

Original Article

Mining Therapeutic Efficacy from Treasure Chest of Biodiversity and Chemodiversity: Pharmacophylogeny of Ranunculales Medicinal Plants*

HAO Da-cheng¹, XU Li-jia², ZHENG Yu-wei¹, LYU Huai-yu¹, and XIAO Pei-gen²

ABSTRACT Ranunculales, comprising of 7 families that are rich in medicinal species frequently utilized by traditional medicine and ethnomedicine, represents a treasure chest of biodiversity and chemodiversity. The phylogenetically related species often have similar chemical profile, which makes them often possess similar therapeutic spectrum. This has been validated by both ethnomedicinal experiences and pharmacological investigations. This paper summarizes molecular phylogeny, chemical constituents, and therapeutic applications of Ranunculales, i.e., a pharmacophylogeny study of this representative medicinal order. The phytochemistry/metabolome, ethnomedicine and bioactivity/pharmacology data are incorporated within the phylogenetic framework of Ranunculales. The most studied compounds of this order include benzyloisoquinoline alkaloid, flavonoid, terpenoid, saponin and lignan, etc. Bisbenzyloisoquinoline alkaloids are especially abundant in Berberidaceae and Menispermaceae. The most frequent ethnomedicinal uses are arthritis, heat-clearing and detoxification, carbuncle-abscess and sore-toxin. The most studied bioactivities are anticancer/cytotoxic, antimicrobial, and anti-inflammatory activities, etc. The pharmacophylogeny analysis, integrated with both traditional and modern medicinal uses, agrees with the molecular phylogeny based on chloroplast and nuclear DNA sequences, in which Ranunculales is divided into Ranunculaceae, Berberidaceae, Menispermaceae, Lardizabalaceae, Circaeasteraceae, Papaveraceae, and Eupteleaceae families. Chemical constituents and therapeutic efficacy of each taxonomic group are reviewed and the underlying connection between phylogeny, chemodiversity and clinical uses is revealed, which facilitate the conservation and sustainable utilization of Ranunculales pharmaceutical resources, as well as developing novel plant-based pharmacotherapy.

KEYWORDS Ranunculales, chemical constituent, bioactivity, diversity, pharmacophylogeny

Ranunculales is an order of angiosperms, containing 7 families, i.e., Ranunculaceae (at least 50 genera, >2,000 species), Berberidaceae (17 genera, 650 species), Menispermaceae (65 genera, >350 species), Lardizabalaceae (9 genera, 50 species), Circaeasteraceae (2 species), Papaveraceae (38 genera, >700 species), and Eupteleaceae (2 species, Appendix 1). Ranunculales belongs to the basal eudicots, in which it is the most basal clade; its widely known members include poppies, barberries, monseed, akebia and buttercupss. The chemodiversity of Ranunculaceae, the flagship family of Ranunculales, and their correlation with the biodiversity and pharmacotherapy, have been systematically reviewed by us in the context of pharmacophylogeny.⁽¹⁾ In recent years, the research enthusiasm for the other 4 major Ranunculales families has been growing rapidly, as plants of these families also meet human needs for food, flavours, fragrances, and not the least, medicines during long history. Ranunculales plants form the basis of sophisticated traditional medicine systems, e.g., Chinese medicine (CM) and Ayurveda medical system.⁽²⁾ These systems of medicine give rise to some important Ranunculales-based drugs still in use today.^(3,4) According to incomplete statistics, 573 Ranunculaceae and 308 Berberidaceae species

were in medicinally hot nodes;⁽⁴⁾ the therapeutic efficacy of 2 families differed substantially. For instance, up to 361 Chinese Ranunculaceae species are used against musculoskeletal diseases, followed by 321 against hepatic diseases, 242 against circulatory diseases, and 210 against skin diseases, etc.; in contrast, as many as 251 Chinese Berberidaceae species are used against eyesight diseases, followed by 249 against hepatic diseases, 242 against digestive diseases and 90 against oral diseases, etc. Nevertheless, except Ranunculaceae, the

©The Chinese Journal of Integrated Traditional and Western Medicine Press and Springer-Verlag GmbH Germany, part of Springer Nature 2022

*Supported by the National Science and Technology Fundamental Resources Investigation Program of China (No. 2018FY100700) and CAMS Innovation Fund for Medical Sciences (No. CIFMS 2021-I2M-1-032)

1. Biotechnology Institute, School of Environment and Chemical Engineering, Dalian Jiaotong University, Dalian (116028), China; 2. Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences, Beijing (100193), China

Correspondence to: Prof. XIAO Pei-gen, E-mail: pgxiao@implad.ac.cn

DOI: <https://doi.org/10.1007/s11655-022-3576-x>

phytochemistry/metabolome, ethnomedicine and bioactivity/ pharmacology data of other Ranunculales families have not been incorporated within the phylogenetic framework of Ranunculales and related basal eudicots. Ethnobotany and ethnopharmacognosy lend support to the nowadays search for new molecules of different sources and classes. The flora of the tropics, Southwest China and Himalayas is fantastic and contains the most diverse Ranunculales taxa.^(5,6) The biodiversity of these hotspot regions plays a significant role in providing new drug leads for the innovative and improved clinical treatments, although the sovereignty and property rights should be addressed along with the convention for biological diversity.

The present study highlights the recent progress of Berberidaceae, Menispermaceae, Lardizabalaceae and Papaveraceae, provides an overview of the classes of molecules present in Ranunculales plants and gives examples of the types of molecules and secondary metabolites that lead to the development of pharmacologically active extracts/ preparations. We assume that the phylogenetic framework is helpful to recognize the distribution law of compounds and efficacy, and to analyze the cryptic link between bioactive compounds and clinical efficacy. Only by mastering the distribution regularity of compound bioactivity/toxicity can Ranunculales plant products be used to develop functional foods. This study simultaneously focuses on the front, middle and back of pharmacophylogeny, trying to find a trade-off. The pharmacological validation of Ranunculales extracts/ preparations is indispensable, and safety, efficacy and quality of phyto-medications should always be emphasized.

Systematics and Evolution of Ranunculales

Around 75% of all angiosperm species belong to the eudicot clade, which is strongly supported by molecular data and the morphology of single synapomorphy-triaperturate pollen. Ranunculales is a basal clade of eudicots, which is sister to all other eudicots (Appendix 2),⁽⁷⁾ e.g., Saxifragales, Caryophyllales, rosids, and asterids. Despite the progress in resolving angiosperm phylogenetic relationships, the branching order among basal eudicots remains an issue; while fossils should be integrated with extant taxa into a comprehensive tree of angiosperm phylogeny, the chemotaxonomic evidence is also essential.⁽⁸⁾ Many phylogenetic studies suggest that Eupteleaceae, an East Asian family of 2 tree species with perianthless flowers, are sister to all remaining Ranunculales, with Papaveraceae branching next (Appendix 1B).⁽⁹⁻¹¹⁾ Lardizabalaceae and Circaeasteraceae cluster together, which are basal to the clade containing Menispermaceae, Berberidaceae and Ranunculaceae; Berberidaceae and Ranunculaceae are closer to each other than to other families.

All the lineages of basal eudicots emerged during the latest part of the Early Cretaceous.⁽¹²⁾ Among early-diverging eudicots, the age of Ranunculales was around 120 my, older than Proteales (119 my), Sabiales (118 my), Buxales (117 my) and Trochodendrales (116 my). Recently

the complete plastome sequences of many early-diverging eudicot taxa, e.g., *Epimedium sagittatum* (Berberidaceae), *Euptelea pleiosperma* (Eupteleaceae), *Akebia trifoliata* (Lardizabalaceae), *Stephania japonica* (Menispermaceae) and *Papaver somniferum* (Papaveraceae), are used to elucidate the evolution of plastome structure and the phylogenetic correlation.⁽¹¹⁾ The maximum likelihood phylogenetic analysis of a 79-gene, 97-taxon data set that included all available early-diverging eudicots largely agreed with previous estimates of phylogenetic relationships (Appendix 2).

Menispermaceae consists of 2 subfamilies: Tinosporoideae and Menispermoideae (Appendix 1A).⁽¹³⁾ Within Tinosporoideae, tribe Coscineae is basal; within Menispermoideae, tribe Menispermeae is basal. Tinosporoideae taxa usually have apical style scars, bilateral curvature, subhemispherical condyles, and foliaceous cotyledons with divaricate or imbricate orientation. Menispermoideae taxa generally have basal or subbasal style scars, dorsoventral curvature, bilaterally and/or dorsoventrally compressed condyles, and subterete or fleshy cotyledons oriented dorsoventrally or laterally. Most of the genera with more pharmaceutical research belong to Menispermoideae. In Papaveraceae (Appendix 1B), the subfamily Fumarioideae (20 genera, 593 species) is characterized by flowers that are either disymmetric (i.e. 2 perpendicular planes of bilateral symmetry) or zygomorphic (i.e. 1 plane of bilateral symmetry). In contrast, the subfamily Papaveroideae (23 genera, 230 species) has actinomorphic flowers (i.e. more than 2 planes of symmetry). Six plastid markers and 1 nuclear marker were used to infer the phylogenetic relationship of *Pteridophyllum*, 73 species of Fumarioideae and 11 species of Papaveroideae.⁽¹⁴⁾ *Pteridophyllum* is not nested in Fumarioideae. Fumarioideae are monophyletic and *Hypecoum* (18 species) is the sister group of the remaining genera. Relationships within the core Fumarioideae are well resolved and supported (Appendix 1B). *Dactylicapnos* and zygomorphic genera, e.g., *Fumaria* and *Corydalis*, form a well-supported clade nested among disymmetric taxa. As compared with Fumarioideae, more Papaveroideae taxa are medicinally studied.

Chemical Composition of Berberidaceae

Phylogenetically and phytochemically, Berberidaceae is closer to Ranunculaceae⁽¹⁾ than to other Ranunculales families (Appendix 1). It is divided into 3 subfamilies: Nandinoideae, Berberidoideae, and Podophylloideae. Nandinoideae is characterized by a rich spectrum of benzyloisoquinoline alkaloids (BIAs). The cyanogenic nandinin, biflavonoid amentoflavone and benzaldehyde-4-O-glucoside in this family indicates its relatively distant relation with other subfamilies.⁽¹⁵⁾ Berberidoideae contains mainly BIAs, particularly a higher content of bisbenzyloisoquinoline (BBI) represented by berbamine and oxyacanthine. Podophylloideae is divided into 2 tribes. The tribe Podophylleae contains extensively various podophyllotoxin lignans, while the tribe Epimediaceae has diverse constituents such as icariin flavonoids.

Gymnospermium, *Leontice*, *Caulophyllum* and *Bongardia* contain β -amyrin triterpenoids and quinolizidine alkaloids.

Nandinoideae Subfamily

Phylogenetically, the genus *Nandina* is basal to *Caulophyllum* and *Gymnospermium* (Appendix 1); it is rich in various BIAs, e.g., berberine, palmatine, jatrorrhizine, coptisine, magnoflorine, domesticine, nandinine and protopine. Steroidal alkaloid, e.g., nandsterine, is also found in the fruit (Appendix 3).⁽¹⁶⁾ Pyrrole alkaloids methyl-E-mangolamide and methyl-Z-mangolamide, megastigmane glycosides nandinamegastigmanes I–IV were isolated from the fruits of *Nandina domestica*.⁽¹⁷⁾ Phenolic 1-benzyl-N-methyltetrahydroisoquinolines are present in cell cultures of *Corydalis*, *Macleaya*, and *Nandina*.⁽¹⁸⁾ The benzyltetrahydroisoquinoline alkaloid higenamine exists in *Aconitum*,⁽¹⁾ *Tinospora crispa*, *N. domestica*,⁽¹⁹⁾ *Gnetum parvifolium*, *Asarum heterotropoides*, and *Nelumbo nucifera*. The existence of the cyanogenic compound nandinin,⁽²⁰⁾ biflavonoid amentoflavone and benzaldehyde-4-O-glucoside in *Nandina* indicates its relatively distant relation with other subfamilies. The cycloartane-type triterpenoid was found in the fruits of *Nandina*.⁽²¹⁾

Magnoflorine, taspine, and boldine, belonging to aporphine alkaloids, are found in *Caulophyllum*.⁽²²⁾ Typical quinolizidine alkaloids are also present in this genus. Alkaloids with piperidine-acetophenone conjugates, rare in the plant kingdom, are only found in *Caulophyllum* and *Boehmeria*. Fluorenone alkaloids and one dihydroazafluoranthene alkaloid were isolated from the roots of *Caulophyllum robustum*.⁽²³⁾ The piperidines and N-containing xanthone derivative were isolated from the roots of *C. robustum*.⁽²⁴⁾ Seven classes of triterpene saponins, including malonyl-triterpene saponins, were isolated from *C. robustum*.^(25,26) Twelve kinds of aglycones are discovered, e.g., oleanolic acid, hederagenin, echinocystic acid, caulophyllogenin, and erythrodiol.

Berberidoideae

Berberis and *Mahonia* contain mainly BIAs, e.g., berberine, palmatine, jatrorrhizine, columbamine (protoberberine type), magnoflorine (aporphine type), particularly a higher content of BBI alkaloid berbamine,⁽²⁷⁾ isotetrandrine⁽²⁸⁾ and oxyacanthine. Phenolics, flavonoids, and tannins are abundant in *Berberis*,⁽²⁹⁾ and the number of type VIII BBI is the most (Appendix 4), followed by types IV, I, VII and XI. In *Mahonia*, the number of type VIII BBI is more than that of type VII. Berberine is the principal component for many medicinal plants, e.g., *Coptis chinensis*, *Phellodendron chinense*, and *Mahonia bealei*. The stems of *Mahonia fortunei* contain multiple bioactive compounds,⁽³⁰⁾ including 3,4,5-trimethoxyphenol-1-O- β -D-glucopyranoside, 5-hydroxypicolinic acid methyl ester, acortatarin A, syringic acid, 9-epi-acortatarin A, vomifoliol, corydaldine, noroxyhydrastinine, columbamine, jatrorrhizine, palmatine, berberine and schisandrin.

