

Article



Novel Sulfamethoxazole Ureas and Oxalamide as Potential Antimycobacterial Agents

Martin Krátký^{1,*}, Jiřina Stolaříková² and Jarmila Vinšová¹

- ¹ Department of Organic and Bioorganic Chemistry, Faculty of Pharmacy in Hradec Králové, Charles University, Heyrovského 1203, 50005 Hradec Králové, Czech Republic; vinsova@faf.cuni.cz
- ² Laboratory for Mycobacterial Diagnostics and Tuberculosis, Regional Institute of Public Health in Ostrava, Partyzánské náměstí 7, 70200 Ostrava, Czech Republic; Jirina.Stolarikova@zu.cz
- * Correspondence: martin.kratky@faf.cuni.cz; Tel.: +420-495-067-343; Fax: +420-495-067-166

Academic Editor: Claudiu T. Supuran Received: 2 March 2017; Accepted: 24 March 2017; Published: 28 March 2017

Abstract: Infections caused by *Mycobacterium tuberculosis* (*Mtb.*) and nontuberculous mycobacteria (NTM) are considered to be a global health problem; current therapeutic options are limited. Sulfonamides have exhibited a wide range of biological activities including those against mycobacteria. Based on the activity of 4-(3-heptylureido)-*N*-(5-methylisoxazol-3-yl)benzenesulfonamide against NTM, we designed a series of homologous sulfamethoxazole-based *n*-alkyl ureas (C_1 - C_{12}), as well as several related ureas and an oxalamide. Fifteen ureas and one oxalamide were synthesized by five synthetic procedures and characterized. They were screened for their activity against *Mtb.* and three NTM strains (*M. avium, M. kansasii*). All of them share antimycobacterial properties with minimum inhibitory concentration (MIC) values starting from 2 μ M. The highest activity showed 4,4'-[carbonylbis(azanediyl)]bis[*N*-(5-methylisoxazol-3-yl)benzenesulfonamide] with MIC of 2–62.5 μ M (i.e., 1.07–33.28 μ g/mL). Among *n*-alkyl ureas, methyl group is optimal for the inhibition of both *Mtb.* and NTM. Generally, longer alkyls led to increased MIC values, heptyl being an exception for NTM. Some of the novel derivatives are superior to parent sulfamethoxazole. Several urea and oxalamide derivatives are promising antimycobacterial agents with low micromolar MIC values.

Keywords: antimycobacterial activity; in vitro activity; oxalamide; sulfamethoxazole; sulfonamides; tuberculosis; ureas

1. Introduction

The progression of resistance to clinically used drugs among human pathogenic bacteria justifies the development of new antimicrobial agents effective against, e.g., *Mycobacterium tuberculosis* (*Mtb.*) nontuberculous (atypical) mycobacteria (NTM), methicillin-resistant *Staphylococcus aureus*, and polyresistant Gram-negative species [1].

The modification of known drugs represents an established and effective approach in drug design. Sulfonamides have been widely used for the treatment of bacterial and protozoan infections. More recently, sulfonamides were recommended as a part of multidrug regimens for some nontuberculous mycobacterioses [2], but not for tuberculosis. Later, e.g., Ameen and Drancourt [3,4] demonstrated that mycobacterial strains (*Mtb., Mycobacterium avium* complex) are susceptible to clinically-achievable concentrations of sulfonamides (sulfadiazine, sulfamethoxazole, including its combination with trimethoprim) and the investigation of sulfonamide derivatives as potential antimycobacterial agents has been initiated [1,5].

Sulfonamides are well-known inhibitors of dihydropteroate synthases involved in the folate pathway [6], β -carbonic anhydrases [7,8], acetohydroxyacid synthase [9] or β -ketoacyl synthases [10]. Obviously, their mechanism of action against mycobacteria seems to be multiple.

Recently, urea derivatives obtained from sulfamethoxazole (SMX) **1** have been proposed as potential inhibitors of human carbonic anhydrases [11], anticancer [12,13], and antimicrobial agents [14,15]. Importantly, we reported the synthesis and antimicrobial activity of nine sulfamethoxazole-based ureas and corresponding cyclic analogues, imidazolidine-2,4,5-triones. They were active especially against NTM, followed by *Mtb*. and Gram-positive bacteria. Gram-negative species and fungi showed almost a complete resistance. 4-(3-Heptylureido)-*N*-(5-methylisoxazol-3-yl)benzenesulfonamide (Scheme 1; **2g**) was found to be the most effective derivative in vitro with minimum inhibitory concentration (MIC) values against NTM from 4 μ M [1]. Moreover, ureas are largely investigated as potential antimycobacterial agents [16–19].

This unique activity of the 3-heptylureido derivative within the series inspired us to synthesize and evaluate a homologous series of sulfamethoxazole-based monoalkylureas (C_1 – C_{12}) to determine possible relationships between the length of the alkyl and antimycobacterial activity. Additionally, we synthesized several other ureas (Scheme 1) and one analogue of 2,4,5-trioxoimidazolidines, oxalamide, as potential antimycobacterial agents.



Scheme 1. Design of novel antimycobacterial sulfamethoxazole ureas based on 3-heptylurea derivative $(n = 0-11; R^1 = isopropyl, cyclopropyl, 4-[N-(5-methylisoxazol-3-yl)sulfamoyl]phenyl; R^2 = H, isopropyl).$

2. Results and Discussion

2.1. Chemistry

Sulfamethoxazole-based ureas **2** were obtained from parent sulfonamide **1** and isocyanates, predominantly (reflux, 3 h; compounds **2b–2l**). Method A involved commercially available isocyanates; decyl isocyanate was generated in situ from decylamine using triphosgene (Method B; Scheme 2). In general, it provided satisfactory yields of 71–85%. Methylurea **2a** was synthesized by treating of **1** with 2,5-dioxopyrrolidin-1-yl methylcarbamate (*N*-succinimidyl *N*-methylcarbamate) as a methyl isocyanate substitute in the presence of Hunig's base (Method C; yield of 89%, Scheme **3**). The last approach (Method D) consisted of a one-pot reaction of **1** with 1,1′-carbonyldiimidazole (CDI) to form the (1*H*-imidazole-1-yl)urea intermediate that undergoes aminolysis by an excess of an amine added into the reaction mixture (Scheme **4**). In this case, yields are varying (58–95%). Method D was favorable for a secondary amine (diisopropylamine) and for primary amines of which isocyanates were not available for us (cyclopropylamine for **2m**, SMX itself for the synthesis of "bis-SMX-urea" **2n**).

Oxalamide derivative **3** was obtained by treatment of SMX **1** with oxalyl chloride (Scheme 5) in stoichiometric amounts.

