ORIGINAL ARTICLE

ALOX12 polymorphisms are associated with fat mass but not peak bone mineral density in Chinese nuclear families

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Objective: Arachidonate 12-lipoxygenase (ALOX12) is a member of the lipoxygenase superfamily, which catalyzes the incorporation of molecular oxygen into polyunsaturated fatty acids. The products of ALOX12 reactions serve as endogenous ligands for peroxisome proliferator-activated receptor γ (PPARG). The activation of the PPARG pathway in marrow-derived mesenchymal progenitors stimulates adipogenesis and inhibits osteoblastogenesis. Our objective was to determine whether polymorphisms in the *ALOX12* gene were associated with variations in peak bone mineral density (BMD) and obesity phenotypes in young Chinese men.

Methods: All six tagging single-nucleotide polymorphisms (SNPs) in the *ALOX12* gene were genotyped in a total of 1215 subjects from 400 Chinese nuclear families by allele-specific polymerase chain reaction. The BMD at the lumbar spine and hip, total fat mass (TFM) and total lean mass (TLM) were measured using dual-energy X-ray absorptiometry. The pairwise linkage disequilibrium among SNPs was measured, and the haplotype blocks were inferred. Both the individual SNP markers and the haplotypes were tested for an association with the peak BMD, body mass index, TFM, TLM and percentage fat mass (PFM) using the quantitative transmission disequilibrium test (QTDT).

Results: Using the QTDT, significant within-family association was found between the rs2073438 polymorphism in the ALOX12 gene and the TFM and PFM (P=0.007 and 0.012, respectively). Haplotype analyses were combined with our individual SNP results and remained significant even after correction for multiple testing. However, we failed to find significant within-family associations between ALOX12 SNPs and the BMD at any bone site in young Chinese men.

Conclusions: Our present results suggest that the rs2073438 polymorphism of *ALOX12* contributes to the variation of obesity phenotypes in young Chinese men, although we failed to replicate the association with the peak BMD variation in this sample. Further independent studies are needed to confirm our findings.

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Keywords: ALOX12; fat mass; lean mass; peak bone mineral density; quantitative transmission disequilibrium test

Introduction

It was once thought that osteoporosis and obesity were unrelated,¹ but there is accumulating evidence confirming their association in recent decades. Weight is a major determinant of bone mineral density (BMD); subjects who are overweight or obese have a relatively higher BMD, and

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weight loss is associated with an increased risk of osteoporotic fracture.^{2,3} With aging, bone mass is reduced, whereas adipocyte volume in bone marrow is increased.⁴ Both osteoporosis and obesity have strong genetic components, as the heritability of obesity is estimated at 40–60%,⁵ and 60–80% of the variance in BMD is genetically determined.⁶ In addition, both osteoblasts and adipocytes are derived from mesenchymal stem cells. Extensive data have shown that both adipogenesis and osteogenesis share multiple common genetic factors and signaling pathways, such as vitamin D receptor, peroxisome proliferator-activated receptor γ (PPARG), transforming growth factor- β and Wnt pathways.^{1,7,8}. Meanwhile, adipocytes can secrete estrogen, leptin and inflammatory cytokines to influence bone metabolism.⁹ Therefore,

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exploring the interface between bone and fat at the molecular and genetic levels may improve our understanding of osteoporosis and obesity, which may lead to new interventions and treatments.

Body weight is determined mostly by fat, muscle and bone mass. Body composition assessment by dual-energy X-ray absorptiometry (DXA) is a readily accessible, inexpensive and non-invasive method. DXA detects total body mineral mass, total fat mass (TFM) and total lean mass (TLM). The relationships between bone mass and TFM and TLM have been examined, although the results are controversial.^{10–12} Studies have also shown several genomic regions that influence both obesity and osteoporosis.^{13,14} Although various candidate genes have been identified that may contribute to BMD, far less is known about genes affecting obesity phenotypes such as BMI, TLM and TFM.

Recently, Klein et al.¹⁵ suggested that mouse arachidonate 15-lipoxygenase (ALOX15) could negatively regulate peak bone mass. Compared with control mice, ALOX15 knockout mice have a higher BMD, and pharmacologic inhibitors of ALOX15 improve the BMD and bone strength in a model of osteoporosis. It has also been reported that overexpression of human ALOX15 in transgenic rabbits protects against bone loss.¹⁶ However, based on the products of their lipoxygenase reactions, mouse ALOX15 is functionally more similar to human ALOX12.^{17,18} Therefore, human ALOX12 might represent a strong candidate gene for susceptibility to osteoporosis. In fact, a positive correlation between ALOX12 gene polymorphisms and BMD in human beings has been established by previous studies,^{19–21} although all the studies were limited to Caucasian samples. So far, no such data have been reported for an Asian population. Moreover, most previous studies of candidate genes have been performed in women, with only one study focusing on men.

