

Phylogenetic and morphometric analyses reveal ecophenotypic plasticity in freshwater mussels *Obovaria jacksoniana* and *Villosa arkansasensis* (Bivalvia: Unionidae)

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Abstract

Freshwater mollusk shell morphology exhibits clinal variation along a stream continuum that has been termed the Law of Stream Distribution. We analyzed phylogenetic relationships and morphological similarity of two freshwater mussels (Bivalvia: Unionidae), *Obovaria jacksoniana* and *Villosa arkansasensis*, throughout their ranges. The objectives were to investigate phylogenetic structure and evolutionary divergence of *O. jacksoniana* and *V. arkansasensis* and morphological similarity between the two species. Our analyses were the first explicit tests of phenotypic plasticity in shell morphologies using a combination of genetics and morphometrics. We conducted phylogenetic analyses of mitochondrial DNA (1416 bp; two genes) and morphometric analyses for 135 individuals of *O. jacksoniana* and *V. arkansasensis* from 12 streams. We examined correlations among genetic, morphological, and spatial distances using Mantel tests. Molecular phylogenetic analyses revealed a monophyletic relationship between *O. jacksoniana* and *V. arkansasensis*. Within this *O. jacksoniana/V. arkansasensis* complex, five distinct clades corresponding to drainage patterns showed high genetic divergence. Morphometric analysis revealed relative differences in shell morphologies between the two currently recognized species. We conclude that morphological differences between the two species are caused by ecophenotypic plasticity. A series of Mantel tests showed regional and local genetic isolation by distance. We observed clear positive correlations between morphological and geographic distances within a single drainage. We did not observe correlations between genetic and morphological distances. Phylogenetic analyses suggest *O. jacksoniana* and *V. arkansasensis* are synonymous and most closely related to a clade composed of *O. retusa*, *O. subrotunda*, and *O. unicolor*. Therefore, the synonymous *O. jacksoniana* and *V. arkansasensis* should be recognized as *Obovaria arkansasensis* (Lea 1862) n. comb. Phylogenetic analyses also showed relative genetic isolation among drainages, suggesting no current gene flow. Further investigation of in-progress speciation and/or cryptic species within *O. arkansasensis* is warranted followed by appropriate revision of conservation management designations.

Introduction

Preservation of biological diversity is the fundamental goal of conservation biology; however, the total biodiversity on Earth is still undetermined. Although freshwater ecosystems represent a small portion of the global area,

they harbor a disproportionately high level of diversity (Dudgeon et al. 2006; Strayer and Dudgeon 2010). Meanwhile, freshwater ecosystems may be the most threatened due to human use and associated activities causing worldwide habitat degradation. Examples of habitat degradation include a decline in water quality by pollution,

over exploitation of biota, introduction of invasive species, and modification of natural flow (Dudgeon *et al.* 2006). These activities caused severe declines in the habitat quality, range, and abundance of many freshwater species. In fact, the biodiversity extinction rate for freshwater ecosystems is increasing at a much faster rate than for terrestrial ecosystems (Ricciardi and Rasmussen 1999). Even limited data on extinction rates in North America freshwater ecosystems are indicative of a global freshwater biodiversity crisis (Abell *et al.* 2008).

Freshwater mussels (Bivalvia: Unionoida) are among the most endangered groups of animals in North America (Williams *et al.* 1993; Neves 1998; Lydeard *et al.* 2004; Strayer *et al.* 2004; Haag 2012). Of the approximately 300 recognized freshwater mussel species in North America, approximately 72% are endangered, threatened, proposed for listing or presumed extinct (Williams *et al.* 1993; Lydeard *et al.* 2004). Conservation efforts are hindered by taxonomic uncertainty and a lack of ecological and evolutionary information on most taxa (Strayer *et al.* 2004).

Accurate species identification is necessary to assess management options for freshwater mussel conservation and protection. External morphology is the primary characteristic for freshwater mussel species identification; however, morphology-based identification is problematic and may lead to misidentifications due to shell erosion and cryptic morphology. Furthermore, shell morphologies are highly variable due to environmental factors, sometimes resulting in extreme ecophenotypic plasticity within single species (Watters 1994). In other organisms, morphology often exhibits trends of clinal variation within single species reflecting positional (e.g., latitude, longitude, altitude, and depth) and/or environmental (e.g., temperature, precipitation, and salinity) variables (Gaston *et al.* 2008). Morphological variations in freshwater mussels are thought to be caused by a combination of hydrodynamic forces, substrate types, burrowing activities, and/or historic environmental conditions (Watters 1994; Peacock and Seltzer 2008; Allen and Vaughn 2009). Several studies have observed morphological variations of the shell among populations in different habitats (Utterback 1917; Ortmann 1920; Ball 1922; Mackie and Topping 1988; Hornbach *et al.* 2010; see Haag 2012 for a summary). An early study reported that a single species could be light and laterally compressed in form in headwaters, but heavier and more laterally inflated in downstream waters (Utterback 1917). Detailed observations on the upper Tennessee River fauna reported differences in shell inflation correlating with river positions (Ortmann 1918). Ortmann (1920) termed this clinal morphological variation the Law of Stream Distribution, where gradual changes in shell morphologies occur from upstream to downstream. Although several studies have observed

phenotypic plasticity in shell morphologies, to date few have explicitly tested the Law of Stream Distribution in freshwater mollusks (e.g., Mackie and Topping 1988; Minton *et al.* 2008, 2011).

We used molecular phylogenetics and shell morphometrics to investigate the relationship between *Obovaria jacksoniana* (Frierson 1912) and *Villosa arkansasensis* (Lea 1862) (Fig. 1). These two freshwater mussel species are sometimes confused (Vaughn *et al.* 1996) and are of conservation concern (Williams *et al.* 1993; Harris *et al.* 2010). *O. jacksoniana* is widely distributed from the Interior Highlands to the Gulf of Mexico and Mobile River basins, and inhabits mid- to large-size rivers (Oesch 1995; Howells *et al.* 1996). Although it is locally abundant in preferred habitat, this species is globally rare (Branson 1984; Vaughn *et al.* 1996; NatureServe 2012). The *O. jacksoniana* type locality is the Pearl and Yalobusha rivers in Mississippi (Frierson 1912; Fig. 1). A recent survey of the lower Pearl River did find any live specimens (Brown *et al.* 2010). *V. arkansasensis* is endemic to the Ouachita Mountains region in Arkansas and Oklahoma, and inhabits headwaters to small-size rivers in the Ouachita and Red River systems (Robison and Allen 1995). The *V. arkansasensis* type locality is the Ouachita River near Hot Springs, Arkansas (Lea 1862; Fig. 1); however, it is presumably extirpated from the type locality due to a series of impoundments along the Ouachita River. Both species are typically small, length usually not exceeding 50–55 mm. Shell of *O. jacksoniana* is moderately thick, laterally inflated, and shape is oval to subtriangular (Williams *et al.* 2008). In contrast, *V. arkansasensis* shell is relatively thin, laterally compressed, and shape is subtriangular (Robison and Allen 1995). Both species exhibit



Figure 1. Type specimens of *Obovaria jacksoniana* (left; Academy of Natural Sciences of Drexel University, ANSP 106063) and *Villosa arkansasensis* (right; Smithsonian National Museum of Natural History, USNM 25710). The pictures were provided by the MUSSEL project (<http://mussel-project.uwsp.edu>).

sexual dimorphism with female *O. jacksoniana* more inflated and broader in the posterior margin and female *V. arkansasensis* with an indentation and sulcus emanating from the posterior ventral margin. As both species are distributed in the southeastern United States and have similar shell characteristics, they can be difficult to distinguish in the field, especially in small specimens.

The objectives of this study were to investigate (1) phylogenetic structure and evolutionary divergence of *O. jacksoniana* and *V. arkansasensis* using two mitochondrial DNA (mtDNA) genes; and (2) morphological similarity between the two species using traditional morphometric analysis. We tested the null hypothesis that phylogenetic relationships between *O. jacksoniana* and *V. arkansasensis* reflect current taxonomic status and the similarity of shell morphology between the species is due to convergence. Furthermore, we investigated correlations between morphological variations and geographic distance along the rivers. As the conservation status of both species is considered special concern (Williams et al. 1993; Harris et al. 2010), and they co-occur in certain river drainages, our aim was to confirm their taxonomic status and provide shell morphological characteristics for accurate identification that is integral to population assessment for conservation management.

