

## Review Article

# Regulatory Effect of Mesenchymal Stem Cells on T Cell Phenotypes in Autoimmune Diseases

Zhiping Wei <sup>1</sup>, Jintao Yuan <sup>2</sup>, Gaoying Wang <sup>1</sup>, Dickson Kofi Wiredu Ocansey <sup>1,3</sup>,  
Zhiwei Xu <sup>1</sup> and Fei Mao <sup>1</sup>

<sup>1</sup>Key Laboratory of Medical Science and Laboratory Medicine of Jiangsu Province, School of Medicine, Jiangsu University, Zhenjiang, 212013 Jiangsu, China

<sup>2</sup>The People's Hospital of Danyang, Affiliated Danyang Hospital of Nantong University, Zhenjiang, Jiangsu 212300, China

<sup>3</sup>Directorate of University Health Services, University of Cape Coast, Cape Coast, Ghana

Correspondence should be addressed to Fei Mao; [maofei2003@ujs.edu.cn](mailto:maofei2003@ujs.edu.cn)

Received 25 January 2021; Revised 3 March 2021; Accepted 11 March 2021; Published 30 March 2021

Academic Editor: Jian Fang

Copyright © 2021 Zhiping Wei et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Research on mesenchymal stem cells (MSCs) starts from the earliest assumption that cells derived from the bone marrow have the ability to repair tissues. Several scientists have since documented the crucial role of bone marrow-derived MSCs (BM-MSCs) in processes such as embryonic bone and cartilage formation, adult fracture and tissue repair, and immunomodulatory activities in therapeutic applications. In addition to BM-MSCs, several sources of MSCs have been reported to possess tissue repair and immunoregulatory abilities, making them potential treatment options for many diseases. Therefore, the therapeutic potential of MSCs in various diseases including autoimmune conditions has been explored. In addition to an imbalance of T cell subsets in most patients with autoimmune diseases, they also exhibit complex disease manifestations, overlapping symptoms among diseases, and difficult treatment. MSCs can regulate T cell subsets to restore their immune homeostasis toward disease resolution in autoimmune conditions. This review summarizes the role of MSCs in relieving autoimmune diseases via the regulation of T cell phenotypes.

## 1. Introduction

There are abundant sources of MSCs including the umbilical cord, placenta, bone marrow, adipose tissue, gums, endometrium, menstrual blood, synovium, periosteum, skeletal muscle, and ligamentum cruciatum, among other tissues [1–5]. Besides, human induced pluripotent stem cells (iPS) also serve as a source of MSCs. Although there are variations in the criteria of surface markers of MSC from different sources, the literature shows that classic MSCs express CD105, CD73, CD90, CD34, and CD44, but not CD45, CD34, CD14, CD11b, CD79a, CD19, and HLA-DR [6, 7]. MSCs have the potential of self-renewal and multidirectional differentiation, and their differentiation potential depends on the tissues from which they originate (Table 1). Because of the low immunogenicity and homing capabilities of MSCs, they are used to treat multiple disease conditions. For instance, human umbilical cord-derived mesenchymal stem cells (UC-MSCs) have the

potential to transform into cardiomyocyte-like cells. According to Peter et al., the cardiomyocyte-like contractile cells are produced in vitro by MSC differentiation and aggregation on the cardiomyocyte feeder layer and that only the young MSCs could maintain their low immunogenicity after differentiation into cardiomyocyte-like cells [8]. Similarly, Yu and colleagues found UC-MSCs have higher liver differentiation potential than bone marrow MSCs (BM-MSCs), hence their superiority to BM-MSCs in the treatment of end-stage liver disease [9]. This indicates the differential ability of MSCs from varying sources towards specific tissue repair and regeneration.

Also, the proliferation and differentiation potential of MSCs can be enhanced by the culture environment via modification techniques; therefore, the establishment of various modified culture systems makes the application of MSCs in regenerative medicine even more promising. These modification approaches can be roughly divided into genetic modification and preconditioning modification (using drugs,

TABLE 1: The surface markers and differentiation potential of different kinds of MSCs.

Sources	Surface marker	Differentiation potential	Reference
BM-MSCs	CD271(+), CD59(+), CD81(+), CD47(+), CD151(+), CD147(+), CD98(+), CD143(-), Lin (-) CD45(-), CD140a (PDGFR $\alpha$ ) (low/-)	Strong adipogenic and osteogenic potential, poor chondrogenic potential, strong differentiation potential of corneal epithelial cells, and cardiac progenitors	[14–18]
UC-MSCs	CD73(+), CD90(+), CD105(+), CD44(+), CDH-1(+), CD29(+), CD34(-), CD45(-)	Muscle, neurogenic cells, hepatocyte-like cells, endothelial lineage	[19–23]
AD-MSCs	CD45(-), HLA-DR(-), CD44(+), CD106(+), CD34(+), CD90(+), CD105(+)	Strong adipogenic and osteogenic potential, poor chondrogenic potential, poor differentiation potential of corneal and muscle	[14, 16, 17, 24–26]
DP-MSCs	TRO-1(+), CD146(+), CD29(+), CD90(+), CD105(+), CD44(+), CD59(+), CD73(+), CD146(-), CD34(-), CD45(-), CD11b(-), CD45(-)	Osteogenic, adipogenic, chondrogenic, fibroblast lineage, neural stem cells	[27, 28]
SD-MSCs	CD9(+), CD10(+), CD13(+), CD44(+), CD54(+), CD55(+), CD90(+), CD105(+), CD166(+), D7-FIB(+), CD14(-), CD20(-), CD45(-), CD133(-)	Strong chondrocyte, osteocyte, and adipocyte differentiation ability, as well as muscle differentiation	[16, 29]

growth factors, and other molecules), which can improve the inherent biological activities concerning migration, homing to target site, adhesion, and survival and reduce premature senescence [10]. Existing research has shown that MSCs communicate with other cells through direct contact and paracrine signaling. In effect, MSCs repair tissue by directly contacting, adhering, and subsequently differentiating into the injured cells. It also exerts its anti-inflammatory, repairing, and immunomodulatory effects by secreting extracellular vesicles (EVs) or paracrine factors and mitochondrial transfer [11].

Autoimmune diseases are caused by imbalanced homeostasis of the autologous environment including T cells. While peripheral regulatory T cell (pTreg) and T helper type 17 (Th17) cell share a common precursor cell (the naïve CD4 T cell) and require a common signal for initial differentiation (tumor growth factor- (TGF-)  $\beta$ ), they turn to elicit opposite functions via terminal differentiation: Treg is anti-inflammatory, inhibits autoimmunity, and maintains immune homeostasis, whereas Th17 cell causes autoimmunity and inflammation [12]. Moreover, the instability within T cell phenotypes such as Treg alongside their cellular plasticity and tissue-specificity also affects the development of autoimmune diseases [13]. Detailed exploration of T cell interaction with both immune and nonimmune cells presents not only deeper insight into disease pathogenesis but new therapeutic strategies as well.

MSC-based therapy is widely used in refractory immune diseases and has achieved encouraging results. They effectively restore T cell balance within the autoimmune environment, enhancing inflammation resolution through the complemented effect of both cells. This paper examines the application of MSCs in autoimmune diseases such as inflammatory bowel disease (IBD), rheumatoid arthritis (RA), and systemic lupus erythematosus (SLE), among others, and particularly highlights their modulatory effects on T cell phenotypes and the resultant contribution towards therapeutic strategies.

BM-MSCs: bone marrow-derived MSCs; UC-MSCs: umbilical cord-derived MSCs; AD-MSCs: adipose-derived

MSCs; DP-MSCs: human dental pulp-derived MSCs; SD-MSCs: synovium-derived MSCs.

## 2. Application of MSCs in Autoimmune Diseases

Autoimmune disease is a diverse kind of complex and heterogeneous abnormal condition caused by immune system disorder. Common examples include IBD, RA, SLE, type 1 diabetes (T1D), and multiple sclerosis (MS), among others. In addition to the difficulties in early detection and poor curative effect, patients with autoimmune diseases are also faced with complicated pathogenesis [30]. At present, due to the low immunogenicity and multidirectional differentiation of MSCs, it is promising to study its therapeutic effect on patients with autoimmune diseases.

*2.1. Application in IBD.* IBD refers to a group of chronic and heterogeneous intestinal inflammatory disorders, including ulcerative colitis (UC) and Crohn's disease (CD). The pathogenesis of IBD is highly complex and not completely clear, but current literature shows that it is related to several factors such as genetic susceptibility, environmental triggers, intestinal flora, diet, psychology, and immunity [31, 32]. The incidence and prevalence of IBD vary from region to region. European countries such as Norway and Germany have the highest global prevalence of 505/100,000 in UC and 322/100,000 in CD, respectively, followed by the North American countries, the United States of America with a UC prevalence of 286/100,000, and Canada with CD prevalence of 319/100,000. Due to the recent modernization of Asian and Latin American countries, the prevalence of IBD in such countries is also constantly rising [33–36].

At present, there are several clinical interventions for IBD patients, including drug therapy such as immunomodulators, steroids, and antibiotics, surgical treatment, and fecal microbiota transplantation (FMT) as a novel therapy. However, these treatment approaches are insufficient in curing IBD. Drug therapies are less effective and often associated with adverse reactions. Surgical procedures have certain

requirements for patients' state, and FMT is relatively safe but linked with many problems such as patient acceptance [37–40]. In the phase of these challenges, autologous or allogeneic MSCs emerged as a potentially effective therapy for IBD because of their anti-inflammatory and tissue repair effects, excellent immunomodulatory properties, and low immunogenicity [41]. Experimental studies illustrate that MSCs can repair dextran sulfate sodium- (DSS-) induced acute and chronic colitis in mouse models and prevent the recurrence of experimental IBD. In this process, MSCs regulate immune response, reduce inflammatory cell infiltration, regenerate intestinal epithelial cells, blood vessels, and lymphatic vessels, and change gut microbiota [42–44]. A myriad of clinical trials also shows that MSC therapy is well tolerated with promising efficacy and safety profile. Most recorded adverse effects described for MSCs are mild and transient [45].

A randomized, double-blind, parallel-group, placebo-controlled trial divided 212 patients having a CD with refractory perianal fistula into two groups, which were treated with either autologous AD-MSCs or placebo. The comprehensive remission rates of patients treated with AD-MSCs were higher than those of the placebo group, 50% and 34%, respectively. The incidence of treatment-related adverse events was 17% in AD-MSCs and 29% in placebo groups [46]. Clinical trials by Park et al. also demonstrated the effectiveness of MSCs for perianal fistula repair. A large number of systematic reviews and meta-analyses have also reported the effectiveness and safety of MSCs as IBD-related treatment, especially with long-term effects [47–49]. A study designed to investigate the occurrence of adverse events related to acute infusion toxicity, long-term adverse events, and efficacy of human amnion-derived MSCs (AMSCs) was carried out among CD patients who only achieve partial symptomatic relief with traditional therapy. The results of this phase I/II trial study will be beneficial to further promote the clinical application of AMSCs in IBD [50]. Different doses of BM-MSCs were injected locally in CD patients with refractory perianal fistula to determine the effective dosage that promotes healing of perianal fistulas. The authors concluded that no severe adverse events were associated with the allogeneic MSCs administered and the injection of  $3 \times 10^7$  MSCs appeared to promote healing of perianal fistulas [51].