All of the compounds (Table 1) were characterized by ¹H-, ¹³C-NMR (nuclear magnetic resonance), and infrared (IR) spectra and melting points. Their purity was checked by thin-layer chromatography (TLC) and elemental analysis.

Based on the previously reported activity of isolated propyl- and heptyl-ureas **2c** and **2g** [1], we completed a homologous series of unbranched alkyl chains from methyl to dodecyl to reveal the dependence of antimycobacterial activity on the alkyl length. To investigate the possible influence of a cycloalkyl and a branched alkyl, cyclopropyl derivative **2m** and isopropyl derivative **2o** were also synthesized. Urea **2n** may be considered as a "double molecule" of SMX **1** connected via a carbonyl bridge or as simplified imidazolidine-2,4,5-trione [1] that has arisen from the removal of a –CO–CO–bridge. Oxalamide **3** was intended as another "open analogue" of imidazolidine-2,4,5-trione designed by the removal of the carbonyl bridge.



Scheme 2. Synthesis of the ureas **2b–2l** (Methods A and B; $Et_3N =$ triethylamine; DCM = dichloromethane; MeCN = acetonitrile; R = C₂ to C₁₂ *n*-alkyl).



Scheme 3. Synthesis of the methyl derivative 2a (Method C; DIPEA = N,N-diisopropylethylamine).



Scheme 4. Synthesis of the urea derivatives 2m-2o (Method D; CDI = 1,1'-carbonyldiimidazole; R^1 = cyclopropyl, isopropyl, 4-[*N*-(5-methylisoxazol-3-yl)sulfamoyl]phenyl; R^2 = H, isopropyl).



Scheme 5. Synthesis of the oxalamide 3.

0	
R _N N H H	2a-m

Table 1. Antimycobacterial activity of sulfamethoxazole-based ureas 2 and oxalamides 3.

Code	R	MIC (µM)										
		Mycobacterium tuberculosis 331/88		Mycobacterium avium 330/88		Mycobacterium kansasii 235/80			Mycobacterium kansasii 6509/96			ClogP
		14 d	21 d	14 d	21 d	7 d	14 d	21 d	7 d	14 d	21 d	_
2a	Methyl	16	32	62.5	125	32	62.5	125	8	16	32	0.48
2b	Ethyl	62.5	62.5	1000	1000	125	250	250	250	500	500	0.82
2c	Propyl ^a	>1000	>1000	1000	1000	500	1000	1000	250	500	500	1.31
2d	Butyl	62.5	62.5	1000	1000	125	250	250	250	500	500	1.72
2e	Pentyl	250	500	500	1000	250	250	500	250	500	500	2.14
2f	Hexyl	250	250 *	250	250	250	250	500	125	250	250	2.56
2g	Heptyl ^a	250	500	32	62.5	4	8	16	4	4	8	2.97
2h	Octyl	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	3.39
2i	Nonyl	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	3.81
2j	Decyl	125	125	125	125	125	125	125	125	125	125	4.23
2k	Undecyl	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	4.64
21	Dodecyl	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	125 *	5.06
2m	Cyclopropyl	2	4	125	125	4	4	8	8	16	16	0.78



			H₃C-√	N O N S N S N O		O N H 3		N-O	CH3			
3	Н	4	4	125 *	125 *	8	8	8	8	8	8	1.23
SMX (1) ^a		32	32	32	62.5	8	16	16	4	4	4	1.5
I	NH	1	1	>250	>250	>250	>250	>250	8	8	8	-0.64

INH: isoniazid; SMX = sulfamethoxazole; ClogP = calculated logP values. Minimum inhibitory concentration (MIC) values were determined after incubation for 14 and 21 days, for *M. kansasii* additionally for seven days. The best MIC values for each strain are shown in bold. Compounds and data ^a were taken from [1]. * At presented concentration, the growth of the strain was observed; at duplex concentration, there was a precipitate present and/or a turbidity; therefore it was not possible to determine exact MIC value.

2.2. Antimycobacterial Activity

All derivatives **2** and **3** together with parent SMX **1** and isoniazid (INH) were evaluated for their in vitro antimycobacterial activity against *Mycobacterium tuberculosis* CNCTC 331/88 (i.e., H₃₇Rv) and three strains of NTM, namely *Mycobacterium avium* CNCTC 330/88, *Mycobacterium kansasii* CNCTC 235/80 and *M. kansasii* 6509/96 (a clinically isolated strain). The minimal inhibitory concentration (MIC) is the lowest concentration of the substance that prevents the growth of *Mycobacterium* species completely.

An escalated lipophilicity of higher homologues abolished their antimycobacterial evaluation. Octyl derivative **2h** (Clog*P* = 3.39), nonyl **2i**, undecyl **2k**, and dodecyl **2l** ureas were soluble in the testing medium only up to the concentration of 125 μ M. The growth of mycobacteria was observed at this concentration but, at higher concentrations, there was a precipitate present and/or turbidity. That is why it was not possible to determine MIC values.

In general, all soluble ureas (2a–2g, 2j, 2m–2o) and oxalamide 3 exhibited MIC values starting from 2 μ M. In this series, *M. avium* showed the highest rate of resistance among mycobacterial strains. In contrast, the susceptibility of *Mtb.* and *M. kansasii* was well marked. MIC values for *M. avium* were within the range of 32–1000 μ M; *N,N*-diisopropylurea 2o was inactive. Heptylurea 2g [1], methylurea 2a, and "bis-SMX-urea" 2n produced an activity comparable to SMX 1; neither of the derivatives 2 and 3 were superior to the parent drug. Three derivatives (cyclopropylurea 2m, "bis-SMX-urea" 2n, and oxalamide 3) exceeded the in vitro activity of SMX against *Mtb.* up to sixteen times (2m and 2n; MIC of 2–4 μ M, i.e., 0.67–1.35 μ g/mL and 1.07 μ g/mL, respectively) and methylurea 2a showed an equal inhibition. Additionally, ethyl 2b, butyl 2d and decyl 2j ureas were superior to original heptylurea 2g. Both strains of *M. kansasii* share a similar susceptibility of 2–1000 μ M, only methylurea 2a exhibited moderately lower MIC values for the clinical isolate. The most active derivatives, 2m, 2n, and 3, share an identical or a slightly superior activity than both parent SMX 1 and heptylurea 2g.