Methods

Sampling

We used 116 individuals of *O. jacksoniana* representing 12 streams from seven major drainages and 19 individuals of *V. arkansasensis* representing three streams from two major drainages with initial species assignments based on shell morphology (Fig. 2; see Table S1). Specimens were either collected specifically for this study or obtained from museum collections. We obtained *O. jacksoniana* paralectotypes (University of Michigan Museum of Zoology, UMMZ 107523) and *V. arkansasensis* (near) topotypic specimens (Arkansas State University Museum of Zoology, ASUMZ 4579-4583) for inclusion. All putative *Obovaria* species (*O. jacksoniana*, *O. olivaria* [Rafinesque 1820], *O. retusa* [LaMarck 1819], *O. subrotunda* [Rafinesque 1820], and *O. unicolor* [Lea 1845]), except *O. hadletoni* (Athearn 1964), and four putative *Villosa* species (*V. fabalis* [Lea 1831], *V. iris* [Lea 1829], *V. vanuxemensis* [Lea 1838], and *V. villosa* [Wright 1898]) were included in the phylogenetic analysis. "*Obovaria*" *rotulata* (Wright 1899) was also included in this analysis, and it has been placed in both *Fusconaia* (Williams et al. 2008) and *Obovaria* (Graf and Cummings 2007), but more recently it has been assigned to the new genus *Reginaia* (Campbell and Lydeard 2012a,b; Haag 2012). It should also be noted

that recent phylogenetic analyses of *Villosa* have shown paraphyly and taxonomic revision is needed (Zanatta and Murphy 2006; Kuehnl 2009). Outgroups in the phylogenetic analysis included representatives from seven tribes in the North American Unionidae (see Table S2).

Newly collected specimens were separated into soft tissues and shells with soft tissues preserved in absolute ethanol in -20°C freezer and shells scrubbed inside and out to remove excess material. In most instances, we used the same specimens for both molecular phylogenetic and morphometric analyses.

DNA Extraction and sequencing

We extracted whole genomic DNA from ethanol-preserved mantle tissue using standard CTAB/chloroform extraction followed by ethanol precipitation (Saghai-Marouf et al. 1984). We obtained dried muscle tissues from adductor scars of paralectotype *O. jacksoniana* specimens. Dried tissues were soaked in CTAB buffer and incubated at 55°C overnight prior to extraction. We amplified two mtDNA genes, cytochrome *c* oxidase subunit I (COI), and NADH dehydrogenase subunit 1 (ND1), following the specifications included with *Taq* DNA polymerase (QIAGEN, Inc., Valencia, CA) in a $20\text{-}\mu\text{L}$ reaction. We used the COI and ND1 primers, reaction conditions, reagent concentrations, and thermal cycles described in Campbell et al. (2005). We cleaned PCR products using a QIAquick gel extraction kit (QIAGEN, Inc.) per $26\text{ }\mu\text{L}$ of PCR product. We used the same primers as PCR for sequencing reactions. Sequencing products were analyzed on a CEQTM 8000 automated sequencer (Beckman Coulter, Inc., Brea, CA). DNA sequences were assembled, edited, and aligned by eye using the program DNADYNAMO (Blue Tractor Software, Ltd., Llanfairfechan, U. K.), and an open reading frame for these genes was verified. Ambiguous sequences of both the 3'- and 5'-ends were trimmed.

Phylogenetic analyses

We conducted the following statistical tests prior to phylogenetic analyses in order to qualify the genetic data. The test for nucleotide saturation was based on Xia et al. (2003) implemented in DAMBE v.5.2.18 (Xia and Xie 2001). Nucleotide saturation is a substantial problem in loss of phylogenetic information contained in sequence data resulting in misleading phylogenetic relationships. Substitution saturations were determined for each gene and each codon of genes.

We analyzed phylogenetic relationships using maximum likelihood analysis (ML) and Bayesian inference analysis (BI). MtDNA sequence data from COI and ND1 gene portions were analyzed as a concatenated dataset of unique

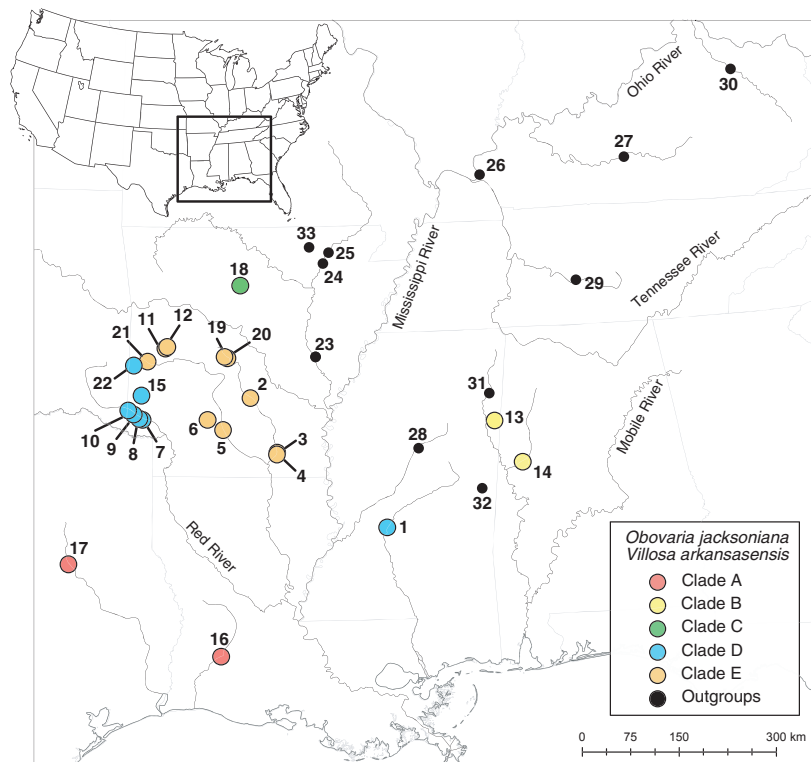


Figure 2. Collection sites in the southern United States. Colors correspond to phylogenetic clades (Fig. 3). Numbers correspond to locality information (see Table S1).

haplotypes only using TCS v.1.21 (Clement et al. 2000). For estimating best-fit models of nucleotide substitution, we used KAKUSAN4 (Tanabe 2011). KAKUSAN4 computes substitution models for multiple loci and each codon partition and estimates likelihoods of substitution models and automatically generates TREEFINDER and NEXUS files to directly analyze data using each phylogenetic program. Based on the Akaike information criteria (AIC), best-fit models for a concatenated dataset were GTR+ Γ for all codon positions, except HKY85 for the second codon of COI. These best-fit models were used in both ML and BI. ML was performed with TREEFINDER (Jobb et al. 2004; Jobb 2008) using the default settings and the files generated by Kakusan4. Support values were generated by pseudo-bootstrapping with 1000 replicates using the expected-likelihood weights with local rearrangements of tree topology (LR-ELW) implemented in TREEFINDER. The LR-ELW edge support can be directly interpreted as confidence in the configuration of branches adjacent to a particular edge. BI was performed with MRBAYES v3.1.2 (Huelsenbeck and Ronquist 2001) by Markov Chain Monte Carlo. Two simultaneous Markov chains (each chain contains three heated chains and one cold chain) were run for four million generations with trees sampled every 1000th generation yielding 4001 trees for each chain in the initial samples. We assessed burn-in by plotting the log likelihood scores for each sampling point using TRACER v1.5 (Rambaut and Drummond 2007), and

we considered the Markov chains stationary when likelihood values reached a plateau. Therefore, we discarded the first 1001 trees (25%) as burn-in for each run, and the remaining 3000 trees were calculated using the 50% majority rule consensus trees. BI trees were compared with the ML tree, and the most credible inferences of relationship were confined to nodes where both the LR-ELW bootstrap support values were >70 and the Bayesian posterior probabilities were >0.95.

Evolutionary divergence

We estimated genetic diversity within *Obovaria* species and clades of the *O. jacksoniana/V. arkansasensis* complex. We calculated the number of substitutions per site by averaging overall sequence pairs within group. We also estimated evolutionary divergence among *Obovaria* species and clades within the *O. jacksoniana/V. arkansasensis* complex using the concatenated dataset and the maximum composite likelihood method in MEGA v5.05 (Tamura et al. 2011).