Podophylloideae

The tribe Podophylleae, consisting of *Podophyllum*

(including *Sinopodophyllum* and *Dysosma*) and *Diphylleia*, is rich in various podophyllotoxin lignans.⁽¹⁵⁾ Deoxypodophyllotoxin (DPPT) is extracted from *Podophyllum peltatum*, *P. pleianthum*, *P. emodi* (*P. hexandrum*) and *Diphylleia grayi*.⁽³¹⁾ The major lignans of *Diphylleia sinensis* are podophyllotoxin, epipodophyllotoxin, and 4'-demethylpodophyllotoxin.⁽³²⁾ Prenylated flavonoids, flavonoid glycoside, and labdane diterpenes were isolated from the fruits of *Sinopodophyllum hexandrum*.^(33,34) Prenylated biflavonoids were isolated from the roots and rhizomes of *Sinopodophyllum*.⁽³⁵⁾ The aporphine alkaloids, magnoflorine, corytuberine and muricinine, were found in the underground and aerial tissues of *Podophyllum hexandrum*.⁽³⁶⁾

Both *Epimedium* and *Vancouveria* contain predominately bioactive icariin flavonoids, the characteristic chemical constituents of this group. Neolignans, e.g., 8-O-4' neolignan, were isolated from *E. pseudowushanense*.⁽³⁷⁾ *Gymnospermium*, *Leontice*, *Caulophyllum* and *Bongardia*, contain mainly β -amyrin triterpenoids and quinolizidine alkaloids. Aporphine BIAs were found in at least 6 Berberidaceae genera (Appendix 1), e.g., *Epimedium*, *Podophyllum*, *Mahonia*, *Berberis*, *Caulophyllum* and *Nandina*.

Chemical Composition of Menispermaceae

Menispermaceae is a medium-sized family of 70 genera and 420 extant species, mostly climbing plants. It has various medicinal properties, which are used in CM and the Ayurvedic system of medicine. Menispermaceae plants are rich in alkaloids, especially BBI type (Appendix 5).⁽³⁸⁾ The number of type VIII is the most, followed by VI, XXIII, XX and XXI (Appendixes 6 and 4). BBI alkaloids, morphine alkaloids, aporphine alkaloid, syringaresinol (lignan) and aristolochic acid I (organic acid) could be marker compounds of this family.⁽³⁹⁾

Stephania

Phylogenetically, *Stephania* is closer to *Cyclea* and *Cissampelos* than to *Cocculus*, *Sinomenium* and *Menispermum* in subfamily Menispermoideae (Appendix 1). Interestingly, *Stephania* and *Cyclea/Cissampelos* share 5 types of BBIs (Appendix 4), while it and *Cocculus* share 4 BBIs; these 4 genera share type VI, VIII and XX BBIs. The alkaloids of *S. rotunda* were classified into 9 structural families: phenanthrene, quinoline, isoquinoline, benzyltetrahydroisoquinoline, protoberberine, tetrahydroprotoberberine, aporphine, oxoapomorphine, and morphinane derivative, etc.⁽⁴⁰⁾ Alkaloids of *S. tetrandra* were classified into 6 structural classes,⁽⁴¹⁾ including monobenzyltetrahydroisoquinoline, BBI, aporphine, protoberberine and tetrahydroprotoberberine, etc. An aporphine glycoside angkorwatine and 8 alkaloids: oblongine, stepharine, asimilobine- β -D-glucopyranoside, isocorydine, tetrahydropalmatine (THP), jatrorrhizine, palmatine, and roemerine were isolated from the tuber of *S. cambodica*.⁽⁴²⁾

Cyclea and Cissampelos

Azafluoranthene alkaloids and phytoecdysone were

isolated from the stems of *Cyclea barbata*.⁽⁴³⁾ Cissampentine A, an enantiomer of cissampentin, and cycleatjehenine-type BBI alkaloids, cissampentine B–D, were isolated from the roots of *C. tonkinensis*.⁽⁴⁴⁾ Curine-type BBI alkaloids are found in the roots of *C. wattii*.⁽⁴⁵⁾ Racemosidine A, isolated from the roots of *C. racemosa*, is a BBI alkaloid that has diphenyl ether bridges at C-11/C-7' and C-8/C-12' and a benzyl-phenyl ether bridge at C-7/C-11'.⁽⁴⁶⁾ Phytosterols and alkaloids are major phytoconstituents in petroleum ether extract of *C. peltata* leaf.⁽⁴⁷⁾ The ethanolic extract of *C. peltata* leaf showed the presence of alkaloids, flavonoids, tannins, diterpenes and saponins.

Ten skeletal types were identified in alkaloids of *Cissampelos*: benzylisoquinoline, BBI, aporphine, morphinandiene, tetrahydroprotoberberine, tropoloisoquinoline, azafluoranthene, pro-aporphine, stephaoxocan, and pyrrolic nucleus-derived alkaloid, etc.⁽⁴⁸⁾ The phylogenetically close *Cissampelos* and *Cyclea* share type V, VI, VIII, XX, XXI, XXII and XXVI BBIs (Appendix 4), and their BBI profiles are closer to *Cocculus* than to *Menispermum*, which is in accordance with their phylogenetic distance (Appendix 1). The composition of BBI may have chemotaxonomic significance. The roots of *Cissampelos pareira* have high concentration of alkaloids especially the high concentration of berberine, which was present in very low concentration in *Stephania japonica* and absent in roots of *C. peltata*.⁽⁴⁹⁾ The roots of *C. peltata* contain high concentration of saponins, compared with low concentration in *C. pareira* and absence in roots of *S. japonica*.

Cocculus

At least 8 structural types are identified in alkaloids of *Cocculus*; BBI are the most, followed by erythrinane, aporphine, isoquinolone, morphinane, benzyltetrahydroisoquinoline, protoberberine, and others. In *C. hirsutus*, benzyltetrahydroisoquinoline alkaloid, aporphine, and BBI alkaloid were identified,⁽⁵⁰⁾ so was the pentacyclic triterpene hirsudiol. The curine-type BBI Wattisine A, O-methylcoccoline, (+) coccoline, (+) coccoline, magnoflorine, sino-coccoline, isosinococcoline, (-) coccolaurine, daucosterol, β -sitosterol and 1-oleoyl-3-(9Z, 12Z-arachoyl) glycerol were isolated from the root of *C. orbiculatus* var. *mollis*.⁽⁵¹⁾

Sinomenium and Menispermum

Sinomenine, magnoflorine, coccolaurine, acutumine, and higenamine are proposed as chemical markers for quality control of *Sinomenium acutum* stem,⁽⁵²⁾ higenamine and coccolaurine are recommended as chemical markers for safety control. The oxoisoaporphine, benzo[h]quinoline, aporphine, protoberberine, hasubanane, and proaporphine alkaloids were identified from stems and rhizomes of *S. acutum*.⁽⁵³⁾ Alkaloids laudanosoline-1-O-xylopyranose, 6-O-methyl-laudanosoline-1-O-glucopyranoside, sinomenine, menisperine, laurifoline, magnoflorine and norsinoacutin were identified in *S. acutum*.⁽⁵⁴⁾ Pyrrolo[2,1-a]isoquinoline and pyrrole alkaloids are isolated from *S. acutum*,⁽⁵⁵⁾ which are not found in other genera of Menispermaceae. The major difference between

Sinomenium and *Menispermum* might lie in BBI, possibly due to the lack of BBI study on the former. Numerous BBIs, mainly tail-to-tail with one diphenyl ether linkage, are identified from the latter. N-oxides of dauricine-type BBIs and rare tail-to-tail quaternary alkaloids were identified from the rhizome of *Menispermum dauricum*.⁽⁵⁶⁾ N-methylcorydaldine, thalifoline, stepholidine, acutumine, daurisolone, acutumidine, dauricolone, bianfugecine, 6-O-demethylmenisporphine, bianfugedine, dauricoside, eleutheroside D, aristolactone, aristoloterpenate I and aristolochic acid were identified from *M. dauricum*.⁽⁵⁷⁾

Tinospora

Alkaloids (including protoberberine, aporphine BIAs), sesquiterpenoids, diterpenoids (mainly clerodane type), phenolics, steroids, aliphatic compounds and polysaccharides were found in *Tinospora cordifolia*.⁽⁵⁸⁾ In *T. sinensis*, the amide alkaloids were the most,⁽⁵⁹⁾ followed by organic amine alkaloids, imidazole alkaloids, guanidine alkaloids, purine alkaloids, pyrrolidine alkaloids, piperidine alkaloids, indole alkaloids, and pyrimidine, etc. Crispene A, B, C and D, clerodane type furanoid diterpenes, and a furanoid diterpene glucoside borapetoside E were isolated from the stem of *Tinospora crispa*.⁽⁶⁰⁾ Clerodane type diterpenoids were also isolated from tuberous roots of *T. sagittata*.⁽⁶¹⁾

Other Genera

Alkaloids reticuline, asimilobine, acutumine, dihydroxyprotoberberine, and stepholidine were isolated from the vine stems of *Diploclisia affinis*.⁽⁶²⁾ Ecdysteroid, (2-nitro ethyl) phenyl and cyanophenyl glycosides, steroidal and triterpenoid saponins were found in the fruits of *D. glaucescens*.⁽⁶³⁾ Ecdysteroid and oleanane glycosides are isolated from the leaves of *D. glaucescens*.⁽⁶⁴⁾

A β -carboline alkaloid sacleximine A, together with palmatine, isotetrandrine (BBI), trans-N-feruloyltyramine, trans-N-caffeoyltyramine, yangambin, syringaresinol, sesamin, (+)epiquercitol, 4-hydroxybenzaldehyde, β -sitosterol, quercetin 3-O-rutinoside and myricetin 3-O- β -glucose (1 \rightarrow 6) α -rhamnoside, were isolated from *Triclisia saclexii* aerial parts.⁽⁶⁵⁾ This genus, unlike *Diploclisia* and *Sinomenium*, could be an excellent resource of BBIs.⁽⁶⁶⁾

Arcangelisia and *Fibraurea* belong to the subfamily Tinosporoideae (Appendix 1). Protoberberine BIAs are isolated from *Arcangelisia gusanlung* and *A. flava*.^(67,68) Fibaruretin B (2 β , 3 α -dihydroxy-2,3,7,8 α -tetrahydro-penianthic acid lactone) was isolated from the roots of *A. flava*.⁽⁶⁹⁾ Furanoditerpenes were found in *A. flava*.⁽⁷⁰⁾

Palmatine, fibrecisine, berberines, tetrahydroberberines and aporphine derivatives are found in *Fibraurea recisa*.^(71,72) Furanoditerpenoids, e.g., epi-8-hydroxycolumbin, fibaruretin B, C, E, and F, were isolated from the stems of *F. tinctoria*.⁽⁷³⁾ Twenty-five protoberberine BIAs were present in both *Tinosporae Radix* and *Fibraurea Caulis*,⁽⁷⁴⁾ while 5 compounds were detected only

in the former; the contents of 4 alkaloids in *Tinosporae Radix* were much higher than those in its adulterant, *Fibraurea Caulis*.

Chemical Composition of Lardizabalaceae and Circaeasteraceae

Oleanane type triterpenoid saponins, as a characteristic chemical component of Lardizabalaceae, have certain chemotaxonomic significance, which not only suggest the phylogenetic relationship between Lardizabalaceae and Ranunculaceae, but also the oxidation level of aglycone skeleton is related to the phylogeny of various genera in the family.

Sargentodoxa

Over 110 chemical constituents have been identified from the stem of *S. cuneata*,⁽⁷⁵⁾ the only species of *Sargentodoxa*, including phenolic acids, phenolic glycosides, lignans, flavones, triterpenoids and others. *Sargentodoxa* is rich in tannins, which are lacking in other genera of Lardizabalaceae. 3,4-Dihydroxyphenylethyl alcohol glycoside, salidroside, chlorogenic acid, and liriiodendrin in the stem of *S. cuneata* are quantified simultaneously by high-performance liquid chromatography (HPLC) coupled with evaporative light scattering detection.⁽⁷⁶⁾ The marker constituents salidroside, chlorogenic acid and 3, 4-dihydroxy-phenylethyl- β -D-glucopyranoside in *Sargentodoxae Caulis* is required by the quality standard of the crude material recorded in the Chinese Pharmacopoeia.⁽⁷⁷⁾

Akebia

Akebiae Caulis refers to a group of herbal medicines with different phylogenetic positions. *A. Caulis* is derived from 8 species of families Lardizabalaceae and Ranunculaceae.⁽⁷⁸⁾ In the Chinese Pharmacopoeia, it has been separated into 2 categories: *A. Caulis* and *Clematidis Armandii Caulis*. *A. Caulis* is one of the newest raw materials officially introduced into European Pharmacopoeia from CM;⁽⁷⁹⁾ triterpenoid and their saponins are dominant specialized metabolites of *Akebia* and could be the major material basis of therapeutic efficacy of *A. Caulis*. The triterpenoid saponins mutongsaponin C and saponin Pj1 was found only in *A. trifoliata*, while the phenolic glycoside 2-(3,4-dihydroxyphenyl)-ethyl-O- β -D-glucopyranoside was found only in *A. quinata*. Diterpene glycoside was isolated from stems of *A. quinata*.⁽⁸⁰⁾ Hederagenin and saponins were isolated from *A. quinata* fruit.^(81,82) *Akebia* saponin D is present in both *Akebia* and Dipsacaceae.⁽⁸³⁾ Pentacyclic triterpene saponins, e.g., akebiaoside K and akebiaoside N, were isolated from leaves of *A. trifoliata*.⁽⁸⁴⁾

Holboellia

Phylogenetically, *Holboellia* is closer to *Stauntonia* than to *Akebia* (Appendix 1), and *Sargentodoxa* is basal to these genera. 30-Noroleanane triterpenoid saponins, e.g., akebonoic acid 28-O- β -d-glucopyranosyl-(1" \rightarrow 6')- β -d-glucopyranosyl ester, holboellside A, and holboellside B, were isolated from the stems of *Holboellia coriacea*.⁽⁸⁵⁾ Triterpenoid saponins, fargosides A, B, C, D, and E, were isolated from the roots of *Holboellia fargesii*.⁽⁸⁶⁾

Stauntonia

Triterpenoid saponins (e.g., oleanane, nor-oleanane and lupane type saponins),⁽⁸⁷⁾ flavonoids, lignanoids, and phenylethanoid glycosides were identified in *S. brachyanthera*,⁽⁸⁸⁾ *S. chinensis*,^(89,90) and *S. obovatifoliola*.⁽⁹¹⁾ Their structural types are much similar to those of *A. quinata*, i.e., either penta-saccharidic or hexa-saccharidic bidesmoside triterpenoid glycosides.⁽⁹²⁾ Calceolarioside B is abundant in both *Akebia* and *Stauntonia*. *S. brachyanthera* could be a succedaneum of *Akebia Caulis*, which undergoes the supply crisis in recent years. Phenolic acids, flavonoids, organic acids, and calceolariosides were isolated from *S. hexaphylla*.⁽⁹³⁾

Circaeasteraceae

The medicinal compounds of *Circaeaster agrestis* and *Kingdonia uniflora* have not been reported.