However, it is important to note that regardless of the success witnessed in MSC therapy in IBD, it is still confronted with several challenges such as severe adverse events, encouraging tumor growth and metastasis, among other reactions as detailed in recent reviews by Ocansey et al. [52, 53]. There is the need to identify supportive or combined therapies of MSC transplantation and also choose the most appropriate stem cell and treatment approach to enhance effectiveness while avoiding the occurrence of serious adverse events [54].

**2.2. Application in RA.** RA is a systemic autoimmune disease, which mainly affects and damages joints and bones. The main clinical manifestations are pain, swelling, deformation of joints, and dyskinesia. In RA, there are unresolved immune cells infiltrating joints and unregulated autoanti-

body levels. Additionally, RA also affects other organs, including the blood vessels, kidneys, heart, and lungs, resulting in severe pain in patients. The complexity of the definition of RA makes it difficult to study its incidence. According to the criteria defined by the American Rheumatology Society 1987, the global prevalence of RA is about 0.24% (95% CI 0.23% to 0.25%), and the number of patients with RA will reach 4.8 million by 2010 (95% CI 3.7 million to 6.1 million). However, after the revision of the diagnostic criteria of RA in 2010, the number of patients with RA increased further [55, 56]. The main risk factors of RA are gender (women usually have a higher risk than men), heredity, environment, and psychological factors [57]. Recent studies have also shown that RA has a strong correlation with periodontal diseases [58, 59].

At present, there is no effective therapy for RA. The use of nonsteroidal anti-inflammatory drugs and cortisol can alleviate the symptoms of pain and stiffness, but cannot delay the progress of the disease. The use of disease-modifying antirheumatic drugs such as methotrexate and sulfasalazine can delay the disease progression. For example, the depletion of B cells and the development of B cell inhibitory antibodies, IL-6 inhibitors, and T cell-targeted drugs can bring a glimmer of hope for the treatment of RA, but the adverse reactions and toxic side effects of these drugs still hinder their application [60]. Presently, the application of MSCs as a treatment option for RA has demonstrated unique advantages in a host of clinical trials.

According to Wang et al., patients with insufficient response to traditional RA drugs were divided into two groups: patients injected with traditional antirheumatic drugs plus culture medium not containing UC-MSCs and patients injected with traditional antirheumatic drugs and culture medium containing UC-MSCs. They found that the group that received UC-MSCs was significantly relieved, and only one infusion capably achieved the relief effect for 3–6 months, with no major adverse reactions [61]. Another group of researchers selected refractory RA patients in phase Ib/IIa clinical trials and reported similar results without serious side effects in the short term after intravenous injection of AD-MSCs [62]. These demonstrated safety and efficacy studies indicate the encouraging development of MSC therapy in RA, regardless of challenges that demand further exploration in areas such as maintaining the therapeutic effect for a long period.

**2.3. Application in SLE.** SLE, a common autoimmune disease, involves an inflammatory disorder of multiple organs and systems of the body such as the kidney, lung, and skin. It mostly affects females, and some patients develop symmetrical butterfly erythema or other rashes on their faces. The main cause of mortality and morbidity of SLE patients at the end stage is lupus nephropathy [63]. SLE also has the characteristics of complex pathogenesis which largely remains unclear. Notwithstanding, it is agreed that the pathogenesis of SLE is related to the imbalance of factors such as heredity, environment, and endocrine and autoimmune system [64]. Although many genes related to SLE have been found, including complement component-related genes

(C1q, C1r, C1s, and C4) and HLA-DR, the specific effects of each gene are still unknown [65, 66]. Besides, estrogen and prolactin have also been identified as risk factors of SLE, which is consistent with the higher prevalence in women than men. Ultraviolet radiation can also increase the risk of SLE. Most SLE patients produce autoantibodies that are related to the clearance defect of apoptotic cells [67]. The global incidence and prevalence of SLE vary with gender, age, race, and time, which are partly explained by the differences in genetic and environmental risk factors [68].

Current treatment of SLE includes immunomodulators and immunosuppressants such as hydroxychloroquine and other drugs that prevent complications. While symptomatic treatment is given to subside various systemic manifestations [69], monoclonal antibody therapy is administered to target and deplete B cells as a proposed treatment of lupus nephritis [70]. Both the traditional and newly developed therapies have limitations related to drug administration, as well as gradual drug reduction until withdrawal, both of which may affect the balance between disease activity control and organ damage caused by long-term and/or unbalanced immunosuppression [71, 72].

MSCs have been used in the treatment of lupus nephritis and refractory SLE patients for more than ten years. Most documented clinical trials are self-controlled studies with only a few being randomized controlled trials. In a meta-analysis study that evaluated the efficacy and safety of MSC treatment in SLE patients, the researchers report that the MSC group showed significantly decreased SLE disease activity index, as well as decreased urine protein, and increased complement C3 [73]. A meta-analysis of the animal model of lupus nephritis also confirmed that MSC treatment resulted in lower levels of disease-associated elements such as double-stranded DNA (ds-DNA), antinuclear antibody (ANA), serum creatinine (Scr), blood urea nitrogen (BUN), proteinuria, and renal sclerosis score, as well as higher albumin levels [53]. Several other studies have reported the promising effect of MSCs in SLE experimental studies and clinical trials [74]. These current pieces of evidence show that MSCs capably improve the disease activity, hypocomplementemia, and proteinuria in SLE patients. However, large-scale and high-quality randomized controlled trials are required to validate the efficacy and safety of MSC treatment in SLE patients. It is also worth noting that allogeneic and autologous MSC treatment of SLE may have opposite effects; hence, allogeneic rather than autologous MSC transplantation could be potentially advantageous for SLE patients [63, 75, 76].

**2.4. Application in Other Autoimmune Diseases.** Recent studies demonstrate the remarkable therapeutic effectiveness of MSCs towards several other autoimmune diseases such as type 1 diabetes [77], multiple sclerosis [78], Hashimoto's autoimmune thyroiditis [79], autoimmune hepatitis, primary biliary cirrhosis [80], and vitiligo [81], among others. For example, MSCs have been shown to prevent inflammation and neurodegeneration in animal models of multiple sclerosis (MS). These experimental studies have set the ground for clinical trials such as a recent randomized, double-blind, placebo-compared phase I/II clinical trial with autologous

BM-MSCs in MS which is currently ongoing (ClinicalTrials.gov NCT01854957) [82]. Autoimmune destruction of insulin-producing B cells in the pancreas results in type 1 diabetes, a disease condition that demands more than a mere administration of exogenous insulin to gently and sensitively regulate blood glucose concentration. MSCs can transdifferentiate into insulin-producing cells, support the regeneration of residual B cells via production of growth and trophic factors, or participate in the suppression of the autoimmune reaction against B cells [83, 84]. Hashimoto's thyroiditis (HT) is a disease wherein lymphocytes mediate the autoimmune damage and destruction of the thyroid gland. MSCs have been demonstrated to improve HT via reducing the level of thyroid autoantibody partly by regulating Th17/Treg interactions [79].

A detailed exposition of research progress on MSC therapy in autoimmune diseases indicating remarkable therapeutic effectiveness has recently been reviewed by Chen and colleagues [85]. MSCs also capably home to the disease site, regulating the balance of T cells through direct contact and secretion of active factors. Table 2 presents some of the documented studies of MSC's role in immune regulation of selected autoimmune diseases.

T1D: type 1 diabetes mellitus; TGF- $\beta$ /MSCs: TGF- $\beta$  engineered MSCs; MS: multiple sclerosis; SS: Sjögren's syndrome; PBC: primary biliary cirrhosis; HT: Hashimoto's thyroiditis.

### 3. Regulatory Effect of MSCs on T Cells in IBD

In the experimental IBD model, MSCs regulate the generation of T cell subsets to alleviate intestinal inflammation [102, 103]. For instance, the coculture of peripheral blood mononuclear cells (PBMCs) and MSCs strongly inhibits the proliferation of CD4+ and CD8+ T cells, as well as natural killer (NK) cells. The researchers found that the mechanism involved is not dependent on cell contact, but rather activated by interferon-gamma (IFN- $\gamma$ ) produced by lymphocytes. IFN- $\gamma$  stimulates MSCs to produce indoleamine 2,3-dioxygenase (IDO), prostaglandin E2 (PGE2), and IL-10, wherein the IDO principally inhibits the proliferation of T lymphocytes [104, 105]. MSCs homing in colon tissue can promote the proliferation of intestinal epithelial cells and the regeneration of intestinal stem cells. This effect has been shown to be related to the downregulation of Th1/Th17. Other molecules reduced in the process owing to the anti-inflammatory effect of IFN- $\gamma$  include interleukin- (IL-) 2, tumor necrosis factor- (TNF-)  $\alpha$ , IFN- $\gamma$ , T-bet, IL-6, IL-17, and retinoic-acid-receptor-related orphan nuclear receptor gamma (ROR $\gamma$ t) [106]. However, in most cases, IFN- $\gamma$  is still considered a proinflammatory factor. It has been reported that MSCs significantly inhibit the secretion of IFN- $\gamma$  and promote the production of IL-10 by T cells. IL-10 acts with dendritic cells (DCs) to promote anti-inflammatory effect [107, 108]. These findings show the complex role of IFN- $\gamma$  in MSC-mediated immune regulation including its role in inducing T cell inhibition via MSC regulation.

Regulatory T cells (Tregs), the special T cell subset for immunosuppression, specifically express transcription factor

TABLE 2: The application of MSCs in other autoimmune diseases.

Disease	Source of MSCs	Effects	Reference
T1D	UC-MSCs TGF- $\beta$ /MSCs	MSCs were safe and tolerable Hyperglycemia was significantly controlled	[77, 86–89]
MS	BM-MSCs	Clinically feasible and relatively safe and could immediately produce immune regulation	[90–94]
SS	UC-MSCs	Effective in treatment	[95, 96]
PBC	BM-MSCs UC-MSCs	MSCs were well tolerated and no obvious side effects were found Symptoms were significantly alleviated	[97–99]
HT	AD-MSCs	MSCs inhibited inflammation and helped recover from injury	[79, 100, 101]

forkhead box P3 (FoxP3) in the nucleus and CD25 and CTLA-4 (cytotoxic T lymphocyte-associated protein 4) on the cell surface [109]. Tregs play a crucial role in the inhibition of inflammation associated with several diseases such as IBD [110]. The combination of MSCs and Tregs in experimental treatment results in longer survival time for exogenous Tregs, upregulation of endogenous Tregs, and downregulation of proinflammatory Th17 cells [111, 112]. It is worth noting that endogenous CD4+CD25+Foxp3+ T cells are differentiated from CD4+ T cells, rather than natural Treg amplification, which involves TGF- $\beta$  and/or programmed cell death- (PD-) 1/PD-L1 mechanism [45, 105].

