

1 Closing the genome of *Teredinibacter turnerae* T7902 by long-read
2 nanopore sequencing
3

4 Mark T. Gasser^a | Annie Liu^a | Ron Flatau^b | Marvin Altamia^b | Claire Marie Filone^a | Dan
5 Distel^{b#}
6

7 ^aJohns Hopkins University Applied Physics Laboratory, Laurel, Maryland, USA 20723

8 ^bOcean Genome Legacy Center, Northeastern University, Nahant, Massachusetts, USA 01908
9

10 Running Title: Closing the genome of *T. turnerae* T7902
11

12 MTG: mark.gasser@jhuapl.edu (ORCID: 0000-0002-9396-4954)

13 AL: annie.liu@jhuapl.edu (ORCID: 0009-0002-3224-6008)

14 RF: r.flatau@northeastern.edu (ORCID: 0000-0002-4357-5870)

15 MAA: marvin.altamia@gmail.com (ORCID: 0000-0002-8625-767X)

16 CMF: claire.marie.filone@jhuapl.edu (ORCID: 0000-0002-4041-3948)

17 DLD: d.distel@northeastern.edu (ORCID: 0000-0002-3860-194X) #Corresponding author
18

19 Abstract

20 We present the complete closed circular genome sequence derived from Oxford Nanopore

21 sequencing of the shipworm endosymbiont *Teredinibacter turnerae* T7902 (DSM 15152, ATCC

22 39867), originally isolated from the shipworm *Lyrodus pedicellatus* (1). This sequence will aid in

23 the comparative genomics of shipworm endosymbionts and the understanding of host-
24 symbiont evolution.

25 [Announcement](#)

26 *Teredinibacter* species are cellulolytic gammaproteobacteria (Cellvibrionaceae) that occur as
27 intracellular endosymbionts of wood-boring bivalves (Teredinidae) (1–4), commonly known as
28 shipworms. Strain T7902 was isolated from the gills of a single specimen of the shipworm
29 *Lyrodus pedicellatus* collected in Long Beach, CA, in 1979 and was the second strain of *T.*
30 *turnerae* brought into pure culture (1). It is the original representative of *T. turnerae* Clade II,
31 one of two distinct clades previously identified among *T. turnerae* strains (5, 6). Gills were
32 dissected, washed in sterile seawater, and homogenized in shipworm basal medium (SBM) (7),
33 and the homogenate was streaked onto a culture plate containing 0.9% agar and SBM at pH 8.0
34 supplemented with 0.2% w/v powdered cellulose (Sigmacell Type 100; Sigma-Aldrich) and 5
35 mM ammonium chloride. Individual colonies were picked and restreaked on fresh plates until a
36 pure clonal isolate was obtained. The original genome sequence of *T. turnerae* T7902 was
37 published to GenBank (GCA_000379165.1) but has not been described previously in peer-
38 reviewed literature. This sequence was completed on 2012-05-22 at the DOE Joint Genome
39 Institute under award 10.46936/10.25585/60001419 using 454 GS FLX Titanium and Illumina
40 HiSeq 2000 sequencing platforms. It was assembled using Velvet v. 1.0.13 (8) and ALLPATHS v.
41 R40295 (9), resulting in an improved high-quality draft assembly comprised of 72 scaffolds with
42 76 contigs. At the time of this work, the genomes of nine strains of *T. turnerae* were publicly
43 available at the National Center for Biotechnology Information (NCBI), and only one strain
44 genome, T7901 (Clade I, Genbank: GCA_000023025.1), was complete and closed. Here, we