Focusing on structure-activity relationship, the investigation of 3-ureido substitution identified 4-(N-(5-methylisoxazol-3-yl)sulfamoyl)phenyl (2n) as the most convenient substituent. Obviously, the link of two SMX molecules to one molecular entity via a carbonyl bridge led to an advantageous augmentation of the antimycobacterial activity; this "double" sulfonamide is the most active within the series, being comparable (M. avium, M. kansasii 6509/96) or considerably superior to the parent drug. The introduction of 3-cyclopropyl moiety also produced highly active urea **2m**. Evidently, the cyclization of *n*-propyl to cyclopropyl increased the activity sharply (**2c** vs. **2m**). The cyclopropylamino fragment is frequent in the molecules of many (fluoro)quinolones, known antimycobacterial agents, or in antimycobacterials in preclinical development [20,21]. Mycolic acids, essential components of the mycobacterial cell wall, also contain a cyclopropane motif [22]; thus, interference with these biomolecules may contribute to excellent antimycobacterial action. Interestingly, a review of Talele [23] emphasized the unique role of cyclopropyl in the structures of drug candidates. Similar MIC values were obtained by symmetrically disubstituted N^1 , N^2 -oxalamide 3; interestingly, this molecule was inactive against *M. avium*. On the other hand, N^3 , N^3 -disubstitution did not improve antimycobacterial potency. Diisopropylurea 20 was ineffective against *M. avium* and its activity against remaining strains was moderate.

Considering homologous *n*-alkyls, we can distinguish a clear trend for the tuberculous strain 331/88. The best activity against *Mtb*. is conferred by the lowest alkyl methyl (**2a**, 16–32 μ M, i.e., 4.97–9.93 μ g/mL) and MIC values are increasing with the length of the alkyl chain (moderate activity of ethyl **2b** and butyl **2d** ureas of 62.5 μ M). The mildest activity is connected with an intermediate length of alkyl (five-, six-, and seven-membered chains, **2e–2g**, 250–500 μ M). Based on decylurea **2j** (MIC = 125 μ M), it may be expected that the antitubercular properties are improved with a further prolongation of alkyl chain, but limited solubility of such compounds in the testing medium interferes with the evaluation. Previously described propylurea **2c** [1] seems to be an anomaly. The superior activity against NTM displayed methyl (**2a**, 8–125 μ M, 2.48–38.75 μ g/mL) and especially heptyl (**2g**, 4–62.5 μ M, i.e., 1.58–24.66 μ g/mL). For *M. avium*, concomitantly with the increasing length of the chain

from methyl, the activity is decreased dramatically (**2a** vs. **2b–2e**) but after pentyl (**2e**), it is enhanced again to urea **2g**. Decyl derivative **2j** exhibited a moderate MIC of 125 μ M. The drop in the activity caused by the lengthening of the alkyl was not too sharp for both *M. kansasii* strains. *N*³-Substituted ureas by from C₂ to C₆ (**2b–2f**) share a comparable activity.

The conversion of primary amino group of sulfonamide **1** to a urea or a carboxamide led to controversial results. In some cases, antimycobacterial properties are enhanced (cyclopropylurea **2m** for *Mtb.*, *M. kansasii* 235/80; "bis-SMX-urea" **2n** for *M. tuberculosis* and *M. kansasii* 235/80; oxalamide **3** for *Mtb.*), retained (generally methylurea **2a**, heptylurea **2g** in the case of NTM, urea **2n** for the clinically-isolated *M. kansasii*) or even diminished (a majority of derivatives against *M. avium* and *M. kansasii* 6509/96). For several compounds, a sharply increased lipophilicity prevented the exact determination of MIC. In general, the formation of cyclopropylurea **2m**, "double" SMX urea **2n**, oxalamide **3** as well as previously reported heptylurea **2g** [1] were found to be the most successful modifications of the sulfa drug **1**.

The main impetus for this subsequent study was a unique activity of heptylurea **2g** against NTM within previous series of SMX-based ureas [1]. Now, we obtained seven compounds (**2a**, **2b**, **2d**, **2j**, **2m**, **2n**, and **3**) that are more efficacious in vitro against *Mtb*. (up to 250 times). However, only two derivatives (methyl derivative **2a** and "bis-SMX-urea" **2n**) produced a comparable inhibition of *M*. *avium*. Urea **2n** is also superior for *M*. *kansasii* 235/80 and comparable in the case of the strain 6509/96. Cyclopropropylurea **2m** and amide **3** inhibited both strains of *M*. *kansasii* at similar concentration levels.

Drawing a comparison to INH, a first-line oral antimycobacterial drug, "bis-SMX-urea" **2n** showed an equivalent in vitro activity (±1 dilution) against *Mtb*. while the growth of *M. avium* and both strains of *M. kansasii* was suppressed more effectively by **2n**. Four additional derivatives (**2a**, **2f**, **2j**, and **2m**) were also effective against INH-resistant *M. avium*, evidently. Focusing on INH-resistant *M. kansasii* 235/80, all of the soluble derivatives displayed lower MIC values with an exception (propylurea **2c** [1]). Four derivatives (**2g**, **2m**, **2n**, **3**) share an activity against *M. kansasii* isolate 6509/96 comparable or superior to that on INH, followed by methylurea **2a**.

The uniform and broad-spectrum activity of **2** and **3** against INH-resistant strains indicate that these derivatives do not share any cross-resistance with this established antitubercular drug. Additionally, NTM are usually more chemoresistant than *Mtb*. Consequently, cellular targets of the ureas and oxalamide, although not elucidated, seem to be different from INH. INH is highly specific for *Mtb*. and its mechanism of action includes multiple effects on various cellular structures and molecules. After metabolic activation, it inhibits enoyl-acyl carrier protein reductase (InhA) involved in the biosynthesis of mycolic acids [24].

3. Materials and Methods

3.1. Chemistry

3.1.1. General

All of the reagents and solvents were purchased from Sigma-Aldrich (St. Louis, MO, USA) or Penta Chemicals (Prague, Czech Republic), and they were used as received. Melting points were determined on a Büchi Melting Point machine B-545 apparatus (BÜCHI, Flawil, Switzerland) using open capillaries, and the reported values are uncorrected. The progress of the reactions and the purity of the products were monitored by TLC using a mixture of ethyl acetate with *n*-hexane (2:1, v/v). Plates were coated with 0.2 mm Merck 60 F254 silica gel (Merck Millipore, Darmstadt, Germany) and were visualized by UV irradiation (254 nm).

Elemental analysis (C, H, N) was performed on a CHNS-O CE automatic microanalyzer instrument (FISONS EA 1110, Milano, Italy). Infrared spectra (ATR) were recorded on a Nicolet 6700 FTIR spectrometer (ThermoFisher Scientific, Waltham, MA, USA) in the range of 400 to 4000 cm⁻¹. The NMR spectra were measured in DMSO- d_6 at ambient temperature on a Varian VNMR S500 instrument (500 MHz for ¹H and 125 MHz for ¹³C; Varian Comp., Palo Alto, CA, USA) or a Varian

Mercury-Vxbb 300 (300 MHz for ¹H and 75.5 MHz for ¹³C; Varian Comp., Palo Alto, CA, USA). The chemical shifts, δ , are given in ppm, with respect to tetramethylsilane as an internal standard. The coupling constants (*J*) are reported in Hz.