Studies by Sarah and others indicate that cytokines secreted by MSCs intensely participate in the immunomodulatory role on T cells. TGF- $\beta$ 1, a soluble cytokine produced by MSCs, can induce Tregs under TCR (T cell receptor) costimulation, promote the activation of monocytes, and enhance monocyte differentiation into type II macrophages. Macrophages produce a large amount of IL-10 and CCL-18 (C-C motif chemokine ligand-18), which has been shown to play an important role in Treg induction; IL-10 further inhibits the pathogenicity of Th17 [113]. Macrophages can significantly inhibit the proliferation of CD4+ T cells and reduce the content of inflammatory factors TNF- $\alpha$  and IFN- $\gamma$  [114]. TGF- $\beta$  can also induce Foxp3 expression, inhibit Th17 differentiation, and stimulate Treg development [103, 115], via its biological activities through transcription regulation of several genes. Activated TGF- $\beta$  binds to TGF- $\beta$ 1 and TGF- $\beta$ 2 receptors, followed by induction of the formation of phosphor-mothers against decapentaplegic homolog 2 (pSmad2), pSmad3, and Smad4 complexes, thus activating the intracellular signal activation of TGF- $\beta$  signaling [105, 116]. Studies have shown that the lack of TGF- $\beta$ 1 leads to severe colonic inflammation, while the restoration of TGF- $\beta$ 1 activity improves the resolution of colitis [117, 118]. MSCs can block the induction of inflammatory-associated TNF and interleukins while promoting T cells to secrete anti-inflammatory cytokine like IL-10. The secretion of polyethylene glycol (PEG) by MSCs in the process of inflammation resolution has also increased significantly, as PEG is an important regulator to maintain immune homeostasis [119].

The immunomodulatory effect of MSCs is not only due to the role of soluble factors but also the effect of intercellular contact. MSCs constitutively express FasL and PD-L1. FasL induces apoptosis of activated T cells, while PD-L1 on the

surfaces of MSCs combine with PD-1 on the surfaces of T cells, exerting immunosuppression through major histocompatibility complex II (MHC II). MSCs can also secrete IFN- $\beta$  to increase the expression of PD-1 on the surface of T cells and strengthen the inhibition of T cells [105, 107, 120]. Mice with PD-1 gene knocked out produce autoimmune diseases, which also prove that MSCs can inhibit the activation, expansion, and cytokine production of T cells through the PD-1/PD-L1 pathway [121]. Experiments prove that tonsil-derived MSCs (T-MSCs) weaken the differentiation of Th17 and directly regulate the phosphorylation of signal transducer and activator of transcription 3 (STAT3) through the PD-L1 expression [122]. The imbalance of the IL6/IL6R-STAT3-SOCS3 (suppressor of cytokine signaling 3) pathway is closely related to IBD-related diseases [123, 124]. MSCs express NOD2 (nucleotide-binding oligomerization domain-containing protein 2), and its binding with ligand MDP (muramyl dipeptide) enhances the production of PEG2 and increases the production of IL-10 and Tregs through NOD2-RIP2 (receptor-interacting protein 2) pathway. In several experimental colitis models in mice, MSCs have been demonstrated to highly express Jagged-1, induce Notch signaling of T lymphocytes, reduce the activity of NF- $\kappa$ B, reduce the production of IL-2 and IFN- $\gamma$ , and hinder the proliferation of T lymphocytes [125–128].

Intercellular adhesion molecule-1 (ICAM-1), also known as CD54, is involved in signal transmission between cells, regulates immune response, and mediates cell differentiation, development related to lymphocyte homing and circulation. Generally, ICAM-1 is not expressed on the surface of MSCs, but ICAM-1 is upregulated in the inflammatory microenvironment. MSCs overexpressing ICAM-1 significantly reduced the percentage of Th1 and Th17 cells in the spleen, increasing the number of Tregs of IBD mice. Further analysis revealed remarkably reduced mRNA levels of INF- $\gamma$  and IL-17A and promoted expression of Foxp3, thus alleviating the experimental colitis [129].

Nitric oxide (NO) produced by MSCs can also inhibit the expression of CD25 in T cells by regulating LKB1- (liver kinase B1-) AMPK- (adenosine 5' monophosphate-activated protein kinase-) mTOR pathway. It is reported that the deletion of LKB1 decreases AMPK phosphorylation level and activates mTORC1, which leads to T cell activation and inflammation, while MSCs can increase LKB1 and AMPK phosphorylation level, thus exerting inhibitory effects on inflammatory T cell proliferation and increasing anti-

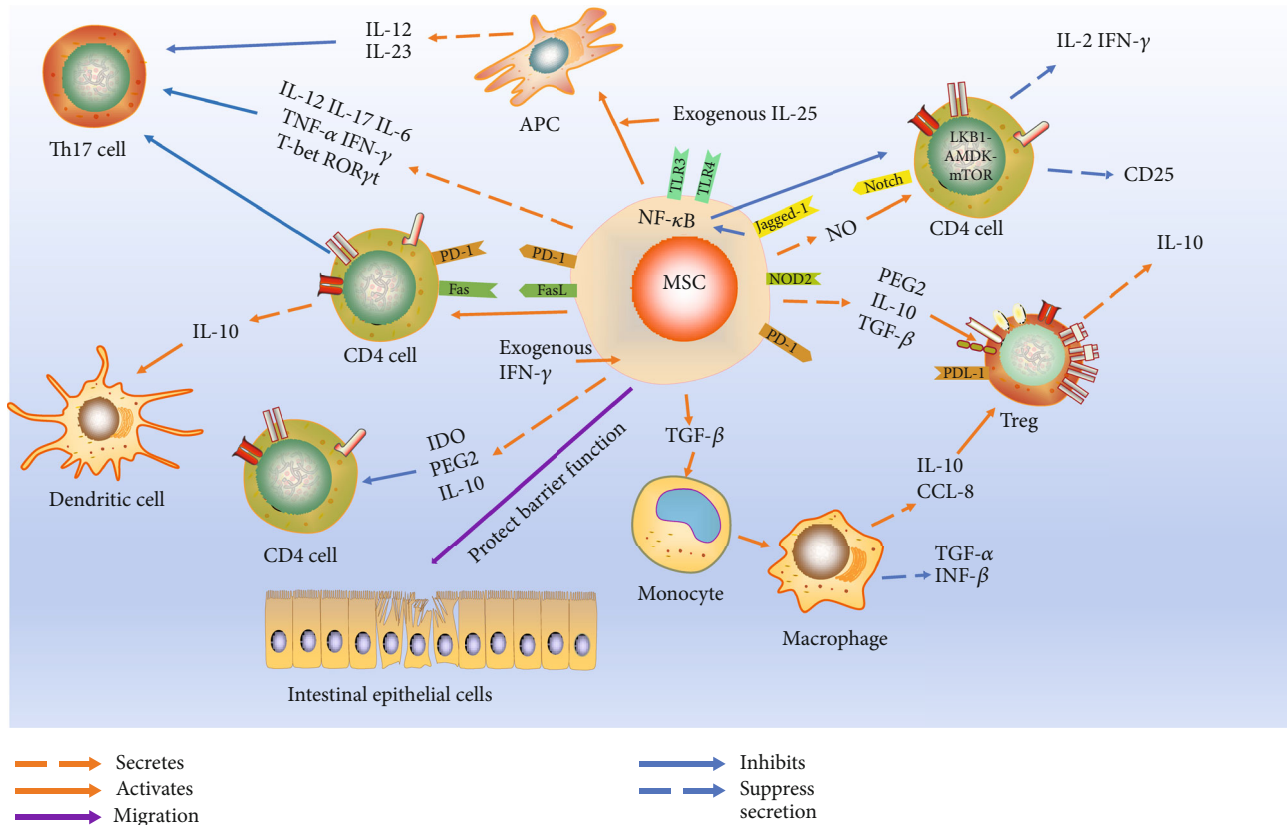


FIGURE 1: MSCs alleviate IBD by regulating T cells. MSCs induce CD4 T cells to differentiate into Treg and maintain Th17/Th1 balance through a series of cytokines and cell-to-cell contact. This results in decreased inflammatory activities to repair intestinal inflammation.

inflammation [130]. IL-25 can inhibit TNF- $\gamma$  and IL-17A produced by CD4<sup>+</sup> T cells of IBD patients, promote the secretion of anti-inflammatory IL-10, and inhibit the differentiation of CD4<sup>+</sup> T cells of IBD into proinflammatory Th1 and Th17 cells [131, 132].

It is worth mentioning that peripheral circulating T lymphocytes play an important role in the MSC treatment mechanism of IBD. There are groups of intestinal intraepithelial lymphocytes (IELs) that are similar to peripheral lymphocytes in the intestinal tract and have complex cell subsets, including TCR-positive and TCR-negative cells. Every ten intestinal epithelial cells (IECs) in the small intestine contain about one IEL, which is lower in the colon. IELs reside in intestinal epithelial cells and do not participate in the circulation. They are associated with the maintenance of the intestinal mucosal immune barrier [133, 134]. The increase of IELs can be observed in the intestinal tract of children with IBD [135]. The imbalance of IEL subsets is related to the pathogenesis of IBD, but it is still unknown whether MSCs can regulate IELs.

IDO plays an important role in the regulation of MSCs on experimental enteritis in mice. MSCs can secrete IDO, which is a rate-limiting enzyme that catalyzes tryptophan metabolism. IDO and its downstream metabolites kynurenine (KYN) and kynurenic acid (KYNA) play a powerful role in inhibiting T cell proliferation and Treg differentiation [136, 137]. IDO can alleviate DSS-induced enteritis by regulating tryptophan metabolites KYN and KYNA in MSCs,

activating transcription factor aryl hydrocarbon receptor (AhR), and upregulating the expression of TNF-stimulated gene 6 (TSG-6) [138]. Under the action of the inflammatory microenvironment, MSCs enhance the glycolytic pathway and upregulate the IDO level through the Janus kinase (JAK)/STAT1 pathway, which plays an immunosuppressive role [139, 140]. The activities of MSCs in IBD as discussed above are summarized in Figure 1.

#### 4. Regulatory Effect of MSCs on T Cells in RA

The mechanism by which MSCs regulate T cells to relieve RA overlaps with the mechanism of regulating IBD. Just as reported in other autoimmune diseases, there is an imbalance of T cell subsets in RA patients too, including Th17/Treg cells which are capably regulated by MSCs. The expression of TNF- $\alpha$  inducible protein 3 (TNFAIP3), also known as A20, from BM-MSCs of RA patients has been found to be reduced. TNFAIP3 is a protective protein of chronic arthritis, which can negatively regulate the NF- $\kappa$ B pathway and reduce the expression of IL-6. MSCs overexpressing A20 can inhibit the expression of IL-6, thus restoring Th17/Treg balance. A20 deficiency also increases Th17 and decreases Tregs, while Th1 and Th2 are not affected. Specifically, inflammatory cytokines induce A20 expression in MSCs. Mechanism studies show that knocking out A20 in MSCs can inhibit the activation of the p38 mitogen-activated protein kinase (MAPK) pathway, effectively promote the production of