Chemical Composition of Papaveraceae and Eupteleaceae

Papaveraceae has at least 38 genera, with more than 700 species. Phylogenetically, Papaveraceae is basal to other Ranunculales families except Eupteleaceae (Appendixes 1 and 2). Its chemical profile is distinct as compared with other Ranunculales families.

Fumarioideae: Corydalis, Fumaria and Hypecoum

Phylogenetically, *Corydalis* is closer to *Fumaria* than to *Hypecoum* (Appendix 5). Among identified *Corydalis* alkaloids, the quaternary isoquinoline type is the most,⁽⁹⁴⁾ followed by benzophenanthridine type, phthalide isoquinoline, simple isoquinoline, aporphine type, lignin amide and protopine type, etc. Isoquinoline alkaloids are the major bioactive ingredients of *Corydalis*. Eight types of alkaloids were found in Chinese *Corydalis*: protopine; protoberberine; phthalide; benzophenanthridine; aporphine; spirobenzylisoquinoline; benzylisoquinoline; etc.⁽⁹⁵⁾ Volatile oils, steroids and flavonoids are found in *Corydalis*. The content of flavonoids and polyphenol was greater than that of alkaloids.⁽⁹⁶⁾

Numerous alkaloids, flavonoids, saponins, and terpenoids are present in *Fumaria*. Phthalide isoquinolines (including secophthalide isoquinoline), protoberberines, and spirobenzylisoquinolines are the major alkaloids,⁽⁹⁷⁾ followed by benzylisoquinolines, aporphines, benzophenanthridines, indenobenzazepines, protopines, etc. The structural types of *Hypecoum* isoquinoline alkaloids can be divided into protoopoid, protoberberine, benzophenanthridine, aporphine, simple isoquinoline, benzylisoquinoline, spirobenzylisoquinoline, etc.⁽⁹⁸⁾ Other alkaloids such as benzazepines, phenylpropanamides, phthalides and phenylethylamines were also identified. Protoopoid alkaloids are not abundant in this genus, but they are widely distributed. Simple isoquinoline, benzylisoquinoline and spirobenzene isoquinoline alkaloids are the most in *Hypecoum*, followed by aporphine and benzophenanthridine. Different from other genera of Papaveraceae, phthalide alkaloids are less reported in *Hypecoum*.

Papaveroideae: *Papaver*, *Meconopsis*, and *Argemone*

The genus *Papaver* is closely related with *Meconopsis* (blue poppy),⁽⁸⁾ and *Argemone* is basal to these genera in the subfamily Papaveroideae (Appendix 1). Isoquinoline alkaloids are chemical markers and bioactive constituents of *Papaver* species.⁽⁹⁹⁾ (S)-reticuline is a pivotal intermediate in the biosynthesis of many BIAs,⁽¹⁰⁰⁾ among which berberine is present in species across the entire Ranunculales. The benzophenanthridine BIA, which includes the antimicrobial sanguinarine, is specific to the Papaveraceae family, whilst the phthalideisoquinoline noscapine and morphinan BIAs are exclusive to *Papaver* (opium poppy).

The number of BIAs identified in *Meconopsis* are less than that of *Papaver*, but the structural type of the former could be as diverse as that of the latter (Appendix 5), and more flavonoids were identified in *Meconopsis* than BIAs.^(8,101) Berberine, protopine, chelirithrine, sanguinarine, coptisine, palmatine, magnoflorine, and galanthamine are identified in *Argemone mexicana* roots.⁽¹⁰²⁾ Terpenoids, flavonoids and phenolics are found in *A. mexicana* whole plant extracts.⁽¹⁰³⁾

Papaveroideae: *Chelidoniaeae*, *Glaucium* and *Eschscholtzia*

The genera *Eomecon*, *Sanguinaria* (bloodroot), *Macleaya*, *Glaucium*, *Chelidonium* (greater celandine) and *Hylomecon* are included in the tribe Chelidoniaeae. *Glaucium* is closer to *Chelidonium* and *Hylomecon* (Appendix 1), and *Macleaya* is basal to these 3 genera. *Eschscholtzia* is closer to *Eomecon* and *Sanguinaria* than to the above 4 genera. *Sanguinaria* contains benzophenanthridine and protopine BIAs at biologically relevant concentrations.⁽¹⁰⁴⁾ Six types of BIAs were identified in *Eomecon*: 6 benzyltetrahydroisoquinolines, 9 protopines, 5 N-methyltetrahydroprotoberberines, 6 protoberberines, 8 benzophenanthridines, and 61 dihydrobenzophenanthridines.⁽¹⁰⁵⁾ Benzo[c]phenanthridine and dihydrobenzo[c]phenanthridine BIAs, i.e., sanguinarine, chelirubine, macarpine, etc., were identified in *Eschscholtzia* cell culture.⁽¹⁰⁶⁾ Six structural types (pavinane, protopine, benzyloisoquinoline, benzophenanthridine, aporphine and protoberberine) were identified in 3 *Argemone* and 4 *Eschscholtzia* species.⁽¹⁰⁷⁾

Protoberberine and quaternary benzo[c]phenanthridine BIAs were identified from *Chelidonium*,^(108,109) as well as lignanamides. Besides the above alkaloids, gypsogenin containing pentacyclic triterpene saponin, flavonoids, and megastigmoids were isolated from *Hylomecon*.⁽¹¹⁰⁾ Differently, the aporphine and protopine BIAs were salient in *Glaucium* (yellow horned poppy).^(111,112) The benzophenanthridines, protopines, protoberberine BIAs were found in *Macleaya*,⁽¹¹³⁾ so were the oleanane containing pentacyclic triterpenes. In Berberidaceae and Papaveraceae families, the cytotoxic aporphine BIA N,N-dimethyl-hernovine was identified in *Meconopsis cambrica*, *Berberis thunbergii*, *M. aquifolium*, *Corydalis cava*, *Glaucium flavum*, and *Chelidonium majus*;⁽¹¹⁴⁾ methyl-hernovine

was identified in *G. flavum*; columbamine was identified in *B. thunbergii*, while methyl-corypalmine, chelidonine, and sanguinarine were identified in *Fumaria officinalis*. The richest source of protopine was *Macleaya cordata* (5,463.64 μ g/g). The highest amounts of chelidonine and sanguinarine were found in *C. majus* (51,040.0 and 7,925.8 μ g/g, respectively), while *B. thunbergii* had the highest amount of berberine (6,358.4 μ g/g).

Eupteleaceae

Nortriterpene and pentacyclic triterpene oligoglycosides were isolated from fresh leaves of *Euptelea polyandra*.⁽¹¹⁵⁾

Ethnopharmacology and Pharmacological Activity

Each taxonomic group has distinctive bioactivities and ethnopharmacological uses, depending on their diverse phytometabolites. During the past decades, more than 30,000 cards have been collected, which record the ethnomedicinal uses of more than 5,000 Chinese medicinal plant species. The traditional remedy index (TRI) = $C_1^2/C_2 \times 100$,⁽¹¹⁶⁾ C_1 is the number of cards on which a specific ethnopharmacological use is recorded in China for a single genus; C_2 is the number of species that has the same ethnomedicinal use in the same genus. TRI ≥ 300 is considered as significant. The higher the TRI, the more reliable that the genus has the specific ethnomedicinal use. Distribution density of ethnomedicinal use (β) = $SP_1/SP_2 \times 100$, $SP_1=C_2$, SP_2 = number of Chinese species in this genus.

Berberidaceae

The ethnomedicinal use of Ranunculaceae has been evaluated.⁽¹⁾ Berberidaceae is phylogenetically close to Ranunculaceae; accordingly, some genera of 2 families share analogous ethnomedicinal uses, partially due to shared material basis. In the subfamily Berberidoideae, the genus *Berberis* is most commonly used in redness swelling and pain of the eye (TRI 4483, β 73), followed by gastroenteritis (TRI 3512, β 65), heat-clearing and detoxification (TRI 3114, β 59), dysentery (TRI 3430, β 56), carbuncle-abscess and sore-toxin (TRI 2604, β 54, Appendix 7), etc. Medicinal properties for all parts of *Berberis* plants include antioxidant, sedative, antimicrobial, anti-leishmaniasis, antiemetic, antipyretic, anti-inflammatory, anti-arrhythmic, anti-cholinergic, cholagogic, anti-malaria.⁽¹¹⁷⁾ The main compounds found in various species of *Berberis* are berberine and berbamine. Lipid-lowering and insulin-resistance improving actions are the most studied properties of berberine in numerous randomized clinical trials.⁽¹¹⁸⁾ There are also clinical trials regarding cardiovascular, anticancer, gastrointestinal, central nervous system (CNS), and endocrine effects (Appendix 8).

The genus *Mahonia* is most commonly used in redness swelling and pain of the eye (TRI 3553, β 100), followed by dysentery (TRI 3095, β 100), tuberculosis (TRI 2625, β 89), gastroenteritis (TRI 2579, β 94), heat-clearing and detoxification (TRI 2178, β 61). The isolated compounds and crude extracts of *Mahonia* exhibit a wide spectrum of *in vitro* and *in vivo*

pharmacological effects,⁽²⁸⁾ including antioxidant, antimicrobial, anti-inflammatory, hepatoprotective, antimutagenic and analgesic properties. Preparations containing *Mahonia* species exert good efficacy for the clinical treatment of dysentery, internal and external hemorrhage, acne vulgaris and chronic pharyngitis.

In the subfamily Nandinoideae, the genus *Caulophyllum* is traditionally used in promoting blood circulation to remove stasis (TRI 909, β 100), dispelling wind and dampness (TRI 582, β 100), and arthritis (TRI 736, β 100). The *Caulophyllum* extracts and compounds showed the antibacterial activity, anti-inflammatory and analgesic effects, antioxidant effects, antiacetylcholinesterase activity, effect on atherosclerosis and myocardial ischemia, antitumor activity, cytochrome p450 inhibition, topoisomerase inhibition, and effect on wound healing.⁽²²⁾

The genus *Nandina* is used in heat-clearing and detoxification (TRI 405, β 100), cough suppression (TRI 605, β 100), bronchitis (TRI 405, β 100), and pertussis (TRI 845, β 100). *Nandina* extract and compounds showed the antibacterial, antitumor and anti-inflammatory effects.⁽¹¹⁹⁻¹²¹⁾ Caffeoyl glucosides of *N. domestica* inhibit lipopolysaccharide (LPS)-induced endothelial inflammatory responses.⁽¹²²⁾ The essential oil of *Nandina* exhibited significant antioxidant activities.⁽¹²³⁾

In the subfamily Podophylloideae, the genus *Diphylleia* is used in carbuncle-abscess and sore-toxin (TRI 500, β 100) and arthritis (TRI 500, β 100). Deoxypodophyllotoxin of *Diphylleia* and *Podophyllum* induced apoptosis of human prostate cancer cells.⁽³¹⁾ The podophyllotoxin of *Diphylleia* and *Podophyllum* could be the major antibacterial lignan,⁽¹²⁴⁾ while flavonoids could be responsible for the antioxidant activity.

The genus *Dysosma* is most commonly used in carbuncle-abscess and sore-toxin (TRI 2669, β 100), followed by snakebite (TRI 2144, β 87), blood-activating and stasis-removing (TRI 1694, β 87), mumps (TRI 903, β 75), dispersing swelling and resolving toxin (TRI 574, β 87). Podophyllotoxins and flavonoids have cytotoxic activities.⁽¹²⁵⁾ Deoxypodophyllotoxin had antineoplastic effects on glioblastoma U-87 MG and SF126 cells.⁽¹²⁶⁾ Podophyllotoxone inhibited human prostate cancer cells.⁽¹²⁷⁾ Kaempferol of *Dysosma versipellis* inhibits angiogenesis through vascular endothelial growth factor (VEGF) and fibroblast growth factor (FGF) pathways.⁽¹²⁸⁾ Flavonol dimers from callus cultures of *D. versipellis* display *in vitro* neuraminidase inhibitory activities.⁽¹²⁹⁾ The coexisting flavonoids alleviate the podophyllotoxin toxicity.⁽¹³⁰⁾

The genus *Sinopodophyllum* is used in cough suppression (TRI 357, β 100), traumatic injury (TRI 514, β 100), stomachache (TRI 514, β 100), arthritis (TRI 357, β 100), and menoxenia (TRI 357, β 100). Flavonoids of *Sinopodophyllum* have anticancer activities against human breast cancer cells.⁽³³⁾ *S. hexundrum* promotes K562 cell apoptosis possibly by

inhibiting BCR/ABL-STAT5 survival signal pathways and activating the mitochondrion-associated apoptotic pathways.⁽¹³¹⁾

The genus *Epimedium* is most commonly used in impotence (TRI 2597, β 100), followed by strengthening yang (TRI 1988, β 70), dispelling wind and dampness (TRI 1838, β 70), arthritis (TRI 1062, β 72), and neurasthenia (TRI 662, β 80). *Epimedium* and its active compounds displayed wide pharmacological activities, e.g., strengthening yang, hormone regulation, anti-osteoporosis, immunological function modulation, anti-oxidation and anti-tumor, anti-aging, anti-atherosclerosis and anti-depressant activities.⁽¹³²⁾ The total flavonoid fraction of the *E. koreanum* extract had neuroprotective effects on dopaminergic neurons.⁽¹³³⁾ Icarin had neuroprotective properties in the mouse model of Parkinson's disease.⁽¹³⁴⁾ The protective mechanisms of icaritin against oxygen-glucose deprivation-induced injury may be related to down-regulating the expression of hypoxia-inducible factor 1 α (HIF-1 α), heat shock protein 60 (HSP-60) and HSP-70.⁽¹³⁵⁾ *Epimedium* improves neuroplasticity and accelerates functional recovery of the diseased brain.⁽¹³⁶⁾

The genus *Plagiorhegma* (*Jeffersonia*) is used in dysentery (TRI 400, β 100) and redness swelling and pain of the eye (TRI 400, β 100). The protoberberine BIA jatrorrhizine, the lignane glucosides dehydrodiconiferyl-alcohol-4- β -D-glucoside and its isomer dehydrodiconiferyl-alcohol- γ - β -D-glucoside, isolated from *Plagiorhegma dubium* cell culture, were anti-inflammatory.⁽¹³⁷⁾ The extract from *Jeffersonia dubia* exhibited the high cytotoxicity *in vitro*,⁽¹³⁸⁾ and suppressed the LPS-induced nitric oxide (NO) production.