The human *ALOX12* gene is composed of 14 exons and 13 introns, mapping to chromosome 17p13, a region that is thought to be associated with body mass index (BMI) (http:// obesitygene.pbrc.edu). Furthermore, *ALOX12* has an important function in obesity-related complex phenotypes, including hypertension, diabetes, atherosclerosis and insulin secretion.^{22–24} However, the relationship between *ALOX12* and obesity has never been studied. In this study, we performed family-based-association analyses on all six tagging single-nucleotide polymorphisms (SNPs) in the *ALXO12* gene using the quantitative transmission disequilibrium test (QTDT) to determine whether SNPs and haplotypes in *ALXO12* were associated with peak BMD variations and obesity-related phenotypes in a large sample of Chinese males in nuclear families.

Materials and methods

Subjects

We recruited 1296 individuals from 427 Chinese nuclear families with male offspring from 2004 to 2007. The average

family size was 3.03, as 402 and 25 families had 1 and 2 children, respectively. Every study subject completed a questionnaire concerning age, sex, medical history, family history, marital status, physical activity, alcohol use, diet habits and smoking history. All male offspring were healthy. The following criteria were used to exclude individuals from the study: (1) serious effects from cerebrovascular disease; (2) diabetes mellitus; (3) chronic renal disease; (4) serious chronic liver disease or alcoholism; (5) significant chronic lung disease; (6) corticosteroid therapy at pharmacologic levels for >3 months; (7) treatment with anticonvulsant therapy for >6 months; (8) evidence of other metabolic or inherited bone diseases such as hyper- or hypoparathyroidism, Paget's disease of the bone, osteomalacia, osteogenesis imperfecta or others; (9) rheumatoid arthritis or collagen disease; (10) recent major gastrointestinal disease (within the past year) such as peptic ulcer, malabsorption, chronic ulcerative colitis, regional enteritis or any significant chronic diarrhea state; (11) significant disease of any endocrine organ that would affect bone mass; (12) hyperthyroidism and (13) any neurological or musculoskeletal condition that would be a non-genetic cause of low bone mass.

All study subjects belonged to the Chinese Han ethnic group. All were residents of Shanghai City, located on the mid-eastern coast of China. The study was approved by the ethics committee of the Shanghai Jiao Tong University Affiliated Sixth People's Hospital. All subjects signed informed consent documents before entering the study.

BMD and body composition measurements

The BMD of the lumbar spine 1-4, the left proximal femur (including the femoral neck and the total hip), as well as TFM and TLM were measured by DXA on a GE-LUNAR Prodigy (Lunar Corp., Madison, WI, USA) in fan-beam mode. All subjects were measured for BMD, and all sons were also measured for TFM and TLM. The machine was calibrated daily, and the coefficient of variability values of the DXA measurements (which were obtained from 15 individuals repeated three times) were 1.39% for the lumbar spine, 2.22% for the femoral neck and 0.70% for the total hip. The coefficient of variabilities were 1.18 and 3.72% for TLM and LFM, respectively. The long-term reproducibility of our DXA data during the trial, based on weekly repeated phantom measurements using standardized equipment, was 0.45%. BMI is defined as body weight/height² ($kg m^{-2}$). The percentage fat mass (PFM) is the ratio of TFM to body weight.

Genotyping

SNPs located within the *ALOX12* gene were selected from the NCBI LocusLink (http://Ncbi.nlm.nih.gov/LocusLink) and HapMap (http://hapmap.org). Polymorphisms spanning the *ALOX12* gene were selected from the SNPs resource based on the estimated pairwise linkage disequilibrium (LD), r^2 , between all common SNPs. The algorithm bins SNPs whose

pairwise LD exceeds a threshold r^2 of 0.8, tagging SNPs are selected from each bin to constitute a minimal set of highly informative markers, whereas minimizing redundant data. We selected SNPs on the basis of the following criteria: (1) degree of heterozygosity (minor allele frequencies > 0.10) and (2) classification as tagging SNPs. In total, six SNPs in the ALOX12 gene were selected: rs2073438, rs2292350, rs312470, rs434473, rs1235805 and rs312462. The study subjects were genotyped for all six SNPs using the TaqMan allelic discrimination assay (Applied Biosystems, Foster City, CA, USA), whereas primer and probe sequences were optimized using the SNP assay-by-design service of Applied Biosystems. Reactions were performed on the Mx3000P Real-Time PCR System (STRATAGENE, La Jolla, CA, USA). One allelic probe was labeled with FAM dye and the other with HEX dye. Genomic DNA (20ng) was amplified on 96-well plates in the presence of a $1 \times$ TaqMan probe assay and $1 \times$ TaqMan Universal PCR Master Mix (Applied Biosystems).

LD and haplotype analyses

Haplotypes were constructed from the population genotype data by the algorithm of Stephens using the Phase program, version 2.0.2. Significant levels of LD between the markers of this gene were assessed based on the observed haplotype and allele frequencies using Haploview 3.2.²⁵ We examined Lewontin's *D* and LD coefficient r^2 between all pairs of biallelic loci. The frequencies of genotypes and haplotypes were calculated using the unrelated parents of nuclear families.

