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Kinetics of arterial carbon dioxide during veno-venous extracorporeal membrane oxygenation support in an apnoeic porcine model

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Abstract

Background: Extracorporeal membrane oxygenation (ECMO) is a technique widely used worldwide to improve gas exchange. Changes in ECMO settings affect both oxygen and carbon dioxide. The impact on oxygenation can be followed closely by continuous pulse oximeter. Conversely, carbon dioxide equilibrates much slower and is not usually monitored directly.

Methods: We investigated the time to stabilization of arterial carbon dioxide partial pressure (PaCO₂) following step changes in ECMO settings in 5 apnoeic porcine models under veno-venous ECMO support with polymethylpentene membranes. We collected sequential arterial blood gases at a pre-specified interval of 50 min using a sequence of standardized blood and sweep gas flow combinations.

Results: Following the changes in ECMO parameters, the kinetics of carbon dioxide was dependent on sweep gas and ECMO blood flow. With a blood flow of 1500 mL/min, PaCO₂ takes longer than 50 min to equilibrate following the changes in sweep gas flow. Furthermore, the sweep gas flow from 3.0 to 10.0 L/min did not significantly affect PaCO₂. However, with a blood flow of 3500 mL/min, 50 min was enough for PaCO₂ to reach the equilibrium and every increment of sweep gas flow (up to 10.0 L/min) resulted in additional reductions of PaCO₂.

Conclusions: Fifty minutes was enough to reach the equilibrium of PaCO₂ after ECMO initiation or after changes in blood and sweep gas flow with an ECMO blood flow of 3500 ml/min. Longer periods may be necessary with lower ECMO blood flows.

Keywords: Acute respiratory distress syndrome, Mechanical ventilation, Swine and extracorporeal membrane oxygenation

Background

Extracorporeal membrane oxygenation (ECMO) has been successfully used to support severely hypoxemic patients [1–7]. Despite the worldwide increase in ECMO support use, there are few studies exploring the physiology of polymethylpentene membranes in the veno-venous configuration [8, 9].

After initiation of extracorporeal support, blood flow and sweep gas flow are adjusted to achieve target values of arterial oxygen (PaO₂), pH, and carbon dioxide (PaCO₂) partial pressures. Titrating these ECMO settings can involve multiple steps because, frequently, the initially chosen parameters will not produce the desired result in terms of blood gases, leading to at least one more cycle of parameter change and blood gas analysis. The whole process has to be repeated whenever the patient condition changes. As a consequence of its lower solubility and lower volume of distribution throughout multiple compartments, PaO₂ reaches equilibrium much sooner than PaCO₂ [10]. Therefore, in order to optimize the frequency of changes in ECMO settings, it would be of utmost importance to know how long it takes for CO₂ to equilibrate after ECMO support initiation or any parameter change.

Based on previous unpublished data, we have hypothesized that a 50-min period would be enough for PaCO₂ equilibrium after each change on ECMO settings. Therefore, we conducted this experimental study to analyze the kinetics of arterial partial pressure of carbon dioxide following the changes in ECMO parameters during apnoeic veno-venous ECMO support.

Methods

This manuscript is part of a sequence of experiments during porcine respiratory ECMO support, some of which were previously published elsewhere [11]. The study was approved by the Institutional Animal Research Ethics Committee from the Hospital Sírio-Libanês in São Paulo, Brazil, and was done according to the National Institutes of Health guidelines for the use of experimental animals. Instrumentation, surgical preparation, and lung injury were performed as previously described [11].