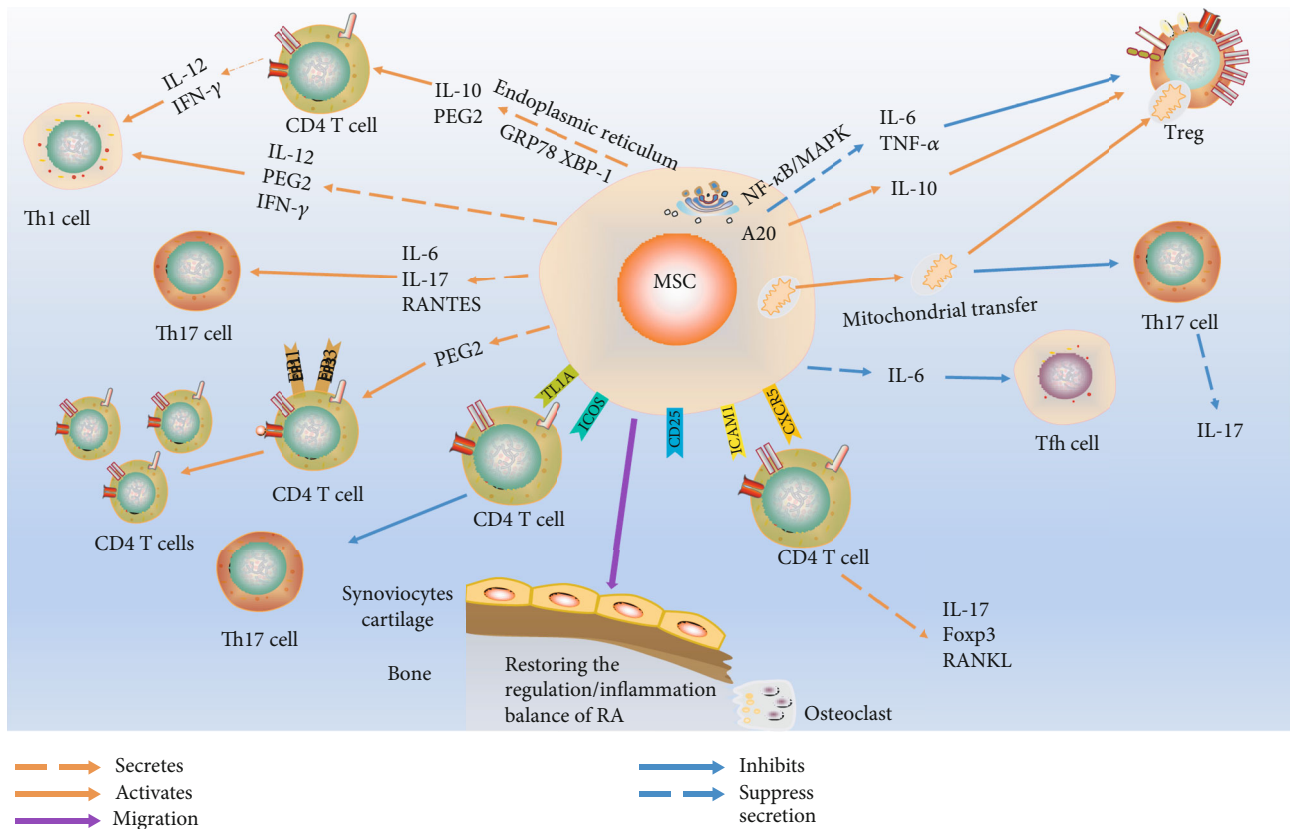


FIGURE 2: MSCs relieve RA by regulating T cells. MSCs can regulate the balance of T cells by homing to the articular cavity and secreting a series of cytokines that increase the anti-inflammatory activity of the environment. T cells are also regulated via the transfer of mitochondria from MSCs to T cells. Additionally, MSCs under endoplasmic reticulum stress can also play a regulatory role in inducing T cells.

TNF- $\alpha$ , and inhibit the production of IL-10. It is worth noting that the therapeutic effect of MSCs on RA may be different due to the different expression of A20 [76, 141]. Mitochondrial transfer from MSCs to Th17 cells, as a mechanism of MSC regulating immunity, can occur through intercellular contact, resulting in increased oxygen consumption of Th17 and reduced production of IL-17. At the same time, the mitochondria of Treg markers on the surface of Th17 cells increase, indicating that mitochondria from MSCs can increase the production of anti-inflammatory phenotype [142]. Inflammatory microenvironment can lead to the activation of the PI3K/Akt/mTORC1 pathway, which is closely related to cell metabolism and can enhance glycolysis and activate lymphocytes. Mitochondrial metastasis can transform energy metabolism into oxidative phosphorylation, and Treg-related markers are upregulated while proinflammatory markers are downregulated [143, 144].

In addition to Th17 cells, T follicular helper (Tfh)/T follicular regulatory (Tfr) cells are also closely related to RA. It is reported that the number of Tfh and Tfr in RA patients is increased, but the ratio of Tfr/Tfh is decreased, with a significantly increased number of circulating B cells related to Tfh [145, 146]. The production of autoantibodies in RA patients, such as antirheumatoid factor (RF) and anticyclic citrullinated peptide (CCP), leads to the deposition of immune complexes, while Tfh cells can migrate to the germinal center (GC) to maintain the differentiation of B cells. Tfh is closely

related to the production of autoantibodies in B cells. Tfh cells express high levels of CXCR5, PD-1, IL-21, and other characteristic markers, and their cellular differentiation is regulated by a complex network of transcription factors, including positive factors (Bcl6, ATF-3, Batf, IRF4, c-Maf, etc.) and negative factors (Blimp-1, STAT5, IRF8, Bach2, etc.) [147, 148]. On the other hand, Tfr is a type of cell in the Treg subgroup, which can inhibit the reaction in GC and the production of high-affinity antibody. As an inflammatory factor, IL-6 plays a role in the pathogenesis of RA, by phosphorylating STAT3 and participating in Tfh differentiation [149]. MSCs can significantly reduce the production of IL-6 in vivo, which may have an alleviating effect by regulating Tfh/Tfr. Whether MSCs can regulate these transcription factors and participate in the regulation of Tfh cells in RA patients is rarely reported at present, which is also the direction of future research.

MSCs can exert their immune function and relieve autoimmune diseases through PEG2, TGF- $\beta$ , HGF, IL-10, and IDO, which are found in RA, IBD, and other autoimmune diseases. Studies have shown that endoplasmic reticulum- (ER-) stressed MSCs can produce higher levels of IL-10 and PEG2 than ordinary MSCs and downregulate CD4+CXCR5+ICOS+ T cells (Tfh) in RA patients. It may be that glucose-regulated protein 78 (GRP78) and X-box binding protein 1 (XBP-1) are strongly induced in ER-stressed MSCs, resulting in a large amount of PEG2 production. PEG2 receptors, membrane-bound G

protein-coupled receptors termed EP1, EP2, EP3, and EP4, are expressed on CD4+ T cell surface. PEG2 can also increase the levels of IL-12 and IFN- $\gamma$ , which triggers Th1 cells to differentiate. The anti-T cell proliferation effect is realized by the EP/COX2/PEG2 axis, wherein COX2 is upregulated under the condition of inflammatory stress [150–152].

In RA patients, MSCs can phagocytize apoptotic cells (ACs) in an actin-dependent way and secrete IL-6, IL-17, and RANTES (regulated upon activation, normal T cell expressed and presumably secreted). In this process, MSCs can express CXCR4, CXCR5, and ICAM-1 and migrate to inflammatory joints through the SDF/CXCR4 pathway, which makes synovial CD4+CD25+CD69+ T cells increase and makes them express IL-17, FoxP3, and RANKL, known to promote the increase of osteoclasts. Th17 cell differentiation depends on IL-6 and IL-1 $\beta$ , and intercellular contact mediated by costimulatory molecules CD25, ICOS (inducible costimulatory), and TL1A (TNF-like ligand 1A) can also participate in Th17 cell differentiation. The MSCs induced by RA can express IL-6 and MHCII and then increase the level of IL-17. Cytokines and intercellular contact promote Th17 cells and osteoclast formation. In other studies, RA-induced MSCs did not change the number of CD4+FOXP3+Treg; therefore, MSCs may enhance the pathogenic effect of RA in patients; hence, MSC therapy for RA should be carefully considered [153, 154].

CD4+ T cells expressed by the granulocyte-macrophage colony-stimulating factor (GM-CSF) play certain roles in RA induction. AD-MSCs can reduce the number of GM-CSF+CD4+ T cells. MSCs participate in regulating immune response by promoting early adaptive regulatory T cell signals, which is characterized by a decrease in the level of T cells secreting pathogenic GM-CSF, an increase in the number of Tregs, and the development of effector Th17 cells towards IL-10-driven anti-inflammatory response, thus restoring the regulation/inflammation balance of RA [155]. The effects of MSCs within the RA environment are illustrated in Figure 2.

## 5. Regulatory Effect of MSCs on T Cells in SLE

Literature indicates that the disorder of AC clearance mechanism may be one of the pathogenesis of SLE [67]. It has been demonstrated that MSCs phagocytize ACs and regulate immune homeostasis in vivo [154], in a time- and dose-dependent manner. MSCs exposed to ACs activate the NF- $\kappa$ B pathway by recognizing phosphatidylserine, which leads to highly expressed COX2, associated with the production of a large amount of PEG2. MSCs activated by ACs can also inhibit the proliferation of CD4+ T cells more strongly than controls, which are related to soluble cytokines IFN- $\gamma$  and IL-17. However, how ACs activate the NF- $\kappa$ B pathway in MSCs is still unknown.

The decreased expression of CD4, CD25, and Foxp3 indicates a reduced number of Tregs in SLE patients. While MSCs recover Tregs through secretion of TGF- $\beta$ , Tregs inhibit the response and the production of autoantibodies by B cells through the induction of apoptosis

related to the expression of granzyme A, granzyme B, and perforin [156, 157].

Abnormal methylation of T cells in SLE patients leads to overexpression of methylation-sensitive autoimmune genes CD70, ITGAL (integrin subunit alpha L) (CD11a), selectin-1, IL-4, and IL-13 in lupus. CD70, a costimulatory factor, activates B cell response. ITGAL is related to the self-activation of T cells. Studies have shown that MEK/ERK pathway defects and T cell methylation changes in SLE patients lead to increased immune disorders. The MEK/ERK pathway of PBMC from SLE patients can be active after coculture with BM-MSCs. After coculture, DNA methyltransferase 1 DNMT1 was upregulated, and CD7, CD70, integrin, ITGAL, selectin-1, and IL-13 were downregulated in PBMC of patients. BM-MSCs downregulate the expression of methylation-related genes and reduce the self-activation of PBMC through the MEK/ERK pathway [158–160].

Several other studies have investigated the Th17 and Treg imbalance found in SLE patients. Stress response and immune regulation molecule heme oxygenase-1 (HO-1) are involved in the induction of Tregs. MSCs can express HO-1 and participate in the induction of Treg cell subsets, but there are big individual differences. Perhaps HO-1 expressed by MSCs can play certain roles in SLE patients, and the specific mechanism needs to be studied [161, 162].

## 6. Conclusion

As a cell-based therapy, MSCs possess the potential of ameliorating injury or possibly offering a cure for patients with immune-mediated conditions. Available document on the contribution of MSCs in restoring T cell balance within the autoimmune environment is promising, as the MSCs exert their immunoregulatory effects via direct contact and secretion of active factors. The mechanism of action of MSCs overlaps and has quite a few differences in various autoimmune diseases, which may be related to the origin of MSCs and the heterogeneity of autoimmune diseases. Additionally, MSCs treated with certain factors to overexpress desired cytokines result in stronger regulation of T cell immunity. Further explorations of key targets of MSCs during T cell regulation and their associated mechanisms in autoimmune diseases are needed to enhance understanding towards improving the therapeutics of MSCs.

## Conflicts of Interest

All authors declare no conflict of interest.

## Authors' Contributions

Zhiping Wei and Jintao Yuan contributed equally to this work. All authors approved the final version of the article.

## Acknowledgments

The study was funded by the National Natural Science Foundation of China (Grant no. 32000903), the Project of Zhenjiang Key Research and Development Plan (social



development) (Grant no. SH2019025), Project Funded by the Priority Academic Program Development of Jiangsu Higher Education Institutions, Zhenjiang Key Laboratory of High Technology Research on Exosomes Foundation and Transformation Application (grant no. SS2018003), and Scientific Research Project of Health Commission of Jiangsu Province (Grant no. Z2019036).