Menispermaceae

In the subfamily Tinosporoideae, the genus *Arcangelisia* is used in heat-clearing and detoxification (TRI 320, β 100), dysentery (TRI 500, β 100), gastroenteritis (TRI 320, β 100), and malaria (TRI 500, β 100). N-trans-feruloyltyramine of *Arcangelisia gusanlung* is an active phenylpropanoid compound. It possesses antioxidant, antimicrobial, anti-melanogenesis, immunomodulative, and anticancer activities.⁽¹³⁹⁾ Gusanlungionosides A–D, the megastigmane glycoside from the stems of *A. gusanlung*, are potential tyrosinase inhibitors.⁽¹⁴⁰⁾ Berberine, extracted from *A. flava*, dose-dependently inhibited *Plasmodium* telomerase activity.⁽¹⁴¹⁾ *A. flava* and *Fibraurea tinctoria* have antiplasmodial activity.⁽¹⁴²⁾ Protoberberine BIAs and 20-hydroxyecdysone of *A. flava* had the antibabesial activity against *Babesia gibsoni* in culture.⁽⁶⁸⁾

As compared with *Arcangelisia*, *Tinospora* has more diverse chemical components (Appendix 5) and is useful as the anti-oxidant, anti-hyperglycemic, antihyperlipidemic, hepatoprotective, cardiovascular protective, neuroprotective, osteoprotective, radioprotective, anti-anxiety, adaptogenic, analgesic, anti-inflammatory, antipyretic, thrombolytic, anti-diarrheal, anti-ulcer, antimicrobial and anticancer agents.⁽⁵⁸⁾ Its anti-diabetic, antipyretic, anti-inflammatory, anti-oxidant, hepato-protective, and immuno-modulatory activities are

prominent.⁽²⁾ The genus *Fibraurea* is most commonly used in heat-clearing and dampness-drying (TRI 500, β 100), and carbuncle-abscess and sore-toxin (TRI 500, β 100), followed by dysentery (TRI 417, β 100), sore throat (TRI 417, β 100), redness swelling and pain of the eye (TRI 417, β 100), heat-clearing and detoxification (TRI 320, β 100). The cognitive-enhancing effects and antidepressant effect of *Fibraurea* alkaloids are noteworthy.^(143,144) The aporphine BIA roemerine from the fresh rattan stem of *Fibraurea recisa* has antifungal activity.⁽¹⁴⁵⁾ The methanol-water fraction of *F. tinctoria* showed higher anti-proliferative activities than its methanolic extract.⁽¹⁴⁶⁾

In the subfamily Menispermaceae, the genus *Cocculus* is most commonly used in dispelling wind and dampness (TRI 1521, β 100), followed by arthritis (TRI 1184, β 100), carbuncle-abscess and sore-toxin (TRI 720, β 100), snakebite (TRI 637, β 100), heat-clearing and detoxification (TRI 605, β 100). Extracts of *Cocculus* have antimicrobial, antidiabetic, immunomodulatory and hepatoprotective activities.⁽⁵⁰⁾

The genus *Cyclea* is most commonly used in heat-clearing and detoxification (TRI 847, β 83), followed by sore throat (TRI 476, β 83), carbuncle-abscess and sore-toxin (TRI 408, β 50), snakebite (TRI 400, β 67) and urinary tract infection (TRI 313, β 50). (-)-Curine, a BBI alkaloid from the roots of *Cyclea wattii*, induced cell cycle arrest and cell death in hepatocellular carcinoma cells in a p53-independent way.⁽¹⁴⁷⁾ Tetrandrine isolated from *Cyclea peltata* induced cytotoxicity and apoptosis through reactive oxygen species (ROS) and caspase pathways in breast and pancreatic cancer cells.⁽¹⁴⁸⁾ The roots of *Cissampelos pareira* var. *hirsuta* are used in the treatment of various diseases like stomach pain, fever, skin disease, etc. in Ayurveda and is commonly known as Patha. Two other species, *Cyclea peltata* and *Stephania japonica* of the same subfamily are being used as the source of Patha in various parts of India.⁽¹⁴⁹⁾

The genus *Stephania* is used in heat-clearing and detoxification (TRI 4102, β 94) and dispelling wind and dampness (TRI 1945, β 44). *S. tetrandra* has a wide range of pharmacological properties,⁽⁴¹⁾ e.g., antimicrobial, anti-inflammatory, anticancer/antiproliferative, CNS activity, cardiovascular activity, immunomodulatory, antifibrotic, antidiabetic, antiplatelet effects, etc. So far, studies focus principally on the alkaloids and extracts of *Stephania*. The most commonly reported molecules are tetrandrine, fangchinoline and cepharanthine. Cepharanthine suppresses nuclear factor- κ B (NF- κ B) activation, lipid peroxidation, NO production, cytokine production, and expression of cyclooxygenase;⁽¹⁵⁰⁾ it could be used against severe acute respiratory syndrome (SARS)-coronavirus (CoV)-2 and homologous viruses, and could be therapeutically important in managing coronavirus disease (COVID)-19 cases.

The genus *Diploclisia* is used in urinary tract infection (TRI 320, β 100), arthritis (TRI 320, β 100), and snakebite (TRI 500, β 100). *D. glaucescens* showed strong antifeedant

activity.⁽¹⁵¹⁾ The scarcity of pharmacological data suggests the asymmetry between phytochemical research, ethnomedicinal experiences and pharmacological research.

The genus *Menispermum* is used in heat-clearing and detoxification (TRI 1108, β 100) and sore throat (TRI 931, β 100). The *M. dauricum* alkaloids exhibited significant anti-hypoxia activity.⁽⁵⁷⁾ Oxoisoaporphine of *M. dauricum* is a potent telomerase inhibitor.⁽¹⁵²⁾ Phenolic alkaloids from *M. dauricum* inhibit BxPC-3 pancreatic cancer cells by blocking of Hedgehog signaling pathway.⁽¹⁵³⁾ They also inhibited gastric cancer *in vivo*.⁽¹⁵⁴⁾ The closely related genus *Sinomenium* is used in dispelling wind and dampness (TRI 514, β 100) and arthritis (TRI 514, β 100). The top 5 predicted significant pathways of *S. acutum* targets include phosphatidylinositol 3 kinase/Akt (PI3K/Akt) signaling, prostate cancer signaling, macrophage migration inhibitory factor (MIF) regulation of innate immunity, guanosine-binding protein coupled receptor (GPCR) signaling, and ataxia telangiectasia mutated protein (ATM) signaling.⁽¹⁵⁵⁾ The mitogen activated protein kinase 1 (MAPK1), MAPK3, p65 nuclear factor κ B (RELA), nuclear factor of κ B inhibitor alpha (NF κ BIA), interleukin 1 β (IL-1 β), prostaglandin G/H synthase 2 (PTGS2) and tumor protein 53 (TP53) were the predicted critical targets in various diseases treated by *S. acutum*. Sinomenine, a main compound from *S. acutum*, showed the activities on the immune system, cardiovascular system, and nervous system.⁽¹⁵⁶⁾

Lardizabalaceae and Circaeasteraceae

There is a lack of various types of alkaloids in Lardizabalaceae, and the ethnomedicinal uses and pharmacological activities of this family are distinct when compared with the above 3 families. In the subfamily Sargentodoxoideae, the genus *Sargentodoxa* is most commonly used in promoting blood circulation to remove stasis (TRI 2181, β 100), followed by dispelling wind and dampness (TRI 1706, β 100), arthritis (TRI 1290, β 100), traumatic injury (TRI 632, β 100), menoxenia (TRI 632, β 100). The extract and compounds of *S. cuneata* have a wide spectrum of pharmacological activities, including antimicrobial, antitumor, anti-inflammatory, antioxidant, anti-sepsis and anti-arthritis effects,⁽⁷⁵⁾ as well as protective activity against cerebrovascular diseases.

In the subfamily Lardizabaloideae, the genus *Akebia* is used in promoting blood circulation to remove stasis (TRI 455, β 100), heat-clearing and dampness-drying (TRI 368, β 100), arthritis (TRI 655, β 100), and urinary tract infection (TRI 455, β 100). Bioactivity studies of *A. quinata* stem, leaf and/or fruit extracts confirmed diuretic, hepatoregenerative, neuroprotective, analgesic, anti-inflammatory, and anti-obesity effects and an influence on ethanol metabolism.⁽⁷⁹⁾ *A. trifoliata* stem, leaf and/or fruit extracts had different action profiles. Both species showed the antibacterial and anticancer (liver and stomach) effects.

The genus *Holboellia* is most commonly used in arthritis

(TRI 800, β 100), followed by urinary tract infection (TRI 613, β 100), beriberi (TRI 613, β 100), breast milk stoppage (TRI 450, β 100), menoxenia (TRI 313, β 100). There is a lack of pharmacological studies of triterpenoid saponins from *Holboellia*. The phylogenetically close genus *Stauntonia* is used in arthritis (TRI 613, β 100) and traumatic injury (TRI 514, β 75). *Stauntonia* extract and triterpenoid showed anti-inflammation and anti-prostate hyperplasia effects.^(157,158) It is speculated that the triterpenoids of *Holboellia* also have anti-inflammatory activity. *S. hexaphylla* leaf constituents inhibit aldose reductase and the formation of advanced glycation end products.⁽⁹³⁾ Triterpenoid saponins from *S. chinensis* ameliorate insulin resistance via the AMP-activated protein kinase (AMPK) and IR/IRS-1/PI3K/Akt pathways in insulin-resistant HepG2 cells.⁽⁸⁹⁾ *S. chinensis* polysaccharide showed protective effect on CCl₄-induced acute liver injuries in mice.⁽¹⁵⁹⁾ Nor-oleanane triterpenoid saponins from *S. brachyanthera* inhibit uridine diphosphate (UDP)-glucuronosyltransferases,⁽¹⁶⁰⁾ and had anti-gout activity.⁽¹⁶¹⁾

Papaveraceae

In the subfamily Fumarioideae, around 428 species of genus *Corydalis* are distributed worldwide, 298 of which are in China, and 10 groups and 219 species are endemic in China. *Corydalis* is widely used as folk medicines in China and adjacent countries, especially in traditional Tibetan medicines, for the treatment of fever, hepatitis, edema, gastritis, cholecystitis, hypertension and other diseases.⁽⁹⁶⁾ According to Chinese Dictionary of Ethnic Medicine,⁽³⁾ up to 62 *Corydalis* species are used by Chinese ethnic minorities against musculoskeletal diseases, followed by 49 species against respiratory diseases, 42 species against skin diseases, 35 against gastrointestinal diseases, 32 against circulatory diseases and hepatobiliary diseases, respectively. *Rhizoma Corydalis* has antinociceptive, sedative, anti-epileptic, antidepressive and anti-anxiety, acetylcholinesterase inhibitory effect, drug abstinence, anti-arrhythmic, antimyocardial infarction, dilated coronary artery, cerebral ischaemia reperfusion (I/R) injury protection, antihypertensive, antithrombotic, antigastrointestinal ulcer, liver protection, antimicrobial, anti-inflammation, antiviral, and anticancer effects.⁽¹⁶²⁾

According to Chinese Dictionary of Ethnic Medicine,⁽³⁾ both *Fumaria officinalis* and *F. schleicheri* are used by Chinese ethnic minorities against skin diseases, and the former is also used against gastrointestinal and urinary ailments. Pharmacological studies revealed diverse bioactivities, e.g., hepatoprotective, anti-inflammatory, antimicrobial, antioxidant, and antitumor activities.⁽⁹⁷⁾ *F. indica* (Fumitory) showed various pharmacological activities, e.g., smooth muscle relaxant, spasmogenic and spasmolytic, analgesic, anti-inflammatory, neuropharmacological, hepatoprotective, antifungal and antibacterial activities.⁽¹⁶³⁾

Hypocoum erectum and *H. leptocarpum* are used by Chinese ethnic minorities against poisoning, gastrointestinal/hepatobiliary diseases, eye/ear/nose/throat diseases,

musculoskeletal diseases, respiratory diseases and skin diseases.⁽³⁾ Isoquinolines of *H. erectum* had anti-inflammatory and analgesic activities.⁽¹⁶⁴⁾ The crude alkaloid mixture of *H. ponticum* containing quaternary isoquinolines showed potent antifungal and antibacterial activity.⁽¹⁶⁵⁾ Leptopidine of *H. leptocarpum* could suppress growth and induce cytotoxicity in breast cancer cells via inhibiting fatty acid synthase expression.⁽¹⁶⁶⁾

In the subfamily Papaveroideae, *Glaucium squamigerum* is used by Chinese ethnic minorities against gastrointestinal and respiratory diseases.⁽³⁾ The most reported bioactivity of *Glaucium* alkaloids is anticancer and anti-cholinesterase effects.⁽¹⁶⁷⁾ Yet, most species have not been investigated either phytochemically or pharmacologically. *G. flavum* (yellow horn poppy) is the most studied species. In the tribe Chelidoneae, *Chelidonium majus* exhibits antiinflammatory, antimicrobial, antitumor, analgesic, and hepatoprotective effects,⁽¹⁶⁸⁾ supporting its traditional uses. However, several important properties, e.g., diuretic, antitussive and eye-regenerative effects, have not been scientifically studied. *C. majus* also has scientifically proven effects, such as anti-osteoporotic activity and radioprotection, which are not recorded in traditional medicine. In addition, the hepatoprotective versus hepatotoxic effects of *C. majus* deserve attention.

Macleaya cordata and *M. microcarpa* are used by Chinese ethnic minorities against poisoning, gastrointestinal, musculoskeletal, oral/ear/nose/throat, pediatric, reproductive, skin and respiratory diseases.⁽³⁾ *Macleaya* compounds and/or extract have antitumor, anti-inflammatory, insecticidal, and antibacterial activities in addition to potential toxicities.⁽¹¹³⁾ Historical uses of *Sanguinaria* can provide valuable information for the research and development of new therapies. *S. canadensis* alkaloids have the anticancer, cardiovascular, anti-inflammatory, anti-infectious, neurotransmitter, local anesthetic, gastrointestinal and coagulation effects.⁽¹⁰⁴⁾ *Eomecon chionantha* is used by Chinese ethnic minorities against poisoning, gastrointestinal, eye, musculoskeletal, oral/ear/nose/throat, pediatric, skin and respiratory diseases.⁽³⁾ The pharmacological study of *Eomecon* is lacking.