Morphometric analysis

We measured three shell characters on only the *O. jacksoniana/V. arkansasensis* complex from all of the locations. We measured each shell to the nearest 0.05 mm for

maximum length (anterior to posterior), height (dorsal to ventral), and width (right to left valve) using dial or digital calipers. To standardize the variables for size, we calculated the height/length, width/length, and width/height ratios for all specimens. The ratio data were normalized using arcsine transformation, and normal distribution was verified for each ratio using Shapiro–Wilk test (Sokal and Rohlf 1995). This method has been used to distinguish morphologies of cryptic species in freshwater mussels (Gangloff *et al.* 2006) and to evaluate morphological variations within species (Hornbach *et al.* 2010). We examined morphological variation within and among species through principal component analysis (PCA) and canonical variates analysis (CVA). PCA simplifies descriptions of variation among individuals, while CVA simplifies descriptions of variation between groups (Zelditch *et al.* 2004). In PCA, no a priori assumptions are needed to group individuals. Meanwhile, an a priori assumption of group membership is required for CVA, as it determines the set of axes that best discriminate between groups. We performed a CVA on a data set with groups assigned by phylogenetic clades. Additionally, we utilized Hotelling's tests for pairwise comparisons between groups assigned by species and by phylogenetic clades, and we conducted discriminant function analysis (DFA) for each pair of groups to determine how frequently PC scores correctly distinguished between species. All statistical analyses were performed using PAST software package (Hammer *et al.* 2001).

Test of correlations

We tested correlations of geographic distance, morphological distance, and genetic distance using Mantel tests (Mantel 1967) to examine the effect of landscape features on genetic and morphological structure in *O. jacksoniana* and *V. arkansasensis*. For geographic distance, we measured total distance between populations along the rivers (river distance) using ArcGIS v9.3 (ESRI, Inc., Redlands, CA) because direct geographic distance is often not the best indicator of isolation by distance in aquatic organisms (Spear *et al.* 2005; Storfer *et al.* 2007). This is especially true of mussels as they are fully aquatic and incapable of migration over terrestrial habitats. When two populations were not connected by rivers, we took the closest distance through the Gulf of Mexico. For morphological distance, we calculated the Euclidean distance between individuals from PCA plots using R (R Development Core Team 2011). We calculated pairwise genetic distances between individuals from the concatenated data set with a maximum composite likelihood model using MEGA v5.05 (Tamura *et al.* 2011). We performed Mantel tests for a combination of three matrices: (1) between river distance

and genetic distance (hereafter $R \times G$); (2) between river distance and morphological distance ($R \times M$); and (3) between morphological distance and genetic distance ($M \times G$). In addition, because we had a number of populations represented in the Saline and Little rivers (see Table S1), we performed Mantel tests for each matrix combination on these rivers separately from the overall data set. Mantel tests were performed in the Microsoft Excel add-in program GENALEX (Peakall and Smouse 2006) using 9999 permutations.

Results

Phylogenetic analyses

We obtained 645 bp of the COI gene from 135 individuals and 771 bp of the ND1 gene from 125 individuals. We concatenated both genes and obtained a total of 116 unique haplotypes. All DNA sequences were deposited in NCBI GenBank (accession numbers: KF035133–KF035280 for COI and KF035281–KF035420 for ND1). The test for substitution saturation showed no indication of saturation among codons (data not shown). Thus, we used all codon positions in our analyses.

Phylogenetic analyses showed that *O. jacksoniana* and *V. arkansasensis* formed a single monophyletic clade (Fig. 3). *Obovaria* species (including *V. arkansasensis*) showed a monophyletic relationship with high support values; however, *Villosa* species included in our study were polyphyletic forming four clades. Among *Obovaria*, *O. olivaria* comprised the basal clade in the ML tree followed by a clade comprised of *O. retusa*, *O. unicolor*, and *O. subrotunda*, and finally a clade of *O. jacksoniana* and *V. arkansasensis*. According to both ML and BI trees, each *Obovaria* species forms a reciprocally monophyletic clade with high support values validating current taxonomic concepts.

Phylogeographic analysis revealed five distinct clades with high support values associated with major drainages for the *O. jacksoniana/V. arkansasensis* complex. Individuals from the Western Gulf drainages (e.g., Calcasieu and Neches rivers) comprised the basal clade of the *O. jacksoniana/V. arkansasensis* complex, which continued to individuals from the Mobile River (Eastern Gulf) drainage and from the White River drainage (Fig. 3). Individuals from the Red and Ouachita River drainages were located as sister clades. Individuals from the Fourche La Fave River, a tributary of the Arkansas River, fell into the same clade as the Ouachita populations (Fig. 3 Clade E). Surprisingly, a paralectotype individual from the Pearl River, MS was more closely related to individuals from the Cosatot River and Mountain Fork of the Little River (Red River drainages). No individuals from the same population were located in more than one major drainage clade.

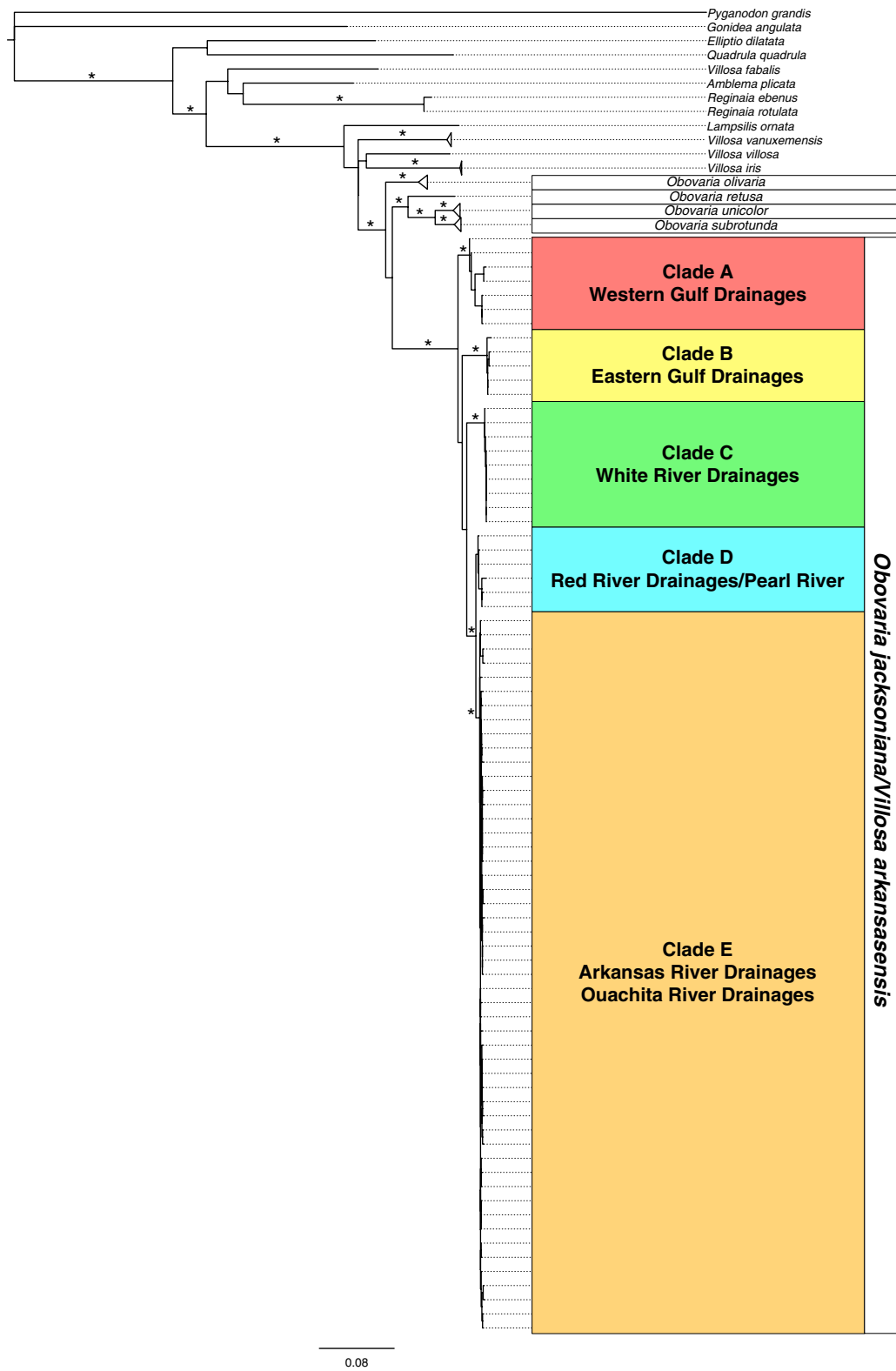


Figure 3. Phylogenetic tree generated by Bayesian and maximum likelihood analyses for concatenated genes. Asterisks indicate branches with bootstrap support values >70 for maximum likelihood analysis and posterior probabilities >0.95 for Bayesian analysis. *Pyganodon grandis* is the outgroup. Colors on *Obovaria jacksoniana/Villosa arkansasensis* clades correspond to Fig. 2.