Instrumentation and surgical preparation

Five domestic female *Agroceres* pigs (80 [79,81] kg) were anesthetized with thionembu-
tal (10 mg.kg⁻¹, thiopental, Abbott, Brazil) and pancuronium bromide (0.1 mg.kg⁻¹, Pavulon, AKZO Nobel, Brazil) and connected to a mechanical ventilator (Evita XL Dräger, Dräger, Lübeck, Germany) with the following parameters: tidal volume 8 mL/kg, end-expiratory pressure 5 cmH₂O, FiO₂ initially set at 100 % and subsequently adjusted to maintain arterial saturation between 94 and 96 %, and respiratory rate titrated to maintain PaCO₂ between 35 and 45 mmHg or an end-tidal CO₂ (NICO, Dixtal Biomedica Ind. Com, São Paulo, Brazil) between 30 and 40 mmHg. The electrocardiogram, heart rate, oxygen saturation, and systemic pressures of the animals were monitored with a multiparametric monitor (Infinity Delta XL, Dräger, Lübeck, Germany). Central venous pressure (CVP), mean pulmonary artery pressure (PAPm), pulmonary artery occluded pressure (PAOP), and cardiac output (CO) were measured with the use of a pulmonary artery catheter. Anesthesia was maintained with midazolam (1–5 mg.kg⁻¹.h⁻¹) and fentanyl (5–10 mcg.kg⁻¹.h⁻¹), and muscular relaxation was maintained with pancuronium bromide (0.2 mg.kg⁻¹.h⁻¹).

A 25-cm ECMO arterial cannula (Edwards Lifesciences, Irvine, CA, USA) was introduced into the right external jugular vein. A 55-cm ECMO drainage cannula (Edwards Lifesciences, Irvine, CA, USA) was positioned close to the right atrium via the right femoral vein with the aid of transhepatic ultrasonographic visualization. Only the

guidewires were inserted until the first baseline measurements, after the stabilization period (Fig. 1). Heparin infusion was then started at 1000 IU/h. After the first baseline, guidewires were replaced by the ECMO cannulas. Cannula diameter was chosen in accordance with vein diameter as measured with the aid of ultrasonography. Four animals had a 20-Fr and one had a 21-Fr drainage cannula. Three animals had a 21-Fr and two had a 20-Fr return cannula. A central venous catheter and an arterial line were placed in the left femoral vein and artery, respectively.

Stabilization and support of the animals

After surgical preparation, we allowed the animals to stabilize for 1 h. A continuous infusion of $3 \text{ mL} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ of lactated Ringer's solution was infused throughout the experiment. Fluid challenges and vasopressors were used to maintain the mean arterial pressure between 65 and 80 mmHg.

ECMO priming, starting, and maintenance

The ECMO system (Permanent Life Support (PLS) System, Jostra–Quadrox D, Maquet Cardiopulmonary, Hirrlingen, Germany) was primed with a 37°C normal saline solution and connected to a centrifugal pump (Rotaflow, Jostra, Maquet Cardiopulmonary, Hirrlingen, Germany). Heparin infusion was titrated to keep the activated coagulation time 1.5–2.5 times its baseline value. The absence of significant re-circulation was confirmed by an oxygen saturation of less than 70 % collected from the pre-membrane port [10, 12] 10 min after the beginning of each new sweep gas flow.

Varying ECMO blood flow and sweep gas flow

The animal was kept in apnoea with 10 cmH_2O of PEEP and a $\text{FiO}_2 = 1.0$ using a concentric coil-resistor PEEP valve (Vital Signs Inc., Totowa, NJ, USA) with a humidified oxygen continuous flow at 10.0 L/min. ECMO blood flow was initially set at 5.0 L/min, and the sweep flow was set at 5.0 L/min. After 10 min, clinical and laboratorial data were collected, and blood flow was reduced to 1500 mL/min, with an initial sweep gas flow = 3.0 L/min. An arterial blood sample was collected every 10 min for up to