Argemone mexicana has antibacterial, anti-cancer, sedative and probably anti-anxiety properties.⁽¹⁶⁹⁾ *A. mexicana* BIAs have antimicrobial, antiparasitic, antimalarial, pesticide, cytotoxic and neurological properties.⁽¹⁷⁰⁾ *A. mexicana* silver nanoparticles are suitable for bio-formulation against mosquitoes and microbes.⁽¹⁷¹⁾ Alkaloids isolated from *A. mexicana* are cytotoxic to the SW480 human colon cancer cell line.⁽¹⁷²⁾ According to Chinese Dictionary of Ethnic Medicine,⁽³⁾ up to 19 *Meconopsis* species are used by Chinese ethnic minorities against respiratory diseases, followed by 18 species against hepatobiliary diseases, 12 against musculoskeletal diseases, 9 against ear/nose/throat diseases, 8 against skin diseases. *Meconopsis* plants have antitumor, hepatoprotective, analgesic, antimicrobial, anti-oxidant, antitussive, and anti-inflammatory activities.⁽¹⁰¹⁾ The crude extracts and metabolites of *Meconopsis* also showed CNS effects, cardiovascular

effects, antibiosis, and antiviral activity.⁽¹⁷³⁾ Six *Papaver* species are used by Chinese ethnic minorities against respiratory, gastrointestinal, musculoskeletal, reproductive and skin diseases.⁽³⁾ *Papaver* species have analgesic, antioxidant, antimutagenic, anticarcinogenic, antiproliferative, antiviral, and cardioprotective activities.⁽⁶⁾ The pharmacological study of *Pteridophyllum* is lacking.

Eupteleaceae

Nortriterpene oligoglycosides, isolated from the fresh leaves of *Euptelea polyandra*, have gastroprotective activity.⁽¹⁷⁴⁾ *E. polyandra* extracts exhibited pro-osteoblastic and anti-osteoclastic activity with low cytotoxicity,⁽¹⁷⁵⁾ suggesting their potential effectiveness against osteoporosis.

DISCUSSION

Relationship between Molecular Phylogeny and Chemical Profile of Ranunculales Families

Phylogenetically, Ranunculales is sister to all other eudicots (Appendix 2), and there is some uncertainty about basal eudicot relationship. The phytochemical, metabolomic and chemotaxonomic studies of 7 Ranunculales families may help resolve the interrelationship of basal eudicots.⁽¹⁷⁶⁾ Ranunculaceae is the largest Ranunculales family with more than 50 genera and more than 2,000 species, followed by Papaveraceae, Berberidaceae, Menispermaceae and Lardizabalaceae, which lead to a significant chemodiversity. The commonly found phytometabolites, e.g., flavonoids, phenolics, terpenoids, steroids and organic acids, etc. (Appendix 5), are also identified from Ranunculales families. Though, the most striking phytometabolites of Ranunculales are various alkaloids, especially diverse types of BIAs.

BIAs can be used as the chemotaxonomic marker, which consist of more than 2,500 diverse structures mainly generated by the order Ranunculales and Eumagnoliids. BIAs are also present in Rutaceae, Lauraceae, Cornaceae and Nelumbonaceae,⁽¹⁷⁷⁾ and sporadically distributed throughout the order Piperales. The limited occurrence of BIAs outside Ranunculales and Eumagnoliids suggests the requirement for a highly specialized, but evolutionarily unstable cellular platform to accommodate or reactivate the BIA biosynthesis pathway in divergent taxa. Some BIAs function in the defense of plants against herbivores and pathogens, thus the BIA biosynthesis might play an important role in the reproductive fitness of certain plants; more importantly, these defense molecules also find a place in the treatment of human diseases (Appendixes 7 and 8).

Large-scale molecular phylogenies provide a framework for interdisciplinary investigations in chemotaxonomy and evolution of phytometabolite biosynthesis,^(5,6) equally importantly, the phylogenetic signal of specific medicinal compounds and/or therapeutic efficacies could be identified.⁽⁴⁾ It is obvious that different types of BIAs are distributed selectively. Aporphine BIAs and protoberberine BIAs are most widely distributed, and are abundant in most genera of Menispermaceae

and Papaveraceae. However, unlike Ranunculaceae, the aporphine BIA magnoflorine was not detected in Papaveraceae genera such as *Meconopsis*, *Macleaya*, *Corydalis* and *Fumaria*.⁽¹⁷⁸⁾ Berberine is abundant in Ranunculaceae subfamilies Thalictroideae and Coptidoideae, Berberidaceae, but absent in Papaveraceae genera except *Chelidonium* and *Macleaya*.⁽¹¹⁴⁾ The protopine BIAs and phenanthridine BIAs are mainly distributed in Papaveraceae, morphinan BIAs and benzyltetrahydroisoquinoline alkaloids are concentrated in some genera of Menispermaceae and Papaveraceae, while BBIs are found in Menispermaceae, *Berberis* and *Mahonia* of Berberidaceae, *Isopyrum* and *Dichocarpum* of Ranunculaceae.^(66,179) The data about the distribution of pavinane alkaloid, quinolizidine alkaloid, pyrrolidine alkaloid and steroidal alkaloid in Ranunculales are relatively few, and their distribution pattern is uncertain. In contrast, alkaloids with certain distribution could be useful in chemotaxonomy, and the distribution patterns provide clues for finding new medicinal compounds purposefully, not just by chance.

Besides alkaloids, steroids, phenolics, flavonoids and their glycosides are scattered in various Ranunculales families (Appendix 5). Lignans are commonly found in Ranunculales families except Papaveraceae. The pentacyclic triterpenoids are distributed throughout Ranunculales, which is relatively devoid of diterpenoids and tetracyclic triterpenoids. Some of these compounds have a variety of pharmacological activities (Appendix 8) and the relevant families and genera should be further excavated.

It was suggested that, from the perspective of pharmacophylogeny, Berberidaceae could be treated as 4 independent families: Nandinaceae, Berberidaceae (s.s.), Podophyllaceae and Leonticaceae.⁽¹⁵⁾ The monotypic family Nandinaceae is rich in various BIAs, e.g., aporphine BIA, protoberberine BIA, protopine BIA, morphinan BIA and benzyltetrahydroisoquinoline alkaloid, which is similar to that of Menispermaceae and Papaveraceae, but BBI, phenanthridine BIA and pavinane alkaloid are absent in *Nandina*. The presence of the cyanogenic compound nandinin, biflavonoid amentoflavone and benzaldehyde-4-O-glucoside in *Nandina* indicates its relatively distant relation with the other Berberidaceae genera. *Nandina indica* is ethnopharmacologically used for clearing heat and counteracting toxins, and as antitussive (Appendix 7). The proposed Berberidaceae (s.s.) consists of *Berberis* and *Mahonia*, containing mainly BIAs, e.g., aporphine BIA, protoberberine BIA and BBI, particularly a higher content of type VIII BBI berbamine and type VI BBI oxyacanthine. Ethnopharmacologically the plants of *Berberis* and *Mahonia* are mainly used as medicines for clearing heat and counteracting toxins. Plants of both genera have long been used as the main sources of berberine and berbamine.^(180,181) The proposed Podophyllaceae can be divided into 2 tribes. The tribe Podophylleae, consisting of *Podophyllum*, *Sinopodophyllum*, *Dysosma* and *Diphylleia*, is rich in various podophyllotoxin lignans, and these plants are used as the most important source for producing the anticancer

podophyllotoxin's derivatives. Ethnopharmacologically, these plants are mainly used for activating blood, revolving stasis, relieving swelling, removing toxin, and clearing heat. The tribe Epimedieae, consisting of *Epimedium*, *Vancouveria*, *Jeffersonia* and *Plagiorhegma*, has diverse chemicals. Both *Epimedium* and *Vancouveria* contain predominantly icariin flavonoids, the characteristic chemical of this group. Ethnopharmacologically the *Epimedium* plants are used as a male sexual tonic, and for dispelling wind and removing dampness. The phytochemistry of *Achlys*, *Jeffersonia* and *Ranzania* has not been thoroughly investigated. *Jeffersonia dubia* is used for dysentery and inflammatory eye pain by the Korean nationality of Northeast China. The proposed Leonticeae, including *Gymnospermium*, *Leontice*, *Caulophyllum* and *Bongardia*, contains mainly β -amyrin triterpenoids and quinolizidine alkaloids; they are used for activating blood, revolving stasis, dispelling wind and removing dampness.

Relationship between Phytometabolites and Therapeutic Efficacy of Ranunculales Families

Among the traditional medicinal uses of Ranunculales, 25 genera, including 8 of Ranunculaceae,⁽¹⁾ 5 of Berberidaceae and Papaveraceae respectively, 4 of Lardizabalaceae, and 3 of Menispermaceae, are used to treat arthritis (Appendix 7). Up to 29 genera, i.e., 13 of Ranunculaceae,⁽¹⁾ 9 of Papaveraceae, 4 of Berberidaceae, and 3 of Menispermaceae, are used to treat carbuncle-abscess and sore-toxin. Twenty-six genera, including 11 of Ranunculaceae,⁽¹⁾ 9 of Papaveraceae, 4 of Berberidaceae, and 2 of Lardizabalaceae, are used to treat traumatic injury. Twenty-three genera of Ranunculaceae, Menispermaceae and Berberidaceae, rather than Lardizabalaceae and Papaveraceae genera, are used in heat-clearing and detoxification; 16 genera of Ranunculaceae,⁽¹⁾ Papaveraceae, Berberidaceae and Menispermaceae, rather than Lardizabalaceae genera, are ethnomedicinally used against dysentery. The greater differences of ethnopharmacological uses between Ranunculaceae and other Ranunculales families are revealed in this study (Appendixes 7 and 9). In addition to the above 5 common uses, there are many plants of Papaveraceae, Berberidaceae, Menispermaceae and Lardizabalaceae used to treat snake bites, gastroenteritis, redness swelling and pain of eyes, sore throat and urinary tract infection; in contrast, there are more Ranunculaceae plants used in wind-dispersing and dampness-eliminating, blood-activating and stasis-removing, swell-reducing and detoxification, or as the pesticide, antitussive and expectorant.⁽¹⁾ Such differences in traditional uses could be partially explained by the distinct material basis of different families/genera (Appendixes 4 and 5). Since the material basis and responsible phytometabolites of some ethnomedicinal utilities are still elusive, it is imperative to further clarify the internal relationship between the two, so as to better harness the traditional wisdom in novel drug discovery and development.

In modern bioactivity studies, 46 genera, including 17 of Ranunculaceae,⁽¹⁾ 12 of Papaveraceae, 8 of Menispermaceae, 7 of Berberidaceae, 2 of Lardizabalaceae, showed the

anticancer/cytotoxic/antiproliferative activities *in vitro* or *in vivo* (Appendix 8). Up to 39 genera, i.e., 15 of Ranunculaceae,⁽¹⁾ 12 of Papaveraceae/Eupteleaceae, 5 of Berberidaceae, 4 of Menispermaceae, and 3 of Lardizabalaceae, displayed the antibacterial/antifungal/antiviral properties, which is followed by anti-inflammatory (36 species), analgesic/sedative (24 species), immunomodulatory and antioxidant (18 respectively), hepatoprotective⁽¹³⁾ and antiparasitic/pesticide⁽¹²⁾ activities, etc. In addition, 6 Ranunculaceae species exhibited the antihypertensive activity,⁽¹⁾ whereas 9, 8 and 7 species of other 4 families had the anti-diabetic, neuroprotective, anti-atherosclerosis and myocardial ischemia properties, respectively. Some Ranunculaceae alkaloids, terpenoids, saponins and polysaccharides have shown anticancer activities *in vitro* and *in vivo*.⁽¹⁸²⁾ Most concerns have been raised for the monoterpene thymoquinone and the isoquinoline alkaloid berberine, and the latter could also partially explain the anticancer properties of Berberidaceae plants. The anticancer compounds of other 3 families are also diverse. For example, both oxoisoaporphine alkaloids and BBIs of Menispermaceae have anticancer effects (Appendix 9).^(66,183,184) The pentacyclic triterpene saponins of Lardizabalaceae such as Akebia saponin E inhibited the proliferation of carcinoma cells.^(79,185) The phenanthridine BIAs of Papaveraceae such as sanguinarine and chelerythrine had strong anticancer activities.⁽¹⁸⁶⁾ At least 17 Ranunculaceae genera are rich in anticancer phytometabolites, and 29 genera of other Ranunculales families have numerous cytotoxic compounds, constituting an enormous chemical space for bioprospecting. Some of these phytometabolites induce the cell cycle arrest and apoptosis of cancer cells or enhance immune activities, while others inhibit the proliferation, invasion, angiogenesis and metastasis, or reverse the multi-drug resistance of cancer cells, thereby regulating hallmarks of cancer.⁽¹⁸²⁾ These phytometabolites could exert their anticancer activities via multiple signaling pathways.

Conclusion

The 7 families of Ranunculales are rich in medicinal species frequently utilized by traditional medical systems and ethnomedicine,⁽¹⁸⁷⁾ which represent a treasure chest of biodiversity and chemodiversity.⁽¹⁸⁸⁾ The phylogenetically related species often have similar phytometabolites, and therefore similar bioactivities. This study gives a systematic review of the molecular phylogeny, chemical components and pharmacological activities of Ranunculales within the framework of pharmacophylogeny. The metabolomic techniques greatly facilitate the systematic mining of both primary and secondary metabolites,⁽¹⁸⁹⁾ the far-reaching metabolite profiling based on multiple analytical platforms enabled a more inclusive picture of overall metabolism occurring in selected species. Traditionally important medicinal species and their close relatives should be subjected to a metabolomics approach.

The surging bioactivity studies of extracts and compounds help clarify the rationale of ethnomedicinal utilities and deepen our understanding of the medicinal potential of Ranunculales.

The most studied compounds of this order include BIAs, flavonoids and phenolics, lignans, terpenoids and saponins, etc. BBI alkaloids, with versatile bioactivities, are especially abundant in Berberidaceae and Menispermaceae. The most frequent ethnomedicinal uses are arthritis, heat-clearing and detoxification, carbuncle-abscess and sore-toxin, etc. The most studied bioactivities are anticancer/cytotoxic, antimicrobial, and anti-inflammatory activities. Some pharmacological activities are in line with traditional uses, but some others are not related to the latter, implying the broad pharmacological space to be explored. Our pharmacophylogeny analysis, integrated with both traditional and contemporary medicinal uses, agrees with the molecular phylogeny based on chloroplast and nuclear DNA sequences. With the increasing evidence of chemotaxonomy and therapeutic efficacy of each taxonomic group, the underlying connection between phylogeny, chemodiversity and clinical uses is revealed, which facilitates the conservation and sustainable utilization of Ranunculales pharmaceutical resources, as well as developing novel plant-based pharmacotherapy. The high-quality chromosome-scale genome assembly and annotation of *Coptis chinensis* have been presented.⁽¹⁹⁰⁾ In the near future, the whole genome sequences of more Ranunculales species could be deciphered; it will be possible to reconstruct the phylogenetic relationship based on transcriptome and/or genome sequences. Decoding the biosynthesis pathways and their regulation based on genome sequences could be conducive to predicting the specific types or classes of compounds, and cross-validating the metabolomic data. With the development of R language software packages, the distribution of therapeutic compounds and efficacy of Ranunculales species on the seed plant Tree of Life^(5,6,191) can be quantitatively investigated.^(4,187)

The absorption, distribution, metabolism, and excretion/toxicity (ADMET) properties of phytometabolites have the significant impact on the therapeutic efficacy;⁽¹⁸⁰⁾ the structurally analogous compounds could have similar (or distinct) metabolism and pharmacokinetic attributes. In the early stage of drug discovery and development, the ADMET properties should also be taken into account in the context of pharmacophylogeny, so as to improve the theoretical basis and application value of pharmacophylogeny. In the mechanism studies, the gene expression profiling and relevant omics platforms (e.g. genomics, transcriptomics, proteomics, and metabolomics) could reveal differential effects of phytometabolites on the phenotypically heterogeneous human cells.