Evolutionary divergence

We estimated intraspecific genetic diversity over a concatenated data set (Table 1). As the phylogenetic analyses showed five *Obovaria* clades representing recognized species, and five additional clades within the *O. jacksoniana/V. arkansasensis* complex, we estimated intraclade genetic diversity as well. Intraspecific genetic diversity ranged from 0.005 (*O. unicolor*) to 0.02 (*O. jacksoniana/V. arkansasensis* complex). Within the *O. jacksoniana/V. arkansasensis* clades, genetic diversity ranged from 0.001 (Clade C) to 0.009 (Clade A). Overall, *O. jacksoniana/V. arkansasensis* had relatively higher genetic diversity when compared to other *Obovaria* species.

We estimated evolutionary divergence between clades of *Obovaria* (Table 1). Evolutionary divergences ranged from 0.038 to 0.109 with a mean of 0.076. We observed the highest divergence between *O. retusa* and *O. jacksoniana/V. arkansasensis* complex (0.109, SE = 0.0092), and the lowest divergence between *O. subrotunda* and *O. unicolor* (0.038, SE = 0.005). Within the *O. jacksoniana/V. arkansasensis* clades, we observed relatively high evolutionary divergences between Clade A and Clade B (0.046, SE = 0.0067), between Clade B and Clade C (0.043, SE = 0.0057), and between Clade B and Clade E (0.043, SE = 0.0059). These evolutionary divergences were higher than between *O. subrotunda* and *O. unicolor*. We observed small divergences between Clade D and Clade E (0.011, SE = 0.0022).

Morphometric analyses

We analyzed a total of 114 individuals identified morphologically as *O. jacksoniana* and 17 individuals identified morphologically as *V. arkansasensis*. PCA yielded two distinct eigenvalues and described >99% of the total variability between species. The PC1 axis described 80.05% and the PC2 axis described 19.94% of the total variation (Fig. 4A and B). The PCA, with group assigned by species, showed a wide morphological range for *O. jacksoniana* and a relatively small morphological range for *V. arkansasensis* (Fig. 4A) with large overlapping cluster portions that included 29.77% of the total individuals. While PCA showed large cluster portions overlapping between the two species, pairwise comparisons (Hotelling's tests) between *O. jacksoniana* and *V. arkansasensis* showed significantly different morphologies (Bonferroni corrected, $P < 0.005$). The DFA scores between two groups revealed 76.34% of individuals were correctly assigned a group, where 29 individuals of *O. jacksoniana* were assigned to *V. arkansasensis* and two individuals of *V. arkansasensis* was assigned to *O. jacksoniana*.

Table 1. Intraspecific nucleotide diversity and evolutionary divergence estimates (SE estimates in parentheses) over concatenated sequence pairs among genus *Obovaria*.

Species	Clade	<i>Obovaria jacksoniana/Villosa arkansasensis</i>				
		<i>Obovaria olivaria</i>	<i>Obovaria retusa</i>	<i>Obovaria subrotunda</i>	<i>Obovaria unicolor</i>	Overall
<i>Obovaria olivaria</i>		0.007 (0.0014)				
<i>Obovaria retusa</i>		0.088 (0.0086)	n.a.			
<i>Obovaria subrotunda</i>		0.087 (0.0085)	0.081 (0.0085)	0.006 (0.0014)		
<i>Obovaria unicolor</i>		0.084 (0.0078)	0.082 (0.0083)	0.038 (0.0050)	0.005 (0.0009)	
<i>Obovaria jacksoniana</i>	Overall	0.093 (0.0073)	0.109 (0.0092)	0.106 (0.0079)	0.100 (0.0079)	0.020 (0.0020)
<i>Villosa arkansasensis</i>	Clade A	0.086 (0.0098)	0.107 (0.0124)	0.092 (0.0097)	0.091 (0.0101)	n.a.
	Clade B	0.095 (0.0091)	0.118 (0.0115)	0.116 (0.0105)	0.107 (0.0097)	n.a.
	Clade C	0.093 (0.0095)	0.107 (0.0109)	0.103 (0.0098)	0.095 (0.0091)	n.a.
	Clade D	0.087 (0.0089)	0.106 (0.0112)	0.100 (0.0094)	0.094 (0.0087)	n.a.
	Clade E	0.095 (0.0090)	0.109 (0.0109)	0.108 (0.0094)	0.102 (0.0091)	n.a.
					0.009 (0.0027)	0.009 (0.0027)
					0.046 (0.0067)	0.046 (0.0067)
					0.040 (0.0070)	0.043 (0.0057)
					0.036 (0.0057)	0.041 (0.0056)
					0.039 (0.0061)	0.043 (0.0059)
					0.043 (0.0059)	0.043 (0.0059)
					0.011 (0.0022)	0.011 (0.0022)
					0.004 (0.0007)	0.004 (0.0007)

Clades in *Obovaria jacksoniana/Villosa arkansasensis* complex correspond to phylogenetic tree (Fig. 3). n.a., not applicable.

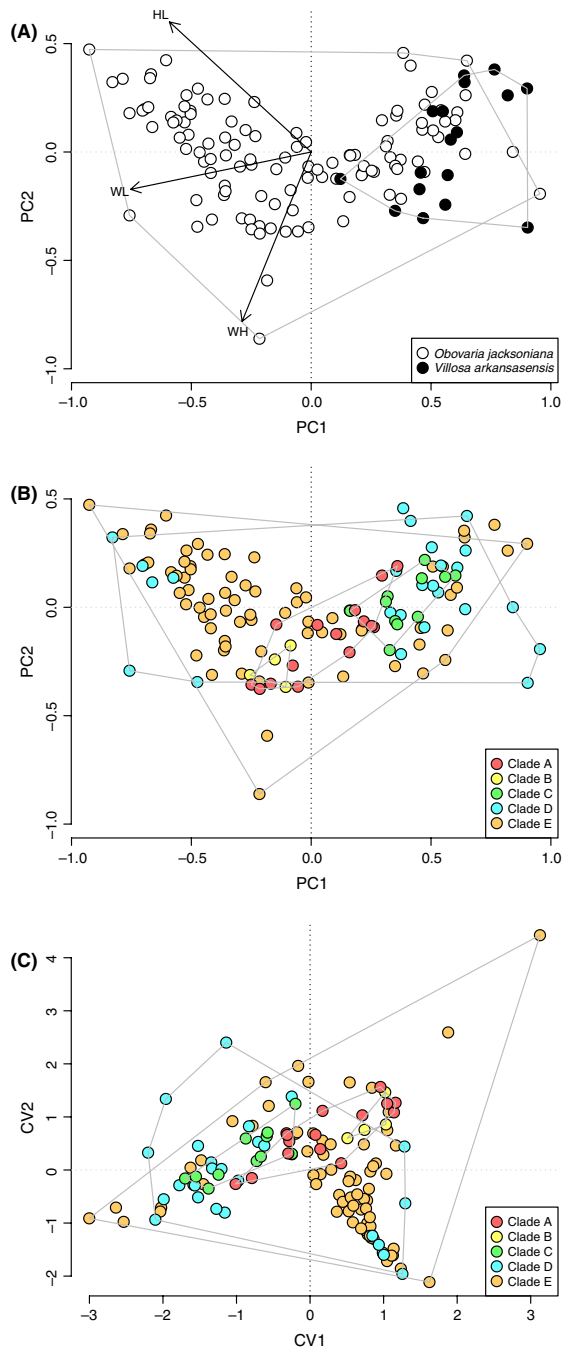


Figure 4. Scatter plots from principal component analysis (PCA; A and B) and canonical variates analysis (CVA; C) of *Obovaria jacksoniana* ($n = 114$) and *Villosa arkansasensis* ($n = 17$). Polygons enclose convex hull of each group assigned. PCA plots were grouped assigned by species (A) and by phylogenetic clades (B). Arrows showed a biplot of variables on PCA (HL, height/length, WL, width/length, and WH, width/height). The PC1 axis described 80.05% and the PC2 axis described 19.94% of total variation. (C) CVA plot was analyzed with groups assigned by phylogenetic clades. The CV1 axis described 68.14% and the CV2 axis described 31.86% of total variation. Colors of phylogenetic clades correspond to Fig. 3.