Statistical analyses

The allele frequencies were estimated by gene counting. Hardy–Weinberg equilibrium was tested by a χ^2 goodnessof-fit statistic. To ensure unrelated individual samples, only genotype data from the parents of each nuclear family were used in the statistical analysis. The heritability estimate was performed using a linear regression of the parents' mean value and the offspring's value of every phenotype (described at http://www.heritability.com). The statistical power was estimated with Piface software (version 1.65) (http:// www.math.uiowa.edu/~rlenth/Power/) on our current sample size according to the minor allele frequency of every genotype and the variation of the BMD and obesity phenotypes. The QTDT program using the orthogonal model was used to test for population stratification, linkage and within-family association between SNPs and haplotypes and BMD phenotypes, TLM, TFM and BMI. The QTDT software package is available online (http://www.sph.umich.edu/csg/ abecasis/QTDT/). This method, as implemented in the QTDT software,²⁶ extends the trio-based TDT to quantitative trait data and uses genotype data from available siblings and parents. As in our nuclear families, all of the children were sons and the effects of parent phenotypes were excluded in the QTDT, sex was not used as a covariate to adjust the variations in the sons' bone phenotypes. The raw BMD values were adjusted by age, height and weight as covariates, and BMI, TFM and TLM were adjusted by age as covariates. As false-positive results might be generated in multiple tests, which were used in this study, permutations (1000 simulations) were performed to generate the empirical *P*-values^{26,27} to assess the reliability of the results. The QTDT program generates *P*-values for various tests through an asymptotic χ^2 distribution. *P*<0.05 was considered significant for all analyses.

In unrelated sons, the differences in the BMD among the genotype and haplotype groups were tested using general linear model ANOVA (GLM-ANOVA) adjustments for the confounding variables of age, height and weight. The differences in the BMI, TFM and TLM among the genotype and haplotype groups were performed using GLM-ANOVA adjusted for age. To evaluate the associations of the TFM and TLM with the BMD, we used Pearson's correlation analyses to examine the coefficient (r) and multiple linear regression to estimate the relative contribution of the TFM and TLM to the BMD at various sites in young men. Statistical analysis was performed using SPSS, version 11.0 (SPSS, Chicago, IL, USA).

Results

Basic characteristics of the study subjects

Of the males sampled from 427 nuclear families, 15 individuals could not be genotyped because of the poor quality of the DNA sample, and 12 sons deviated from Mendelian inheritance. Ultimately, there were 400 male nuclear families composed of 1215 individuals in this study. The basic characteristics of the study subjects are shown in Table 1. As the effects of parent phenotypes were excluded in the statistical analysis (both the QTDT and ANOVA), we only obtained the TFM and TLM of sons in this study. Peak BMD is thought to be under strong genetic control. In our sample, the heritability estimates for the peak BMD in the

Table 1	Basic characteristics	of the subjects	(mean ± s.d.)
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Variables	<i>Father</i> (n = 400)	<i>Mother</i> (n = 400)	Son (n = 415)
Age (years)	61.1 ± 7.1	58.4 ± 6.3	30.4 ± 6.1
Height (cm)	167.8 ± 6.0	155.7 ± 5.5	172.9 ± 5.9
Weight (kg)	69.7 ± 9.5	58.2 ± 8.2	70.7 ± 10.8
BMI (kg m^{-2})	25.2 ± 2.7	24.0 ± 3.1	24.2 ± 3.2
Lumbar spine BMD (g cm ⁻²)	1.139 ± 0.171	0.992 ± 0.168	1.138 ± 0.137
Femoral neck BMD (g cm ⁻²)	0.892 ± 0.132	0.796 ± 0.144	0.995 ± 0.141
Total hip BMD (g cm ^{-2})	0.958 ± 0.138	0.852 ± 0.162	1.008 ± 0.142
TFM (kg)	_	_	16.31 ± 7.56
TLM (kg)	_	_	51.43 ± 5.76
PFM (%)	—	—	21.89 ± 7.17

Abbreviations: BMD, bone mineral density; BMI, body mass index; PFM, percentage fat mass; TFM, total fat mass; TLM, total lean mass.

Relationship between the BMD and the TFM and TLM in young men

As shown in Table 2, Pearson's correlation analyses confirmed that both the TFM and TLM were positively correlated with the BMD, with correlations ranging from 0.198 to 0.508 for the lumbar spine, femoral neck and total hip BMD. According to the Pearson's coefficient (*r*), the TLM was more strongly related to the BMD at a given site. To determine the relative contributions of the TLM and TFM to the BMD, we constructed multiple regression models, which were simultaneously adjusted for age and height. Both the TLM and TFM were positively correlated

Table 2 Pearson's correlation coefficient (r) of the BMD with the TFM and TLM

Variation	Lumbar spine BMD (g cm ⁻²)	Femoral neck BMD (g cm ⁻²)	Total hip BMD (g cm ⁻²)	TFM (kg)	TLM (kg)
Lumbar spine	_	_	_	_	_
BMD ($g cm^{-2}$)	_	_	_	_	_
Femoral neck	0.639	_	_	_	_
BMD (g cm ^{-2})	(0.000)	_	_	_	_
Total hip	0.719	0.922	_	_	_
BMD (g cm ⁻²)	(0.000)	(0.000)	_	_	
Total fat	0.243	0.198	0.242	_	
mass (kg)	(0.000)	(0.000)	(0.000)	_	
Total lean	0.389	0.470	0.508	0.367	
mass (kg)	(0.000)	(0.000)	(0.000)	(0.000)	_

Abbreviations: BMD, bone mineral density; TFM, total fat mass; TLM, total lean mass. Statistical significance is indicated in parentheses with the relevant *P*-values.