Conflict of Interest

The authors declare that they have no competing interests. Author XIAO Pei-gen is a member of the Editorial Board for CJIM. The paper was handled by the other Editor and has undergone rigorous peer review process. Author XIAO Pei-gen was not involved in the journal's review of, or decisions related to, this manuscript.

Author Contributions

Xiao PG, Hao DC and Xu LJ contributed to study concept and design. Hao DC undertook data analysis and interpretation

and wrote the paper. Xu LJ provided financial support. Hao DC and Xu LJ contributed equally to this work. Zheng YW and Lyu HY collected data and Zheng YW drew figures.

Electronic Supplementary Material: Supplementary material (Appendixes 1–9) is available in the online version of this article at <https://doi.org/10.1007/s11655-022-3576-x>.

REFERENCES

- Hao DC, Xiao PG, Ma HY, et al. Mining chemodiversity from biodiversity: pharmacophylogeny of medicinal plants of Ranunculaceae. *Chin J Nat Med* 2015;13:507-520.
- Kumar P, Kamle M, Mahato DK, et al. *Tinospora cordifolia* (Giloy): phytochemistry, ethnopharmacology, clinical application and conservation strategies. *Curr Pharm Biotechnol* 2020;21:1165-1175.
- Jia MR, Zhang Y, eds. Dictionary of Chinese ethnic medicine. Beijing: China Medical Science Press;2016:1-886.
- Zaman W, Ye JF, Saqib S, et al. Predicting potential medicinal plants with phylogenetic topology: inspiration from the research of traditional Chinese medicine. *J Ethnopharmacol* 2021;281:114515.
- Chen ZD, Yang T, Lin L, et al. Tree of life for the genera of Chinese vascular plants. *J Syst Evol* 2016;54:277-306.
- Hu HH, Liu B, Liang YS, et al. An updated Chinese vascular plant tree of life: phylogenetic diversity hotspots revisited. *J Syst Evol* 2020;58:663-672.
- Soltis PS, Soltis DE. The origin and diversification of angiosperms. *Am J Bot* 2004;91:1614-1626.
- Hao DC, Gu X, Xiao PG, eds. Medicinal plants: chemistry, biology and omics. Oxford: Elsevier-Woodhead;2015:1-400.
- Wang W, Lu AM, Ren Y, et al. Phylogeny and classification of Ranunculales evidence from four molecular loci and morphological data. *Persp Plant Ecol Evol Syst* 2009;11:81-110.
- Soltis DE, Smith SA, Cellinese N, et al. Angiosperm phylogeny: 17 genes, 640 taxa. *Am J Bot* 2011;98:704-730.
- Sun Y, Moore MJ, Zhang S, et al. Phylogenomic and structural analyses of 18 complete plastomes across nearly all families of early-diverging eudicots, including an angiosperm-wide analysis of IR gene content evolution. *Mol Phylogenet Evol* 2016;96:93-101.
- Anderson CL, Bremer K, Friis EM. Dating phylogenetically basal eudicots using rbcL sequences and multiple fossil reference points. *Am J Bot* 2005;92:1737-1748.
- Weferling KM, Hoot SB, Neves SS. Phylogeny and fruit evolution in Menispermaceae. *Am J Bot* 2013;100:883-905.
- Sauquet H, Carrive L, Poullain N, et al. Zygomorphy evolved from disymmetry in Fumarioideae (Papaveraceae, Ranunculales): new evidence from an expanded molecular phylogenetic framework. *Ann Bot* 2015;115:895-914.
- Peng Y, Chen SB, Liu Y, et al. A pharmacophylogenetic study of the Berberidaceae (s.l.). *Acta Phytotax Sin (Chin)* 2006;44:241-257.
- Peng CY, Liu JQ, Zhang R, et al. A new alkaloid from the fruit of *Nandina domestica* Thunb. *Nat Prod Res* 2014;28:1159-1164.
- Imahori D, Matsumoto T, Saito Y, et al. Cell death-inducing activities via P-glycoprotein inhibition of the constituents isolated from fruits of *Nandina domestica*. *Fitoterapia* 2021;154:105023.
- Iwasa K, Doi Y, Takahashi T, et al. Enantiomeric separation of racemic 1-benzyl-N-methyltetrahydroisoquinolines on chiral columns and chiral purity determinations of the O-methylated metabolites in plant cell cultures by HPLC-CD on-line coupling in combination with HPLC-MS. *Phytochemistry* 2009;70:198-206.
- Zhang N, Lian Z, Peng X, et al. Applications of higenamine in pharmacology and medicine. *J Ethnopharmacol* 2017;196:242-252.
- Forrester MB. Pediatric *Nandina domestica* ingestions reported to poison centers. *Hum Exp Toxicol* 2018;37:338-342.
- Kodai T, Horiuchi Y, Nishioka Y, et al. Novel cycloartane-type triterpenoid

- from the fruits of *Nandina domestica*. J Nat Med 2010;64:216-218.
22. Xia YG, Li GY, Liang J, et al. Genus *Caulophyllum*: an overview of chemistry and bioactivity. Evid Based Complement Alternat Med 2014;2014:684508.
 23. Wang XL, Liu BR, Chen CK, et al. Four new fluorenone alkaloids and one new dihydroazafluoranthene alkaloid from *Caulophyllum robustum* Maxim. Fitoterapia 2011;82:793-797.
 24. Yang HX, Li W, Li Q, et al. Piperidine alkaloids and xanthone from the roots of *Caulophyllum robustum* Maxim. Fitoterapia 2019;132:22-25.
 25. Xia YG, Liang J, Li GY, et al. Energy-resolved technique for discovery and identification of malonyl-triterpene saponins in *Caulophyllum robustum* by UHPLC-electrospray fourier transform mass spectrometry. J Mass Spectrom 2016;51:947-958.
 26. Xia YG, Li GY, Liang J, et al. A strategy for characterization of triterpene saponins in *Caulophyllum robustum* hairy roots by liquid chromatography with electrospray ionization quadrupole-time-of-flight mass spectrometry. J Pharm Biomed Anal 2014;100:109-122.
 27. Jia XJ, Li X, Wang F, et al. Berberine exerts anti-inflammatory effects via inhibition of NF- κ B and MAPK signaling pathways. Cell Physiol Biochem 2017;41:2307-2318.
 28. He JM, Mu Q. The medicinal uses of the genus *Mahonia* in traditional Chinese medicine: an ethnopharmacological, phytochemical and pharmacological review. J Ethnopharmacol 2015;175:668-683.
 29. Belwal T, Giri L, Bhatt ID, et al. An improved method for extraction of nutraceutically important polyphenolics from *Berberis jaeschkeana* C.K. Schneid. fruits. Food Chem 2017;230:657-666.
 30. Liu L, Cui ZX, Yang XW, et al. Simultaneous characterisation of multiple *Mahonia fortunei* bioactive compounds in rat plasma by UPLC-MS/MS for application in pharmacokinetic studies and anti-inflammatory activity *in vitro*. J Pharm Biomed Anal 2020;179:113013.
 31. Hu S, Zhou Q, Wu WR, et al. Anticancer effect of deoxydopodophyllotoxin induces apoptosis of human prostate cancer cells. Oncol Lett 2016;12:2918-2923.
 32. Zhao C, Zhang N, He W, et al. Simultaneous determination of three major lignans in rat plasma by LC-MS/MS and its application to a pharmacokinetic study after oral administration of *Diphylleia sinensis* extract. Biomed Chromatogr 2014;28:463-467.
 33. Wang QH, Guo S, Yang XY, et al. Flavonoids isolated from *Sinopodophylli Fructus* and their bioactivities against human breast cancer cells. Chin J Nat Med 2017;15:225-233.
 34. Sun YJ, Gao ML, Zhang YL, et al. Labdane diterpenes from the fruits of *Sinopodophyllum emodi*. Molecules 2016;21:434.
 35. Sun YJ, Pei LX, Wang KB, et al. Preparative isolation of two prenylated biflavonoids from the roots and rhizomes of *Sinopodophyllum emodi* by Sephadex LH-20 column and high-speed counter-current chromatography. Molecules 2015;21:E10.
 36. Marques JV, Dalisay DS, Yang H, et al. A multi-omics strategy resolves the elusive nature of alkaloids in *Podophyllum* species. Mol Biosyst 2014;10:2838-2849.
 37. Ti H, Wu P, Xu L, et al. Anti-inflammatory neolignans from *Epimedium pseudowushanense*. Nat Prod Res 2017;31:2621-2628.
 38. Thavamani BS, Mathew M, Palaniswamy DS. Anticancer activity of *Cocculus hirsutus* against Dalton's lymphoma ascites (DLA) cells in mice. Pharm Biol 2014;52:867-872.
 39. Sim HJ, Kim JH, Lee KR, et al. Simultaneous determination of structurally diverse compounds in different Fangchi species by UHPLC-DAD and UHPLC-ESI-MS/MS. Molecules 2013;18:5235-5250.
 40. Desgrouas C, Taudon N, Bun SS, et al. Ethnobotany, phytochemistry and pharmacology of *Stephania rotunda* Lour. J Ethnopharmacol 2014;154:537-563.
 41. Jiang Y, Liu M, Liu H, et al. A critical review: traditional uses, phytochemistry, pharmacology and toxicology of *Stephania tetrandra* S. Moore (Fen Fang Ji). Phytochem Rev 2020;2020:1-41.
 42. Dary C, Bun SS, Herbet G, et al. Chemical profiling of the tuber of *Stephania cambodica* Gagnep. (Menispermaceae) and analytical control by UHPLC-DAD. Nat Prod Res 2017;31:802-809.
 43. Wang XJ, Zhang Q, Peng YR, et al. Two azafluoranthene alkaloids and a phytoecdysone from the stems of *Cyclea barbata*. J Asian Nat Prod Res 2019;21:217-226.
 44. Wang JZ, Liao J, Xu WL, et al. Bisbenzylisoquinoline alkaloids from the roots of *Cyclea tonkinensis*. Planta Med 2015;81:600-605.
 45. Wang JZ, Liu XY, Wang FP. Two new curine-type bisbenzylisoquinoline alkaloids from the roots of *Cyclea watti* with cytotoxic activities. Chem Pharm Bull 2010;58:986-988.
 46. Wang JZ, Chen QH, Wang FP. Cytotoxic bisbenzylisoquinoline alkaloids from the roots of *Cyclea racemosa*. J Nat Prod 2010;73:1288-1293.
 47. Hullatti KK, Gopikrishna UV, Kuppatt IJ. Phytochemical investigation and diuretic activity of *Cyclea peltata* leaf extracts. J Adv Pharm Technol Res 2011;2:241-244.
 48. da Silva Mendes JW, Cunha WEM, Rodrigues FFG, et al. *Cissampelos* genus: biological activities, ethnobotanical and phytochemical aspects. Phytochem Rev 2020;19:955-982.
 49. Hullatti KK, Sharada MS. Comparative phytochemical investigation of the sources of ayurvedic drug patha: a chromatographic fingerprinting analysis. Indian J Pharm Sci 2010;72:39-45.
 50. Logesh R, Das N, Adhikari-Devkota A, et al. *Cocculus hirsutus* (L.) W. Theob. (Menispermaceae): a review on traditional uses, phytochemistry and pharmacological activities. Medicines 2020;7:69.
 51. Liao J, Lei Y, Wang JZ. Chemical constituents of *Cocculus orbiculatus* var. *mollis* root. J Chin Med Mater (Chin) 2014;37:254-257.
 52. Huang YF, He F, Wang CJ, et al. Discovery of chemical markers for improving the quality and safety control of *Sinomenium acutum* stem by the simultaneous determination of multiple alkaloids using UHPLC-QQQ-MS/MS. Sci Rep 2020;10:14182.
 53. Lyu HN, Zeng KW, Cao NK, et al. Alkaloids from the stems and rhizomes of *Sinomenium acutum* from the Qinling Mountains, China. Phytochemistry 2018;156:241-249.
 54. Sun M, Wang J, Zhou Y, et al. Isotetrandrine reduces astrocyte cytotoxicity in neuromyelitis optica by blocking the binding of NMO-IgG to aquaporin 4. Neuroimmunomodulation 2016;23:98-108.
 55. Lv HN, Zeng KW, Zhao MB, et al. Pyrrolo[2,1-a]isoquinoline and pyrrole alkaloids from *Sinomenium acutum*. J Asian Nat Prod Res 2018;20:195-200.
 56. Wei H, Han Y, Wang J, et al. Analgesic bisbenzylisoquinoline alkaloids from the rhizoma of *Menispermum dauricum* DC. Bioorg Chem 2021;107:104517.
 57. Shao J, Shi CF, Wei JX, et al. Chemical constituents from rhizome of *Menispermum dauricum* and their anti-hypoxic activities. China J Chin Mater Med (Chin) 2019;44:723-729.
 58. Singh D, Chaudhuri PK. Chemistry and pharmacology of *Tinospora cordifolia*. Nat Prod Commun 2017;12:299-308.
 59. Zhu XF. Study on the separation of chemical constituents and determination of active components in Tibetan medicine *Tinospora sinensis* [dissertation]. Nanchang: Jiangxi University of Traditional Chinese Medicine;2019.
 60. Hossen F, Ahasan R, Haque MR, et al. Crispene A, B, C and D, four new clerodane type furanoid diterpenes from *Tinospora crispa* (L.). Pharmacogn Mag 2016;12(S1):37-41.
 61. Zhang G, Ma H, Hu S, et al. Clerodane-type diterpenoids from tuberous roots of *Tinospora sagittata* (Oliv.) Gagnep. Fitoterapia 2016;110:59-65.
 62. Wang EJ, Ma YB, Zhang XM, et al. Five alkaloids from vine stems of *Diploclisia affinis*. China J Chin Mater Med (Chin) 2008;33:2503-2505.
 63. Jayasinghe UL, Hara N, Fujimoto Y. (2-Nitro ethyl)phenyl and cyanophenyl glycosides from the fruits of *Diploclisia glaucescens*. Nat Prod Res 2007;21:260-264.
 64. Jayasinghe L, Jayasooriya CP, Oyama K, et al. 3-Deoxy-1 β ,20-dihydroxyecdysone from the leaves of *Diploclisia glaucescens*. Steroids 2002;67:555-558.
 65. Samita F, Ochieng CO, Owuor PO, et al. Isolation of a new β -carboline alkaloid from aerial parts of *Triclisia saclouxii* and its antibacterial and cytotoxicity effects. Nat Prod Res 2017;31:529-536.

