

# Native Chondrocyte Viability during Cartilage Lesion Progression: Normal to Surface Fibrillation

Cartilage  
1(4) 306–311  
© The Author(s) 2010  
Reprints and permission:  
sagepub.com/journalsPermissions.nav  
DOI: 10.1177/1947603510373918  
http://cart.sagepub.com  


Kumkum Ganguly<sup>1</sup>, Ian D. McRury<sup>2</sup>, Peter M. Goodwin<sup>3</sup>, Roy E. Morgan<sup>2</sup>, and Wayne K. Augé II<sup>2,4</sup>

## Abstract

**Objective:** Early surgical intervention for articular cartilage disease is desirable before full-thickness lesions develop. As early intervention treatments are designed, native chondrocyte viability at the treatment site before intervention becomes an important parameter to consider. The purpose of this study is to evaluate native chondrocyte viability in a series of specimens demonstrating the progression of articular cartilage lesions to determine if the chondrocyte viability profile changes during the evolution of articular cartilage disease to the level of surface fibrillation. **Design:** Osteochondral specimens demonstrating various degrees of articular cartilage damage were obtained from patients undergoing knee total joint replacement. Three groups were created within a patient harvest based on visual and tactile cues commonly encountered during surgical intervention: group 1, visually and tactilely intact surfaces; group 2, visually intact, tactilely soft surfaces; and group 3, surface fibrillation. Confocal laser microscopy was performed following live/dead cell viability staining. **Results:** Groups 1 to 3 demonstrated viable chondrocytes in all specimens, even within the fibrillated portions of articular cartilage, with little to no evidence of dead chondrocytes. Chondrocyte viability profile in articular cartilage does not appear to change as disease lesion progresses from normal to surface fibrillation. **Conclusions:** Fibrillated partial-thickness articular cartilage lesions are a good therapeutic target for early intervention. These lesions retain a high profile of viable chondrocytes and are readily diagnosed by visual and tactile cues during surgery. Early intervention should be based on matrix failure rather than on more aggressive procedures that further corrupt the matrix and contribute to chondrocyte necrosis of contiguous untargeted cartilage.

## Keywords

cartilage, chondrocyte, viability, fibrillated, partial thickness

## Introduction

Articular cartilage disease defines a large disease burden afflicting our population.<sup>1–5</sup> Significant efforts continue in developing and tracking treatment solutions for articular cartilage lesions at various stages of their progression.<sup>6</sup> A considerable interest exists for effective surgical interventions that address an earlier stage of disease progression rather than waiting for full-thickness articular cartilage lesions to develop. Early surgical intervention is appealing for an aging population to mitigate downstream disease burden associated with full-thickness defects. Surgical treatment of fibrillated partial-thickness articular cartilage lesions remains an important category because these lesions can be readily diagnosed and characterized by visual and tactile cues during surgery and hence remain an attractive therapeutic target for early surgical intervention modalities.

Surgical treatment for fibrillated partial-thickness lesions has been traditionally limited to debridement chondroplasty techniques developed to remove the damaged articular cartilage that causes a mechanical and inflammatory impairment of joint function and leads to a deterioration of joint health. Smoothing the articular surface can eliminate the mechanical