The calculated log*P* values (Clog*P*), which are the logarithms of the partition coefficients for octan-1-ol/water, were determined using the program CS ChemOffice Ultra version 15.0 (CambridgeSoft, Cambridge, MA, USA).

3.1.2. Synthesis

Synthesis of Urea Derivatives 2

Method A

Sulfamethoxazole 1 (1 mmol) was dissolved in dry acetonitrile (MeCN, 6 mL) and appropriate isocyanate (1.1 mmol) was added in one portion. The solution was heated under reflux for 3 h and then stirred at ambient temperature overnight. The resulting crystals were filtered, washed with a small volume of MeCN and dried. The product was recrystallized from ethyl acetate (EtOAc) if necessary [1].

Method B

Triphosgene (0.4 mmol) was dissolved in anhydrous dichloromethane (DCM; 5 mL) under argon atmosphere and decylamine (1.01 mmol) in dry DCM (3 mL) was added dropwise. This mixture was stirred for 30 min at room temperature and then treated with triethylamine (2.1 mmol) in dry DCM (3 mL). After 30 min, a solution of sulfamethoxazole **1** (1.01 mmol) in 5 mL of dry MeCN was added. The reaction mixture was stirred for 12 h at room temperature. Then 1 mL of glacial acetic acid was added and the reaction mixture was stirred for additional 2 h. The mixture was evaporated to dryness. After addition of 10 mL of MeCN, the suspension was let to stay overnight. The resulting crystals were filtered, washed with a small volume of diethylether and dried to obtain the final product.

Method C

Sulfamethoxazole **1** (1 mmol) was dissolved in dry acetonitrile (5 mL), then *N*,*N*-diisopropylethylamine (2 mmol) followed by 2,5-dioxopyrrolidin-1-yl methylcarbamate (1.5 mmol) was added. The solution was stirred for 24 h. The resulting crystals were filtered, washed with a small volume of MeCN and dried. The product was recrystallized from EtOAc if necessary.

Method D

Sulfamethoxazole **1** (1 mmol) was dissolved in dry acetonitrile (6 mL) and 1,1'-carbonyldiimidazole (1.2 mmol) was added. The mixture was refluxed for 12 h and then treated with amine (1.2 mmol; 3 h refluxing, then overnight at room temperature). The resulting crystals were filtered, washed with a small amount of MeCN and dried to obtain the final product.

N-(5-*Methylisoxazol-3-yl*)-4-(3-*methylureido*)*benzenesulfonamide* (**2a**) [25]. White solid; yield 89% (Method C); m.p. 242.5–244 °C. IR: 3375 (N–H), 1662 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.22 (1H, s, SO₂NH), 9.04 (1H, s, N¹–H ureido), 7.70–7.66 (2H, m, H2, H6), 7.57–7.54 (2H, m, H3, H5), 6.20 (1H, q, J = 4.5 Hz, N³-H ureido), 6.11 (1H, d, J = 1.0 Hz, isoxazole), 2.63 (3H, d, J = 4.6 Hz, CH₃), 2.28 (3H, d, J = 1.0 Hz, CH₃ isoxazole). ¹³C-NMR (126 MHz, DMSO): δ 170.36, 157.88, 155.44, 145.38, 130.70, 128.25, 117.16, 95.53, 26.44, 12.26. Anal. Calcd. for C₁₂H₁₄N₄O₄S (310.33): C, 46.45; H, 4.55; N, 18.05. Found: C, 46.54; H, 4.88; N, 17.99.

4-(3-*Ethylureido*)-*N*-(5-*methylisoxazol*-3-*yl*)*benzenesulfonamide* (**2b**). White solid; yield 80% (Method A); m.p. 240–241.5 °C. IR: 3374 (N–H), 1684 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.19 (1H, s, SO₂NH), 8.93 (1H, s, N¹–H ureido), 7.70–7.66 (2H, m, H2, H6), 7.57–7.53 (2H, m, H3, H5), 6.29 (1H, t, *J* = 5.6 Hz, N³–H ureido), 6.10 (1H, d, *J* = 0.9 Hz, isoxazole), 3.09 (2H, td, *J* = 7.2 Hz, *J* = 5.6 Hz, CH₂), 2.28 (3H, d, J = 1.0 Hz, CH₃ isoxazole), 1.03 (3H, t, J = 7.1 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.32, 157.84, 154.72, 145.32, 130.72, 128.24, 117.15, 95.52, 34.19, 15.45, 12.22. Anal. Calcd. for C₁₃H₁₆N₄O₄S (324.36): C, 48.14; H, 4.97; N, 17.27. Found: C, 48.29; H, 4.87; N, 17.44.

N-(5-*Methylisoxazol*-3-*yl*)-4-(3-*propylureido*)*benzenesulfonamide* (2c). Compound 2c was synthesized following a procedure described by Krátký et al. [1].

4-(3-Butylureido)-N-(5-methylisoxazol-3-yl)benzenesulfonamide (2d). White solid; yield 73% (Method A); m.p. 216–218 °C. IR: 3369 (N–H), 1683 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.19 (1H, s, SO₂NH), 8.90 (1H, s, N¹–H ureido), 7.70–7.66 (2H, m, H2, H6), 7.56–7.51 (2H, m, H3, H5), 6.30 (1H, t, *J* = 5.7 Hz, N³–H ureido), 6.10 (1H, d, *J* = 1.0 Hz, isoxazole), 3.07 (2H, td, *J* = 6.9 Hz, *J* = 5.6 Hz, C¹H₂), 2.28 (3H, d, *J* = 1.0 Hz, CH₃ isoxazole), 1.43–1.36 (2H, m, C²H₂), 1.33–1.24 (2H, m, C³H₂), 0.87 (3H, t, *J* = 7.3 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.31, 157.85, 154.79, 145.31, 130.71, 128.24, 117.11, 95.52, 38.89, 31.86, 19.65, 13.83, 12.22. Anal. Calcd. for C₁₅H₂₀N₄O₄S (352.41): C, 51.12; H, 5.72; N, 15.90. Found: C, 51.29; H, 5.90; N, 15.74.