## References

- [1] J. L. Spees, R. H. Lee, and C. A. Gregory, "Mechanisms of mesenchymal stem/stromal cell function," *Stem Cell Research & Therapy*, vol. 7, no. 1, p. 125, 2016.
- [2] P. T. Sharpe, "Dental mesenchymal stem cells," *Development*, vol. 143, no. 13, pp. 2273–2280, 2016.
- [3] Y. Sakaguchi, I. Sekiya, K. Yagishita, and T. Muneta, "Comparison of human stem cells derived from various mesenchymal tissues: superiority of synovium as a cell source," *Arthritis and Rheumatism*, vol. 52, no. 8, pp. 2521–2529, 2005.
- [4] S. Ranga Rao and R. Subbarayan, "Passage-dependent expression of STRO-1 in human gingival mesenchymal stem cells," *Journal of Cellular Biochemistry*, vol. 120, no. 3, pp. 2810–2815, 2018.
- [5] S. P. Bruder, D. J. Fink, and A. I. Caplan, "Mesenchymal stem cells in bone development, bone repair, and skeletal regeneration therapy," *Journal of Cellular Biochemistry*, vol. 56, no. 3, pp. 283–294, 1994.
- [6] F. J. Lv, R. S. Tuan, K. M. C. Cheung, and V. Y. L. Leung, "Concise review: the surface markers and identity of human mesenchymal stem cells," *Stem Cells*, vol. 32, no. 6, pp. 1408–1419, 2014.
- [7] F. Cakiroglu, J. W. Osbahr, J. Kramer, and J. Rohwedel, "Differences of cell surface marker expression between bone marrow- and kidney-derived murine mesenchymal stromal cells and fibroblasts," *Cellular and Molecular Biology*, vol. 62, no. 12, pp. 11–17, 2016.
- [8] P. Szaraz, Y. S. Gratch, F. Iqbal, and C. L. Librach, "In vitro differentiation of human mesenchymal stem cells into functional cardiomyocyte-like cells," *Journal of Visualized Experiments*, vol. 126, article e55757, 2017.
- [9] Y. B. Yu, Y. Song, Y. Chen, F. Zhang, and F. Z. Qi, "Differentiation of umbilical cord mesenchymal stem cells into hepatocytes in comparison with bone marrow mesenchymal stem cells," *Molecular Medicine Reports*, vol. 18, no. 2, pp. 2009–2016, 2018.
- [10] D. K. Ocansey, B. Pei, Y. Yan et al., "Improved therapeutics of modified mesenchymal stem cells: an update," *Journal of Translational Medicine*, vol. 18, no. 1, p. 42, 2020.
- [11] R. Ramezanifard, M. Kabiri, and H. H. Ahvaz, "Effects of platelet rich plasma and chondrocyte co-culture on MSC chondrogenesis, hypertrophy and pathological responses," *EXCLI Journal*, vol. 16, pp. 1031–1045, 2017.
- [12] G. R. Lee, "The balance of Th17 versus Treg cells in autoimmunity," *International Journal of Molecular Sciences*, vol. 19, no. 3, p. 730, 2018.
- [13] M. Dominguez-Villar and D. A. Hafler, "Regulatory T cells in autoimmune disease," *Nature Immunology*, vol. 19, no. 7, pp. 665–673, 2018.
- [14] K. A. Russell, N. H. Chow, D. Dukoff et al., "Characterization and immunomodulatory effects of canine adipose tissue- and bone marrow-derived mesenchymal stromal cells," *PLoS One*, vol. 11, no. 12, article e0167442, 2016.
- [15] H. Li, R. Ghazanfari, D. Zacharaki, H. C. Lim, and S. Scheduling, "Isolation and characterization of primary bone marrow mesenchymal stromal cells," *Annals of the New York Academy of Sciences*, vol. 1370, no. 1, pp. 109–118, 2016.
- [16] F. Djouad, C. Bony, T. Häupl et al., "Transcriptional profiles discriminate bone marrow-derived and synovium-derived mesenchymal stem cells," *Arthritis Research & Therapy*, vol. 7, no. 6, pp. R1304–R1315, 2005.
- [17] C. Brown, C. McKee, S. Bakshi et al., "Mesenchymal stem cells: cell therapy and regeneration potential," *Journal of Tissue Engineering and Regenerative Medicine*, vol. 13, no. 9, pp. 1738–1755, 2019.
- [18] E. Amati, O. Perbellini, G. Rotta et al., "High-throughput immunophenotypic characterization of bone marrow- and cord blood-derived mesenchymal stromal cells reveals common and differentially expressed markers: identification of angiotensin-converting enzyme (CD143) as a marker differentially expressed between adult and perinatal tissue sources," *Stem Cell Research & Therapy*, vol. 9, no. 1, p. 10, 2018.
- [19] H. Pham, R. Tonai, M. Wu, C. Birtolo, and M. Chen, "CD73, CD90, CD105 and cadherin-11 RT-PCR screening for mesenchymal stem cells from cryopreserved human cord tissue," *International Journal of Stem Cells/International Journal of Stem Cells*, vol. 11, no. 1, pp. 26–38, 2018.
- [20] S. Mishra, J. K. Sevak, A. Das, G. A. Arimbasseri, S. Bhatnagar, and S. D. Gopinath, "Umbilical cord tissue is a robust source for mesenchymal stem cells with enhanced myogenic differentiation potential compared to cord blood," *Scientific Reports*, vol. 10, no. 1, p. 18978, 2020.
- [21] P. Li, C. Zhou, L. Yin, X. Meng, and L. Zhang, "Role of hypoxia in viability and endothelial differentiation potential of UC-MSCs and VEGF interference," *Zhong Nan Da Xue Xue Bao. Yi Xue Ban*, vol. 38, no. 4, pp. 329–340, 2013.
- [22] Z. Chen, Q. Kuang, X. J. Lao, J. Yang, W. Huang, and D. Zhou, "Differentiation of UC-MSCs into hepatocyte-like cells in partially hepatectomized model rats," *Experimental and Therapeutic Medicine*, vol. 12, no. 3, pp. 1775–1779, 2016.
- [23] S. R. Ali, W. Ahmad, N. Naeem, A. Salim, and I. Khan, "Small molecule 2'-deoxycytidine differentiates human umbilical cord-derived MSCs into cardiac progenitors in vitro and their in vivo xeno-transplantation improves cardiac function," *Molecular and Cellular Biochemistry*, vol. 470, no. 1–2, pp. 99–113, 2020.
- [24] C. Peng, L. Lu, Y. Li, and J. Hu, "Neurospheres induced from human adipose-derived stem cells as a new source of neural progenitor cells," *Cell Transplantation*, vol. 28, 1\_suppl, pp. 66s–75s, 2019.
- [25] F. Y. Meligy, K. Shigemura, H. M. Behnsawy, M. Fujisawa, M. Kawabata, and T. Shirakawa, "The efficiency of in vitro isolation and myogenic differentiation of MSCs derived from adipose connective tissue, bone marrow, and skeletal muscle tissue," *In Vitro Cellular & Developmental Biology - Animal*, vol. 48, no. 4, pp. 203–215, 2012.
- [26] M. el-Sayed, M. A. el-Feky, M. I. el-Amir et al., "Immunomodulatory effect of mesenchymal stem cells: cell origin and cell quality variations," *Molecular Biology Reports*, vol. 46, no. 1, pp. 1157–1165, 2019.
- [27] N. Nuti, C. Corallo, B. M. F. Chan, M. Ferrari, and B. Gerami-Naini, "Multipotent differentiation of human dental pulp