<sup>1</sup>B-Division, Los Alamos National Laboratory, Los Alamos, NM, USA

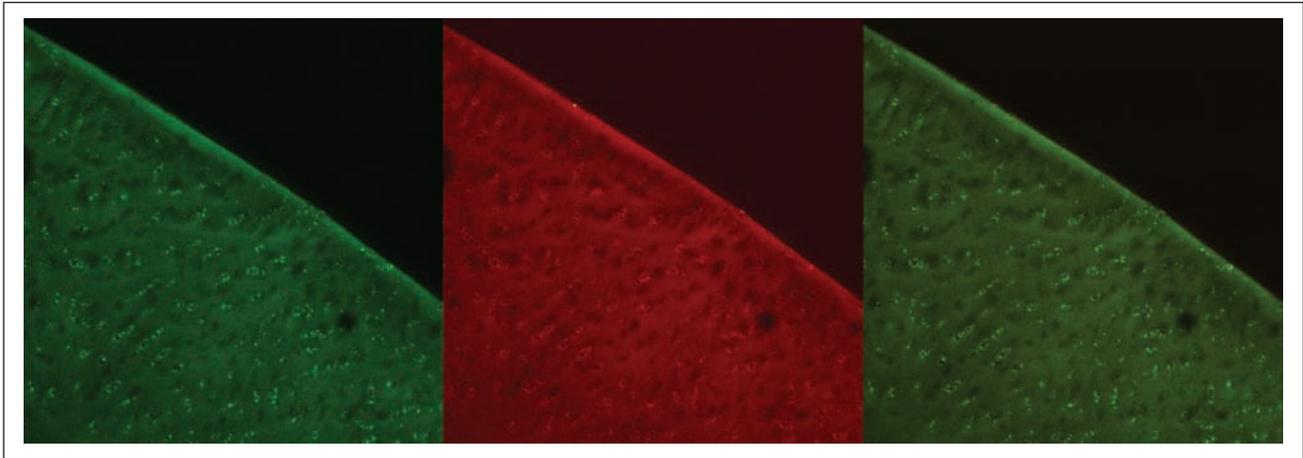
<sup>2</sup>NuOrtho Surgical, Inc., Fall River, MA, USA

<sup>3</sup>Center for Integrated Nanotechnologies, Los Alamos National Laboratory, Los Alamos, NM, USA

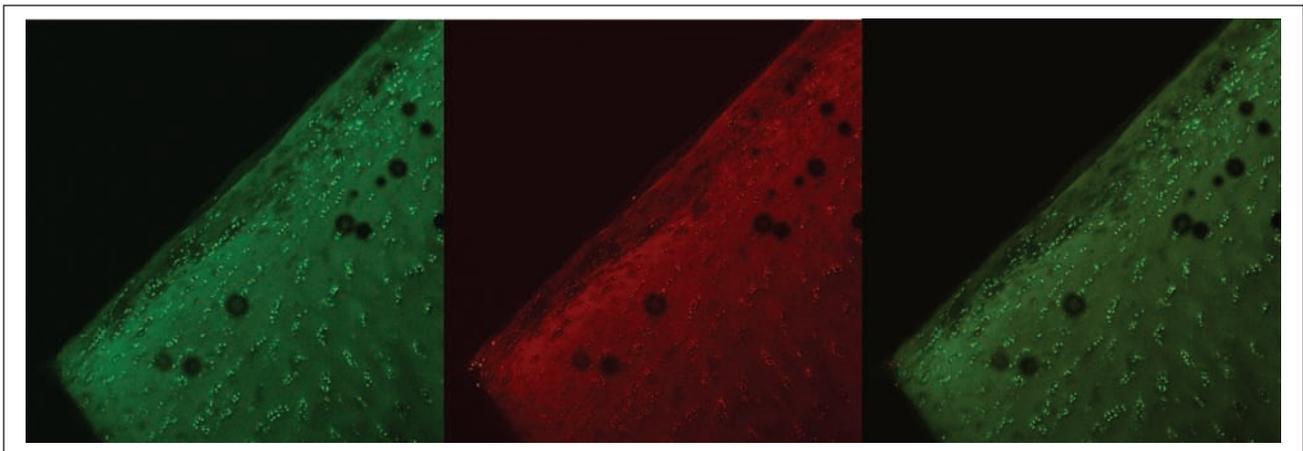
<sup>4</sup>Center for Orthopaedic and Sports Performance Research, Inc., Santa Fe, NM, USA

### Corresponding Author:

Wayne K. Augé II, MD, Center for Orthopaedic and Sports Performance Research, Inc., 936 Vista Jemez Court, Santa Fe, NM 87505, USA  
Email: infocospr@aol.com; waug@nuorthosurgical.com



**Figure 1.** Confocal laser microscopy images, group 1. Representative images depicting live cell stain (green), dead cell stain (red), and a combined image with both live cell and dead cell stain. Original magnification 10x.