The PCA, with group assigned by clades, showed wide morphological ranges for Clade D and Clade E, while the rest of the clades showed relatively small morphological ranges overlapping with Clade D and Clade E (Fig. 4B). The CVA, with group assigned by clades, yielded two distinct axes and described 96% of the total variability among clades (Fig. 4C). The CV1 axis described 67.41% and the CV2 axis described 29.58% of the total variation. Similar to PCA results, CVA clusters also showed a wide morphological range of Clade D and Clade E and large overlap among clades. Pairwise comparisons among clades showed no significant difference between Clade A, Clade B, and Clade E (Bonferroni corrected, $P = 1-0.369$, see Table S3), while the rest of the pairwise comparisons was significantly different (Bonferroni corrected, $P < 0.05$).

Test of correlations

Mantel tests examining correlations of three distance matrices in overall populations indicated significant correlations of $R \times G$ and $R \times M$ (Fig. 5). $R \times G$ showed strong positive correlations indicating strong isolation by distance (Mantel's $r = 0.837$, $P < 0.001$; Fig. 5A). Although $R \times M$ correlation was statistically significant, correlation was only slightly positive (Mantel's $r = 0.079$, $P = 0.017$; Fig. 5B). Correlation of $M \times G$ was not statistically significant (Mantel's $r = 0.012$, $P = 0.404$; Fig. 5C) but indicated two distinct clusters. The bottom cluster shows that shell morphologies are highly variable within genetically closely related individuals. Meanwhile, the top cluster shows that less related individuals possessed both similar and different shell morphologies.

Results for the Saline and Little rivers showed similar trends to the overall data set; however, only correlations of $R \times M$ for both rivers were statistically significant (Mantel's $r = 0.761$, $P < 0.001$ for Saline River; $r = 0.929$, $P = 0.002$ for Little River; Fig. 6B and E). Correlations of $R \times G$ were only significant for the Saline River and marginally significant for the Little River (Mantel's $r = 0.151$, $P = 0.021$ for the Saline River; $r = 0.772$, $P = 0.057$ for the Little River; Fig. 6A and D). This indicates that isolation by distance relationships are weakened at smaller spatial scales. Similar to the overall data set, neither river showed correlations of $M \times G$ (Mantel's $r = 0.038$, $P = 0.337$ for Saline River; $r = 0.783$, $P = 0.143$ for Little River; Fig. 6C and F).

Discussion

Genetic structure

Phylogenetic analysis did not support the hypothesis that phylogenetic relationships between *O. jacksoniana* and

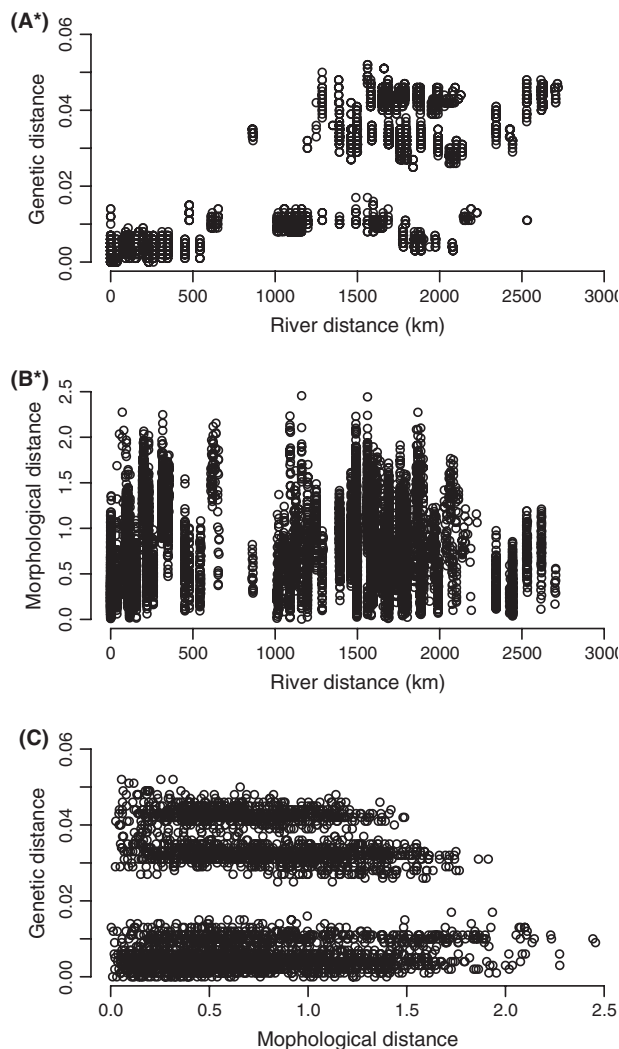


Figure 5. Results of Mantel tests between (A) pairwise river distance among collection sites (km) and pairwise genetic distance, (B) pairwise river distance and pairwise morphological distance, and (C) pairwise morphological distance and pairwise genetic distance. * $P < 0.05$.

V. arkansasensis reflect current taxonomic status; rather, these two species shared a single clade. These two nominal species are morphological variants of the same phylogenetic species. The *O. jacksoniana/V. arkansasensis* complex split into five reciprocally monophyletic clades indicating five phylogeographic isolations primarily corresponding to major river drainages. Phylogeographic patterns associated with geographic barriers and/or historical vicariant events are reported in many freshwater organisms such as fish (Mayden 1985, 1988; Turner and Robinson 2006; Berendzen *et al.* 2007), crayfish (Crandall and Templeton 1999; Fetzner and Crandall 2003), and mussels (King *et al.* 1999; Turner *et al.* 2000; Roe *et al.* 2001; Serb 2006; Burdick and White 2007; Elderkin *et al.* 2008).

In our study, the five *O. jacksoniana/V. arkansasensis* clades are distributed in the western Gulf of Mexico (Calcasieu and Neches rivers), eastern Gulf of Mexico (Mobile River), and three major Mississippi River drainage systems (White, Arkansas/Ouachita, and Red rivers), which correspond to the recently established Sabine-Trinity, Mobile Basin, and Interior Highlands mussel provinces (Haag 2009), respectively. It is noteworthy that topotypic Pearl River *O. jacksoniana* grouped with the Red River clade. Our results suggest that large rivers such as the Mississippi River and the marine environment of the Gulf of Mexico are substantial barriers to gene flow for this species complex. This is evidenced by a lack of haplotype sharing among river drainages, relatively high levels of evolutionary divergence among drainages [*O. jacksoniana/V. arkansasensis* complex (3.4%) compared to *Potamilus* (1.32%) and *Quadrula* (3.65%) (Roe and Lydeard 1998; Serb *et al.* 2003)], and significant positive isolation by distance patterns. These patterns of isolation by distance have been observed in other freshwater mussel species; *Actinonaias ligamentina* (LaMarck 1819) (Elderkin *et al.* 2008), *Elliptio dilatata* (Rafinesque 1820) (Berg *et al.* 2007; Elderkin *et al.* 2008), *Epioblasma torulosa rangiana* (Lea 1838) (Zanatta and Murphy 2007), and *Unio pictorum* (Linnaeus 1758) (Zieritz *et al.* 2010). Another potential cause of isolation by drainage is the relative immobility of host fishes. While the life history of *O. jacksoniana* is not completely known, specifically with regards to host fish, we make the assumption that *O. jacksoniana* uses the same hosts as *V. arkansasensis*, which are centrarchids (*Ambloplites ariommus* [Viosca 1936] and *Lepomis cyanellus* [Rafinesque 1819]) and benthic percids (*Etheostoma blennioides* [Rafinesque 1819] and *Etheostoma collettei* [Birdsong and Knapp 1969]) (Christian *et al.* 2007). All of these species have small home ranges and poor dispersal abilities (Greenberg 1991; Gatz 2007).

Shell morphologies and correlations with landscapes

Various taxa have displayed phenotypic plasticity in response to biotic and abiotic factors (Brönmark and Miner 1992; Via *et al.* 1995). Both environmental and landscape factors can influence phenotypic variation (e.g., Hinch *et al.* 1986; Shepherd 1998; Minton *et al.* 2008). In freshwater mussels, gradual changes in shell morphologies from upstream to downstream have been observed in several taxa (Ortmann 1920; Graf 1998; Hornbach *et al.* 2010). Similar to these studies, our results showed clear evidence that spatial and river characteristics strongly influenced morphological variations of these freshwater mussels.