- stem cells: a literature review,” *Stem Cell Reviews and Reports*, vol. 12, no. 5, pp. 511–523, 2016.
- [28] M. Lei, K. Li, B. Li, L. N. Gao, F. M. Chen, and Y. Jin, “Mesenchymal stem cell characteristics of dental pulp and periodontal ligament stem cells after *in vivo* transplantation,” *Biomaterials*, vol. 35, no. 24, pp. 6332–6343, 2014.
- [29] J. Fan, R. R. Varshney, L. Ren, D. Cai, and D. A. Wang, “Synovium-derived mesenchymal stem cells: a new cell source for musculoskeletal regeneration,” *Tissue Engineering. Part B, Reviews*, vol. 15, no. 1, pp. 75–86, 2009.
- [30] I. Sanz and F. Lund, “Complexity and heterogeneity - the defining features of autoimmune disease,” *Current Opinion in Immunology*, vol. 61, pp. iii–vi, 2019.
- [31] Y. Z. Zhang and Y. Y. Li, “Inflammatory bowel disease: pathogenesis,” *World Journal of Gastroenterology*, vol. 20, no. 1, pp. 91–99, 2014.
- [32] A. N. Ananthakrishnan, “Epidemiology and risk factors for IBD,” *Nature Reviews Gastroenterology & Hepatology*, vol. 12, no. 4, pp. 205–217, 2015.
- [33] J. W. Windsor and G. G. Kaplan, “Evolving epidemiology of IBD,” *Current Gastroenterology Reports*, vol. 21, no. 8, p. 40, 2019.
- [34] S. C. Ng, H. Y. Shi, N. Hamidi et al., “Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies,” *The Lancet*, vol. 390, no. 10114, pp. 2769–2778, 2017.
- [35] I. Hilmi, F. Jaya, A. Chua, W. C. Heng, H. Singh, and K. L. Goh, “A first study on the incidence and prevalence of IBD in Malaysia—results from the Kinta Valley IBD Epidemiology Study,” *Journal of Crohn's & Colitis*, vol. 9, no. 5, pp. 404–409, 2015.
- [36] G. G. Kaplan, “The global burden of IBD: from 2015 to 2025,” *Nature Reviews Gastroenterology & Hepatology*, vol. 12, no. 12, pp. 720–727, 2015.
- [37] A. R. Weingarden and B. P. Vaughn, “Intestinal microbiota, fecal microbiota transplantation, and inflammatory bowel disease,” *Gut Microbes*, vol. 8, no. 3, pp. 238–252, 2017.
- [38] A. B. Pithadia and S. Jain, “Treatment of inflammatory bowel disease (IBD),” *Pharmacological Reports*, vol. 63, no. 3, pp. 629–642, 2011.
- [39] D. Y. Jeong, S. Kim, M. J. Son et al., “Induction and maintenance treatment of inflammatory bowel disease: a comprehensive review,” *Autoimmunity Reviews*, vol. 18, no. 5, pp. 439–454, 2019.
- [40] F. Gomollón, A. Dignass, V. Annese et al., “3rd European evidence-based consensus on the diagnosis and management of Crohn's disease 2016: part 1: diagnosis and medical management,” *Journal of Crohn's & Colitis*, vol. 11, no. 1, pp. 3–25, 2017.
- [41] X. Shi, Q. Chen, and F. Wang, “Mesenchymal stem cells for the treatment of ulcerative colitis: a systematic review and meta-analysis of experimental and clinical studies,” *Stem Cell Research & Therapy*, vol. 10, no. 1, p. 266, 2019.
- [42] H. Liu, Z. Liang, F. Wang et al., “Exosomes from mesenchymal stromal cells reduce murine colonic inflammation via a macrophage-dependent mechanism,” *JCI Insight*, vol. 4, no. 24, 2019.
- [43] S. Soontarak, L. Chow, V. Johnson et al., “Mesenchymal stem cells (MSC) derived from induced pluripotent stem cells (iPSC) equivalent to adipose-derived MSC in promoting intestinal healing and microbiome normalization in mouse inflammatory bowel disease model,” *Stem Cells Translational Medicine*, vol. 7, no. 6, pp. 456–467, 2018.
- [44] V. B. F. Alves, B. C. de Sousa, M. T. C. Fonseca et al., “A single administration of human adipose tissue-derived mesenchymal stromal cells (MSC) induces durable and sustained long-term regulation of inflammatory response in experimental colitis,” *Clinical and Experimental Immunology*, vol. 196, no. 2, pp. 139–154, 2019.
- [45] R. I. Azevedo, E. Minskaia, A. Fernandes-Platzgummer et al., “Mesenchymal stromal cells induce regulatory T cells via epigenetic conversion of human conventional CD4 T cells *in vitro*,” *Stem Cells*, vol. 38, no. 8, pp. 1007–1019, 2020.
- [46] J. Panés, D. García-Olmo, G. van Assche et al., “Expanded allogeneic adipose-derived mesenchymal stem cells (Cx601) for complex perianal fistulas in Crohn's disease: a phase 3 randomised, double-blind controlled trial,” *The Lancet*, vol. 388, no. 10051, pp. 1281–1290, 2016.
- [47] F. Cheng, Z. Huang, and Z. Li, “Mesenchymal stem-cell therapy for perianal fistulas in Crohn's disease: a systematic review and meta-analysis,” *Techniques in Coloproctology*, vol. 23, no. 7, pp. 613–623, 2019.
- [48] A. L. Lightner, Z. Wang, A. C. Zubair, and E. J. Dozois, “A systematic review and meta-analysis of mesenchymal stem cell injections for the treatment of perianal Crohn's disease: progress made and future directions,” *Diseases of the Colon and Rectum*, vol. 61, no. 5, pp. 629–640, 2018.
- [49] Y. B. Cho, K. J. Park, S. N. Yoon et al., “Long-term results of adipose-derived stem cell therapy for the treatment of Crohn's fistula,” *Stem Cells Translational Medicine*, vol. 4, no. 5, pp. 532–537, 2015.
- [50] S. Otagiri, S. Ohnishi, A. Miura et al., “Evaluation of amnion-derived mesenchymal stem cells for treatment-resistant moderate Crohn's disease: study protocol for a phase I/II, dual-centre, open-label, uncontrolled, dose-response trial,” *BMJ Open Gastroenterology*, vol. 5, no. 1, article e000206, 2018.
- [51] I. Molendijk, B. A. Bonsing, H. Roelofs et al., “Allogeneic bone marrow-derived mesenchymal stromal cells promote healing of refractory perianal fistulas in patients with Crohn's disease,” *Gastroenterology*, vol. 149, no. 4, pp. 918–927, 2015.
- [52] D. K. Ocansey, W. Qiu, J. Wang et al., “The achievements and challenges of mesenchymal stem cell-based therapy in inflammatory bowel disease and its associated colorectal cancer,” *Stem Cells International*, vol. 2020, Article ID 7819824, 18 pages, 2020.
- [53] D. K. W. Ocansey, L. Zhang, Y. Wang et al., “Exosome-mediated effects and applications in inflammatory bowel disease,” *Biological Reviews*, vol. 95, no. 5, pp. 1287–1307, 2020.
- [54] A. Jauregui-Amezaga, M. Rovira, P. Marin et al., “Improving safety of autologous haematopoietic stem cell transplantation in patients with Crohn's disease,” *Gut*, vol. 65, no. 9, pp. 1456–1462, 2016.
- [55] M. Cross, E. Smith, D. Hoy et al., “The global burden of rheumatoid arthritis: estimates from the global burden of disease 2010 study,” *Annals of the Rheumatic Diseases*, vol. 73, no. 7, pp. 1316–1322, 2014.
- [56] T. Otón and L. Carmona, “The epidemiology of established rheumatoid arthritis,” *Best Practice & Research Clinical Rheumatology*, vol. 33, no. 5, article 101477, 2019.
- [57] J. A. Pradeepkiran, “Insights of rheumatoid arthritis risk factors and associations,” *Journal of Translational Autoimmunity*, vol. 2, article 100012, 2019.

- [58] R. S. de Molon, C. Rossa Jr., R. M. Thurlings, J. A. Cirelli, and M. I. Koenders, "Linkage of periodontitis and rheumatoid arthritis: current evidence and potential biological interactions," *International Journal of Molecular Sciences*, vol. 20, no. 18, p. 4541, 2019.
- [59] J. Potempa, P. Mydel, and J. Koziel, "The case for periodontitis in the pathogenesis of rheumatoid arthritis," *Nature Reviews Rheumatology*, vol. 13, no. 10, pp. 606–620, 2017.
- [60] Q. Guo, Y. Wang, D. Xu, J. Nossent, N. J. Pavlos, and J. Xu, "Rheumatoid arthritis: pathological mechanisms and modern pharmacologic therapies," *Bone Research*, vol. 6, no. 1, p. 15, 2018.
- [61] L. Wang, L. Wang, X. Cong et al., "Human umbilical cord mesenchymal stem cell therapy for patients with active rheumatoid arthritis: safety and efficacy," *Stem Cells and Development*, vol. 22, no. 24, pp. 3192–3202, 2013.
- [62] J. M. Álvaro-Gracia, J. A. Jover, R. García-Vicuña et al., "Intravenous administration of expanded allogeneic adipose-derived mesenchymal stem cells in refractory rheumatoid arthritis (Cx611): results of a multicentre, dose escalation, randomised, single-blind, placebo-controlled phase Ib/IIa clinical trial," *Annals of the Rheumatic Diseases*, vol. 76, no. 1, pp. 196–202, 2016.
- [63] A. Fathollahi, N. B. Gabalou, and S. Aslani, "Mesenchymal stem cell transplantation in systemic lupus erythematosus, a mesenchymal stem cell disorder," *Lupus*, vol. 27, no. 7, pp. 1053–1064, 2018.
- [64] A. A. Justiz Vaillant, *Systemic lupus erythematosus*, in *StatPearls*, StatPearls Publishing LLC, Treasure Island (FL), 2020.
- [65] Y. Deng and B. P. Tsao, "Advances in lupus genetics and epigenetics," *Current Opinion in Rheumatology*, vol. 26, no. 5, pp. 482–492, 2014.
- [66] Z. Liu, Y. Yu, Y. Yue et al., "Genetic alleles associated with SLE susceptibility and clinical manifestations in Hispanic patients from the Dominican Republic," *Current Molecular Medicine*, vol. 19, no. 3, pp. 164–171, 2019.
- [67] A. Mahajan, M. Herrmann, and L. E. Muñoz, "Clearance deficiency and cell death pathways: a model for the pathogenesis of SLE," *Frontiers in Immunology*, vol. 7, p. 35, 2016.
- [68] F. Rees, M. Doherty, M. J. Grainge, P. Lanyon, and W. Zhang, "The worldwide incidence and prevalence of systemic lupus erythematosus: a systematic review of epidemiological studies," *Rheumatology*, vol. 56, no. 11, pp. 1945–1961, 2017.
- [69] A. Fava and M. Petri, "Systemic lupus erythematosus: diagnosis and clinical management," *Journal of Autoimmunity*, vol. 96, pp. 1–13, 2019.
- [70] S. Almaani and B. H. Rovin, "B-cell therapy in lupus nephritis: an overview," *Nephrology, Dialysis, Transplantation*, vol. 34, no. 1, pp. 22–29, 2019.
- [71] M. Gatto, M. Zen, L. Iaccarino, and A. Doria, "New therapeutic strategies in systemic lupus erythematosus management," *Nature Reviews Rheumatology*, vol. 15, no. 1, pp. 30–48, 2019.
- [72] A. Jones, P. Müller, C. J. Dore et al., "Belimumab after B cell depletion therapy in patients with systemic lupus erythematosus (BEAT Lupus) protocol: a prospective multicentre, double-blind, randomised, placebo-controlled, 52-week phase II clinical trial," *BMJ Open*, vol. 9, no. 12, article e032569, 2019.
- [73] S. Liu, Y. L. Guo, J. Y. Yang, W. Wang, and J. Xu, "Efficacy of mesenchymal stem cells on systemic lupus erythematosus: a meta-analysis," *Beijing Da Xue Xue Bao Yi Xue Ban*, vol. 50, no. 6, pp. 1014–1021, 2018.
- [74] T. Zhou, H. Y. Li, C. Liao, W. Lin, and S. Lin, "Clinical efficacy and safety of mesenchymal stem cells for systemic lupus erythematosus," *Stem Cells International*, vol. 2020, Article ID 6518508, 11 pages, 2020.
- [75] R. J. Cheng, A. J. Xiong, Y. H. Li et al., "Mesenchymal stem cells: allogeneic MSC may be immunosuppressive but autologous MSC are dysfunctional in lupus patients," *Frontiers in Cell and Development Biology*, vol. 7, p. 285, 2019.
- [76] Z. Feng, Y. Zhai, Z. Zheng et al., "Loss of A20 in BM-MSCs regulates the Th17/Treg balance in rheumatoid arthritis," *Scientific Reports*, vol. 8, no. 1, p. 427, 2018.
- [77] B. Chandravanshi and R. R. Bhonde, "Human umbilical cord-derived stem cells: isolation, characterization, differentiation, and application in treating diabetes," *Critical Reviews in Biomedical Engineering*, vol. 46, no. 5, pp. 399–412, 2018.
- [78] F. X. Cuascut and G. J. Hutton, "Stem cell-based therapies for multiple sclerosis: current perspectives," *Biomedicine*, vol. 7, no. 2, p. 26, 2019.
- [79] Y. Cao, X. Jin, Y. Sun, and W. Wen, "Therapeutic effect of mesenchymal stem cell on Hashimoto's thyroiditis in a rat model by modulating Th17/Treg cell balance," *Autoimmunity*, vol. 53, no. 1, pp. 35–45, 2020.
- [80] S. Khan, R. S. Khan, and P. N. Newsome, "Cellular therapies for the treatment of immune-mediated GI and liver disease," *British Medical Bulletin*, vol. 136, no. 1, pp. 127–141, 2020.
- [81] D. Esquivel, R. Mishra, and A. Srivastava, "Stem cell therapy offers a possible safe and promising alternative approach for treating vitiligo: a review," *Current Pharmaceutical Design*, vol. 26, no. 37, pp. 4815–4821, 2020.
- [82] on behalf of the MESEMS study group, A. Uccelli, A. Laroni et al., "MEsenchymal StEm cells for Multiple Sclerosis (MESEMS): a randomized, double blind, cross-over phase I/II clinical trial with autologous mesenchymal stem cells for the therapy of multiple sclerosis," *Trials*, vol. 20, no. 1, p. 263, 2019.
- [83] L. Vija, D. Farge, J. F. Gautier et al., "Les cellules souches mésenchymateuses comme nouvelle approche thérapeutique du diabète de type 1," *Diabetes & Metabolism*, vol. 35, no. 2, pp. 85–93, 2009.
- [84] P. Boháčová and V. Holán, "Mesenchymal stem cells and type 1 diabetes treatment," *Vnitřní Lékařství*, vol. 64, no. 7–8, pp. 725–728, 2018.
- [85] Y. Chen, Q. Yu, Y. Hu, and Y. Shi, "Current research and use of mesenchymal stem cells in the therapy of autoimmune diseases," *Current Stem Cell Research & Therapy*, vol. 14, no. 7, pp. 579–582, 2019.
- [86] J. Cai, Z. Wu, X. Xu et al., "Umbilical cord mesenchymal stromal cell with autologous bone marrow cell transplantation in established type 1 diabetes: a pilot randomized controlled open-label clinical study to assess safety and impact on insulin secretion," *Diabetes Care*, vol. 39, no. 1, pp. 149–157, 2015.
- [87] R. R. Bhonde, P. Sheshadri, S. Sharma, and A. Kumar, "Making surrogate  $\beta$ -cells from mesenchymal stromal cells: perspectives and future endeavors," *The International Journal of Biochemistry & Cell Biology*, vol. 46, pp. 90–102, 2014.
- [88] S. Daneshmandi, M. H. Karimi, and A. A. Pourfathollah, "TGF- $\beta$  engineered mesenchymal stem cells (TGF- $\beta$ /MSCs) for treatment of type 1 diabetes (T1D) mice model," *International Immunopharmacology*, vol. 44, pp. 191–196, 2017.