**Figure 2.** Confocal laser microscopy images, group 2. Representative images depicting live cell stain (green), dead cell stain (red), and a combined image with both live cell and dead cell stain. Original magnification 10x.

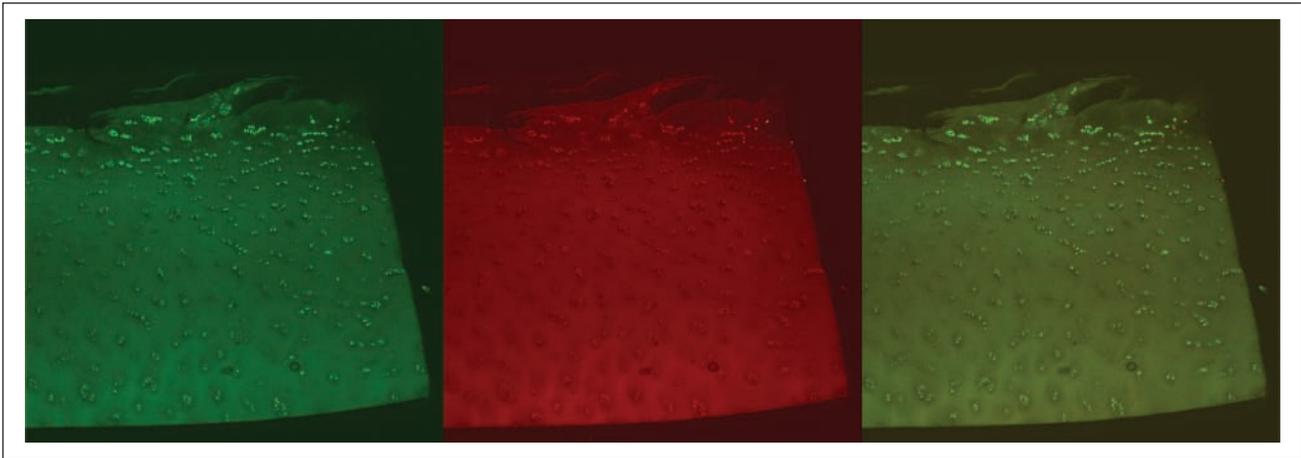
stress risers that cause symptoms and propagate cartilage damage; removal of the loose surface debris associated with loss of cartilage function decreases the biologic load the joint needs to address.<sup>7-11</sup> Although an attractive surgical approach, current debridement chondroplasty techniques are imprecise, are aggressive, and induce necrosis and collateral damage to contiguous untargeted cartilage tissue at the treatment site.<sup>12-23</sup> Accordingly, widespread adoption of current debridement chondroplasty techniques as an early surgical intervention modality to treat fibrillated partial-thickness lesions has not emerged due to the fear of contributing to disease progression resulting from an attempt to provide disease burden relief.

Because it is difficult to imagine an early surgical intervention for articular cartilage that does not include *in situ* removal of damaged tissue present at the lesion locale, researchers have sought to create more targeted interventions that are based on lesion progression. As early surgical intervention techniques become more precise and allow tissue preservation of articular cartilage, native chondrocyte viability at the treatment site

before intervention becomes an important parameter to consider. The purpose of this study is to evaluate native chondrocyte viability in a series of specimens demonstrating the progression of articular cartilage lesions to determine if the chondrocyte viability profile changes during the evolution of articular cartilage disease to the level of surface fibrillation.