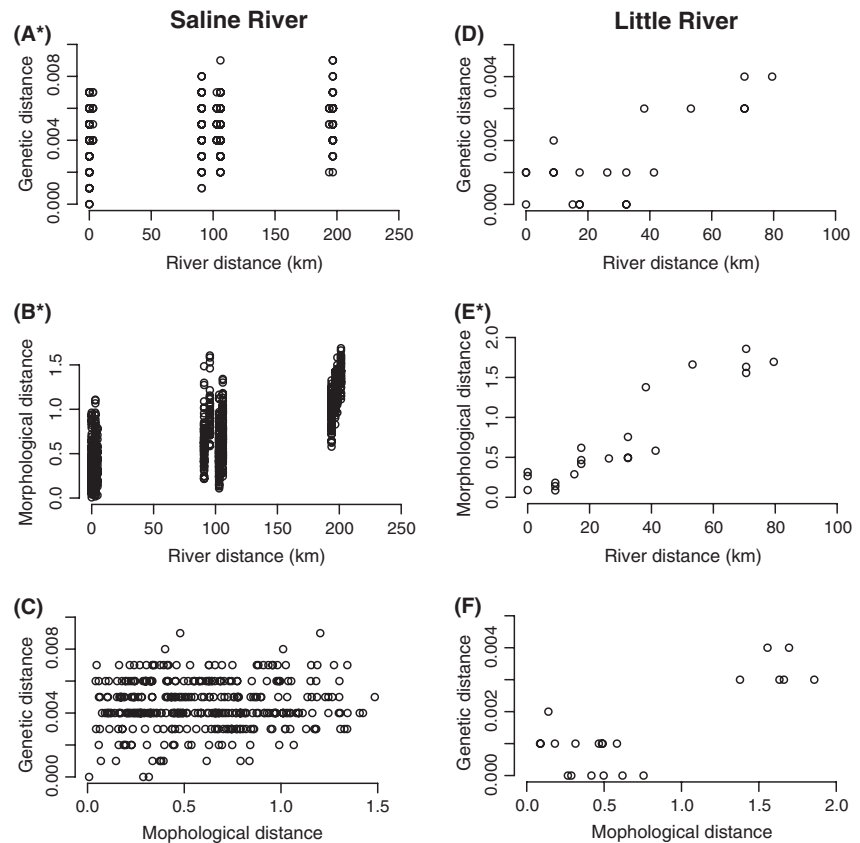


Figure 6. Results of Mantel tests between (A and D) pairwise river distance among collection sites (km) and pairwise genetic distance, (B and E) pairwise river distance and pairwise morphological distance, and (C and F) pairwise morphological distance and pairwise genetic distance for the Saline River and Little River, respectively. * $P < 0.05$.

Although Hotelling's test showed significant morphological differences between *O. jacksoniana* and *V. arkansasensis*, 29 individuals (22.8%) were incorrectly assigned to initially identified species, and morphometric analysis through PCA showed significant morphological overlap between the species. The PCA biplot showed that PC1 is mainly weighted by a proportion of lateral inflation and shell length, and some *O. jacksoniana* individuals exhibit less lateral inflation, similar to *V. arkansasensis* shell morphologies. Correlations of $R \times M$ at local scales (individual rivers) showed that similarity of shell morphologies gradually declined with increasing river distance between sites; however, this relationship was only slightly positive at the regional scale where similarity of shell morphologies did not decline with increasing distance between rivers. Additionally, we did not observe significant correlations of $M \times G$ in either local or regional scales indicating that genetic characters were not the primary control of shell morphological variation, and genetically similar individuals can exhibit either the *O. jacksoniana* or *V. arkansasensis* morphotype. Thus, for this species complex, it is likely that shell morphology is primarily determined by environmental factors, such as river position. Given the evidence, it appears *V. arkansasensis* is the headwater

morphotype and *O. jacksoniana* is the larger river morphotype in the *O. jacksoniana/V. arkansasensis* complex.

Conservation implications

Based on our findings, *V. arkansasensis* and *O. jacksoniana* are synonymous and most closely related to a clade composed of *O. retusa*, *O. subrotunda*, and *O. unicolor*. Therefore, *V. arkansasensis* and *O. jacksoniana* should be recognized as *Obovaria arkansasensis* (Lea 1862) n. comb. As *O. jacksoniana* and *V. arkansasensis* are thought to occupy different longitudinal reaches along the stream continuum, synonymy will expand the overall distribution of the taxon within a specific river drainage from headwaters to large river habitats. However, we must be cautious in reassessing the conservation status of *O. arkansasensis*. The mtDNA phylogeny showed five reciprocally monophyletic clades and high levels of evolutionary divergence among clades indicating in-progress speciation and/or cryptic species within the *O. jacksoniana/V. arkansasensis* complex. These lineages are evolutionarily significant and require separate conservation management. We should note, however, that the mtDNA phylogeny represents only part of species' evolutionary history. Therefore, further

study using additional specimens from throughout the geographic range and additional genetic markers such as the nuclear DNA genes and microsatellite loci is required to elucidate genetic divergences and to refine species limits. Finally, biological and ecological assessments, such as host fish and habitat specificities among the *O. jacksoniana/V. arkansasensis* complex, are highly recommended, and we suggest host fish species investigations for the *O. jacksoniana/V. arkansasensis* complex from multiple drainages to further assess ecological variance of the species complex.

Recent molecular phylogenetic analyses elucidated cryptic diversity within freshwater mussels (King et al. 1999; Gangloff et al. 2006). However, discovery of potential synonymous species possessing different phenotypes due to (eco)phenotypic plasticity is rare compared to other taxa. For example, a recent study of deep-sea macroinvertebrates revealed synonymous species in tubeworms and shrimps previously assigned to different species or genera due to developmental plasticity (Vrijenhoek 2009). We believe there may be other examples in freshwater mussels, which have yet to be discovered.

Conclusions

Although shell morphology is often used as the key characteristic for identification and taxonomy of freshwater mussel species, morphologies can be extremely variable in different habitats. Our study shows evidence of ecophenotypic plasticity in freshwater mussel shell morphologies within the same drainages and highlights the difficulty of shell morphology-based identification. Our study was unique in that genetic information provided evidence for synonymy of species, while morphometric data supported two distinct taxa. Based on our study, we conclude that *O. jacksoniana* and *V. arkansasensis* are different morphotypes in the same species complex due to ecophenotypic plasticity and should be synonymized under the nomen *Obovaria arkansasensis* (Lea 1862) n. comb. However, we recommend this species retain the conservation status of special concern due to its relative rarity over a large portion of its overall range. Moreover, our study suggests that correlations among distance matrices are of great utility to investigate not only patterns of genetic isolation by distance but also phenotypic plasticity within the species. Genetic isolation by distance has been frequently used to examine dispersal abilities, population fragmentation, and gene flow among populations for other freshwater mussel species (Elderkin et al. 2007, 2008; Zieritz et al. 2010) and other aquatic organisms (Costello et al. 2003; Finn et al. 2006; Alp et al. 2012). However, combinations between genetic and morphological distances have been rarely used to investigate

phenotypic plasticity in freshwater mussels (Zieritz et al. 2010). Our study revealed cryptic diversity within the *O. jacksoniana/V. arkansasensis* complex that has prompted additional investigations of genetic diversity, geographic distribution, and relative abundance to clearly define the taxonomic status of these clades. We believe that the results of this study encourage researchers and conservation agencies to use combinations of genetic, biological, and morphological information for developing conservation strategies of these highly imperiled animals.

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Conflict of Interest

None declared.

References

- Abell, R., M. L. Thieme, C. Revenga, M. Bryer, M. Kottelat, N. Bogutskaya, et al. 2008. Freshwater ecoregions of the world: a new map of biogeographic units for freshwater biodiversity conservation. *BioScience* 58:403–414.
- Allen, D. C., and C. C. Vaughn. 2009. Burrowing behavior of freshwater mussels in experimentally manipulated communities. *J. North Am. Benthol. Soc.* 28:93–100.