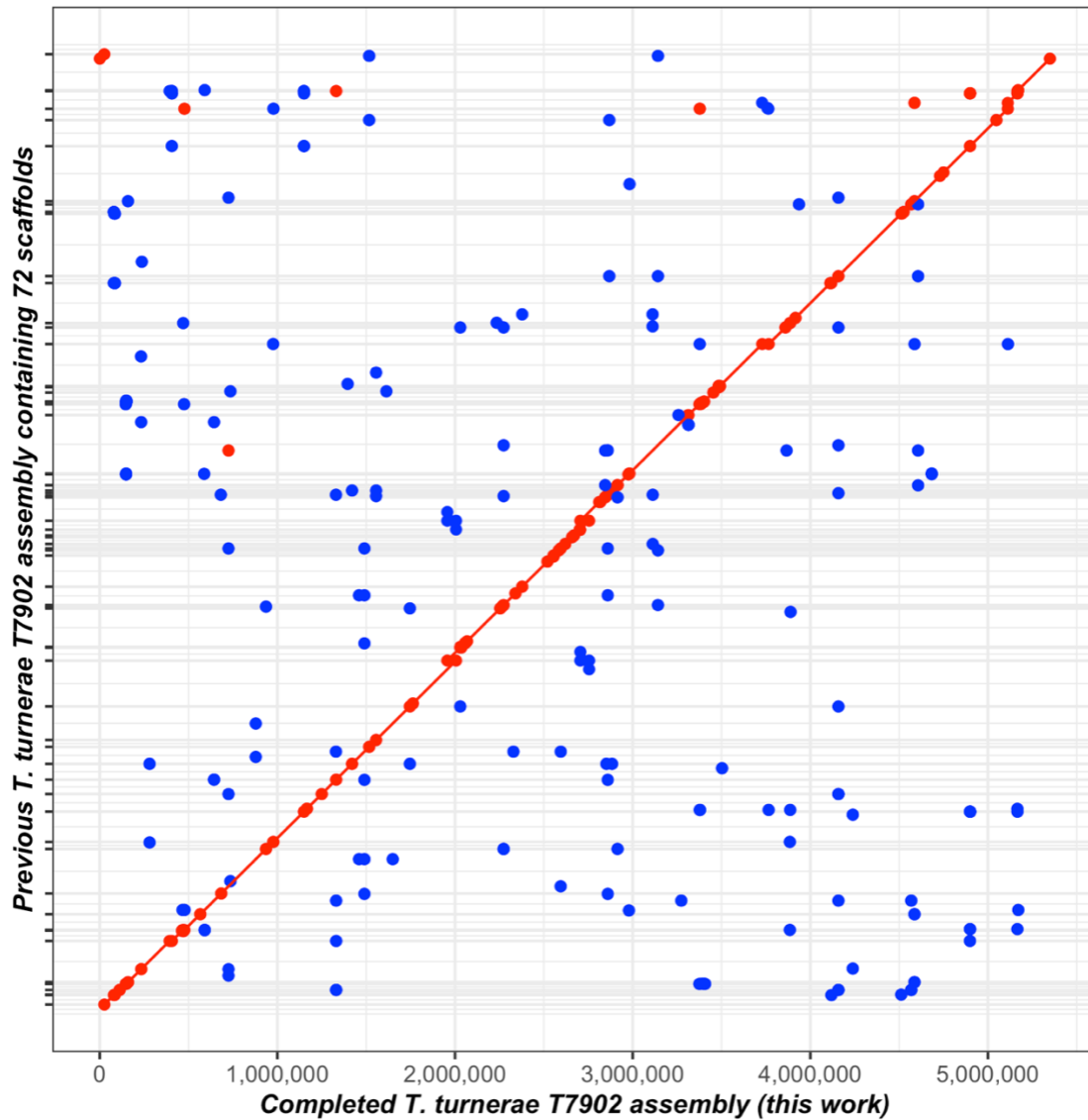
45 present the re-sequencing and completed genome of strain T7902 from nanopore-only
46 sequencing (Table 1).

47 *Table 1 Assemblies of Teredinibacter turnerae T7902*

GenBank Assembly	Scaffolds (contigs)	Size (bp)	GC%	CDS
GCA_000379165.1	72 (76)	5,387,817	50.8	4268
GCA_037935975.1 (this work)	1 (1)	5,348,823	50.9	4212