## Materials and Methods

Osteochondral specimens were harvested from patients undergoing total knee replacement under an approved Institutional Review Board protocol. The total knee replacement procedures were performed by a single surgeon in the normal course of his practice. The tissue to be normally discarded during the procedure was examined prior to harvest once the knee joint was entered surgically to determine if it met the requirements for study inclusion. Specimens were included that demonstrated an area of uniform normal or partial-thickness damage of sufficient size to obtain test samples wherein harvest margin



**Figure 3.** Confocal laser microscopy images, group 3. Representative images depicting live cell stain (green), dead cell stain (red), and a combined image with both live cell and dead cell stain. Original magnification 10x.

artifact would not be a confounding variable.<sup>24</sup> Three groups were created within each patient tissue harvest based on visual and tactile characteristics of the specimens as customarily assessed during surgery: group 1 included visually and tactilely normal cartilage surfaces, group 2 included visually normal but tactilely soft (as judged by indentation pressure) cartilage surfaces, and group 3 included surface fibrillation of the cartilage surfaces.

Immediately after harvest, three 0.5-mm coronal sections of each cartilage sample were obtained referencing the center of the sample after removal of subchondral bone. The sections were prepared for staining by washing in HEPES buffered saline solution. Live/Dead<sup>®</sup> Reduced Biohazard Cell Viability Kit 1 “green and red fluorescence,” SKU #L-7013 (Invitrogen, Carlsbad, California), was used per the manufacturer’s specification to stain specimens. Specimens were glutaraldehyde fixed, transferred to standard flat glass slides, and flooded with VectaShield<sup>®</sup> fluorescence protection oil prior to the placement of #1.5 borosilicate glass coverslips over each specimen section.

The cartilage tissue and the articular surface were assessed by confocal fluorescence laser microscopy analysis performed by personnel blinded to the identity of the samples. Confocal imaging was performed with an Olympus IX-81 inverted microscope coupled to an Olympus FV300 confocal laser scanning unit (Center Valley, Pennsylvania) using 488-nm laser excitation. Live chondrocytes were captured under green fluorescent channel (505-525 nm), and dead chondrocytes were captured under red fluorescent channel (577-634 nm), generating a live image, a dead image, and an integrated image.

## Results

Six separate osteochondral specimens originating from femoral condyle resection were included for study ( $n = 6$ ), with 2 specimens per group. **Figures 1 to 3** demonstrate representative images of the specimens. Interspecimen comparisons did

not reveal significant differences in relative chondrocyte population densities, chondron orientation, or cellular distribution patterns. Comparison of specimens within a patient harvest indicated a distinct progression of lesion from normal to fibrillated.

Group 1 specimens (**Figure 1**) demonstrated intact and normal articular surfaces consistent with gross visual and tactile inspection of the harvested tissues. The superficial, transitional, and deep zones remained structurally intact with chondrocytes and chondron appearances typical of normal articular cartilage. Live chondrocytes were observed residing throughout the tissue abundantly and present in zonal density patterns typical of healthy cartilage. Dead chondrocytes were not observed within the substance of the cartilage tissue. The lamina splendens region appeared congruous without evidence of disruption.

Group 2 specimens (**Figure 2**) demonstrated intact but softened articular surfaces consistent with gross visual and tactile inspection of the harvested tissues. The superficial, transitional, and deep zones remained structurally intact, but the superficial zone demonstrated areas with loss of surface cellularity and lacunar emptying within the superficial zone deep to the lamina splendens. Live chondrocytes were abundantly present in zonal density patterns typical of healthy cartilage around and below the surface changes. Dead chondrocytes were not observed within the substance of the cartilage tissue. Adjacent to the softened segments, a more typical normal appearance was observed without softening, loss of surface cellularity, or lacunar emptying, as noted in group 1 specimens indicating lesion transition.