N-(5-*Methylisoxazol-3-yl*)-4-(3-*pentylureido*)*benzenesulfonamide* (**2e**). White solid; yield 71% (Method A); m.p. 213.5–214.5 °C. IR: 3370 (N–H), 1682 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.19 (1H, s, SO₂NH), 8.90 (1H, s, N¹–H ureido), 7.70–7.66 (2H, m, H2, H6), 7.56–7.52 (2H, m, H3, H5), 6.30 (1H, t, J = 5.7 Hz, N³-H ureido), 6.10 (1H, d, J = 1.0 Hz, isoxazole), 3.06 (2H, td, J = 6.9 Hz, J = 5.6 Hz, C¹H₂), 2.28 (3H, d, J = 0.9 Hz, CH₃ isoxazole), 1.45–1.37 (2H, m, C²H₂), 1.32–1.20 (4H, m, C³H₂, C⁴H₂), 0.86 (3H, t, J = 7.0 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.31, 157.83, 154.77, 145.30, 130.69, 128.24, 117.10, 95.51, 39.19, 29.41, 28.71, 22.02, 14.10, 12.21. Anal. Calcd. for C₁₆H₂₂N₄O₄S (366.44): C, 52.44; H, 6.05; N, 15.29. Found: C, 52.32; H, 5.92; N, 14.99.

4-(3-Hexylureido)-N-(5-methylisoxazol-3-yl)benzenesulfonamide (**2f**). White solid; yield 82% (Method A); m.p. 218.5–220 °C. IR: 3373 (N–H), 1683 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.08 (1H, s, SO₂NH), 8.80 (1H, s, N¹–H ureido), 7.59–7.56 (2H, m, H2, H6), 7.46–7.42 (2H, m, H3, H5), 6.20 (1H, t, *J* = 5.7 Hz, N³–H ureido), 6.00 (1H, d, *J* = 0.9 Hz, isoxazole), 2.96 (2H, td, *J* = 6.9 Hz, *J* = 5.6 Hz, C¹H₂), 2.18 (3H, d, *J* = 0.9 Hz, CH₃ isoxazole), 1.35–1.26 (2H, m, C²H₂), 1.32–1.20 (6H, m, C³H₂, C⁴H₂, C⁵H₂), 0.74 (3H, t, *J* = 7.1 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.30, 157.83, 154.77, 145.30, 130.70, 128.23, 117.10, 95.51, 39.19, 31.16, 29.70, 26.17, 22.24, 14.08, 12.21. Anal. Calcd. for C₁₇H₂₄N₄O₄S (380.46): C, 53.67; H, 6.36; N, 14.73. Found: C, 54.00; H, 6.58; N, 14.86.

4-(3-*Heptylureido*)-*N*-(5-*methylisoxazol*-3-*yl*)*benzenesulfonamide* (**2g**). Compound **2g** was synthesized following a procedure described by Krátký et al. [1].

N-(5-*Methylisoxazol*-3-*yl*)-4-(3-octylureido)benzenesulfonamide (**2h**). White solid; yield 83% (Method A); m.p. 214–216 °C. IR: 3368 (N–H), 1677 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.18 (1H, s, SO₂NH), 8.89 (1H, s, N¹–H ureido), 7.69–7.65 (2H, m, H2, H6), 7.56–7.52 (2H, m, H3, H5), 6.30 (1H, t, *J* = 5.7 Hz, N³-H ureido), 6.10 (1H, d, *J* = 1.1 Hz, isoxazole), 3.06 (2H, td, *J* = 6.8 Hz, *J* = 5.6 Hz, C¹H₂), 2.28 (3H, d, *J* = 1.0 Hz, CH₃ isoxazole), 1.44–1.36 (2H, m, C²H₂), 1.32–1.20 (10H, m, C³H₂, C⁴H₂, C⁵H₂, C⁶H₂, C⁷H₂), 0.84 (3H, t, *J* = 6.9 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.28, 157.85, 154.76, 145.28, 130.71, 128.22, 117.09, 95.50, 39.20, 31.40, 29.72, 28.88, 28.84, 26.50, 22.24, 14.11, 12.21. Anal. Calcd. for C₁₉H₂₈N₄O₄S (408.52): C, 55.86; H, 6.91; N, 13.71. Found: C, 56.02; H, 6.92; N, 14.02.

N-(5-*Methylisoxazol*-3-*yl*)-4-(3-*nonylureido*)*benzenesulfonamide* (**2i**). White solid; yield 84% (Method A); m.p. 211–212.5 °C. IR: 3382 (N–H), 1679 (C=O) cm⁻¹. ¹H-NMR (300 MHz, DMSO): δ 11.20 (1H, s, SO₂NH), 8.91 (1H, s, N¹–H ureido), 7.70–7.65 (2H, m, H2, H6), 7.57–7.51 (2H, m, H3, H5), 6.30 (1H, t, J = 5.7 Hz, N³–H ureido), 6.10 (1H, d, J = 0.9 Hz, isoxazole), 3.05 (2H, q, J = 6.5 Hz, C¹H₂), 2.28 (3H, d, J = 0.9 Hz, CH₃ isoxazole), 1.44–1.36 (2H, m, C²H₂), 1.30–1.18 (12H, m, C³H₂, C⁴H₂, C⁵H₂, C⁶H₂, C⁷H₂, C⁸H₂), 0.84 (3H, t, J = 6.9 Hz, CH₃ alkyl). ¹³C-NMR (75 MHz, DMSO): δ 170.35, 157.88, 154.81, 145.35, 130.70, 128.28, 117.13, 95.53, 38.88, 31.49, 29.77, 29.20, 28.97, 28.86, 26.53, 22.30, 14.15, 12.24. Anal. Calcd. for C₂₀H₃₀N₄O₄S (422.54): C, 56.85; H, 7.16; N, 13.26. Found: C, 56.97; H, 6.95; N, 13.40.

10 of 13

4-(3-Decylureido)-N-(5-methylisoxazol-3-yl)benzenesulfonamide (**2j**). White solid; yield 85% (Method B); m.p. 212–213.5 °C. IR: 3377 (N–H), 1682 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.19 (1H, s, SO₂NH), 8.90 (1H, s, N¹–H ureido), 7.70–7.66 (2H, m, H2, H6), 7.56–7.52 (2H, m, H3, H5), 6.30 (1H, t, *J* = 5.7 Hz, N³–H ureido), 6.10 (1H, d, *J* = 1.0 Hz, isoxazole), 3.06 (2H, td, *J* = 6.9 Hz, *J* = 5.6 Hz, C¹H₂), 2.28 (3H, d, *J* = 0.8 Hz, CH₃ isoxazole), 1.44–1.37 (2H, m, C²H₂), 1.29–1.17 (14H, m, C³H₂, C⁴H₂, C⁵H₂, C⁶H₂, C⁷H₂, C⁸H₂, C⁹H₂), 0.84 (3H, t, *J* = 7.0 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.30, 157.85, 154.79, 145.31, 130.73, 128.24, 117.11, 95.51, 39.21, 31.47, 29.73, 29.20, 29.14, 28.93, 28.87, 26.50, 22.27, 14.12, 12.22. Anal. Calcd. for C₂₁H₃₂N₄O₄S (436.57): C, 57.77; H, 7.39; N, 12.83. Found: C, 55.65; H, 7.52; N, 12.84.

