# Two cases of neonatal hyperglycemia caused by a homozygous COQ9 stop-gain variant

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## Keywords

Coenzyme Q10, Mitochondrial disease, Neonatal diabetes

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## **ABSTRACT**

Neonatal diabetes mellitus (NDM) is a monogenic condition diagnosed <6 months of age with >40 genetic causes. International guidelines recommend referral for genetic testing immediately after diagnosis since the genetic result guides clinical management. We used next-generation sequencing to identify a homozygous pathogenic variant, p.(Arg244\*), in COQ9 in 2 individuals referred for NDM testing. Both had insulin-treated hyperglycemia, severe structural brain defects, dysmorphic features, and lactic acidosis. Recessive loss-of-function variants in COQ9 cause Coenzyme Q10 deficiency-5, a multi-system mitochondrial disease, with 7 cases reported. Neonatal hyperglycemia has not been reported in any of these cases but has been described for two other Coenzyme Q10 disorders caused by variants in COQ2 and COQ4. Our report shows that individuals with COQ9-related disease can present with neonatal hyperglycemia, expanding the clinical spectrum of this disorder. We recommend the inclusion of COQ9, as well as COQ2 and COQ4, to gene panels used for NDM testing.

# **INTRODUCTION**

Neonatal diabetes mellitus (NDM) is a monogenic condition diagnosed in the first 6 months of life. Over 40 different genetic aetiologies have been described in individuals with NDM<sup>1</sup>, explaining >85% of cases<sup>2</sup>.

A genetic diagnosis guides clinical management in individuals with NDM and determines which treatment is most appropriate (for example, sulphonylureas in individuals with activating variants in the potassium channel genes *KCNJ11*<sup>3,4</sup> and *ABCC8*<sup>5,6</sup>), and whether the diabetes occurs in isolation (isolated NDM) or is one of the first manifestations of a multiorgan disease (syndromic NDM). The genes involved in the pathogenesis of syndromic NDM have essential roles in biological pathways; for example, *GATA6*, crucial in organ development<sup>7</sup>, and *EIF2AK3*, crucial for the endoplasmic reticulum stress response<sup>8</sup>.

Recent findings have highlighted mitochondrial dysfunction as another mechanism leading to NDM with the identification of novel NDM aetiological genes, such as  $NARS2^9$  and  $TARS2^{10}$ , both encoding proteins essential for mitochondrial protein translation. The importance of mitochondrial function

analysis of a panel including all the known NDM genes<sup>16</sup> is the most common approach.

Here, we describe two individuals who were referred for NDM genetic testing following presentation with neonatal hyperglycemia and extra-pancreatic features. In both individuals, a homozygous stop-gain variant was detected in *COQ9*, a

tinely included on NDM gene testing panels.

to beta cells is further supported by previous studies identifying

recessive variants in CYC1, which encodes a subunit of Complex III of the mitochondrial respiratory chain 11,12 and mito-

chondrial DNA deletions causing Pearson syndrome in 3 and 6

individuals with NDM, respectively<sup>13</sup>. Furthermore, the

m.3243A>G variant of the mitochondrial DNA is a cause of

later onset diabetes in individuals affected by Maternally inher-

International consensus guidelines recommend that all

infants diagnosed with diabetes in the first 6 months of life be immediately referred for genetic testing 15. Targeted sequencing

## **MATERIALS AND METHODS**

ited diabetes and deafness (MIDD)<sup>14</sup>.

Probands 1 and 2 had hyperglycaemia detected on the first and third days of life, respectively, which required insulin treatment.

gene essential for mitochondrial function, which is not rou-

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Additional clinical details are summarized in Table 1 and in the Appendix S1.

DNA samples from both probands and the parents of proband 1 were sent to the Exeter genomics laboratory for NDM genetic testing. Samples from proband 2's parents and deceased sibling were not available for testing. All known genetic causes of NDM were tested in both probands using a combination of Sanger sequencing, targeted next generation sequencing (tNGS<sup>16</sup>) and methylation-specific PCR to assess the imprinted chr6q24 transient neonatal diabetes region. Nine years after the original referral, whole exome sequencing (WES) was performed on the samples from proband 1 and her parents to identify potential novel genetic causes of NDM. The study complied with the Declaration of Helsinki, with informed consent obtained from the parents.