- Alp, M., I. Keller, A. M. Westram, and C. T. Robinson. 2012. How river structure and biological traits influence gene flow: a population genetic study of two stream invertebrates with differing dispersal abilities. *Freshw. Biol.* 57:969–981.
- Ball, G. H. 1922. Variation in fresh-water mussels. *Ecology* 3:93–121.
- Berendzen, P. B., A. M. Simons, R. M. Wood, T. E. Dowling, and C. L. Secor. 2007. Recovering cryptic diversity and ancient drainage patterns in eastern North America: historical biogeography of the *Notropis rubellus* species group (Teleostei: Cypriniformes). *Mol. Phylogenet. Evol.* 46:721–737.
- Berg, D. J., A. D. Christian, and S. I. Guttman. 2007. Population genetic structure of three freshwater mussel (Unionidae) species within a small stream system: significant variation at local spatial scales. *Freshw. Biol.* 52:1427–1439.
- Branson, B. A. 1984. The mussels (Unionacea: Bivalvia) of Oklahoma—part 3: Lampsilini. *Proc. Okla. Acad. Sci.* 64:20–36.
- Brönmark, C., and J. G. Miner. 1992. Predator-induced phenotypical change in body morphology in crucian carp. *Science* 258:1348–1350.
- Brown, K. M., W. Daniel, and G. George. 2010. The effect of Hurricane Katrina on the mussel assemblage of the Pearl River, Louisiana. *Aquat. Ecol.* 44:223–231.
- Burdick, R. C., and M. M. White. 2007. Phylogeography of the Wabash pigtoe, *Fusconaia flava* (Rafinesque, 1820) (Bivalvia: Unionidae). *J. Molluscan Stud.* 73:367–375.
- Campbell, D. C., and C. Lydeard. 2012a. Molecular systematics of *Fusconaia* (Bivalvia: Unionidae: Ambleminae). *Am. Malacol. Bull.* 30:1–17.
- Campbell, D. C., and C. Lydeard. 2012b. The genera of Pleurobemini (Bivalvia: Unionidae: Ambleminae). *Am. Malacol. Bull.* 30:19–38.
- Campbell, D. C., J. M. Serb, J. E. Buhay, K. J. Roe, R. L. Minton, and C. Lydeard. 2005. Phylogeny of North American amblesines (Bivalvia: Unionoida): prodigious polyphyly proves pervasive across genera. *Invertebr. Biol.* 124:131–164.
- Christian, A. D., J. L. Farris, J. L. Harris, and S. E. Seagraves. 2007. P. 67 in Life history and population biology of the state special concern Ouachita creekshell, *Villosa arkansasensis* (I. Lea 1862). Final report to the Arkansas Game and Fish Commission, Perrytown, Arkansas.
- Clement, M., D. Posada, and K. A. Crandall. 2000. TCS: a computer program to estimate gene genealogies. *Mol. Ecol.* 9:1657–1659.
- Costello, A. B., T. E. Down, S. M. Pollard, C. J. Pacas, and E. B. Taylor. 2003. The influence of history and contemporary stream hydrology on the evolution of genetic diversity within species: an examination of microsatellite DNA variation in bull trout, *Salvelinus confluentus* (Pisces: Salmonidae). *Evolution* 57:328–344.
- Crandall, K. A., and A. R. Templeton. 1999. The zoogeography and centers of origin of the crayfish subgenus *Procericambarus* (Decapoda: Cambaridae). *Evolution* 53:123–134.
- Dudgeon, D., A. H. Arthington, M. O. Gessner, Z.-I. Kawabata, D. J. Knowler, C. Leveque, et al. 2006. Freshwater biodiversity: importance, threats, status and conservation challenges. *Biol. Rev.* 81:163–182.
- Elderkin, C. L., A. D. Christian, C. C. Vaughn, J. L. Metcalfe-Smith, and D. J. Berg. 2007. Population genetics of the freshwater mussel, *Amblema plicata* (Say 1817) (Bivalvia: Unionidae): evidence of high dispersal and post-glacial colonization. *Conserv. Genet.* 8:355–372.
- Elderkin, C. L., A. D. Christian, J. L. Metcalfe-Smith, and D. J. Berg. 2008. Population genetics and phylogeography of freshwater mussels in North America, *Elliptio dilatata* and *Actinonaias ligamentina* (Bivalvia: Unionidae). *Mol. Ecol.* 17:2149–2163.
- Fetzner, J. W. Jr, and K. A. Crandall. 2003. Linear habitats and the nested clade analysis: an empirical evaluation of geographic versus river distances using an Ozark crayfish (Decapoda: Cambaridae). *Evolution* 57:2101–2118.
- Finn, D. S., D. M. Theobald, W. C. Black IV, and N. L. Poff. 2006. Spatial population genetic structure and limited dispersal in a Rocky Mountain alpine stream insect. *Mol. Ecol.* 15:3553–3566.
- Frierson, L. S. 1912. *Unio (Obovaria) jacksoniana*, new species. *Nautilus* 26:23–24.
- Gangloff, M. M., J. D. Williams, and J. W. Feminella. 2006. A new species of freshwater mussel (Bivalvia: Unionidae), *Pleurobema athearni*, from the Coosa River Drainage of Alabama, USA. *Zootaxa* 1118:43–56.
- Gaston, K. J., S. L. Chown, and K. L. Evans. 2008. Ecogeographical rules: elements of synthesis. *J. Biogeogr.* 35:483–500.
- Gatz, A. J. J. 2007. Homing ability of displaced stream-dwelling green sunfish. *J. Freshw. Ecol.* 22:403–409.
- Graf, D. L. 1998. Freshwater pearly mussels: pigtoes and Ortmann's Law. *Am. Conchol.* 26:20–21.
- Graf, D. L., and K. S. Cummings. 2007. Review of the systematics and global diversity of freshwater mussel species (Bivalvia: Unionoida). *J. Molluscan Stud.* 73:291–314.
- Greenberg, L. A. 1991. Habitat use and feeding behavior of thirteen species of benthic stream fishes. *Environ. Biol. Fishes* 31:389–401.
- Haag, W. R. 2009. A hierarchical classification of freshwater mussel diversity in North America. *J. Biogeogr.* 37:12–26.
- Haag, W. R. 2012. North American freshwater mussels: natural history, ecology, and conservation. Cambridge Univ. Press, Cambridge, U. K..
- Hammer, Ø., D. A. T. Harper, and P. D. Ryan. 2001. PAST: paleontological statistics software package for education and data analysis. *Palaeontol. Electronica* 4:1–9.
- Harris, J. L., W. R. Posey II, C. L. Davidson, J. L. Farris, S. R. Oetker, J. N. Stoeckel, et al. 2010. Unionoida (Mollusca: Margaritiferidae, Unionidae) in Arkansas, third status review. *J. Ark. Acad. Sci.* 63:50–86.