- [89] L. Zazzeroni, G. Lanzoni, G. Pasquinelli, and C. Ricordi, "Considerations on the harvesting site and donor derivation for mesenchymal stem cells-based strategies for diabetes," *CellR4 Repair Replacement Regeneration & Reprogramming*, vol. 5, no. 5, 2017.
- [90] D. Karussis, C. Karageorgiou, A. Vaknin-Dembinsky et al., "Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis," *Archives of Neurology*, vol. 67, no. 10, pp. 1187–1194, 2010.
- [91] S. Llifriu, M. Sepúlveda, Y. Blanco et al., "Randomized placebo-controlled phase II trial of autologous mesenchymal stem cells in multiple sclerosis," *PLoS One*, vol. 9, no. 12, article e113936, 2014.
- [92] M. Mohyeddin Bonab, M. Ali Sahraian, A. Aghsaie et al., "Autologous mesenchymal stem cell therapy in progressive multiple sclerosis: an open label study," *Current Stem Cell Research & Therapy*, vol. 7, no. 6, pp. 407–414, 2012.
- [93] S. Dahbour, F. Jamali, D. Alhattab et al., "Mesenchymal stem cells and conditioned media in the treatment of multiple sclerosis patients: clinical, ophthalmological and radiological assessments of safety and efficacy," *CNS Neuroscience & Therapeutics*, vol. 23, no. 11, pp. 866–874, 2017.
- [94] V. K. Harris, R. Faroqui, T. Vyshkina, and S. A. Sadiq, "Characterization of autologous mesenchymal stem cell-derived neural progenitors as a feasible source of stem cells for central nervous system applications in multiple sclerosis," *Stem Cells Translational Medicine*, vol. 1, no. 7, pp. 536–547, 2012.
- [95] G. Yao, J. Qi, J. Liang et al., "Mesenchymal stem cell transplantation alleviates experimental Sjögren's syndrome through IFN- $\beta$ /IL-27 signaling axis," *Theranostics*, vol. 9, no. 26, pp. 8253–8265, 2019.
- [96] B. Shi, J. Qi, G. Yao et al., "Mesenchymal stem cell transplantation ameliorates Sjögren's syndrome via suppressing IL-12 production by dendritic cells," *Stem Cell Research & Therapy*, vol. 9, no. 1, p. 308, 2018.
- [97] D. Wang, H. Zhang, J. Liang et al., "Effect of allogeneic bone marrow-derived mesenchymal stem cells transplantation in a polyI:C-induced primary biliary cirrhosis mouse model," *Clinical and Experimental Medicine*, vol. 11, no. 1, pp. 25–32, 2011.
- [98] J. Fan, X. Tang, Q. Wang et al., "Mesenchymal stem cells alleviate experimental autoimmune cholangitis through immunosuppression and cytoprotective function mediated by galectin-9," *Stem Cell Research & Therapy*, vol. 9, no. 1, p. 237, 2018.
- [99] L. Wang, J. Li, H. Liu et al., "Pilot study of umbilical cord-derived mesenchymal stem cell transfusion in patients with primary biliary cirrhosis," *Journal of Gastroenterology and Hepatology*, vol. 28, Suppl 1, pp. 85–92, 2013.
- [100] B. Tan, W. Yuan, J. Li et al., "Therapeutic effect of human amniotic epithelial cells in murine models of Hashimoto's thyroiditis and systemic lupus erythematosus," *Cytotherapy*, vol. 20, no. 10, pp. 1247–1258, 2018.
- [101] K. Che, X. Liu, J. Chi et al., "The effects of adipose-derived mesenchymal stem cells combined with sodium selenite on Hashimoto's thyroiditis," *American Journal of Translational Research*, vol. 12, no. 10, pp. 6422–6433, 2020.
- [102] Y. Li, K. Ma, L. Zhang, H. Xu, and N. Zhang, "Human umbilical cord blood derived-mesenchymal stem cells alleviate dextran sulfate sodium-induced colitis by increasing regulatory T cells in mice," *Frontiers in Cell and Development Biology*, vol. 8, article 604021, 2020.
- [103] M. Heidari, S. Pouya, K. Baghaei et al., "The immunomodulatory effects of adipose-derived mesenchymal stem cells and mesenchymal stem cells-conditioned medium in chronic colitis," *Journal of Cellular Physiology*, vol. 233, no. 11, pp. 8754–8766, 2018.
- [104] M. Krampera, L. Cosmi, R. Angeli et al., "Role for interferon-gamma in the immunomodulatory activity of human bone marrow mesenchymal stem cells," *Stem Cells*, vol. 24, no. 2, pp. 386–398, 2006.
- [105] M. Bulati, V. Miceli, A. Gallo et al., "The immunomodulatory properties of the human amnion-derived mesenchymal stromal/stem cells are induced by INF- $\gamma$  produced by activated lymphomonocytes and are mediated by cell-to-cell contact and soluble factors," *Frontiers in Immunology*, vol. 11, p. 54, 2020.
- [106] Q. Q. Chen, L. Yan, C. Z. Wang et al., "Mesenchymal stem cells alleviate TNBS-induced colitis by modulating inflammatory and autoimmune responses," *World Journal of Gastroenterology*, vol. 19, no. 29, pp. 4702–4717, 2013.
- [107] Y. Z. Gu, Q. Xue, Y. J. Chen et al., "Different roles of PD-L1 and FasL in immunomodulation mediated by human placenta-derived mesenchymal stem cells," *Human Immunology*, vol. 74, no. 3, pp. 267–276, 2013.
- [108] S. Aggarwal and M. F. Pittenger, "Human mesenchymal stem cells modulate allogeneic immune cell responses," *Blood*, vol. 105, no. 4, pp. 1815–1822, 2005.
- [109] N. Ohkura and S. Sakaguchi, "Transcriptional and epigenetic basis of Treg cell development and function: its genetic anomalies or variations in autoimmune diseases," *Cell Research*, vol. 30, no. 6, pp. 465–474, 2020.
- [110] I. Tindemans, M. E. Joosse, and J. N. Samsom, "Dissecting the heterogeneity in T-cell mediated inflammation in IBD," *Cell*, vol. 9, no. 1, p. 110, 2020.
- [111] E. S. Lee, J. Y. Lim, K. I. Im et al., "Adoptive transfer of Treg cells combined with mesenchymal stem cells facilitates repopulation of endogenous Treg cells in a murine acute GVHD model," *PLoS One*, vol. 10, no. 9, article e0138846, 2015.
- [112] J. Y. Lim, M. J. Park, K. I. Im et al., "Combination cell therapy using mesenchymal stem cells and regulatory T-cells provides a synergistic immunomodulatory effect associated with reciprocal regulation of TH1/TH2 and th17/treg cells in a murine acute graft-versus-host disease model," *Cell Transplantation*, vol. 23, no. 6, pp. 703–714, 2014.
- [113] J. Guo, L. Y. Wang, J. Wu, L. F. Xu, and M. Sun, "The JAK2 inhibitor AG490 regulates the Treg/Th17 balance and alleviates DSS-induced intestinal damage in IBD rats," *Clinical and Experimental Pharmacology & Physiology*, vol. 47, no. 8, pp. 1374–1381, 2020.
- [114] S. M. Melief, E. Schrama, M. H. Brugman et al., "Multipotent stromal cells induce human regulatory T cells through a novel pathway involving skewing of monocytes toward anti-inflammatory macrophages," *Stem Cells*, vol. 31, no. 9, pp. 1980–1991, 2013.
- [115] Y. Goto, C. Panea, G. Nakato et al., "Segmented filamentous bacteria antigens presented by intestinal dendritic cells drive mucosal Th17 cell differentiation," *Immunity*, vol. 40, no. 4, pp. 594–607, 2014.
- [116] M. Pierau, S. Engelmann, D. Reinhold, T. Lapp, B. Schraven, and U. H. Bommhardt, "Protein kinase B/Akt signals impair

