity, respectively, with those of MCV. With respect to other caliciviruses, SMSV-8 has 55%, 39%, and 34% identity with the three ORFs of SMSV-1 (Genbank accession no. U15301), 55%, 38%, and 35% identity to VESV A48 (Genbank accession no. U76874), and 61%, 46%, and 48% identity to calicivirus Allston 2008/US (Genbank accession no. GQ475302). These data indicate that SMSV-8 is a novel member of the *Vesivirus* genus. This supports Seal et al. (6) and Reid et al. (7), who suggested that SMSV-8 belongs to a separate calicivirus group based on a lack of reactivity using immunologic and genetic screening methods.

**Nucleotide sequence accession number.** The genomic sequence of San Miguel sea lion virus-8 has been deposited in GenBank with the accession no. [KM244552](https://www.ncbi.nlm.nih.gov/nuccore/KM244552).

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## REFERENCES

- Smith AW, Akers TG, Madin SH, Vedros NA. 1973. San Miguel sea lion virus isolation, preliminary characterization and relationship to vesicular exanthema of swine virus. *Nature* 244:108–110. <http://dx.doi.org/10.1038/244108a0>.
- Schaffer FL, Soergel ME. 1973. Biochemical and biophysical characterization of calicivirus isolates from pinnipeds. *Intervirology* 1:210–219. <http://dx.doi.org/10.1159/000148848>.
- Smith AW, Skilling DE, Latham AB. 1981. Isolation and identification of five new serotypes of calicivirus from marine mammals. *Am. J. Vet. Res.* 42:693–694.

4. Victoria JG, Kapoor A, Dupuis K, Schnurr DP, Delwart EL. 2008. Rapid identification of known and new RNA viruses from animal tissues. *PLoS Pathog.* 4:e1000163. <http://dx.doi.org/10.1371/journal.ppat.1000163>.
5. Neill JD, Bayles DO, Ridpath JF. 2014. Simultaneous rapid sequencing of multiple RNA virus genomes. *J. Virol. Methods* 201:68–72. <http://dx.doi.org/10.1016/j.jviromet.2014.02.016>.
6. Seal BS, House JA, Whetstone CA, Neill JD. 1995. Analysis of the serologic relationship among San Miguel sea lion virus and vesicular exanthema of swine virus isolates. Application of the western blot assay for detection of antibodies in swine sera to these virus types. *J. Vet. Diagn. Invest.* 7:190–195. <http://dx.doi.org/10.1177/104063879500700204>.
7. Reid SM, Ansell DM, Ferris NP, Hutchings GH, Knowles NJ, Smith AW. 1999. Development of a reverse transcription polymerase chain reaction procedure for the detection of marine caliciviruses with potential application for nucleotide sequencing. *J. Virol. Methods* 82:99–107. [http://dx.doi.org/10.1016/S0166-0934\(99\)00088-9](http://dx.doi.org/10.1016/S0166-0934(99)00088-9).