- Hinch, S. G., R. C. Bailey, and R. H. Green. 1986. Growth of *Lampsilis radiata* (Bivalvia: Unionidae) in sand and mud: a reciprocal transplant experiment. *Can. J. Fish. Aquat. Sci.* 43:548–552.
- Hornbach, D. J., V. J. Kurth, and M. C. Hove. 2010. Variation in freshwater mussel shell sculpture and shape along a river gradient. *Am. Midl. Nat.* 164:22–36.
- Howells, R. G., R. W. Neck, and H. D. Murray. 1996. Freshwater mussels of Texas. Texas Park and Wildlife Department, Austin, TX.
- Huelsenbeck, J. P., and F. Ronquist. 2001. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* 17:754–755.
- Jobb, G. 2008. TREEFINDER. Available at <http://www.treefinder.de/>. (accessed June 6, 2013)
- Jobb, G., A. von Haeseler, and K. Strimmer. 2004. TREEFINDER: a powerful graphical analysis environment for molecular phylogenetics. *BMC Evol. Biol.* 4:18.
- King, T. L., M. S. Eackles, B. Gjetvaj, and W. R. Hoeh. 1999. Intraspecific phylogeography of *Lasmigona subviridis* (Bivalvia: Unionidae): conservation implications of range discontinuity. *Mol. Ecol.* 9:S65–S78.
- Kuehnl, K. F. 2009. Exploring levels of genetic variation in the freshwater mussel genus *Villosa* (Bivalvia Unionidae) at different spatial and systematic scales: implications for biogeography, taxonomy, and conservation. Ohio State University, Columbus, OH.
- Lea, I. 1862. Descriptions of ten new species of Unionidae of the United States. *Proc. Acad. Nat. Sci. Philadelphia* 14:168–169.
- Lydeard, C., R. H. Cowie, W. F. Ponder, A. E. Bogan, P. Bouchet, S. A. Clark, et al. 2004. The global decline of nonmarine mollusks. *BioScience* 54:321–330.
- Mackie, G. L., and J. M. Topping. 1988. Historical changes in the unionid fauna of the Sydenham River watershed and downstream changes in shell morphometrics of tree common species. *Can. Field-Nat.* 102:617–626.
- Mantel, N. 1967. The detection of disease clustering and a generalized regression approach. *Cancer Res.* 27:209–220.
- Mayden, R. L. 1985. Biogeography of Ouachita highland fishes. *Southwest Nat.* 30:195–211.
- Mayden, R. L. 1988. Vicariance biogeography, parsimony, and evolution in North America freshwater fishes. *Syst. Zool.* 37:329–355.
- Minton, R. L., A. P. Norwood, and D. M. Hayes. 2008. Quantifying phenotypic gradients in freshwater snails: a case study in *Lithasia* (Gastropoda: Pleuroceridae). *Hydrobiologia* 605:173–182.
- Minton, R. L., E. M. Lewis, B. Netherland, and D. M. Hayes. 2011. Large differences over small distances: plasticity in the shells of *Elimia potosiensis* (Gastropoda: Pleuroceridae). *Int. J. Biol.* 3:23–32.
- NatureServe (2012) An online encyclopedia of life. Available at: <http://www.natureserve.org/explorer/> (accessed December 3, 2012).
- Neves, R. J. 1998. National strategy for the conservation of native freshwater mussels. *J. Shellfish Res.* 17:1419–1428.
- Oesch, R. D. 1995. Missouri naiades: a guide to the mussels of Missouri. Missouri Department of Conservation, Jefferson City, MO.
- Ortmann, A. E. 1918. The nayades (freshwater mussels) of the upper Tennessee drainage. With notes on synonymy and distribution. *Proc. Am. Philos. Soc.* 57:521–626.
- Ortmann, A. E. 1920. Correlation of shape and station in fresh-water mussels (Naiades). *Proc. Am. Philos. Soc.* 59:268–312.
- Peacock, E., and J. Seltzer. 2008. A comparison of multiple proxy data sets for paleoenvironmental conditions as derived from freshwater bivalve (unionid) shell. *J. Archaeol. Sci.* 35:2557–2565.
- Peakall, R., and P. E. Smouse. 2006. GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Mol. Ecol. Notes* 6:288–295.
- R Development Core Team. 2011. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rambaut, A., and A. J. Drummond. 2007. Tracer v1.4. Available at <http://beast.bio.ed.ac.uk/Tracer>. (accessed June 6, 2013).
- Ricciardi, A., and J. B. Rasmussen. 1999. Extinction rates of North American freshwater fauna. *Conserv. Biol.* 13:1220–1222.
- Robison, H. W., and R. T. Allen. 1995. Only in Arkansas: a study of the endemic plants and animals of the state. University of Arkansas Press, Fayetteville, AR.
- Roe, K. J., and C. Lydeard. 1998. Molecular systematics of the freshwater mussel genus *Potamilus* (Bivalvia: Unionidae). *Malacologia* 39:195–206.
- Roe, K. J., P. D. Hartfield, and C. Lydeard. 2001. Phylogeographic analysis of the threatened and endangered superconglutinate-producing mussels of the genus *Lampsilis* (Bivalvia: Unionidae). *Mol. Ecol.* 10:2225–2234.
- Saghai-Marouf, M. A., K. M. Soliman, R. A. Jorgensen, and R. W. Allard. 1984. Ribosomal DNA spacer-length polymorphisms in barley: Mendelian inheritance, chromosomal location, and population dynamics. *Proc. Natl Acad. Sci. USA* 81:8014–8018.
- Serb, J. M. 2006. Discovery of genetically distinct sympatric lineages in the freshwater mussel *Cyprogenia aberti* (Bivalvia: Unionidae). *J. Molluscan Stud.* 72:425–434.
- Serb, J. M., J. E. Buhay, and C. Lydeard. 2003. Molecular systematics of the North American freshwater bivalve genus *Quadrula* (Unionidae: Ambleminae) based on mitochondrial ND1 sequences. *Mol. Phylogenet. Evol.* 28:1–11.
- Shepherd, U. L. 1998. A comparison of species diversity and morphological diversity across the North American latitudinal gradient. *J. Biogeogr.* 25:19–29.
- Sokal, R. R., and F. L. Rohlf. 1995. Biometry: the principles and practice of statistics in biological research. W.H. Freeman and Company, New York, NY.

- Spear, S. F., C. R. Peterson, M. D. Matocq, and A. Storfer. 2005. Landscape genetics of the blotched tiger salamander (*Ambystoma tigrinum melanostictum*). *Mol. Ecol.* 14:2553–2564.
- Storfer, A., M. A. Murphy, J. S. Evans, C. S. Goldberg, S. Robinson, S. F. Spear, et al. 2007. Putting the 'landscape' in landscape genetics. *Heredity* 98:128–142.
- Strayer, D. L., and D. Dudgeon. 2010. Freshwater biodiversity conservation: recent progress and future challenges. *J. North Am. Benthol. Soc.* 29:344–358.
- Strayer, D. L., J. A. Downing, W. R. Haag, T. L. King, J. B. Layzer, T. J. Newton, et al. 2004. Changing perspectives on pearly mussels, North America's most imperiled animals. *BioScience* 54:429–439.
- Tamura, K., D. Peterson, N. Peterson, G. Stecher, M. Nei, and S. Kumar. 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum Parsimony methods. *Mol. Biol. Evol.* 28:2731–2739.
- Tanabe, A. S. 2011. Kakusan4 and Aminosan: two programs for comparing nonpartitioned, proportional and separate models for combined molecular phylogenetic analyses of multilocus sequence data. *Mol. Ecol. Resour.* 11:914–921.
- Turner, T. F., and H. W. Robison. 2006. Genetic diversity of the Caddo madtom, *Noturus taylori*, with comments on factors that promote genetic divergence in fishes endemic to the Ouachita highlands. *Southwest Nat.* 51:338–345.
- Turner, T. F., J. C. Trexler, J. L. Harris, and J. L. Haynes. 2000. Nested cladistic analysis indicates population fragmentation shapes genetic diversity in a freshwater mussel. *Genetics* 154:777–785.
- Utterback, W. I. 1917. Naiadogeography of Missouri. *Am. Midl. Nat.* 5:26–30.
- Vaughn, C. C., C. M. Mather, M. Pyron, P. Mehlhop, and E. K. Miller. 1996. The current and historical mussel fauna of the Kiamichi River, Oklahoma. *Southwest Nat.* 41:325–328.
- Via, S., R. Gomulkiewicz, G. De Jong, S. M. Scheiner, C. D. Schlichting, and P. V. Tienderen. 1995. Adaptive phenotypic plasticity: consensus and controversy. *Trends Ecol. Evol.* 10:212–217.
- Vrijenhoek, R. C. 2009. Cryptic species, phenotypic plasticity, and complex life histories: Assessing deep-sea faunal diversity with molecular markers. *Deep Sea Res. Part II* 56:1713–1723.
- Watters, G. T. 1994. Form and function of unionoidean shell sculpture and shape (Bivalvia). *Am. Malacol. Bull.* 11:1–20.
- Williams, J. D., M. L. Warren Jr, K. S. Cummings, J. L. Harris, and R. J. Neves. 1993. Conservation status of freshwater mussels of the United States and Canada. *Fisheries* 18:6–22.
- Williams, J. D., A. E. Bogan, and J. T. Garner. 2008. Freshwater mussels of Alabama and the mobile basin in Georgia, Mississippi & Tennessee. The University of Alabama Press, Tuscaloosa, AL.
- Xia, X., and Z. Xie. 2001. DAMBE: Software package for data analysis in molecular biology and evolution. *J. Hered.* 92:371–373.
- Xia, X., Z. Xie, M. Salemi, L. Chen, and Y. Wang. 2003. An index of substitution saturation and its application. *Mol. Phylogenet. Evol.* 26:1–7.
- Zanatta, D. T., and R. W. Murphy. 2006. Evolution of active host-attraction strategies in the freshwater mussel tribe Lampsilini (Bivalvia: Unionidae). *Mol. Phylogenet. Evol.* 41:195–208.
- Zanatta, D. T., and R. W. Murphy. 2007. Range-wide population genetic analysis of the endangered northern riffleshell mussel, *Epioblasma torulosa rangiana* (Bivalvia: Unionoida). *Conserv. Genet.* 8:1393–1404.
- Zelditch, M. L., D. L. Swiderski, H. D. Sheets, and W. L. Fink. 2004. Geometric morphometrics for biologists: a primer. Elsevier Academic Press, San Diego, CA.
- Zieritz, A., J. I. Hoffman, W. Amos, and D. C. Aldridge. 2010. Phenotypic plasticity and genetic isolation-by-distance in the freshwater mussel *Unio pictorum* (Mollusca: Unionoida). *Evol. Ecol.* 24:923–938.

Supporting Information

Additional Supporting Information may be found in the online version of this article:

Table S1. List of taxa and specimens utilized in this study. Numbers on map correspond to Fig. 2.

Table S2. List of published sequences with NCBI accession numbers and references utilized in this study.

Table S3. Bonferroni corrected *P*-values of the Hotelling's pairwise comparison. Clades in *Obovaria jacksoniana/Villosa arkansasensis* complex correspond to phylogenetic tree (Fig. 3).