Exome sequencing and targeted next-generation sequencing were performed as previously described<sup>7,16</sup>. An in-house script was used to prioritize rare (allele frequency in GnomADv4<sup>17</sup> <0.001%) *de novo*, homozygous, and compound heterozygous coding variants. *De novo* variants were considered only if the ratio between the reads supporting each allele was >0.2. Variant classification was carried out using the ACMG guidelines<sup>18</sup>.

After the identification of a likely genetic cause in Proband 1, replication studies were undertaken to search for additional cases. RNA baits targeting the 9 coding exons and intron/exon boundaries of the *COQ9* gene (NM\_020312.3) were added to our custom-designed tNGS panel<sup>16</sup>. Details of the RNA baits are available upon request. The replication cohort consisted of 168 individuals with genetically unsolved NDM, which included proband 2. Due to privacy concerns, the research data supporting this publication are not publicly available.

#### **RESULTS**

Exome sequencing data in Proband 1 identified only 1 variant which matched all the filtering criteria: the homozygous c.730C>T, p.(Arg244\*) stop-gain variant in COQ9. Both unaffected parents were heterozygous for the variant. Replication studies performed on 168 individuals with genetically unsolved NDM identified the same homozygous p.(Arg244\*) variant in the sample from proband 2. Both probands were reported to be of Pakistani origin but were not known to be related. No further disease-causing variants in COQ9 were identified in the remaining 167 individuals.

The COQ9 gene encodes Coenzyme Q9, a mitochondrial protein involved in the biosynthesis of Coenzyme Q10 (CoQ10), a critical component of the electron transport chain in the synthesis of ATP. Biallelic loss-of-function variants in COQ9 cause Primary Coenzyme Q10 deficiency 5 (COQ10D5; OMIM: 614654), a multiorgan condition with features including intrauterine growth retardation (IUGR), microcephaly, neurological and cardiovascular disease, and metabolic lactic acidosis.

The p.(Arg244\*) variant identified in the two probands in this study was classified as pathogenic according to the ACMG

guidelines. This variant results in the insertion of a premature stop codon in exon 7 of 9 of the COQ9 gene, and it is therefore predicted to cause loss of the COQ9 protein through nonsense-mediated decay of the mRNA transcript. The p.(Arg244\*) variant has been previously reported in 1 individual with COQ10D5<sup>19</sup> and it is listed in ClinVar as 'Pathogenic' (Variation ID: 431, 3 submissions). Furthermore, studies performed on an open muscle biopsy and cultured fibroblasts from a previously reported individual with the same homozygous p.(Arg244\*) COQ9 variant showed a strong quantitative deficit of CoQ10 and significantly reduced CoQ10 biosynthesis<sup>19</sup>. This result confirmed a diagnosis of COQ10D5 in Probands 1 and 2.

#### DISCUSSION

We report the identification of a homozygous p.(Arg244\*) loss-of-function variant in *COQ9* in two individuals referred for NDM genetic testing, following presentation with hyperglycemia in the first 3 days of life. Both probands had severe structural brain defects, IUGR, and arthrogryposis. The clinical features observed in both individuals were consistent with the genetic diagnosis of COQ10D5.

Our results expand the phenotypic spectrum caused by recessive COQ9 variants. Hyperglycemia has not been previously reported in any of the 7 (3 probands and 4 siblings) individuals with COQ10D5 caused by biallelic loss-of-function COQ9 variants described in the literature (Table 1). Probands 1 and 2 in our study had hyperglycaemia on the first and third day of life, respectively, which required insulin treatment. In proband 2, the hyperglycaemia was transient, requiring insulin treatment until the age of 5 weeks. This individual was off insulin for 1 week before their death at 6 weeks. In proband 1, the hyperglycaemia persisted until their death at the age of 3 weeks. Since both individuals died in the neonatal period, we do not know whether the difference in insulin requirement reflects a genuine variability in the phenotype between these two patients, or whether Proband 1's insulin requirement might have also diminished with age. It is not known whether hyperglycaemia was measured in the 7 previously reported patients with COQ9 variants, nor is it known if the one surviving individual who was 3 years of age at the time of reporting had hyperglycaemia.