48
49 *Teredinibacter turnerae* strain T7902 colonies were maintained at 30°C on shipworm basal
50 medium (SBM) (7) plates supplemented with 0.025% NH₄Cl and 0.2% cellulose (Sigmacell Type
51 101; Sigma-Aldrich). A colony was selected and used to inoculate a 6 mL liquid culture of SBM
52 supplemented with 0.025% NH₄Cl and 0.2% carboxymethyl cellulose (CMC) medium and grown
53 at 30°C, 100 rpm for 4 days. Bacterial cells were harvested by centrifugation (10 minutes, 4°C,
54 4,000 × g), and high-molecular-weight DNA was isolated from the cell pellet using the Wizard
55 HMW DNA Extraction Kit (Promega, US) according to the manufacturer's protocol. DNA quality
56 and length were assessed on TapeStation (Agilent Technologies, US). Nanopore (Oxford
57 Nanopore Technologies, UK) sequencing was performed on unsheared gDNA with no size
58 selection and prepared using Nanopore Q20+ chemistry kit v14 and sequenced on a MinION
59 instrument using an R10.4 (FLO-MIN112) flow cell. The standard ligation protocol for the
60 selected kit was followed with the selection of the long fragment buffer (LFB). Bases were
61 called using Guppy v6.5.7 with the super-accurate (SUP) algorithm with default quality read
62 filtering, generating 300,309 reads and an N_{50} read length of 8,763 bp. De novo assembly was
63 performed with Flye v2.9.2 (<https://github.com/fenderglass/Flye>) (10) followed by contig
64 correction and consensus generation with Medaka v1.8.0
65 (<https://github.com/nanoporetech/medaka>). To generate a circular chromosome, overlaps

66 were identified and removed before the assembly was rotated to the gene predicted by
67 prodigal v2.6.3 (11) nearest the middle of the contig with Circlator v1.5.5
68 (<https://github.com/sanger-pathogens/circlator>) (12). A closed, circular chromosomal assembly
69 with a genome coverage of 278.0x was produced and annotated using the NCBI Prokaryotic
70 Genome Annotation Pipeline (PGAP) (13). The new assembly is 99.99% (ANI) (14) identical in
71 primary sequence and highly syntenic (15) with the original (Fig. 1). For example, the new
72 assembly and annotation identify 9 ribosomal RNAs, similar to *T. turnerae* strain T7901
73 (Genbank: GCA_000023025.1, ASM2302v1). However, the new assembly reduces the genome
74 size by 38,994 bp to 5,348,823 bp, contains 56 fewer predicted CDS, and resolves several
75 assembly errors. For all software, default parameters were used except where otherwise noted.



76

77 *Figure 1. Synteny plot comparing the previously published genome of T. turnerae T7902 (GCA_000379165.1) and the new*
78 *genome sequence and assembly presented here (GCA_037935975.1). A MUMmer3 plot was generated with NUCmer v3.1 (15)*
79 *using default settings to assess synteny and completion. Minimum exact matches of 20bp are represented as a dot with lines*
80 *representing exact match lengths >20bp. Forward matches are displayed in red, while reverse matches are shown in blue.*

81

82 [Data availability](#)

83 The complete genome sequence of T7902 has been deposited in GenBank under the accession

84 number [CP149817](#). The Oxford Nanopore sequencing reads are available from the NCBI

85 Sequence Read Archive (SRA) under the accession number [SRR28421272](#).

86 Acknowledgments

87 Research reported in this publication was supported by the following awards to DLD: National
88 Oceanic and Atmospheric Administration (NA19OAR0110303), Gordon and Betty Moore
89 Foundation (GBMF 9339), National Institutes of Health (1R01AI162943-01A1, subaward:
90 10062083-NE), and Johns Hopkins University Applied Physics Laboratory internal research and
91 development funds. The National Science Foundation (DBI 1722553) also funded some
92 equipment used in this research. The funders had no role in study design, data collection and
93 interpretation, or the decision to submit the work for publication.