 Table 3
 Multivariate regression analysis of the BMD against body composition components

BMD	Total lea	ın mass	Total fat mass		
	β	<i>R</i> ²	β	R ²	
Lumbar spine	0.010**	0.149	0.005**	0.056	
Femoral neck	0.012**	0.210	0.004**	0.051	
Total hip	0.012**	0.249	0.005**	0.068	

Abbreviations: BMD, bone mineral density; TFM, total fat mass; TLM, total lean mass. Models were adjusted for age and height. **P < 0.001.

Table 4	Information	on th	e ALOX12	SNPs	analyzed	in this	study

with the BMD, whereas the TLM had a significantly greater effect on the BMD than the TFM per kilogram of tissue mass (Table 3).

SNP genotyping and LD

Six SNPs in the *ALOX12* gene were genotyped; rs1235805 was excluded from further analysis because of the existence of homologous gene sequences, and too many families failed to pass the Mendelian inheritance check. The remaining five SNPs had a minor allele frequency of at least 0.15 and were in Hardy–Weinberg equilibrium (Table 4).

To gain insight into the pattern of LD between alleles at polymorphic loci, pairwise disequilibria measures (*D*) were calculated (Figure 1). We found that each of these SNPs was in strong LD (D > 0.80), and one block with high LD was identified. We inferred that 19 different haplotypes were present in our population using a likelihood method based on PHASE. The most common haplotype (AGAGG) had a frequency of 31.8%, and four haplotypes (AGAGG, GAAAG,



Figure 1 LD patterns for the *ALOX12* gene. The increasing degree of the darkness of the cells from white to black represents the increasing strength of the LD. The values in the cells are the pairwise degree of LD indicated by $D \times 100$ when D < 1.

Physical position	Location and function	Allele change	HWE P-value	MAF in European ^a	MAF in Asian ^a	MAF in this study
6 840 800	Intron 1	A>G	0.43	0.308	0.333	0.353
6842396	Intron 2	A > G	0.98	0.645	0.263	0.338
6842903	Intron 4	A>G	0.57	0.045	0.256	0.159
6845658	Exon 8 (Asn322Ser)	A>G	0.44	0.467	0.444	0.494
6 854 376	Exon 14 (Leu634Leu)	G > A	0.24	0.133	0.111	0.151
	Physical position 6 840 800 6 842 396 6 842 903 6 845 658 6 854 376	Physical position Location and function 6 840 800 Intron 1 6 842 396 Intron 2 6 842 903 Intron 4 6 845 658 Exon 8 (Asn322Ser) 6 854 376 Exon 14 (Leu634Leu)	Physical position Location and function Allele change 6 840 800 Intron 1 A>G 6 842 396 Intron 2 A>G 6 842 903 Intron 4 A>G 6 845 658 Exon 8 (Asn322Ser) A>G 6 854 376 Exon 14 (Leu634Leu) G>A	Physical position Location and function Allele change HWE P-value 6 840 800 Intron 1 A>G 0.43 6 842 396 Intron 2 A>G 0.98 6 842 903 Intron 4 A>G 0.57 6 845 658 Exon 8 (Asn322Ser) A>G 0.44 6 854 376 Exon 14 (Leu634Leu) G>A 0.24	Physical position Location and function Allele change HWE P-value MAF in European ^a 6 840 800 Intron 1 A>G 0.43 0.308 6 842 396 Intron 2 A>G 0.98 0.645 6 842 903 Intron 4 A>G 0.57 0.045 6 845 658 Exon 8 (Asn322Ser) A>G 0.44 0.467 6 854 376 Exon 14 (Leu634Leu) G>A 0.24 0.133	Physical position Location and function Allele change HWE P-value MAF in European ^a MAF in Asian ^a 6 840 800 Intron 1 A>G 0.43 0.308 0.333 6 842 396 Intron 2 A>G 0.98 0.645 0.263 6 842 903 Intron 4 A>G 0.57 0.045 0.256 6 845 658 Exon 8 (Asn322Ser) A>G 0.44 0.467 0.444 6 854 376 Exon 14 (Leu634Leu) G>A 0.24 0.133 0.111

Abbreviations: HWE, Hardy–Weinberg equilibrium; MAF, minor allele frequency; SNP, single-nucleotide polymorphism. ^aAccording to public information available in the dbSNP (http://www.ncbi.nlm.nih.gov) and the HapMap (http://www.Hapmap.org) databases.



GGGAG and GGAGA) accounted for 91.8% of the unrelated parents.