Neonatal hyperglycaemia has been reported as a feature in 7 individuals with COQ10D resulting from COQ2 (n = 5) or COQ4 (n = 2) pathogenic variants<sup>20–23</sup>. In all these cases, neonatal hyperglycemia was a presenting feature of the disorder detected before the age of 6 months (mean age of diagnosis: 63.9 days, range 1–150 days). A transgenic mouse model harboring a homozygous Coq9 truncating variant showed significant loss of ATP and respiratory complex I activity in neuronal cells leading to encephalomyopathy<sup>24</sup>, which is consistent with the phenotype observed in COQ10D5 patients. No functional studies have been performed to assess the effect of pathogenic variants in COQ2, COQ4, or COQ9 in pancreatic  $\beta$ -cells.

 Table 1 | Clinical features of individuals reported with pathogenic COQ9 variants

	Proband 1 (this study)	Proband 1 (this Proband 2 (this Proband 3 <sup>19</sup> Proband 4 <sup>27</sup> study)	Proband 3 <sup>19</sup>	Proband 4 <sup>27</sup>	Proband 5.1 <sup>28</sup>	Proband 5.2 <sup>28</sup>	Proband 5.3 <sup>28</sup>	Proband 5,4 <sup>28</sup>	Proband 6 <sup>29</sup>
COQ9 variant	p.(Arg244*) c.730C>T homozygous	p.(Arg244*) c730C>T homozygous	p.(Arg244*) c.730C>T homozygous	p.(Ser127_Arg202del) c.521 + 1del homozygous	p(Ser127_Arg202del) c.521+27>C p(Ala203_Asp237del) c.711+3G>C compound beteroxorous	p(Sert 27_Arg202del) c521+27>C p(Aa203_Asp237del) c711+3G>C compound betenoxorous	p.(Ser127_Arg202del) c.521+27>C p.(Ala203_Asp237del) c.711+3G>C compound heteroporous	p(Ser127_Arg202del) c521+27>C p(Ala203_Asp237del) c711+3G>C compound heteroxycous	p,(Gly129Valfs) c. 384delG homozygous
Age at clinical	1 day	1 day	6 h	Birth	19 weeks gestation	23 weeks gestation	22 weeks gestation	19 weeks gestation	Birth
Age and cause of death	Before 3 weeks, respiratory failure	6 weeks, seizures and cyanosis	2 years, chest infection	18 days, cardio-respiratory failure	3 days	Pregnancy termination at 28 weeks	12 h at extubation	Pregnancy termination at 19 weeks	Alive, 3 years
IUGR (bwt, gestation)	Yes (1,940 g 37 weeks)	Yes (1,660 g 39 weeks)	Unknown	Yes (1,440 g 36 weeks)	Yes (1,460 g 33 weeks)	Yes (bwt unknown, 28 weeks)	Yes (1,580 g 34 weeks)	Yes (bwt unknown, 19 weeks)	Yes (2000 g 38 weeks)
Oligohydramnios	Not reported	Yes	Not reported	Yes	Yes	Yes	Yes	Not reported	Yes
Hyperglycemia	Detected day 1	Detected day 3	Not reported	Not reported	Not reported	n/a 2/3	Not reported	n/a	Not reported
				lactic acidosis		5		5	elevated lactate
Microcephaly	Not reported	Yes	Yes	Yes	Yes	n/a	Yes	n/a	Yes
Arthrogryposis	Yes	Yes	Not reported	Yes	Yes	n/a	Not reported	n/a	Yes
Brain imaging findings	Cerebellar and	Dandy-Walker	Cerebral and	Leigh-like syndrome	Hydrocephalus,	n/a	n/a	n/a	Cerebellar
	optic	malformation	cerebellar		lobulated cysts in				hypoplasia
	пуроріама		attopriy		II OFFICIAL TODGES				verrins and brain
									stem, enlarged lateral ventricles.
									corpus callosum
									agenesis, cortical
									atrophy
Neuro-developmental features	Seizures	Seizures	selzures, developmental delay, dystonia	sezures, nypotonia	Not reported	Not reported	Abnormal Jerking movements, autopsy showed signs of laich sandrome laich sandrome	Not reported	seizures, developmental delay, hypotonia
Additional features	Hypertrophic cardiomyopathy	Umbilical hemia, dysmorphic features, increased cortical	Poor feeding, hypothermia, increased tone, renal	Poor respiratory efforts, generalized cyanosis, recurrent apnea and	Pulmonary hypertension, respiratory depression, fixed	Echogenic bladder, large cystic kidneys	Hypotension, echogenic bowel, dysmorphic features	Echogenic bowel and kidneys	Respiratory distress, cardiomyopathy, cystic kidneys, dysmorphic
		echogenicity in kidneys, pulmonary hypertension, small ventricular septal defect	tubulopathy, left ventricular hypertrophy	bradycardia	dilated pupils, brisk reflexes, dilated ventricles heart, bradycardia, cardiomegaly, dysmorphic features				features

bwt, birthweight; n/a, not applicable.