66. Gao GY, Xiao PG. Review of studies of bisbenzylisoquinoline alkaloid (BBI) on distribution in higher plant and physiological activities. *Nat Prod Res Develop* 1999;11:96-103.
67. Yu LL, Li RT, Ai YB, et al. Protoberberine isoquinoline alkaloids from *Arcangelisia gusanlung*. *Molecules* 2014;19:13332-13341.
68. Subeki, Matsuura H, Takahashi K, et al. Antibabesial activity of protoberberine alkaloids and 20-hydroxyecdysone from *Arcangelisia flava* against *Babesia gibsoni* in culture. *J Vet Med Sci* 2005;67:223-227.
69. Fun HK, Salae AW, Razak IA, et al. Absolute configuration of fibareuretin B. *Acta Crystallogr Sect E Struct Rep Online* 2011;67(Pt 5):1246-1247.
70. Kawakami Y, Nagai Y, Nezu Y, et al. Indonesian medicinal plants. I. New furanoditerpenes from *Arcangelisia flava* MERR. (2). Stereostructure of furanoditerpenes determined by nuclear magnetic resonance analysis. *Chem Pharm Bull* 1987;35:4839-4845.
71. Qiao W, Wang L, Ye B, et al. Electrochemical behavior of palmatine and its sensitive determination based on an electrochemically reduced L-methionine functionalized graphene oxide modified electrode. *Analyst* 2015;140:7974-7983.
72. Rao GX, Zhang S, Wang HM, et al. Antifungal alkaloids from the fresh rattan stem of *Fibraurea recisa* Pierre. *J Ethnopharmacol* 2009;123:1-5.
73. Su CR, Chen YF, Liou MJ, et al. Anti-inflammatory activities of furanoditerpenoids and other constituents from *Fibraurea tinctoria*. *Bioorg Med Chem* 2008;16:9603-9609.
74. Zhao X, Zhang Y, Wang Q, et al. An integrated strategy for the establishment of a protoberberine alkaloid profile: exploration of the differences in composition between *Tinosporae Radix* and *Fibraurea Caulis*. *Phytochem Anal* 2021;32:1131-1140.
75. Zhang W, Sun C, Zhou S, et al. Recent advances in chemistry and bioactivity of *Sargentodoxa cuneata*. *J Ethnopharmacol* 2021;270:113840.
76. Li DH, Lv YS, Liu JH, et al. Simultaneous determination of four active ingredients in *Sargentodoxa cuneata* by HPLC coupled with evaporative light scattering detection. *Int J Anal Chem* 2016;2016:8509858.
77. Li H, Zhao FC, Yuan XD, et al. Determination of phenols and triterpenoid saponins in stems of *Sargentodoxa cuneata*. *China J Chin Mater Med (Chin)* 2015;40:1865-1871.
78. Li X, Xia Y, Li G, et al. Traditional uses, phytochemistry, pharmacology, and toxicology of *Akebiae Caulis* and its synonyms: a review. *J Ethnopharmacol* 2021;277:114245.
79. Maciag D, Dobrowolska E, Sharafan M, et al. *Akebia quinata* and *Akebia trifoliata*-a review of phytochemical composition, ethnopharmacological approaches and biological studies. *J Ethnopharmacol* 2021;280:114486.
80. An JP, Ha TK, Kim J. Protein tyrosine phosphatase 1B inhibitors from the stems of *Akebia quinata*. *Molecules* 2016;21:1091.
81. Kim DR, Lee JE, Shim KJ, et al. Effects of herbal *Epimedium* on the improvement of bone metabolic disorder through the induction of osteogenic differentiation from bone marrow-derived mesenchymal stem cells. *Mol Med Rep* 2017;15:125-130.
82. Ling Y, Zhang Q, Zhu DD, et al. Identification and characterization of the major chemical constituents in *Fructus Akebiae* by high-performance liquid chromatography coupled with electrospray ionization-quadrupole-time-of-flight mass spectrometry. *J Chromatogr Sci* 2016;54:148-157.
83. Gong LL, Li GR, Zhang W, et al. Akebia saponin D decreases hepatic steatosis through autophagy modulation. *J Pharmacol Exp Ther* 2016;359:392-400.
84. Xu QL, Wang J, Dong LM, et al. Two new pentacyclic triterpene saponins from the leaves of *Akebia trifoliata*. *Molecules* 2016;21:962.
85. Ding W, Li Y, Li G, et al. New 30-noroleanane triterpenoid saponins from *Holboellia coriacea* Diels. *Molecules* 2016;21:734.
86. Fu H, Koike K, Zheng Q, et al. Fargosides A-E, triterpenoid saponins from *Holboellia fargesii*. *Chem Pharm Bull* 2001;49:999-1002.
87. Vinh LB, Jang HJ, Phong NV, et al. Isolation, structural elucidation, and insights into the anti-inflammatory effects of triterpene saponins from the leaves of *Stauntonia hexaphylla*. *Bioorg Med Chem Lett* 2019;29:965-969.
88. Liu XL, Wang DD, Wang ZH, et al. Diuretic properties and chemical constituent studies on *Stauntonia brachyanthera*. *Evid Based Complement Alternat Med* 2015;2015:432419.
89. Hu X, Wang S, Xu J, et al. Triterpenoid saponins from *Stauntonia chinensis* ameliorate insulin resistance via the AMP-activated protein kinase and IR/IRS-1/PI3K/Akt pathways in insulin-resistant HepG2 cells. *Int J Mol Sci* 2014;15:10446-10458.
90. Wang D, Tian J, Zhou GP, et al. Triterpenoid glycosides from *Stauntonia chinensis*. *J Asian Nat Prod Res* 2010;12:150-156.
91. Lu X, Qiu F, Pan X, et al. Simultaneous quantitative analysis of nine triterpenoid saponins for the quality control of *Stauntonia obovatifoliola* Hayata subsp. intermedia stems. *J Sep Sci* 2014;37:3632-3640.
92. Gao H, Zhao F, Chen GD, et al. Bidesmoside triterpenoid glycosides from *Stauntonia chinensis* and relationship to anti-inflammation. *Phytochemistry* 2009;70:795-806.
93. Hwang SH, Kwon SH, Kim SB, et al. Inhibitory activities of *Stauntonia hexaphylla* leaf constituents on rat lens aldose reductase and formation of advanced glycation end products and antioxidant. *Biomed Res Int* 2017;2017:4273257.
94. Deng AP, Zhang Y, Zhou L, et al. Systematic review of the alkaloid constituents in several important medicinal plants of the genus *Corydalis*. *Phytochemistry* 2021;183:112644.
95. Mao Z, Wang X, Liu Y, et al. Simultaneous determination of seven alkaloids from *Rhizoma Corydalis Decumbentis* in rabbit aqueous humor by LC-MS/MS: application to ocular pharmacokinetic studies. *J Chromatogr B Analyt Technol Biomed Life Sci* 2017;1057:46-53.
96. Shang WQ, Chen YM, Gao XL, et al. Phytochemical and pharmacological advance on Tibetan medicinal plants of *Corydalis*. *China J Chin Mater Med (Chin)* 2014;39:1190-1198.
97. Zhang RF, Guo Q, Kennelly EJ, et al. Diverse alkaloids and biological activities of *Fumaria* (Papaveraceae): an ethnomedicinal group. *Fitoterapia* 2020;146:104697.
98. Zhang RF, Zha S, Yin X, et al. Phytochemical and pharmacological progress on Tibetan medicine *Hypecoi Erecti Herba* and plants of *Hypecoum* L. *Chin Tradit Herb Drugs (Chin)* 2016;47:1217-1224.
99. Song K, Oh JH, Lee MY, et al. Molecular network-guided alkaloid profiling of aerial parts of *Papaver nudicaule* L. Using LC-HRMS. *Molecules* 2020;25:2636.
100. Li Y, Winzer T, He Z, et al. Over 100 million years of enzyme evolution underpinning the production of morphine in the Papaveraceae family of flowering plants. *Plant Commun* 2020;1:100029.
101. Guo Q, Bai R, Zhao B, et al. An ethnopharmacological, phytochemical and pharmacological review of the genus *Meconopsis*. *Am J Chin Med* 2016;44:439-462.
102. Kukula-Koch W, Mroczek T. Application of hydrostatic CCC-TLC-HPLC-ESI-TOF-MS for the bioguided fractionation of anticholinesterase alkaloids from *Argemone mexicana* L. roots. *Anal Bioanal Chem* 2015;407:2581-2589.
103. Ghosh S, Tiwari SS, Kumar B, et al. Identification of potential plant extracts for anti-tick activity against acaricide resistant cattle ticks, *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Exp Appl Acarol* 2015;66:159-171.
104. Croaker A, King GJ, Pyne JH, et al. *Sanguinaria canadensis*: traditional medicine, phytochemical composition, biological activities and current uses. *Int J Mol Sci* 2016;17:1414.
105. Lu M, Li K, He H, et al. Systematic characterization of alkaloids in *Eomecon chionantha* Hance using ultrahigh-performance liquid chromatography-tandem quadrupole exactive orbitrap mass spectrometry with a four-step screening strategy. *Rapid Commun Mass Spectrom* 2020;34:e8880.
106. Son SY, Rhee HS, Lee MW, et al. Analysis of benzo[c]phenanthridine alkaloids in *Eschscholtzia californica* cell culture using HPLC-DAD and HPLC-ESI-MS/MS. *Biosci Biotechnol Biochem* 2014;78:1103-1111.
107. Cahliková L, Kucera R, Hostálková A, et al. Identification of pavinane alkaloids in the genera *Argemone* and *Eschscholtzia* by GC-MS. *Nat Prod Commun* 2012;7:1279-1281.
108. Guo W, Lu X, Liu B, et al. Anti-TMV activity and mode of action of three alkaloids isolated from *Chelidonium majus*. *Pest Manag Sci* 2021;77:510-517.