Association of the peak BMD and obesity phenotypes with SNPs in nuclear families

ANOVA and QTDT analyses. Using 400 nuclear families has a power of >80% to test a candidate gene as a QTL, which can explain $\sim 10\%$ of the variation in the BMD or obesity phenotypes. We investigated the association of the SNP genotypes with the BMD and obesity phenotypes using ANOVA in 400 unrelated sons randomly selected from 415 sons. Two SNPs (rs2073438 and rs312462) showed significant association with the TFM and PFM (P < 0.05). The sons with the AA genotype at rs2073438 had a higher adjusted TFM (P = 0.001) and a higher PFM (P = 0.014) compared with the GG genotype. This SNP contributed to 3.8 and 2.6% of the FM and PFM variation, respectively, in our sample. Individuals carrying the GG genotype at rs312462 had a higher TFM and PFM compared with GA carriers (P = 0.011 and 0.003, respectively). The contributions of rs312462 to the phenotypic variation of TFM and PFM were 3.0 and 3.7%,

respectively. However, no significant association was found between any SNP and the BMD, BMI or TLM variation in young men (Table 5).

The associations among the peak BMD, obesity phenotypes and SNPs in ALOX12 were analyzed using the QTDT for our nuclear families. As the effects of parent phenotypes were excluded in the analyses, all 400 families were included. There were 284, 273, 195, 310 and 189 informative families for the TDT analysis at rs2073438, rs2292350, rs312470, rs434473 and rs312462, respectively. We did not detect significant population stratification in our sample. The SNPs rs2073438 and rs312462 were significantly associated with the TFM and PFM in the total-association analysis (P < 0.01). For the within-family analysis, significant associations were found between rs2073438 and the TFM, PFM and BMI (*P*=0.007, 0.012 and 0.038, respectively), whereas rs312462 showed only marginal association with the TFM (P = 0.049). In the multiple parameters tests, permutations of 1000 tests were performed. However, only rs2073438 retained significant with-family associations with both the TFM and the PFM (P = 0.033 and 0.015, respectively). Similar to the results of the ANOVA, we failed to find significant associations

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Genotype	n	Lumbar spine BMD (g cm ⁻²)	Femoral neck BMD (g cm ⁻²)	Total hip BMD (g cm ⁻²)	ВМІ (kg m ⁻²)	TFM (kg)	PFM (%)	TLM (kg)
rs2073438								
GG	173	1.140 ± 0.142	1.002 ± 0.137	1.018 ± 0.137	23.5 ± 3.5	15.06 ± 6.83	20.90 ± 7.01	51.15 ± 5.32
AG	180	1.121 ± 0.132	0.995 ± 0.149	1.013 ± 0.143	23.7 ± 3.3	16.32 ± 6.85	22.13 ± 7.15	51.37 ± 5.99
AA	47	1.184 ± 0.136	0.983 ± 0.137	1.019 ± 0.124	24.1 ± 3.1	$18.37 \pm 6.93^{\#}$	$24.46 \pm 6.67^{\#}$	50.96 ± 5.33
P-value		0.076	0.579	0.696	0.597	0.001	0.015	0.918
rs2292350								
GG	168	1.139 ± 0.139	0.994 ± 0.139	1.017 ± 0.131	23.7 ± 3.3	16.49 ± 7.98	21.94 ± 6.99	51.47±5.45
AG	183	1.140 ± 0.133	1.000 ± 0.148	1.020 ± 0.149	23.8 ± 3.5	16.06 ± 7.11	21.99 ± 7.38	51.06 ± 5.86
AA	49	1.124 ± 0.150	0.989 ± 0.119	0.986 ± 0.107	22.8 ± 2.6	15.10 ± 6.42	21.16±7.18	51.01 ± 5.32
P-value		0.857	0.802	0.592	0.171	0.598	0.843	0.807
rs312470								
AA	278	1.138 ± 0.137	0.994 ± 0.139	1.011 ± 0.132	23.5 ± 3.2	16.23 ± 7.87	21.83 ± 7.53	51.31 ± 5.54
AG	111	1.146 ± 0.139	1.003 ± 0.149	1.026 ± 0.150	23.9 ± 3.6	16.14 ± 6.41	22.18 ± 6.36	51.06 ± 5.82
GG	11	1.138 ± 0.137	0.994 ± 0.139	1.011 ± 0.132	24.8 ± 3.4	14.12 ± 5.82	20.28 ± 6.41	50.97 ± 3.21
P-value		0.803	0.532	0.469	0.180	0.731	0.768	0.925
rs434473								
AA	104	1.138 ± 0.147	1.001 ± 0.139	1.007 ± 0.140	23.7 ± 3.3	15.69 ± 6.88	21.66 ± 6.92	50.98 ± 5.88
AG	210	1.135 ± 0.132	0.999 ± 0.144	1.016 ± 0.136	23.6 ± 3.3	15.85 ± 6.85	21.74 ± 7.22	51.25 ± 5.55
GG	86	$1.156 \pm \pm 0.138$	0.987 ± 0.144	1.014 ± 0.138	23.8 ± 3.6	17.45 ± 9.19	22.57 ± 7.42	51.47±5.57
P-value		0.560	0.519	0.535	0.874	0.203	0.608	0.822
rs312462								
GG	287	1.145 ± 0.140	0.998 ± 0.145	1.016 ± 0.139	23.9 ± 3.3	16.93 ± 7.61	22.67 ± 7.01	51.06 ± 5.71
GA	103	1.122 ± 0.126	0.985 ± 0.135	1.010 ± 0.134	23.2 ± 3.3	14.26 ± 6.69*	19.62 ± 7.01**	51.70 ± 5.52
AA	10	1.123 ± 0.178	1.107 ± 0.117	1.161 ± 0.139	22.1 ± 3.8	13.54 ± 6.88	24.10 ± 4.36	50.51 ± 3.21
P-value		0.234	0.254	0.144	0.632	0.003	0.001	0.570