N-(5-*Methylisoxazol*-3-*yl*)-4-(3-*undecylureido*)*benzenesulfonamide* (**2k**). White solid; yield 79% (Method A); m.p. 218.5–220 °C. IR: 3374 (N–H), 1680 (C=O) cm⁻¹. ¹H-NMR (300 MHz, DMSO): δ 11.20 (1H, s, SO₂NH), 8.90 (1H, s, N¹–H ureido), 7.70–7.65 (2H, m, H2, H6), 7.56–7.51 (2H, m, H3, H5), 6.30 (1H, t, J = 5.6 Hz, N³–H ureido), 6.10 (1H, d, J = 1.0 Hz, isoxazole), 3.05 (2H, q, J = 6.5 Hz, C¹H₂), 2.28 (3H, d, J = 0.9 Hz, CH₃ isoxazole), 1.45–1.35 (2H, m, C²H₂), 1.29–1.18 (16H, m, C³H₂, C⁴H₂, C⁵H₂, C⁶H₂, C⁷H₂, C⁸H₂, C⁹H₂, C¹⁰H₂), 0.84 (3H, t, J = 6.7 Hz, CH₃ alkyl). ¹³C-NMR (75 MHz, DMSO): δ 170.31, 157.85, 154.77, 145.31, 130.67, 128.25, 117.09, 95.51, 39.21, 31.48, 29.74, 29.22, 29.20, 29.18, 28.94, 28.90, 26.51, 22.28, 14.14, 12.23. Anal. Calcd. for C₂₂H₃₄N₄O₄S (450.59): C, 58.64; H, 7.61; N, 12.43. Found: C, 58.45; H, 7.50; N, 12.51.

4-(3-Dodecylureido)-N-(5-methylisoxazol-3-yl)benzenesulfonamide (**2l**). White solid; yield 82% (Method A); m.p. 211.5–213 °C. IR: 3377 (N–H), 1683 (C=O) cm⁻¹. ¹H-NMR (500 MHz, DMSO): δ 11.18 (1H, s, SO₂NH), 8.89 (1H, s, N¹–H ureido), 7.69–7.65 (2H, m, H2, H6), 7.56–7.52 (2H, m, H3, H5), 6.29 (1H, t, *J* = 5.7 Hz, N³–H ureido), 6.10 (1H, s, isoxazole), 3.05 (2H, q, *J* = 6.6 Hz, C¹H₂), 2.28 (3H, s, CH₃ isoxazole), 1.44–1.36 (2H, m, C²H₂), 1.30–1.15 (18H, m, C³H₂, C⁴H₂, C⁵H₂, C⁶H₂, C⁷H₂, C⁸H₂, C⁹H₂, C¹⁰H₂, C¹¹H₂), 0.83 (3H, t, *J* = 6.9 Hz, CH₃ alkyl). ¹³C-NMR (126 MHz, DMSO): δ 170.27, 157.82, 154.75, 145.29, 130.69, 128.22, 117.08, 95.49, 39.20, 31.46, 29.72, 29.21, 29.18, 29.17, 29.16, 28.91, 28.87, 26.49, 22.26, 14.10, 12.20. Anal. Calcd. for C₂₃H₃₆N₄O₄S (464.62): C, 59.46; H, 7.81; N, 12.06. Found: C, 59.58; H, 7.63; N, 11.91.

4-(3-*Cyclopropylureido*)-*N*-(5-*methylisoxazol*-3-*yl*)*benzenesulfonamide* (**2m**). White solid; yield 95% (Method D); m.p. 144–146 °C. IR: 3352 (N–H), 1662 (C=O). ¹H-NMR (500 MHz, DMSO): δ 11.19 (1H, s, SO₂NH), 9.05 (1H, s, N¹–H ureido), 7.66–7.59 (2H, m, H2, H6), 7.50–7.46 (2H, m, H3, H5), 6.28 (1H, d, *J* = 5.7 Hz, N³–H ureido), 6.10 (1H, d, *J* = 1.0 Hz, isoxazole), 2.53–2.44 (1H, m, CH), 2.26 (3H, d, *J* = 0.9 Hz, CH₃ isoxazole), 0.65–0.55 (2H, m, CH₂CH₂), 0.43–0.33 (2H, m, CH₂CH₂). ¹³C-NMR (126 MHz, DMSO): δ 167.31, 163.80, 152.30, 141.71, 138.17, 127.31, 117.61, 96.55, 22.79, 12.34, 4.36. Anal. Calcd. for C₁₄H₁₆N₄O₄S (336.37): C, 49.99; H, 4.79; N, 16.66. Found: C, 50.14; H, 4.90; N, 16.58.

4,4'-[*Carbonylbis(azanediyl)*]*bis*[*N*-(5-*methylisoxazol-3-yl)benzenesulfonamide*] (**2n**) [26]. White solid; yield 58% (Method D); m.p. 262–263.5 °C. IR: 3333 (N–H), 1659 (C=O). ¹H-NMR (500 MHz, DMSO): δ 11.28 (2H, s, SO₂NH), 9.30 (2H, s, NH ureido), 7.79–7.75 (4H, m, H2, H6), 7.66–7.62 (4H, m, H3, H5), 6.12 (2H, d, J = 1.1 Hz, isoxazole), 2.29 (6H, d, J = 1.0 Hz, CH₃). ¹³C-NMR (126 MHz, DMSO): δ 170.39, 157.75, 152.01, 143.92, 132.24, 128.31, 118.19, 95.54, 12.22. Anal. Calcd. for C₂₁H₂₀N₆O₇S₂ (532.55): C, 47.36; H, 3.79; N, 15.78. Found: C, 47.48; H, 3.91; N, 15.66.

4-(3,3-Diisopropylureido)-N-(5-methylisoxazol-3-yl)benzenesulfonamide (**20**). White solid; yield 62% (Method D); m.p. 234.5–236.5 °C. IR: 3361 (N–H), 1613 (C=O). ¹H-NMR (300 MHz, DMSO): δ 11.20 (1H, s, SO₂NH), 9.25 (1H, s, NH ureido), 7.68–7.61 (2H, m, H2, H6), 7.54–7.48 (2H, m, H3, H5), 5.92 (1H, d, *J* = 1.1 Hz, isoxazole), 3.34 (2H, sep, *J* = 6.4 Hz, CH), 2.18 (3H, d, *J* = 0.9 Hz, CH₃ isoxazole), 1.20 (12H, d, *J* = 6.4 Hz, isopropyl CH₃). ¹³C-NMR (75 MHz, DMSO): δ 167.94, 162.68, 152.29, 142.19, 137.10, 127.52, 117.74, 96.42, 46.46, 19.04, 12.37. Anal. Calcd. for C₁₇H₂₄N₄O₄S (380.46): C, 53.67; H, 6.36; N, 14.73. Found: C, 53.43; H, 6.54; N, 15.00.