109. Huang XY, Shao ZX, An LJ, et al. New lignanamides and alkaloids from *Chelidonium majus* and their anti-inflammation activity. *Fitoterapia* 2019;139:104359.
110. Qu YF, Gao JY, Wang J, et al. New triterpenoid saponins from the herb *Hylomecon japonica*. *Molecules* 2017;22:1731.
111. Bournine L, Bensalem S, Wauters JN, et al. Identification and quantification of the main active anticancer alkaloids from the root of *Glaucium flavum*. *Int J Mol Sci* 2013;14:23533-23544.
112. Chang L, Hagel JM, Facchini PJ. Isolation and characterization of O-methyltransferases involved in the biosynthesis of glaucine in *Glaucium flavum*. *Plant Physiol* 2015;169:1127-1140.
113. Lin L, Liu YC, Huang JL, et al. Medicinal plants of the genus *Macleaya* (*Macleaya cordata*, *Macleaya microcarpa*): a review of their phytochemistry, pharmacology, and toxicology. *Phytother Res* 2018;32:19-48.
114. Och A, Szewczyk K, Pecio L, et al. UPLC-MS/MS profile of alkaloids with cytotoxic properties of selected medicinal plants of the Berberidaceae and Papaveraceae families. *Oxid Med Cell Longev* 2017;2017:9369872.
115. Murakami T, Oominami H, Matsuda H, et al. Bioactive saponins and glycosides. XVIII. Nortriterpene and triterpene oligoglycosides from the fresh leaves of *Euptelea polyandra* Sieb. et Zucc. (2): Structures of eupteleasaponins VI, VI acetate, VII, VIII, IX, X, XI, and XII. *Chem Pharm Bull* 2001;49:741-746.
116. Xiao PG, Wang LW, Lv SJ, et al. Statistical analysis of the ethnopharmacologic data based on Chinese medicinal plants by electronic computer I. Magnoliidae. *Chin J Integr Tradit West Med* (Chin) 1986;6:253-256.
117. Mokhber-Dezfuli N, Saeidnia S, Gohari AR, et al. Phytochemistry and pharmacology of berberis species. *Pharmacogn Rev* 2014;8:8-15.
118. Imenshahidi M, Hosseinzadeh H. Berberine and barberry (*Berberis vulgaris*): a clinical review. *Phytother Res* 2019;33:504-523.
119. Guo ZY, Zhang ZY, Xiao JQ, et al. Antibacterial effects of leaf extract of *Nandina domestica* and the underlined mechanism. *Evid Based Complement Alternat Med* 2018;2018:8298151.
120. Jo A, Yoo HJ, Lee M. Robustaflavone isolated from *Nandina domestica* using bioactivity-guided fractionation downregulates inflammatory mediators. *Molecules* 2019;24:1789.
121. Son Y, An Y, Jung J, et al. Protopine isolated from *Nandina domestica* induces apoptosis and autophagy in colon cancer cells by stabilizing p53. *Phytother Res* 2019;33:1689-1696.
122. Kulkarni RR, Lee W, Jang TS, et al. Caffeoyl glucosides from *Nandina domestica* inhibit LPS-induced endothelial inflammatory responses. *Bioorg Med Chem Lett* 2015;25:5367-5371.
123. Bi SF, Zhu GQ, Wu J, et al. Chemical composition and antioxidant activities of the essential oil from *Nandina domestica* fruits. *Nat Prod Res* 2016;30:362-365.
124. Rocha MP, Campana PRV, Scoaris DO, et al. Combined *in vitro* studies and *in silico* target fishing for the evaluation of the biological activities of *Diphylleia cymosa* and *Podophyllum hexandrum*. *Molecules* 2018;23:3303.
125. Yang Z, Wu Y, Wu S, et al. A combination strategy for extraction and isolation of multi-component natural products by systematic two-phase solvent extraction-(13)C nuclear magnetic resonance pattern recognition and following conical counter-current chromatography separation: podophyllotoxins and flavonoids from *Dysosma versipellis* (Hance) as examples. *J Chromatogr A* 2016;1431:184-196.
126. Guerram M, Jiang ZZ, Sun L, et al. Antineoplastic effects of deoxypodophyllotoxin, a potent cytotoxic agent of plant origin, on glioblastoma U-87 MG and SF126 cells. *Pharmacol Rep* 2015;67:245-252.
127. Li J, Feng J, Luo C, et al. Absolute configuration of podophyllotoxone and its inhibitory activity against human prostate cancer cells. *Chin J Nat Med* 2015;13:59-64.
128. Liang F, Han Y, Gao H, et al. Kaempferol identified by zebrafish assay and fine fractionations strategy from *Dysosma versipellis* inhibits angiogenesis through VEGF and FGF pathways. *Sci Rep* 2015;5:14468.
129. Chen R, Duan R, Wei Y, et al. Flavonol dimers from callus cultures of *Dysosma versipellis* and their *in vitro* neuraminidase inhibitory activities. *Fitoterapia* 2015;107:77-84.
130. Li J, Sun H, Jin L, et al. Alleviation of podophyllotoxin toxicity using coexisting flavonoids from *Dysosma versipellis*. *PLoS One* 2013;8:e72099.
131. Zhou FZ, Wang X, Dai AY, et al. Effects of *Sinopodophyllum hexandrum* on apoptosis in K562 cells. *J Southern Med Univ (Chin)* 2016;37:226-231.
132. Ma H, He X, Yang Y, et al. The genus *Epimedium*: an ethnopharmacological and phytochemical review. *J Ethnopharmacol* 2011;134:519-541.
133. Wu L, Du ZR, Xu AL, et al. Neuroprotective effects of total flavonoid fraction of the *Epimedium koreanum* Nakai extract on dopaminergic neurons: *in vivo* and *in vitro*. *Biomed Pharmacother* 2017;91:656-663.
134. Chen WF, Wu L, Du ZR, et al. Neuroprotective properties of icariin in MPTP-induced mouse model of Parkinson's disease: involvement of PI3K/Akt and MEK/ERK signaling pathways. *Phytomedicine* 2017;25:93-99.
135. Mo ZT, Li WN, Zhai YR, et al. The effects of icariin on the expression of HIF-1 α , HSP-60 and HSP-70 in PC12 cells suffered from oxygen-glucose deprivation-induced injury. *Pharm Biol* 2017;55:848-852.
136. Cho JH, Jung JY, Lee BJ, et al. *Epimedium Herba*: a promising herbal medicine for neuroplasticity. *Phytother Res* 2017;31:838-848.
137. Arens H, Fischer H, Leyck S, et al. Antiinflammatory compounds from *Plagiorhegma dubium* cell culture. *Planta Med* 1985;51:52-56.
138. Kim JM, Jung HA, Choi JS, et al. Comparative analysis of the anti-inflammatory activity of Huang-lian extracts in lipopolysaccharide-stimulated RAW264.7 murine macrophage-like cells using oligonucleotide microarrays. *Arch Pharm Res* 2010;33:1149-1157.
139. Jiang Y, Yu L, Wang MH. N-trans-feruloyltyramine inhibits LPS-induced NO and PGE₂ production in RAW 264.7 macrophages: involvement of AP-1 and MAP kinase signalling pathways. *Chem Biol Interact* 2015;235:56-62.
140. Yu LL, Hu WC, Ding G, et al. Gusanlungionosides A-D, potential tyrosinase inhibitors from *Arcangelisia gusanlung*. *J Nat Prod* 2011;74:1009-1014.
141. Sriwilajareon N, Petmitr S, Mutirangura A, et al. Stage specificity of *Plasmodium falciparum* telomerase and its inhibition by berberine. *Parasitol Int* 2002;51:99-103.
142. Nguyen-Pouplin J, Tran H, Tran H, et al. Antimalarial and cytotoxic activities of ethnopharmacologically selected medicinal plants from South Vietnam. *J Ethnopharmacol* 2007;109:417-427.
143. Yu H, He ZM, Shi K, et al. Study on antidepressant effect of total alkaloids of *Fibraurea recisa*. *China J Chin Mater Med* (Chin) 2021;46:3678-3686.
144. Zhang M, Chen W, Zong Y, et al. Cognitive-enhancing effects of fibrauretinone on A β ₁₋₄₂-induced Alzheimer's disease by compatibilization with ginsenosides. *Neuropeptides* 2020;82:102020.
145. Ma C, Du F, Yan L, et al. Potent activities of roemerine against *Candida albicans* and the underlying mechanisms. *Molecules* 2015;20:17913-17928.
146. Manosroi A, Akazawa H, Akihisa T, et al. *In vitro* anti-proliferative activity on colon cancer cell line (HT-29) of Thai medicinal plants selected from Thai/Lanna medicinal plant recipe database "MANOSROI III". *J Ethnopharmacol* 2015;161:11-17.
147. Gong S, Xu D, Zou F, et al. (-)-Curine induces cell cycle arrest and cell death in hepatocellular carcinoma cells in a p53-independent way. *Biomed Pharmacother* 2017;89:894-901.
148. Bhagya N, Chandrashekar KR, Prabhu A, et al. Tetrandrine isolated from *Cyclea peltata* induces cytotoxicity and apoptosis through ROS and caspase pathways in breast and pancreatic cancer cells. *In Vitro Cell Dev Biol Anim* 2019;55:331-340.
149. Vijayan D, Cheethaparambil A, Pillai GS, et al. Molecular authentication of *Cissampelos pareira* L. var. *hirsuta* (Buch.-Ham. Ex DC.) Forman, the genuine source plant of ayurvedic raw drug 'Patha', and its other source plants by ISSR markers. *3 Biotech* 2014;4:559-562.

150. Rogosnitzky M, Okediji P, Koman I. Cepharanthine: a review of the antiviral potential of a Japanese-approved alopecia drug in COVID-19. *Pharmacol Rep* 2020;72:1509-1516.
151. Jayasinghe UL, Kumarihamy BM, Bandara AG, et al. Antifeedant activity of some Sri Lankan plants. *Nat Prod Res* 2003;17:5-8.
152. Wei ZZ, Qin QP, Chen JN, et al. Oxoisoaporphine as potent telomerase inhibitor. *Molecules* 2016;21:1534.
153. Zhou ZG, Zhang CY, Fei HX, et al. Phenolic alkaloids from *Menispermum dauricum* inhibits BxPC-3 pancreatic cancer cells by blocking of Hedgehog signaling pathway. *Pharmacogn Mag* 2015;11:690-697.
154. Zhang HF, Wu D, Du JK, et al. Inhibitory effects of phenolic alkaloids of *Menispermum dauricum* on gastric cancer *in vivo*. *Asian Pac J Cancer Prev* 2014;15:10825-10830.
155. Li YY, Zheng G, Liu L. Bioinformatics based therapeutic effects of *Sinomenium acutum*. *Chin J Integr Med* 2019;25:122-130.
156. Zhao XX, Peng C, Zhang H, et al. *Sinomenium acutum*: a review of chemistry, pharmacology, pharmacokinetics, and clinical use. *Pharm Biol* 2012;50:1053-1061.
157. Hong GL, Park SR, Jung DY, et al. The therapeutic effects of *Stauntonia hexaphylla* in benign prostate hyperplasia are mediated by the regulation of androgen receptors and 5 α -reductase type 2. *J Ethnopharmacol* 2020;250:112446.
158. Li J, Du K, Liu D, et al. New nor-oleanane triterpenoids from the fruits of *Stauntonia brachyanthera* with potential anti-inflammation activity. *Nat Prod Res* 2020;34:915-922.
159. Yang J, Xiong Q, Zhang J, et al. The protective effect of *Stauntonia chinensis* polysaccharide on CCl₄-induced acute liver injuries in mice. *Int J Biomed Sci* 2014;10:16-20.
160. Liu D, Li S, Qi JQ, et al. The inhibitory effects of nor-oleanane triterpenoid saponins from *Stauntonia brachyanthera* towards UDP-glucuronosyltransferases. *Fitoterapia* 2016;112:56-64.
161. Liu XL, Li S, Meng DL. Anti-gout nor-oleanane triterpenoids from the leaves of *Stauntonia brachyanthera*. *Bioorg Med Chem Lett* 2016;26:2874-2879.
162. Tian B, Tian M, Huang SM. Advances in phytochemical and modern pharmacological research of *Rhizoma Corydalis*. *Pharm Biol* 2020;58:265-275.
163. Gupta PC, Sharma N, Rao ChV. A review on ethnobotany, phytochemistry and pharmacology of *Fumaria indica* (Fumitory). *Asian Pac J Trop Biomed* 2012;2:665-669.
164. Yuan HL, Zhao YL, Qin XJ, et al. Diverse isoquinolines with anti-inflammatory and analgesic bioactivities from *Hypecoum erectum*. *J Ethnopharmacol* 2021;270:113811.
165. Doncheva T, Kostova N, Valcheva V, et al. Hypepontine, a new quaternary alkaloid with antimicrobial properties. *Nat Prod Res* 2020;34:668-674.
166. Zhang Q, Luan G, Ma T, et al. Application of chromatography technology in the separation of active alkaloids from *Hypecoum leptocarpum* and their inhibitory effect on fatty acid synthase. *J Sep Sci* 2015;38:4063-4070.
167. Akaberi T, Shourgashti K, Emami SA, et al. Phytochemistry and pharmacology of alkaloids from *Glaucium spp.* *Phytochemistry* 2021;191:112923.
168. Gilca M, Gaman L, Panait E, et al. *Chelidonium majus*—an integrative review: traditional knowledge versus modern findings. *Forsch Komplementmed* 2010;17:241-248.
169. Arcos-Martínez AI, Muñoz-Muñiz OD, Domínguez-Ortiz MÁ, et al. 2016. Anxiolytic-like effect of ethanolic extract of *Argemone mexicana* and its alkaloids in Wistar rats. *Avicenna J Phytomed* 2016;6:476-488.
170. Rubio-Pina J, Vazquez-Flota F. Pharmaceutical applications of the benzylisoquinoline alkaloids from *Argemone mexicana* L. *Curr Top Med Chem* 2013;13:2200-2207.
171. Kamalakannan S, Ananth S, Murugan K, et al. Bio fabrication of silver nanoparticle from *Argemone mexicana* for the control of *Aedes albopictus* and their antimicrobial activity. *Curr Pharm Biotechnol* 2016;17:1285-1294.
172. Singh S, Verma M, Malhotra M, et al. Cytotoxicity of alkaloids isolated from *Argemone mexicana* on SW480 human colon cancer cell line. *Pharm Biol* 2016;54:740-745.
173. Li MX, Wang JH, He XR, et al. Phytochemical and biological studies of plants from the genus *Meconopsis*. *Chem Biodivers* 2010;7:1930-1948.
174. Yoshikawa M, Murakami T, Oomiiami H, et al. Bioactive saponins and glycosides. VII. Nortriterpene oligoglycosides with gastroprotective activity from the fresh leaves of *Euptelea polyandra* Sieb. et Zucc. (1): Structures of Eupteleasaponins I, II, III, IV, V, and V acetate. *Chem Pharm Bull* 2000;48:1045-1050.
175. Suzuki R, Fukami S, Tomomura M, et al. Screening for natural medicines effective for the treatment of osteoporosis. *J Nat Med* 2019;73:331-337.
176. Hao DC, ed. *Ranunculales medicinal plants: biodiversity, chemodiversity and pharmacotherapy*. London: Academic Press;2018:1-350.
177. Liscombe DK, MacLeod BP, Loukanina N, et al. Evidence for the monophyletic evolution of benzylisoquinoline alkaloid biosynthesis in angiosperms. *Phytochemistry* 2005;66:2501-2520.
178. Petruczynik A, Plech T, Tuzimski T, et al. Determination of selected isoquinoline alkaloids from *Mahonia aquifolia*; *Meconopsis cambrica*; *Corydalis lutea*; *Dicentra spectabilis*; *Fumaria officinalis*; *Macleaya cordata* extracts by HPLC-DAD and comparison of their cytotoxic activity. *Toxins* 2019;11:575.
179. Li P, Shen J, Li Y, et al. Metabolite profiling based on UPLC-Q-TOF-MS/MS and the biological evaluation of medicinal plants of Chinese *Dichocarpum* (Ranunculaceae). *Chem Biodivers* 2021;18:e2100432.
180. Hao DC, Yang L. Drug metabolism and disposition diversity of *Ranunculales phytometabolites*: a systems perspective. *Expert Opin Drug Metab Toxicol* 2016;12:1047-1065.
181. Xiao PG, Xiao W, Xu LJ, et al. *Coptidis Rhizoma* and Chinese herbal medicines which contain berberine-type alkaloids. *Modern Chin Med* 2016;18:1381-1385.
182. Hao DC, He CN, Shen J, et al. Anticancer chemodiversity of Ranunculaceae medicinal plants: molecular mechanisms and functions. *Curr Genomics* 2017;18:39-59.
183. Weber C, Opatz T. Bisbenzylisoquinoline alkaloids. *Alkaloids Chem Biol* 2019;81:1-114.
184. Zhang J, Chen L, Sun J. Oxoisoaporphine alkaloids: prospective anti-Alzheimer's disease, anticancer, and antidepressant agents. *Chem Med Chem* 2018;13:1262-1274.
185. Peng P, Jia D, Cao L, et al. Akebia saponin E, as a novel PI3K inhibitor, induces lysosome-associated cytoplasmic vacuolation to inhibit proliferation of hepatocellular carcinoma cells. *J Ethnopharmacol* 2021;266:113446.
186. Almeida IV, Fernandes LM, Biazzi BI, et al. Evaluation of the anticancer activities of the plant alkaloids sanguinarine and chelerythrine in human breast adenocarcinoma cells. *Anticancer Agents Med Chem* 2017;17:1586-1592.
187. Hao DC, Zhang Y, He CN, et al. Distribution of therapeutic efficacy of Ranunculales plants used by ethnic minorities on the phylogenetic tree of Chinese species. *Evid Based Complement Alternat Med* 2022;2022:9027727.
188. Hao DC, Li P, Xiao PG, et al. Dissection of full-length transcriptome and metabolome of *Dichocarpum* (Ranunculaceae): implications in evolution of specialized metabolism of Ranunculales medicinal plants. *Peer J* 2021;9:e12428.
189. Hagel JM, Mandal R, Han B, et al. Metabolome analysis of 20 taxonomically related benzylisoquinoline alkaloid-producing plants. *BMC Plant Biol* 2015;15:220.
190. Liu Y, Wang B, Shu S, et al. Analysis of the *Coptis chinensis* genome reveals the diversification of protoberberine-type alkaloids. *Nat Commun* 2021;12:3276.
191. Angiosperm Phylogeny Group. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Bot J Linnean Soc* 2009;161:105-121.

(Accepted March 7, 2022)

Edited by YU Ming-zhu