Abbreviations: BMD, bone mineral density; BMI, body mass index; PFM, percentage fat mass; SNP, single-nucleotide polymorphism; TFM, total fat mass; TLM, total lean mass. The *P*-values are the results of analysis of variance (ANOVA) tests for the least-square mean of the BMD among various genotypes after adjusting for the significant covariates of age, weight and height. The *P*-values are the results of ANOVA tests for the least-square mean of the BMI, TFM and TLM among various genotypes after adjusting for age as a significant covariate. Bold indicates significant *P*-values (P<0.05). #P=0.001, AA genotype versus GG genotype; #P=0.014, AA genotype versus GG genotype; *P=0.011, GG genotype versus GA genotype; *P=0.003, GG genotype.

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(including the within-family association and 1000-permutation tests) between any SNPs and the BMD at the lumbar spine and hip, although rs2073438 was associated with the femoral neck BMD in the total-population analysis (P = 0.029) (Table 6).

 Table 6
 P-values of tests for population stratification, total association and within-family association using the QTDT.

	rs2073438	rs2292350	rs312470	rs434473	rs312462
Tests of population stra	atification				
Lumbar spine BMD	0.409	0.170	0.558	0.531	0.957
Femoral neck BMD	0.335	0.190	0.783	0.760	0.923
Total hip BMD	0.535	0.303	0.957	0.828	0.712
BMI	0.110	0.569	0.828	0.894	0.870
TFM	0.408	0.456	0.438	0.268	0.751
PFM	0.377	0.351	0.457	0.135	0.421
TLM	0.989	0.206	0.817	0.349	0.471
Test of total association	1				
Lumbar spine BMD	0.683	0.738	0.554	0.542	0.501
Femoral neck BMD	0.029	0.172	0.279	0.081	0.655
Total hip BMD	0.420	0.851	0.405	0.915	0.276
BMI	0.146	0.669	0.289	0.369	0.929
TFM	0.002	0.434	0.942	0.472	0.0012
PFM	0.007	0.668	0.758	0.744	0.001
TLM	0.331	0.405	0.853	0.977	0.284
Test of within-family as	sociation				
Lumbar spine BMD	0.359	0.293	0.845	0.399	0.814
Femoral neck BMD	0.826	0.585	0.479	0.585	0.909
Total hip BMD	0.892	0.411	0.700	0.809	0.844
BMI	0.038	0.489	0.781	0.624	0.907
TFM	0.007	0.283	0.608	0.192	0.049
PFM	0.012	0.323	0.755	0.184	0.117
TLM	0.492	0.669	0.766	0.474	0.846
P-value of 1000 permu	tations of the	within-family	, association		
Lumbar spine BMD	0.344	0.267	0.812	0.365	0.770
Femoral neck BMD	0.830	0.628	0.457	0.610	0.925
Total hip BMD	0.887	0.446	0.705	0.832	0.801
BMI	0.122	0.599	0.838	0.692	0.927
TFM	0.033	0.385	0.673	0.294	0.126
PFM	0.015	0.312	0.754	0.163	0.125
TLM	0.153	0.696	0.793	0.531	0.854

Abbreviations: BMD, bone mineral density; BMI, body mass index; PFM, percentage fat mass; QTDT, quantitative transmission disequilibrium test; TFM, total fat mass; TLM, total lean mass. The BMD values are adjusted for age, height and weight. The BMI, TFM and TLM values are adjusted for age. Bold indicates significant *P*-values (P<0.05).

Relationship of haplotypes and phenotypes

We conducted haplotype analyses using genotype data from all five SNPs, and four common haplotypes accounted for 91.8% of the unrelated parents. We further evaluated the association of haplotypes and the peak BMD with the obesity phenotypes using the QTDT. There were 252, 246, 164 and 146 informative families for the TDT analysis at haplotype 1 (AGAGG), haplotype 2 (GAAAG), haplotype 3 (GGGAG) and haplotype 4 (GGAGA), respectively. We failed to find any evidence of population stratification for all haplotypes (P>0.05). For the total and within-family associations, a significant association was found between the most common haplotype (haplotype 1) and the TFM and PFM (both P < 0.05). Furthermore, haplotype 1 was still significantly associated with the PFM after 1000 permutations (corrected $P_c = 0.033$), but not with the TFM (corrected $P_c = 0.092$). The results of the haplotype 1-association analysis are presented in Table 7.