An early genetic diagnosis of COQ10D is crucially important as CoQ10 replacement therapy may be beneficial in some patients<sup>25</sup>. The genetic cause of NDM had remained undetermined in our two patients following routine testing, as COQ9 was not included on the tNGS gene panel, resulting in a delay in their genetic diagnosis. Many laboratories are now adopting whole exome and whole genome sequencing as first-line genetic tests for conditions such as NDM, and rapid genome sequencing has proven particularly beneficial for individuals with mitochondrial disorders<sup>26</sup>. These approaches will benefit patients such as those reported in this study as they will allow comprehensive testing of a wider panel of genes such as COQ9.

In conclusion, we report neonatal hyperglycaemia as a presenting feature of COQ10D5 in two individuals with a homozygous loss-of-function variant in COQ9. We recommend that analysis of the  $CoQ_{10}$  genes previously associated with NDM, COQ2, COQ4, and now COQ9, is included in genetic testing panels for NDM.

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## **DISCLOSURE**

The authors declare no conflict of interest.

Approval of the research protocol: The protocol for this research project has been approved by the Genetic Beta-Cell Research Bank at the University of Exeter (Ethics approval 517/WA/0327).

Informed consent: The study conforms to the provisions of the Declaration of Helsinki, with written, informed consent obtained from the parents of the subjects. Only anonymized data was used in this report.

Registry and the registration number of the study/trial: N/A. Animal studies: N/A.

#### REFERENCES

- 1. Barbetti F, Deeb A, Suzuki S. Neonatal diabetes mellitus around the world: Update 2024. *J Diabetes Investig* 2024; 15: 1711–1724.
- 2. De Franco E, Flanagan SE, Houghton JA, *et al.* The effect of early, comprehensive genomic testing on clinical care in neonatal diabetes: An international cohort study. *Lancet* 2015: 386: 957–963
- 3. Bowman P, Day J, Torrens L, *et al.* Cognitive, neurological, and behavioral features in adults with KCNJ11 neonatal diabetes. *Diabetes Care* 2019: 42: 215–224.
- 4. Pearson ER, Flechtner I, Njolstad PR, et al. Switching from insulin to oral sulfonylureas in patients with diabetes due to Kir6.2 mutations. *N Engl J Med* 2006; 355: 467–477.
- Bowman P, Mathews F, Barbetti F, et al. Long-term follow-up of glycemic and neurological outcomes in an international series of patients with sulfonylurea-treated ABCC8 permanent neonatal diabetes. *Diabetes Care* 2021; 44: 35–42.
- 6. Rafiq M, Flanagan SE, Patch AM, et al. Effective treatment with oral sulfonylureas in patients with diabetes due to sulfonylurea receptor 1 (SUR1) mutations. *Diabetes Care* 2008; 31: 204–209.
- 7. Allen HL, Flanagan SE, Shaw-Smith C, et al. GATA6 haploinsufficiency causes pancreatic agenesis in humans. *Nat Genet* 2011; 44: 20–22.
- 8. Delepine M, Nicolino M, Barrett T, et al. EIF2AK3, encoding translation initiation factor 2-alpha kinase 3, is mutated in patients with Wolcott-Rallison syndrome. *Nat Genet* 2000; 25: 406–409.
- Yagasaki H, Sano F, Narusawa H, et al. Compound heterozygous variants of the NARS2 gene in siblings with developmental delay, epilepsy, and neonatal diabetes syndrome. Am J Med Genet A 2022; 188: 2466– 2471.
- 10. Donis R, Patel KA, Wakeling MN, *et al.* A homozygous TARS2 variant is a novel cause of syndromic neonatal diabetes. *Diabet Med* 2024; 42: e15471.
- 11. Anastasio N, Tarailo-Graovac M, Al-Khalifah R, *et al.* Mitochondrial complex III deficiency with ketoacidosis and hyperglycemia mimicking neonatal diabetes. *JIMD Rep* 2017; 31: 57–62.
- 12. Gaignard P, Menezes M, Schiff M, et al. Mutations in CYC1, encoding cytochrome c1 subunit of respiratory chain complex III, cause insulin-responsive hyperglycemia. Am J Hum Genet 2013; 93: 384–389.
- 13. Ying Y, Liang Y, Luo X, et al. Case report: Clinical and genetic characteristics of Pearson syndrome in a Chinese boy and 139 patients. Front Genet 2022; 13: 802402.
- 14. Perucca-Lostanlen D, Narbonne H, Hernandez JB, et al. Mitochondrial DNA variations in patients with maternally inherited diabetes and deafness syndrome. Biochem Biophys Res Commun 2000; 277: 771–775.