94 References

- 95 1. Distel DL, Morrill W, MacLaren-Toussaint N, Franks D, Waterbury J. 2002. *Teredinibacter*
96 *turnerae* gen. nov., sp. nov., a dinitrogen-fixing, cellulolytic, endosymbiotic gamma-
97 proteobacterium isolated from the gills of wood-boring molluscs (Bivalvia: Teredinidae).
98 *International Journal of Systematic and Evolutionary Microbiology* 52:2261–2269.
- 99 2. Altamia MA, Shipway JR, Stein D, Betcher MA, Fung JM, Jospin G, Eisen J, Haygood MG,
100 Distel DL. 2020. *Teredinibacter waterburyi* sp. nov., a marine, cellulolytic endosymbiotic
101 bacterium isolated from the gills of the wood-boring mollusc *Bankia setacea* (Bivalvia:
102 Teredinidae) and emended description of the genus *Teredinibacter*. *International Journal of*
103 *Systematic and Evolutionary Microbiology* 70:2388–2394.
- 104 3. Altamia MA, Shipway JR, Stein D, Betcher MA, Fung JM, Jospin G, Eisen J, Haygood MG,
105 Distel DL. 2021. *Teredinibacter haidensis* sp. nov., *Teredinibacter purpureus* sp. nov. and
106 *Teredinibacter franksiae* sp. nov., marine, cellulolytic endosymbiotic bacteria isolated from
107 the gills of the wood-boring mollusc *Bankia setacea* (Bivalvia: Teredinidae) and emended

- 108 description of the genus *Teredinibacter*. International Journal of Systematic and
109 Evolutionary Microbiology 71.
- 110 4. Yang JC, Madupu R, Durkin AS, Ekborg NA, Pedomallu CS, Hostetler JB, Radune D, Toms BS,
111 Henrissat B, Coutinho PM, Schwarz S, Field L, Trindade-Silva AE, Soares CAG, Elshahawi S,
112 Hanora A, Schmidt EW, Haygood MG, Posfai J, Benner J, Madinger C, Nove J, Anton B,
113 Chaudhary K, Foster J, Holman A, Kumar S, Lessard PA, Luyten YA, Slatko B, Wood N, Wu B,
114 Teplitski M, Mougous JD, Ward N, Eisen JA, Badger JH, Distel DL. 2009. The Complete
115 Genome of *Teredinibacter turnerae* T7901: An Intracellular Endosymbiont of Marine Wood-
116 Boring Bivalves (Shipworms). PLoS ONE 4:e6085.
- 117 5. Altamia MA, Lin Z, Trindade-Silva AE, Uy ID, Shipway JR, Wilke DV, Concepcion GP, Distel DL,
118 Schmidt EW, Haygood MG. 2020. Secondary Metabolism in the Gill Microbiota of
119 Shipworms (Teredinidae) as Revealed by Comparison of Metagenomes and Nearly
120 Complete Symbiont Genomes. mSystems 5:e00261-20.
- 121 6. Altamia MA, Wood N, Fung JM, Dedrick S, Linton EW, Concepcion GP, Haygood MG, Distel
122 DL. 2014. Genetic differentiation among isolates of *Teredinibacter turnerae*, a widely
123 occurring intracellular endosymbiont of shipworms. Molecular Ecology 23:1418–1432.
- 124 7. Waterbury JB, Calloway CB, Turner RD. 1983. A Cellulolytic Nitrogen-Fixing Bacterium
125 Cultured from the Gland of Deshayes in Shipworms (Bivalvia: Teredinidae). Science
126 221:1401–1403.

- 127 8. Zerbino DR, Birney E. 2008. Velvet: Algorithms for de novo short read assembly using de
128 Bruijn graphs. *Genome Res* 18:821–829.
- 129 9. Butler J, MacCallum I, Kleber M, Shlyakhter IA, Belmonte MK, Lander ES, Nusbaum C, Jaffe
130 DB. 2008. ALLPATHS: De novo assembly of whole-genome shotgun microreads. *Genome*
131 *Res* 18:810–820.
- 132 10. Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using
133 repeat graphs. *Nat Biotechnol* 37:540–546.
- 134 11. Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic
135 gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119.
- 136 12. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated
137 circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294.
- 138 13. Li W, O’Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F,
139 Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J,
140 Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq:
141 expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model
142 curation. *Nucleic Acids Research* 49:D1020–D1028.
- 143 14. Rodriguez-R LM, Konstantinidis KT. 2016. The enveomics collection: a toolbox for
144 specialized analyses of microbial genomes and metagenomes. preprint. *PeerJ Preprints*.

145 15. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004.

146 Versatile and open software for comparing large genomes. *Genome Biol* 5:R12.

147