In addition, we also investigated the association between haplotype 1 and the BMD and obesity phenotypes in 400 unrelated sons using GLM-ANOVA. The subjects carrying at least one copy of haplotype 1 had a higher TFM and PFM than non-carriers (P=0.006 and 0.004, respectively). This haplotype accounted for 2.5 and 2.7% of the variation in the TFM and PFM, respectively, in this sample of younger males. However, no significant association was found between any haplotype and the BMI, TLM or peak BMD at any site (data not shown).

Discussion

To the best of our knowledge, this is the first study to investigate the possible influence of SNPs and haplotypes in the *ALOX12* gene on variations in obesity-related phenotypes. Our results showed that the polymorphism rs2073438 of the *ALOX12* gene is significantly associated with the TFM and PFM, but not with the BMI or TLM in young men. Furthermore, haplotype-based analyses supported this significant association from the single-locus analyses. The SNP rs2073438, with a polymorphism of G > T, is located in the

Table 7	The QTDT r	results for the	associations	between	haplotype 1	and	phenotypic variations
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Haplotype 1 (AGAGG)	Test of population stratification	Test of total association	Test of within-family association	P-value of 1000 permutations of the within-family association
Lumbar spine BMD	0.508	0.853	0.502	0.456
Femoral neck BMD	0.347	0.019	0.835	0.840
Total hip BMD	0.630	0.353	0.909	0.912
BMI	0.848	0.431	0.856	0.853
TFM	0.892	0.0001	0.021	0.092
PFM	0.721	0.003	0.034	0.033
TLM	0.284	0.632	0.256	0.275

Abbreviations: BMD, bone mineral density; BMI, body mass index; PFM, percentage fat mass; QTDT, quantitative transmission disequilibrium test; TFM, total fat mass; TLM, total lean mass. The BMD values are adjusted for age, height and weight. The BMI, TFM and TLM values are adjusted for age. Bold indicates significant *P*-values (*P*<0.05).

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intron1/exon2 boundary of ALOX12, only 639 base pairs from the 5' promoter region. Moreover, based on the information from HapMap, the criterion for tagging was set at $r^2 > 0.5$, and rs2073438 as a tagging SNP captured seven additional SNPs. Therefore, the significant association of rs2073438 with the TFM and PFM suggests that the SNP itself or the region around it may be important for transcription initiation efficiency or may be in strong LD with certain functional variants influencing the expression of the ALOX12 protein. Further study is needed to clarify how this intronic SNP affects protein function. Although significant associations were observed between rs312462 and the TFM and PFM using ANOVA, we failed to find significant results in our QTDT analyses (both within-family and 1000-permutation tests). As the QTDT analysis is free of confounding population-substructure effects compared with the traditional-association approach, the results of our study should be more valuable because of the robustness of the TDT approach.²⁸ A possible reason for our result is that the heterozygosity of rs312462 is relatively low, and the statistical power of a family-based-association study depends directly on the degree of allelic heterozygosity.²⁹

Interestingly, we found a significant association between rs2073438 and the TFM, but not the BMI. The BMI has been widely used as a surrogate phenotype for the assessment of obesity status. However, the BMI alone may not accurately reflect the percentage of body fat and the relative contributions of muscle and fat,³⁰ and as an index of body fatness, BMI is significantly influenced by age and sex.³¹ The subjects of this study were healthy young men aged 20-40 years. The TLM in our sample accounted for $\sim 70\%$ of the body mass: we speculate that the BMI in this study may be skewed by the TLM so that the BMI values of these young Chinese men do not accurately reflect body fat mass. Our results are in accordance with the study of Liu et al., 32 in which the polymorphisms of the MTHFR gene were significantly associated with the BMI; however, the association diminished after adjusting for the LM. Therefore, the FM as measured by DXA is a more refined measure of obesity.

ALOX12 belongs to the lipoxygenase superfamily, which catalyzes the insertion of molecular oxygen into polyunsaturated fatty acids. It has become increasingly clear that polyunsaturated fatty acids have an important function in the control of adipogenesis and reduce the expression of many enzymes involved in lipid metabolism.^{33,34} Although the exact mechanisms by which polyunsaturated fatty acids exert their effects are not completely understood, several transcription factors and nuclear receptors have been identified as critical regulators of several important genes of lipid metabolism. PPARG, a nuclear transcription factor, is a dominant regulator of adipocyte-specific genes contributing to adipocyte differentiation, susceptibility to obesity and insulin sensitivity.^{35,36} The substrates of ALOX12 (arachidonic acids) and the products of the lipoxygenase reaction are both effective PPARG activators, and ligand activation of PPARy promotes the differentiation of mesenchymal stem cells into adipocytes. Studies have shown that ALOX12 has an important function in obesity-related diseases, such as hypertension, diabetes and atherosclerosis.^{22–24} More recently, Mehrabian *et al.*³⁷ showed that ALOX5, another member of the lipoxy-genase superfamily, has pleiotropic effects on adiposity and type 2 diabetes-related traits. Our results, taken together with the above studies, suggest the involvement of SNP rs2073438 and the AGAGG haplotype in *ALOX12* in the variation of obesity, perhaps through a PPARG-dependent pathway.