Group 3 specimens (**Figure 3**) demonstrated typical fibrillated articular surfaces consistent with gross visual and tactile inspection of the harvested tissues. The superficial zone was clearly disrupted by the fibrillation, but chondrocytes with a flattened appearance typical of this zone remained present even toward the base of the fibrillation. The fibrillation did not penetrate deep to the transitional zone of the articular

cartilage tissue in any specimen. Within the fibrillated tissue itself, live chondrocytes were observed residing within the tissue at varying distances from the surface of the fibrillation. Live chondrocytes were abundantly present in zonal density patterns typical of healthy cartilage around and below the surface fibrillation. Occasional dead chondrocytes were observed to reside in a more extruded position at the margins of the fibrillation itself but not within the substance of the morphologically intact cartilage tissue. Adjacent to the fibrillated segments, a more typical softened appearance was observed without fibrillation but with loss of surface cellularity and lacunar emptying within the superficial zone deep to the lamina splendens, as noted in group 2 specimens indicating lesion transition.

## Discussion

This study explored chondrocyte viability in tissue specimens demonstrating various levels of articular cartilage lesion progression in patients undergoing joint replacement surgery. Although a small sample size, the results indicate that articular cartilage retains a very high percentage of native viable chondrocytes even when lesion progression has led to matrix failure and subsequent surface fibrillation within the same patient. Because of the predominant chondrocyte viability noted, surface fibrillation appears to be an important stage to consider for early surgical intervention—a stage that can be readily diagnosed by visual and tactile cues during surgery.

Matrix failure–based early intervention for articular cartilage disease should be considered a therapeutic target because preserving functioning cartilage tissue is important. Injury and loss of the superficial zone has been strongly associated with the progression of cartilage disease,<sup>25-29</sup> and treatments that contribute to superficial zone injury accelerate the natural progression and disease burden of osteoarthritis.<sup>25,29-33</sup> Once articular cartilage lesions have progressed to matrix failure leading to surface fibrillation, those chondrocytes within the fibrillation, although noted to be viable in this study, may be not be useful to retain considering that fibrillation matrix stress risers can cause additional chondrocytes to become extruded as untreated fibrillation lesions propagate in response to additional loading.<sup>34-41</sup> Stabilizing superficial zone lamina flaps, cleavage planes, and the peeling or breaking of denatured collagen fibrils can be a reasonable matrix failure–based therapeutic target. From an early surgical intervention perspective, precisely removing this fibrillated tissue as a means to stabilize lesions is preferable, as long as the underlying intact chondrons are not injured, because it has been noted that spatial reorganization and proliferation of superficial zone chondrocytes occur in response to distant partial-thickness lesions and may serve as a mechanism to recruit metabolically active units to address focal disease.<sup>42</sup>

Although many studies have supported the efficacy of debridement chondroplasty in relieving patient symptoms by smoothing the articular surface and decreasing the biologic

load of joint cartilage debris,<sup>7-11</sup> these benefits have yet to be shown to contribute to long-term joint health, to promote cartilage longevity, or to mitigate joint replacement surgery. This may be due to the nature of current interventions that are excessively damaging to these lesions and notably contribute to disease progression.<sup>12-23</sup> If iatrogenic cartilage damage can be eliminated during debridement chondroplasty, whereby superficial zone chondrocytes at or around the lesion are preserved, the opportunity to effect beneficial changes in chondrocyte function remains in that resident cell function may be recruited to aid healing.<sup>42-47</sup> Even though other experimental techniques have failed to show a significant repair response of native chondrocytes *within* partial-thickness lesions,<sup>48-50</sup> further study is required to determine whether early surgical intervention treatments that preserve and stabilize tissue permit or even induce a normal healing response.

## Acknowledgments and Funding

The work was performed at the Center for Integrated Nanotechnologies, United States Department of Energy, Office of Basic Energy Sciences User Facility, Los Alamos National Laboratory, Los Alamos, New Mexico (Contract DE-AC52-06NA25396) and Sandia National Laboratories (Contract DE-AC04-94AL85000) and Physicians Medical Center, Santa Fe, New Mexico. This study was supported by the New Mexico Small Business Grant Program WNM700, RO31, Los Alamos National Laboratory, Los Alamos, New Mexico, and by NuOrtho Surgical, Inc., Fall River, Massachusetts.

## Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interests with respect to the authorship and/or publication of this article.

## References

1. Curl WW, Krome J, Gordon ES, Rushing J, Smith BP, Poehling GG. Cartilage injuries: a review of 31,516 knee arthroscopies. *Arthroscopy* 1997;13:456-60.
2. Hjelle K, Solheim E, Strand T, Muri R, Brittberg M. Articular cartilage defects in 1,000 knee arthroscopies. *Arthroscopy* 2002;18:730-4.
3. Moskowitz RW. The burden of osteoarthritis: clinical and quality-of-life issues. *Am J Manag Care* 2009;15(suppl):S223-9.
4. Widuchowski W, Widuchowski J, Trzaska T. Articular cartilage defects: study of 25,124 knee arthroscopies. *Knee* 2007;14:177-82.
5. Woolf AD, Pfleger B. Burden of major musculoskeletal conditions. *Bull World Health Organ* 2003;81:646-56.
6. Burstein D. Tracking longitudinal changes in knee degeneration and repair. *J Bone Joint Surg Am* 2009;91(suppl 1):51-3.
7. Aaron RK, Skolnick AH, Reinert SE, Ciombor DM. Arthroscopic débridement for osteoarthritis of the knee. *J Bone Joint Surg Am* 2006;88:936-43.
8. Cameron-Donaldson M, Holland C, Hungerford DS, Frondoza CG. Cartilage debris increases the expression of chondrodestructive tumor necrosis factor- $\alpha$  by articular chondrocytes. *Arthroscopy* 2004;20:1040-3.