Synthesis of Oxalamide 3

Sulfamethoxazole 1 (1 mmol) was dissolved in MeCN (6 mL) and oxalyl chloride (0.5 mmol) was added in one portion. The mixture was stirred for 2 h at room temperature. The resulting crystals were filtered, washed with a small volume of MeCN and dried. The product was recrystallized from tetrahydrofuran/hexane.

 N^1 , N^2 -*Bis*{4-[N-(5-*methylisoxazol-3-yl)sulfamoyl]phenyl}oxalamide* (**3**). White solid; yield 84%; m.p. > 300 °C (decomp.). IR: 3344 (N–H), 1690 (C=O). ¹H-NMR (300 MHz, DMSO): δ 11.42 (2H, s, CONH), 11.26 (2H, s, SO₂NH), 8.07–8.01 (4H, m, H2, H6), 7.89–7.82 (4H, m, H3, H5), 6.13 (2H, d, *J* = 1.0 Hz, isoxazole), 2.28 (6H, d, *J* = 1.1 Hz, CH₃). ¹³C-NMR (75 MHz, DMSO): δ 170.54, 158.88, 157.69, 142.04, 134.93, 128.08, 120.82, 95.61, 12.28. Anal. Calcd. for C₂₂H₂₀N₆O₈S₂ (560.56): C, 47.14; H, 3.60; N, 14.99. Found: C, 47.40; H, 3.51; N, 15.11.

3.2. Antimycobacterial Activity

The antimycobacterial activity against *Mycobacterium tuberculosis* 331/88 (H_{37} Rv; dilution of this strain was 10^{-3}), *Mycobacterium avium* 330/88 (dilution of 10^{-5}), and two strains of *Mycobacterium kansasii*, namely 235/80 (dilution of 10^{-4}) and the clinically-isolated strain 6509/96 (dilution of 10^{-5}), was evaluated using a previously described method [27]. The following concentrations were used: 1000, 500, 250, 125, 62.5, 32, 16, 8, 4, 2, and 1 μ M. MIC is the lowest concentration at which complete inhibition of mycobacterial growth was observed. The MICs were determined after incubation at 37 °C for 14 and 21 days and, for *M. kansasii*, additionally for 7 days. INH and parent sulfamethoxazole **1** were chosen as the reference drugs. The experiments were prepared in quadruplicates and the determination was repeated twice.

4. Conclusions

In this work, fifteen sulfamethoxazole-based ureas and one oxalamide were synthesized using various methods, characterized, and evaluated for their antimycobacterial activity. All of these compounds inhibited the growth of both *M. tuberculosis* and nontuberculous mycobacteria in vitro with MIC values starting from 2 μ M. Several derivatives exhibited an antimycobacterial activity comparable or superior to sulfamethoxazole and isoniazid, thus, being potentially promising for further investigation. Regarding structure-activity relationships, methyl, cyclopropyl and 4-(*N*-(5-methylisoxazol-3-yl)sulfamoyl)phenyl are favored as the 3-ureido substituents of sulfamethoxazole-based urea.

Acknowledgments: This work was supported by the Czech Science Foundation project No. 17-27514Y. We would like to thank to Jaroslava Urbanová, M.A., for the language help provided and to the staff of the Department of Organic and Bioorganic Chemistry, Faculty of Pharmacy in Hradec Králové, for the technical assistance.

Author Contributions: Martin Krátký designed experiments and synthesized all the presented compounds. Jiřina Stolaříková conducted the antimycobacterial activity determination. Martin Krátký and Jarmila Vinšová wrote the article, and made the discussion and conclusions.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Krátký, M.; Mandíková, J.; Trejtnar, F.; Buchta, V.; Stolaříková, J.; Vinšová, J. Synthesis and antimicrobial activity of sulphamethoxazole-based ureas and imidazolidin-2,4,5-triones. *Chem. Pap.* 2015, 69, 1108–1117. [CrossRef]
- Brown-Elliott, B.A.; Nash, K.A.; Wallace, R.J. Antimicrobial Susceptibility Testing, Drug Resistance Mechanisms, and Therapy of Infections with Nontuberculous Mycobacteria. *Clin. Microbiol. Rev.* 2012, 25, 545–582. [CrossRef] [PubMed]
- 3. Ameen, S.M.; Drancourt, M. In Vitro Susceptibility of *Mycobacterium tuberculosis* to Trimethoprim and Sulfonamides in France. *Antimicrob. Agents Chemother.* **2013**, *57*, 6370–6371. [CrossRef] [PubMed]