To date, >200 different candidate genes have been associated with obesity-related phenotypes, 38 and most of those genes regulate the energy balance through the complex central nervous system.³⁹ The mRNA and protein of ALOX12 has been described mainly in neurons and also in some glial cells throughout the cerebrum, basal ganglia and hippocampus.^{40,41} Recently, Pratico *et al.*⁴² showed that the protein levels and activity of ALOX12 in affected frontal and temporal regions of Alzheimer's disease brains were significantly increased. The levels of the metabolic products of ALOX12 were also markedly elevated in cerebrospinal fluid from mild cognitive impairment and Alzheimer's disease brains.⁴³ In these studies, they believed that ALOX12 metabolic pathway increased oxidative stress and inflammatory responses, which is likely to have a more active function in the pathogenesis of Alzheimer's disease. Obesity was determined as a state of chronic oxidative stress and oxidative stress might be a major mechanism underlying the development of obesity-related co-morbidities.^{44,45} We suspected that there may exist ALOX12 overexpression in central nervous system similar to Alzheimer's disease, to affect the energy balance. However, all of the above are just hypotheses and wait to be tested by further functional analyses.

Polymorphism in ALOX12 has been associated with the BMD in several independent studies. Xiong et al.19 first performed an association analysis of 20 osteoporosis candidate genes on the BMD variation at three clinically important skeletal sites: the spine, hip and ultradistal radius. Their findings show that ALOX12 is a suggestive gene $(0.01 < P \le 0.05)$ for the BMD at the hip and ultradistal radius in the total-population sample. However, the significant association was mainly driven by the female subjects, suggesting the possibility of a sex-specific association of ALOX12 with the BMD. Extensive evidence suggests that the loci that regulate the BMD do so in a sex-specific manner in both human beings and experimental animals.^{19,46} The sex specificity of the associations between genes and the BMD may involve the regulation of sex steroids. In addition, Mullin *et al.*²⁰ also found that three SNPs from *ALOX12* were significantly associated with the BMD at the spine and hip in postmenopausal Caucasian women. In this study, we studied 400 Chinese nuclear families with male offspring and used TDT to simultaneously test the linkage and association of ALOX12 gene polymorphisms with the peak BMD. No significant within-family associations were found between ALOX12 SNPs or haplotypes and the peak BMD in young

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men (aged 20–40 years). However, Ichikawa *et al.*²¹ showed evidence of an association between the spine BMD and six SNPs in the *ALOX12* gene in both White men (aged 18–61 years) and women (aged 20–50 years). The conflicting results may be due to the differences between the participants, especially their ages, ethnicities or some other confounding issue. This is the first study to investigate the possible influence of *ALOX12* SNPs and haplotypes on the BMD variation in a non-Caucasian population. Therefore, our results should be interpreted cautiously, and further studies should be conducted to determine the strength of these associations in various populations.

We also examined the relationship between the BMD and the TFM and TLM in young men. Our results showed that both the TFM and the TLM had significant positive associations with the BMD, and the TLM was a stronger predictor of the BMD than the TFM, consistent with previous studies.^{11,47} However, Cui *et al.*¹⁰ reported that the LM was the only independent factor contributing to the BMD, and the FM was negatively related to the BMD in younger men. In contrast, other independent studies have suggested that the TFM has a stronger effect on the BMD in both pre- and post-menopausal women.⁴⁸ The influence of body composition on the BMD may be confounded by gender, age, menopausal status and other environmental factors.⁴⁹

Our study has several strengths. First, we chose all six tagging SNPs of the *ALOX12* gene. The extensive LD between SNPs guaranteed that the entire gene had been tested for an association with the BMD and obesity phenotypes. Second, our sample was relatively large, composed of 1215 individuals from 400 nuclear families. Therefore, adequate statistical power was ensured to find the genetic variants of modest effect sizes. Third, we performed the QTDT and 1000-permutation analyses to avoid sample heterogeneity and false-positive results because of multiple tests. Our study has several limitations as well. Our nuclear families contained few sibling pairs, and no linkage was detected in this study. Environmental factors such as dietary fat intake were not considered in this study.

In conclusion, we have shown that a genetic variation (rs2073438 and haplotype AGAGG) in *ALOX12* is associated with the TFM in young Chinese men, but we failed to observe a significant association between SNPs or haplotypes and the peak BMD at any site. Our findings suggest that *ALOX12* is potentially important in the pathogenesis of human obesity. Although the mechanisms underlying this relationship are largely unknown, we speculate that ALOX12 may exert its effect on fat mass variation by activating the PPARG pathway or affecting central nervous system to regulate energy homeostasis.

Conflict of interest

The authors declare no conflict of interest.

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