- 15. Greeley SAW, Polak M, Njølstad PR, et al. ISPAD clinical practice consensus guidelines 2022: The diagnosis and management of monogenic diabetes in children and adolescents. *Pediatr Diabetes* 2022; 23: 1188–1211.
- Ellard S, Lango Allen H, De Franco E, et al. Improved genetic testing for monogenic diabetes using targeted next-generation sequencing. *Diabetologia* 2013; 56: 1958– 1963.
- 17. Chen S, Francioli LC, Goodrich JK, *et al.* A genomic mutational constraint map using variation in 76,156 human genomes. *Nature* 2024; 625: 92–100.
- 18. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med 2015; 17: 405–424.
- 19. Duncan AJ, Bitner-Glindzicz M, Meunier B, et al. A nonsense mutation in COQ9 causes autosomal-recessive neonatal-onset primary coenzyme Q10 deficiency: A potentially treatable form of mitochondrial disease. Am J Hum Genet 2009; 84: 558–566.
- 20. Dinwiddie DL, Smith LD, Miller NA, *et al.* Diagnosis of mitochondrial disorders by concomitant next-generation sequencing of the exome and mitochondrial genome. *Genomics* 2013; 102: 148–156.
- 21. Eroglu FK, Ozaltin F, Gönç N, et al. Response to early coenzyme Q10 supplementation is not sustained in CoQ10 deficiency caused by CoQ2 mutation. *Pediatr Neurol* 2018; 88: 71–74.
- 22. Lu M, Zhou Y, Wang Z, et al. Clinical phenotype, in silico and biomedical analyses, and intervention for an east Asian

- population-specific c.370G>a (p.G124S) COQ4 mutation in a Chinese family with CoQ10 deficiency-associated Leigh syndrome. *J Hum Genet* 2019; 64: 297–304.
- 23. Mollet J, Giurgea I, Schlemmer D, et al. Prenyldiphosphate synthase, subunit 1 (PDSS1) and OH-benzoate polyprenyltransferase (COQ2) mutations in ubiquinone deficiency and oxidative phosphorylation disorders. *J Clin Invest* 2007; 117: 765–772.
- 24. García-Corzo L, Luna-Sánchez M, Doerrier C, et al. Dysfunctional Coq9 protein causes predominant encephalomyopathy associated with CoQ deficiency. *Hum Mol Genet* 2013: 22: 1233–1248.
- 25. Wang Y, Hekimi S. The efficacy of coenzyme Q(10) treatment in alleviating the symptoms of primary coenzyme Q(10) deficiency: A systematic review. *J Cell Mol Med* 2022; 26: 4635–4644.
- 26. Ball M, Bouffler SE, Barnett CB, et al. Critically unwell infants and children with mitochondrial disorders diagnosed by ultrarapid genomic sequencing. *Genet Med* 2025; 27: 101293.
- 27. Danhauser K, Herebian D, Haack TB, et al. Fatal neonatal encephalopathy and lactic acidosis caused by a homozygous loss-of-function variant in COQ9. Eur J Hum Genet 2016; 24: 450–454.
- 28. Smith AC, Ito Y, Ahmed A, et al. A family segregating lethal neonatal coenzyme Q(10) deficiency caused by mutations in COQ9. J Inherit Metab Dis 2018; 41: 719–729.
- 29. Olgac A, Öztoprak Ü, Kasapkara ÇS, et al. A rare case of primary coenzyme Q10 deficiency due to COQ9 mutation. *J Pediatr Endocrinol Metab* 2020; 33: 165–170.

## **SUPPORTING INFORMATION**

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Appendix S1. Supplementary Material.