9. Day B. The indications for arthroscopic débridement for osteoarthritis of the knee. *Orthop Clin North Am* 2005;36:413-7.
10. Hunt SA, Jazrawi LM, Sherman OH. Arthroscopic management of osteoarthritis of the knee. *J Am Acad Orthop Surg* 2003;11:290.
11. Jackson RW, Dieterichs C. The results of arthroscopic lavage and debridement of osteoarthritic knees based on the severity of degeneration: a 4- to 6-year symptomatic follow-up. *Arthroscopy* 2003;19:13-20.
12. Amiel D, Ball ST, Tasto JP. Chondrocyte viability and metabolic activity after treatment of bovine articular cartilage with bipolar radiofrequency: an *in vitro* study. *Arthroscopy* 2004;20:503-10.
13. Caffey S, McPherson E, Moore B, Hedman T, Vangsness CT. Effects of radiofrequency energy on human articular cartilage. *Am J Sports Med* 2005;33:1035-9.
14. Cook JL, Kuroki K, Kenter K, Marberry K, Brawner T, Geiger T, et al. Bipolar and monopolar radiofrequency treatment of osteoarthritic knee articular cartilage: acute and temporal effects on cartilage compressive stiffness, permeability, cell synthesis, and extracellular matrix composition. *J Knee Surg* 2004;17:99-108.
15. Edwards RB III, Lu Y, Uthamanthil RK, Bogdanske JJ, Muir P, Athanasiou KA, et al. Comparison of mechanical débridement and radiofrequency energy for chondroplasty in an *in vivo* equine model of partial thickness cartilage injury. *Osteoarthritis Cartilage* 2007;15:169-78.
16. Edwards RB, Lu Y, Cole BJ, Muir P, Markel MD. Comparison of radiofrequency treatment and mechanical débridement of fibrillated cartilage in an equine model. *Vet Comp Orthop Traumatol* 2008;21:41-8.
17. Kang RW, Gomoll AH, Nho SJ, Pylawka TK, Cole BJ. Outcomes of mechanical débridement and radiofrequency ablation in the treatment of chondral defects: a prospective randomized study. *J Knee Surg* 2008;21:116-21.
18. Kaplan LD, Royce B, Meier B, Hoffmann JM, Barlow JD, Lu Y, et al. Mechanical chondroplasty: early metabolic consequences *in vitro*. *Arthroscopy* 2007;23:923-9.
19. Kaplan LD, Chu CR, Bradley JP, Fu FH, Studer RK. Recovery of chondrocyte metabolic activity after thermal exposure. *Am J Sports Med* 2003;31:392-8.
20. Lotto ML, Wright EJ, Appleby D, Zelicof SB, Lemos MJ, Lubowitz JH. *Ex vivo* comparison of mechanical versus thermal chondroplasty: assessment of tissue effect at the surgical endpoint. *Arthroscopy* 2008;24:410-5.
21. Lu Y, Edwards RB III, Nho S, Cole BJ, Markel MD. Lavage solution temperature influences depth of chondrocyte death and surface contouring during thermal chondroplasty with temperature-controlled monopolar radiofrequency energy. *Am J Sports Med* 2002;30:667-73.
22. Owens BD, Stickles BJ, Balikian P, Busconi BD. Prospective analysis of radiofrequency versus mechanical débridement of isolated patellar chondral lesions. *Arthroscopy* 2002;18:151-5.
23. Spahn G, Kahl E, Mückley T, Hofmann GO, Klinger HM. Arthroscopic knee chondroplasty using a bipolar radiofrequency-based device compared to mechanical shaver: results of a prospective, randomized, controlled study. *Knee Surg Sports Traumatol Arthrosc* 2008;16:565-73.
24. Huntley JS, Simpson AH, Hall AC. Use of non-degenerate human osteochondral tissue and confocal laser scanning microscopy for the study of chondrocyte death at cartilage surgery. *Eur Cell Mater* 2005;9:13-22.
25. Broom ND, Ngo T, Tham E. Traversing the intact/fibrillated joint surface: a biomechanical interpretation. *J Anat* 2005;206:55-67.
26. Choi JB, Youn I, Cao L, Leddy HA, Gilchrist CL, Setton LA, et al. Zonal changes in the three-dimensional morphology of the chondron under compression: the relationship among cellular, pericellular, and extracellular deformation in articular cartilage. *J Biomech* 2007;40:2596-603.
27. Glaser C, Putz R. Functional anatomy of articular cartilage under compressive loading: quantitative aspects of global, local and zonal reactions of the collagenous network with respect to the surface integrity. *Osteoarthritis Cartilage* 2002;10:83-99.
28. Setton LA, Zhu W, Mow VC. The biphasic poroviscoelastic behavior of articular cartilage: role of the surface zone in governing the compressive behavior. *J Biomech* 1993;26:581-92.
29. Silver FH, Bradica G, Tria A. Do changes in the mechanical properties of articular cartilage promote catabolic destruction of cartilage and osteoarthritis? *Matrix Biol* 2004;23:467-76.
30. Uthamanthil RK, Edwards RB, Lu Y, Manley PA, Athanasiou KA, Markel MD. *In vivo* study on the short-term effect of radiofrequency energy on chondromalacic patellar cartilage and its correlation with calcified cartilage pathology in an equine model. *J Orthop Res* 2006;24:716-24.
31. Kääh MJ, Bail HJ, Rotter A, Mainil-Varlet P, apGwynn I, Weiler A. Monopolar radiofrequency treatment of partial-thickness cartilage defects in the sheep knee joint leads to extended cartilage injury. *Am J Sports Med* 2005;33:1472-8.
32. Lu Y, Hayashi K, Hecht P, Fanton GS, Thabit G III, Cooley AJ, et al. The effect of monopolar radiofrequency energy on partial-thickness defects of articular cartilage. *Arthroscopy* 2000;16:527-36.
33. Voss JR, Lu Y, Edwards RB, Bogdanske JJ, Markel MD. Effects of thermal energy on chondrocyte viability. *Am J Vet Res* 2006;67:1708-12.
34. Alexopoulos LG, Williams GM, Upton ML, Setton LA, Guilak F. Osteoarthritic changes in the biphasic mechanical properties of the chondrocyte pericellular matrix in articular cartilage. *J Biomech* 2005;38:509-17.
35. Flachsmann R, Kim W, Broom N. Vulnerability to rupture of the intact articular surface with respect to age and proximity to site of fibrillation: a dynamic and static-investigation. *Connect Tissue Res* 2005;46:159-69.
36. Kerin AJ, Coleman A, Wisnom MR, Adams MA. Propagation of surface fissures in articular cartilage in response to cyclic loading *in vitro*. *Clin Biomech (Bristol, Avon)* 2003;18:960-8.
37. Lewis JL, Johnson SL. Collagen architecture and failure processes in bovine patellar cartilage. *J Anat* 2001;199(pt 4):483-92.