- 4. Ameen, S.M.; Drancourt, M. In vitro susceptibility of *Mycobacterium avium* complex mycobacteria to trimethoprim and sulfonamides. *Int. J. Antimicrob. Agents* **2013**, *42*, 281–288. [CrossRef] [PubMed]
- Krátký, M.; Vinšová, J.; Volková, M.; Buchta, V.; Trejtnar, F.; Stolaříková, J. Antimicrobial activity of sulfonamides containing 5-chloro-2-hydroxybenzaldehyde and 5-chloro-2-hydroxybenzoic acid scaffold. *Eur. J. Med. Chem.* 2012, *50*, 433–440. [CrossRef] [PubMed]
- 6. Chio, L.C.; Bolyard, L.A.; Nasr, M.; Queener, S.F. Identification of a Class of Sulfonamides Highly Active against Dihydropteroate Synthase from *Toxoplasma gondii*, *Pneumocystis carinii*, and *Mycobacterium avium*. *Antimicrob. Agents Chemother.* **1996**, *40*, 727–733. [PubMed]
- Ceruso, M.; Vullo, D.; Scozzafava, A.; Supuran, C.T. Sulfonamides incorporating fluorine and 1,3,5-triazine moieties are effective inhibitors of three β-class carbonic anhydrases from *Mycobacterium tuberculosis*. *J. Enzyme Inhib. Med. Chem.* 2014, 29, 686–689. [CrossRef] [PubMed]
- Maresca, A.; Scozzafava, A.; Vullo, D.; Supuran, C.T. Dihalogenated sulfanilamides and benzolamides are effective inhibitors of the three β-class carbonic anhydrases from *Mycobacterium tuberculosis*. *J. Enzyme Inhib. Med. Chem.* 2013, 28, 384–387. [CrossRef] [PubMed]
- 9. Patil, V.; Kale, M.; Raichurkar, A.; Bhaskar, B.; Prahlad, D.; Balganesh, M.; Nandan, S.; Hameed, P.S. Design and synthesis of triazolopyrimidine acylsulfonamides as novel anti-mycobacterial leads acting through inhibition of acetohydroxyacid synthase. *Bioorg. Med. Chem. Lett.* **2014**, 24, 2222–2225. [CrossRef] [PubMed]
- Abrahams, K.A.; Chung, C.W.; Ghidelli-Disse, S.; Rullas, J.; Rebollo-López, M.J.; Gurcha, S.S.; Cox, J.A.G.; Mendoza, A.; Jiménez-Navarro, E.; Martínez-Martínez, M.S.; et al. Identification of KasA as the cellular target of an anti-tubercular scaffold. *Nat. Commun.* 2016, *7*, 12581. [CrossRef] [PubMed]
- Mishra, C.B.; Kumari, S.; Angeli, A.; Monti, S.M.; Buonanno, M.; Prakash, A.; Tiwari, M.; Supuran, C.T. Design, synthesis and biological evaluation of *N*-(5-methyl-isoxazol-3-yl/1,3,4-thiadiazol-2-yl)-4-(3substitutedphenylureido) benzenesulfonamides as human carbonic anhydrase isoenzymes I, II, VII and XII inhibitors. *J. Enzyme Inhib. Med. Chem.* 2016, *31*, 174–179. [CrossRef] [PubMed]
- Luzina, E.L.; Popov, A.V. Synthesis, evaluation of anticancer activity and COMPARE analysis of *N*-bis(trifluoromethyl)alkyl-*N*'-substituted ureas with pharmacophoric moieties. *Eur. J. Med. Chem.* 2012, 53, 364–373. [CrossRef] [PubMed]
- Luzina, E.L.; Popov, A.V. Synthesis of 3,3,3-trifluoroethyl isocyanate, carbamate and ureas. Anticancer activity evaluation of *N*-(3,3,3-trifluoroethyl)-*N*'-substituted ureas. *J. Fluorine Chem.* 2015, 176, 82–88. [CrossRef] [PubMed]
- 14. Sunduru, N.; Salin, O.; Gylfe, A.; Elofsson, M. Design, synthesis and evaluation of novel polypharmacological antichlamydial agents. *Eur. J. Med. Chem.* **2015**, *101*, 595–603. [CrossRef] [PubMed]
- 15. Aly, M.R.E.; Gobouri, A.A.; Hafez, S.H.A.; Saad, H.A. Synthesis, Reactions, and Biological Activity of Some Triazine Derivatives Containing Sulfa Drug Moieties. *Russ. J. Bioorg. Chem.* **2015**, *41*, 437–450. [CrossRef]
- Brown, J.R.; North, E.J.; Hurdle, J.G.; Morisseau, C.; Scarborough, J.S.; Sun, D.; Kordulakova, J.; Scherman, M.S.; Jones, V.; Grzegorzewicz, A.; et al. The structure-activity relationship of urea derivatives as anti-tuberculosis agents. *Bioorg. Med. Chem.* 2011, *19*, 5585–5595. [CrossRef] [PubMed]
- 17. Medapi, B.; Renuka, J.; Saxena, S.; Sridevi, J.P.; Medishetti, R.; Kulkarni, P.; Yogeeswari, P.; Sriram, D. Design and synthesis of novel quinoline–aminopiperidine hybrid analogues as *Mycobacterium tuberculosis* DNA gyraseB inhibitors. *Bioorg. Med. Chem.* **2015**, *23*, 2062–2078. [CrossRef] [PubMed]
- Madaiah, M.; Prashanth, M.K.; Revanasiddappa, H.D.; Veeresh, B. Synthesis and evaluation of novel imidazo[4,5-c]pyridine derivatives as antimycobacterial agents against *Mycobacterium tuberculosis*. *New J. Chem.* 2016, 40, 9194–9204. [CrossRef]
- 19. Brunner, K.; Maric, S.; Reshma, R.S.; Almqvist, H.; Seashore-Ludlow, B.; Gustavsson, A.L.; Poyraz, O.; Yogeeswari, P.; Lundback, T.; Vallin, M.; et al. Inhibitors of the Cysteine Synthase CysM with Antibacterial Potency against Dormant *Mycobacterium tuberculosis. J. Med. Chem.* **2016**, *59*, 6848–6859. [CrossRef] [PubMed]
- Dömling, A.; Achatz, S.; Beck, B. Novel anti-tuberculosis agents from MCR libraries. *Bioorg. Med. Chem. Lett.* 2007, 17, 5483–5486. [CrossRef] [PubMed]
- Chopra, S.; Koolpe, G.A.; Tambo-ong, A.A.; Matsuyama, K.N.; Ryan, K.J.; Tran, T.B.; Doppalapudi, R.S.; Riccio, E.S.; Iyer, L.V.; Green, C.E.; et al. Discovery and Optimization of Benzotriazine Di-N-Oxides Targeting Replicating and Nonreplicating *Mycobacterium tuberculosis*. J. Med. Chem. 2012, 55, 6047–6060. [CrossRef] [PubMed]

- 22. Marrakchi, H.; Lanéelle, M.A.; Daffé, M. Mycolic Acids: Structures, Biosynthesis, and Beyond. *Chem. Biol.* **2014**, *21*, 67–85. [CrossRef] [PubMed]
- 23. Talele, T.T. The "Cyclopropyl Fragment" is a Versatile Player that Frequently Appears in Preclinical/Clinical Drug Molecules. *J. Med. Chem.* **2016**, *59*, 8712–8756. [CrossRef] [PubMed]
- Rychtarčíková, Z.; Krátký, M.; Gazvoda, M.; Komlóová, M.; Polanc, S.; Kočevar, M.; Stolaříková, J.; Vinšová, J. N-Substituted 2-Isonicotinoylhydrazinecarboxamides—New Antimycobacterial Active Molecules. *Molecules* 2014, 19, 3851–3868. [CrossRef] [PubMed]
- 25. Chiang, H.C.; Yao, K.M.; Huang, K.F. Synthesis of nitrosourea analogs of some sulfa drugs. *Proc. Natl. Sci. Counc. Repub. China B* **1984**, *8*, 18–22.
- 26. Gluncic, B.; Junasevic-Holjevac, A.; Grguric, D.; Gustak, E. Note on the preparation on some disubstituted derivatives of p'p'-bis(sulfamoyl)carbanilides and their hydrolysis to corresponding sulphonamides. *Croat. Chem. Acta* **1965**, *37*, 111–114.
- 27. Krátký, M.; Vinšová, J.; Novotná, E.; Mandíková, J.; Trejtnar, F.; Stolaříková, J. Antibacterial Activity of Salicylanilide 4-(Trifluoromethyl)benzoates. *Molecules* **2013**, *18*, 3674–3688. [CrossRef] [PubMed]

Sample Availability: Samples of the compounds 2a–2o and 3 are available from the authors.



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).