- Th17 differentiation and support natural regulatory T cell function and induced regulatory T cell formation," *Journal of Immunology*, vol. 183, no. 10, pp. 6124–6134, 2009.
- [117] D. Kotlarz, B. Marquardt, T. Barøy et al., "Human TGF- $\beta$ 1 deficiency causes severe inflammatory bowel disease and encephalopathy," *Nature Genetics*, vol. 50, no. 3, pp. 344–348, 2018.
- [118] I. Marafini, F. Zorzi, S. Codazza, F. Pallone, and G. Monteleone, "TGF-Beta signaling manipulation as potential therapy for IBD," *Current Drug Targets*, vol. 14, no. 12, pp. 1400–1404, 2013.
- [119] F. Y. Yang, R. Chen, X. Zhang et al., "Preconditioning enhances the therapeutic effects of mesenchymal stem cells on colitis through PGE2-mediated T-cell modulation," *Cell Transplantation*, vol. 27, no. 9, pp. 1352–1367, 2018.
- [120] I. K. Jang, H. H. Yoon, M. S. Yang et al., "B7-H1 inhibits T cell proliferation through MHC class II in human mesenchymal stem cells," *Transplantation Proceedings*, vol. 46, no. 5, pp. 1638–1641, 2014.
- [121] Y. Rui, T. Honjo, and S. Chikuma, "Programmed cell death 1 inhibits inflammatory helper T-cell development through controlling the innate immune response," *Proceedings of the National Academy of Sciences*, vol. 110, no. 40, pp. 16073–16078, 2013.
- [122] J. Y. Kim, M. Park, Y. H. Kim et al., "Tonsil-derived mesenchymal stem cells (T-MSCs) prevent Th17-mediated autoimmune response via regulation of the programmed death-1/programmed death ligand-1 (PD-1/PD-L1) pathway," *Journal of Tissue Engineering and Regenerative Medicine*, vol. 12, no. 2, pp. e1022–e1033, 2018.
- [123] X. Gui, M. Iacucci, and S. Ghosh, "Dysregulation of IL6/IL6R-STAT3-SOCS3 signaling pathway in IBD-associated colorectal dysplastic lesions as compared to sporadic colorectal adenomas in non-IBD patients," *Pathology, Research and Practice*, vol. 216, no. 11, article 153211, 2020.
- [124] C. Soendergaard, F. H. Bergenheim, J. T. Bjerrum, and O. H. Nielsen, "Targeting JAK-STAT signal transduction in IBD," *Pharmacology & Therapeutics*, vol. 192, pp. 100–111, 2018.
- [125] H. S. Kim, T. H. Shin, B. C. Lee et al., "Human umbilical cord blood mesenchymal stem cells reduce colitis in mice by activating NOD2 signaling to COX2," *Gastroenterology*, vol. 145, no. 6, pp. 1392–403.e1-8, 2013.
- [126] F. Liotta, R. Angeli, L. Cosmi et al., "Toll-like receptors 3 and 4 are expressed by human bone marrow-derived mesenchymal stem cells and can inhibit their T-cell modulatory activity by impairing notch signaling," *Stem Cells*, vol. 26, no. 1, pp. 279–289, 2008.
- [127] D. Shi, L. Liao, B. Zhang et al., "Human adipose tissue-derived mesenchymal stem cells facilitate the immunosuppressive effect of cyclosporin A on T lymphocytes through Jagged-1-mediated inhibition of NF- $\kappa$ B signaling," *Experimental Hematology*, vol. 39, no. 2, pp. 214–224.e1, 2011.
- [128] Y. Qiu, J. Guo, R. Mao et al., "TLR3 preconditioning enhances the therapeutic efficacy of umbilical cord mesenchymal stem cells in TNBS-induced colitis via the TLR3-Jagged-1-Notch-1 pathway," *Mucosal Immunology*, vol. 10, no. 3, pp. 727–742, 2017.
- [129] X. Li, Q. Wang, L. Ding et al., "Intercellular adhesion molecule-1 enhances the therapeutic effects of MSCs in a dextran sulfate sodium-induced colitis models by promoting MSCs homing to murine colons and spleens," *Stem Cell Research & Therapy*, vol. 10, no. 1, pp. 267–267, 2019.
- [130] H. S. Yoo, K. Lee, K. Na et al., "Mesenchymal stromal cells inhibit CD25 expression via the mTOR pathway to potentiate T-cell suppression," *Cell Death & Disease*, vol. 8, no. 2, 2017.
- [131] W. Cheng, J. Su, Y. Hu et al., "Interleukin-25 primed mesenchymal stem cells achieve better therapeutic effects on dextran sulfate sodium-induced colitis via inhibiting Th17 immune response and inducing T regulatory cell phenotype," *American Journal of Translational Research*, vol. 9, no. 9, pp. 4149–4160, 2017.
- [132] T. Shi, Y. Xie, Y. Fu et al., "The signaling axis of microRNA-31/interleukin-25 regulates Th1/Th17-mediated inflammation response in colitis," *Mucosal Immunology*, vol. 10, no. 4, pp. 983–995, 2017.
- [133] D. Olivares-Villagómez and L. Van Kaer, "Intestinal intraepithelial lymphocytes: sentinels of the mucosal barrier," *Trends in Immunology*, vol. 39, no. 4, pp. 264–275, 2018.
- [134] H. Ma, Y. Qiu, and H. Yang, "Intestinal intraepithelial lymphocytes: maintainers of intestinal immune tolerance and regulators of intestinal immunity," *Journal of Leukocyte Biology*, vol. 109, no. 2, pp. 339–347, 2021.
- [135] D. Abuquteish and J. Putra, "Upper gastrointestinal tract involvement of pediatric inflammatory bowel disease: a pathological review," *World Journal of Gastroenterology*, vol. 25, no. 16, pp. 1928–1935, 2019.
- [136] G. Wang, K. Cao, K. Liu et al., "Kynurenic acid, an IDO metabolite, controls TSG-6-mediated immunosuppression of human mesenchymal stem cells," *Cell Death and Differentiation*, vol. 25, no. 7, pp. 1209–1223, 2018.
- [137] Y. Yan, G. X. Zhang, B. Gran et al., "IDO upregulates regulatory T cells via tryptophan catabolite and suppresses encephalitogenic T cell responses in experimental autoimmune encephalomyelitis," *Journal of Immunology*, vol. 185, no. 10, pp. 5953–5961, 2010.
- [138] S. Zhang, J. Fang, Z. Liu et al., "Inflammatory cytokines-stimulated human muscle stem cells ameliorate ulcerative colitis via the IDO-TSG6 axis," *Stem Cell Research & Therapy*, vol. 12, no. 1, p. 50, 2021.
- [139] R. Contreras-Lopez, R. Elizondo-Vega, N. Luque-Campos et al., "The ATP synthase inhibition induces an AMPK-dependent glycolytic switch of mesenchymal stem cells that enhances their immunotherapeutic potential," *Theranostics*, vol. 11, no. 1, pp. 445–460, 2021.
- [140] R. Jitschin, M. Böttcher, D. Saul et al., "Inflammation-induced glycolytic switch controls suppressivity of mesenchymal stem cells via STAT1 glycosylation," *Leukemia*, vol. 33, no. 7, pp. 1783–1796, 2019.
- [141] R. J. Dang, Y. M. Yang, L. Zhang et al., "A20 plays a critical role in the immunoregulatory function of mesenchymal stem cells," *Journal of Cellular and Molecular Medicine*, vol. 20, no. 8, pp. 1550–1560, 2016.
- [142] P. Luz-Crawford, J. Hernandez, F. Djouad et al., "Mesenchymal stem cell repression of Th17 cells is triggered by mitochondrial transfer," *Stem Cell Research & Therapy*, vol. 10, no. 1, p. 232, 2019.
- [143] R. G. Jones and E. J. Pearce, "MenTORing immunity: mTOR signaling in the development and function of tissue-resident immune cells," *Immunity*, vol. 46, no. 5, pp. 730–742, 2017.
- [144] M. J. Park, S. H. Lee, S. H. Lee et al., "IL-1 Receptor Blockade Alleviates Graft-versus-Host Disease through Downregulation of an Interleukin-1-Dependent Glycolytic Pathway in

- Th17 Cells,” *Mediators of Inflammation*, vol. 2015, Article ID 631384, 12 pages, 2015.
- [145] X. Wang, C. Yang, F. Xu, L. Qi, J. Wang, and P. Yang, “Imbalance of circulating Tfr/Tfh ratio in patients with rheumatoid arthritis,” *Clinical and Experimental Medicine*, vol. 19, no. 1, pp. 55–64, 2019.
- [146] T. Ding, H. Niu, X. Zhao, C. Gao, X. Li, and C. Wang, “T-follicular regulatory cells: potential therapeutic targets in rheumatoid arthritis,” *Frontiers in Immunology*, vol. 10, p. 2709, 2019.
- [147] S. Crotty, “Follicular helper CD4 T cells (TFH),” *Annual Review of Immunology*, vol. 29, no. 1, pp. 621–663, 2011.
- [148] L. S. Ji, X. H. Sun, X. Zhang et al., “Mechanism of follicular helper T cell differentiation regulated by transcription factors,” *Journal of Immunology Research*, vol. 2020, Article ID 1826587, 9 pages, 2020.
- [149] Q. Niu, Z. C. Huang, X. J. Wu et al., “Enhanced IL-6/phosphorylated STAT3 signaling is related to the imbalance of circulating T follicular helper/T follicular regulatory cells in patients with rheumatoid arthritis,” *Arthritis Research & Therapy*, vol. 20, no. 1, p. 200, 2018.
- [150] J. Wei, X. Ouyang, Y. Tang et al., “ER-stressed MSC displayed more effective immunomodulation in RA CD4(+)CXCR5(+)ICOS(+) follicular helper-like T cells through higher PGE2 binding with EP2/EP4,” *Modern Rheumatology*, vol. 30, no. 3, pp. 509–516, 2020.
- [151] H. Zhang, J. Li, L. Li, P. Liu, Y. Wei, and Z. Qian, “Ceramide enhances COX-2 expression and VSMC contractile hyperre-activity via ER stress signal activation,” *Vascular Pharmacology*, vol. 96-98, pp. 26–32, 2017.
- [152] J. Jiang and R. Dingleline, “Role of prostaglandin receptor EP2 in the regulations of cancer cell proliferation, invasion, and inflammation,” *The Journal of Pharmacology and Experimental Therapeutics*, vol. 344, no. 2, pp. 360–367, 2013.
- [153] C. Dong, “T<sub>H</sub>17 cells in development: an updated view of their molecular identity and genetic programming,” *Nature Reviews. Immunology*, vol. 8, no. 5, pp. 337–348, 2008.
- [154] G. H. Tso, H. K. Law, W. Tu, G. C. Chan, and Y. L. Lau, “Phagocytosis of apoptotic cells modulates mesenchymal stem cells osteogenic differentiation to enhance IL-17 and RANKL expression on CD4+ T cells,” *Stem Cells*, vol. 28, no. 5, pp. 939–954, 2010.
- [155] M. Lopez-Santalla, P. Mancheño-Corvo, R. Menta et al., “Human adipose-derived mesenchymal stem cells modulate experimental autoimmune arthritis by modifying early adaptive T cell responses,” *Stem Cells*, vol. 33, no. 12, pp. 3493–3503, 2015.
- [156] A. Xu, Y. Liu, W. Chen et al., “TGF- $\beta$ -induced regulatory T cells directly suppress B cell responses through a noncytotoxic mechanism,” *Journal of Immunology*, vol. 196, no. 9, pp. 3631–3641, 2016.
- [157] D. M. Darlan, D. Munir, A. Putra, and N. K. Jusuf, “MSCs-released TGF $\beta$ 1 generate CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> in T-reg cells of human SLE PBMC,” *Journal of the Formosan Medical Association*, vol. 120, no. 1, pp. 602–608, 2021.
- [158] M. Jeffries, M. Dozmorov, Y. Tang, J. T. Merrill, J. D. Wren, and A. H. Sawalha, “Genome-wide DNA methylation patterns in CD4+ T cells from patients with systemic lupus erythematosus,” *Epigenetics*, vol. 6, no. 5, pp. 593–601, 2011.
- [159] H. Xiong, Z. Guo, Z. Tang et al., “Mesenchymal stem cells activate the MEK/ERK signaling pathway and enhance DNA methylation via DNMT1 in PBMC from systemic lupus erythematosus,” *BioMed Research International*, vol. 2020, 4174082 pages, 2020.
- [160] K. Sunahori, K. Nagpal, C. M. Hedrich, M. Mizui, L. M. Fitzgerald, and G. C. Tsokos, “The catalytic subunit of protein phosphatase 2A (PP2Ac) promotes DNA hypomethylation by suppressing the phosphorylated mitogen-activated protein kinase/extracellular signal-regulated kinase (ERK) kinase (MEK)/phosphorylated ERK/DNMT1 protein pathway in T-cells from controls and systemic lupus erythematosus patients,” *The Journal of Biological Chemistry*, vol. 288, no. 30, pp. 21936–21944, 2013.
- [161] D. Mougiakakos, R. Jitschin, C. C. Johansson, R. Okita, R. Kiessling, and K. le Blanc, “The impact of inflammatory licensing on heme oxygenase-1-mediated induction of regulatory T cells by human mesenchymal stem cells,” *Blood*, vol. 117, no. 18, pp. 4826–4835, 2011.
- [162] D. Wang, S. Huang, X. Yuan et al., “The regulation of the Treg/Th17 balance by mesenchymal stem cells in human systemic lupus erythematosus,” *Cellular & Molecular Immunology*, vol. 14, no. 5, pp. 423–431, 2017.