38. Lu Y, Markel MD, Swain C, Kaplan LD. Development of partial thickness articular cartilage injury in an ovine model. *J Orthop Res* 2006;24:1974-82.
39. Mobasheri A, Lewis R, Maxwell JE, Hill C, Womack M, Barrett-Jolley R. Characterization of a stretch-activated potassium channel in chondrocytes. *J Cell Physiol* 2010;223:511-8.
40. Papaioannou G, Demetropoulos CK, King YH. Predicting the effects of knee focal articular surface injury with a patient-specific finite element model. *Knee* 2010;17:61-8.
41. Temple-Wong MM, Bae WC, Chen MQ, Bugbee WD, Amiel D, Coutts RD, et al. Biomechanical, structural, and biochemical indices of degenerative and osteoarthritic deterioration of adult human articular cartilage of the femoral condyle. *Osteoarthritis Cartilage* 2009;17:1469-76.
42. Rolauffs B, Williams JM, Aurich M, Grodzinsky AJ, Kuettner KE, Cole AA. Proliferative remodeling of the spatial organization of human superficial chondrocytes distant from focal early osteoarthritis. *Arthritis Rheum* 2010;62:489-498.
43. Hayes AJ, Hall A, Cheung I, Brown L, Tubo R, Caterson B. Surface zone but not deep zone chondrocytes reorganize zonal architecture of articular cartilage grafts grown *in vitro*. Vol. 30, Poster 1774. Transactions of the 51st Annual Meeting of the Orthopaedic Research Society. Washington, DC: Orthopaedic Research Society; 2005.
44. Hayes AJ, Hall A, Brown L, Tubo R, Caterson B. Macromolecular organization and *in vitro* growth characteristics of scaffold-free neocartilage grafts. *J Histochem Cytochem* 2007;55:853-66.
45. Khan IM, Gilbert SJ, Singhrao SK, Duance VC, Archer CW. Cartilage integration: evaluation of the reasons for failure of integration during cartilage repair: a review. *Eur Cell Mater* 2008;16:26-39.
46. Schuurman W, Gawlitta D, Klein TJ, ten Hoop W, van Rijen MH, Dhert WJ, et al. Zonal chondrocyte subpopulations reacquire zone-specific characteristics during *in vitro* redifferentiation. *Am J Sports Med* 2009;37(suppl 1):97S-104S.
47. Shieh AC, Athanasiou KA. Biomechanics of single zonal chondrocytes. *J Biomech* 2006;39:1595-602.
48. Gelse K, von der Mark K, Schneider H. Cartilage regeneration by gene therapy. *Curr Gene Ther* 2003;3:305-17.
49. Quinn TM, Hunziker EB. Controlled enzymatic matrix degradation for integrative cartilage repair: effects on viable cell density and proteoglycan deposition. *Tissue Eng* 2002;8:799-806.
50. Sato M, Ishihara M, Furukawa K, Kaneshiro N, Nagai T, Mitani G, et al. Recent technological advancements related to articular cartilage regeneration. *Med Biol Eng Comput* 2008;46:735-43.