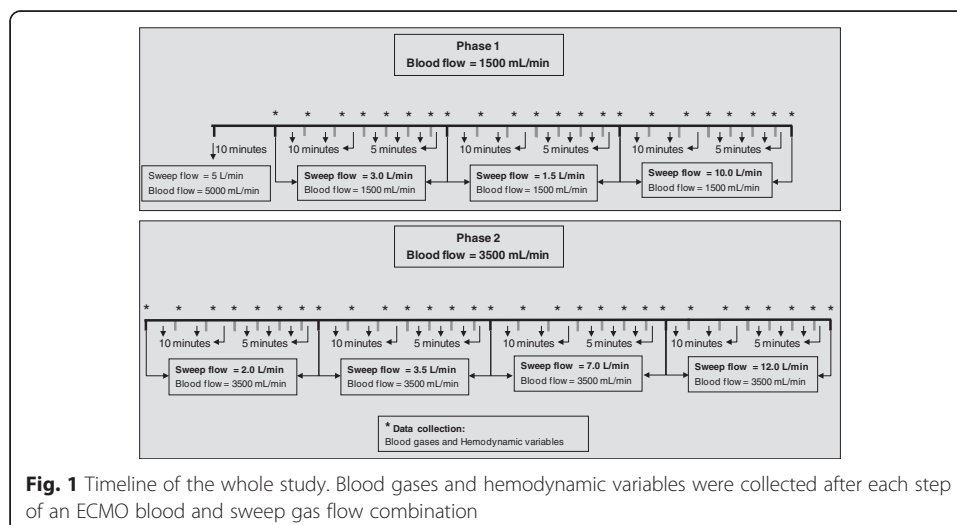


Fig. 1 Timeline of the whole study. Blood gases and hemodynamic variables were collected after each step of an ECMO blood and sweep gas flow combination

30 min; afterwards, an arterial blood sample was collected every 5 min until the new ECMO adjustment was in place for 50 min. Carbon dioxide equilibrium was defined as two blood gas analysis with a PaCO₂ variation less than 3 %. Hemodynamic data were also collected along with blood samples. After this step, the blood flow was kept at 1500 mL/min, and the sweep gas flow was set at 1.5 L/min and subsequently at 10.0 L/min in 50-min intervals. Blood flow was then elevated to 3500 mL/min, and a sequence of sweep gas flows of 2.0, 3.5, 7.0, and 12.0 L/min for 50 min each was applied (see Fig. 1). Two different blood flows were chosen in order to evaluate the impact of blood flow in PaCO₂ equilibrium. For each ECMO blood flow, sweep gas flow sequence was chosen arbitrarily. However, for a blood flow of 1.5 L/min, we chose an alternating sequence of 3.0, 1.5, and 10 L/min in order to minimize a possible carbon dioxide carry-over phenomenon.

Time to equilibrium–time constant (tau) determination

To calculate the time constant for each sweep gas and blood flow combination, data corresponding to the PaCO₂ value along the time were fitted into the following equation [13]:

$$\text{Partial pressure of CO}_2 = a + b \times \left(1 - e^{-t/\text{tau}}\right)$$

where t is the time in minutes, e corresponds to Euler's number, tau is the time constant obtained for each flow combination, and a and b correspond to constants used in the model.

Calculations

Calculations were done using standard formulas:

- Blood oxygen content C_bO₂ [mL O₂/100 mL of blood] = 1.36 × Hb × Sat_bO₂ + 0.0031 × P_bO₂
- O₂ transfer [mL/min] = (1.36 × Hb × (After - pre-membrane SatO₂) + 0.0031 × (After - pre-membrane PO₂)) × ECMO blood flow
- CO₂ transfer [mL/min] = (CO₂ partial pressure of the exhalation port of the membrane/barometric pressure) × sweep flow in mL/min
- Blood CO₂ content [mL/min] [14] = (1 - ((0.0289 × Hb)/(3.352 - 0.456 × (Sat_bO₂/100) × (8.142 - pH_b)))) × 2.226 × 0.0307 + (0.00057 × (37 - temperature)) + (0.00002 × (37 - temperature)²) × P_bCO₂ × (1 + 10^(pH_b - 6.086) + (0.042 × (7.4 - pH_b))) + ((38 - temperature) × 0.00472 + (0.00139 × (7.4 - pH_b)))
- Oxygen delivery DO₂ [mL O₂/min] = cardiac output × CaO₂ × 10
- Oxygen consumption VO₂ [mL O₂/min] = C(a - v)O₂ × cardiac output × 10
- Standard base excess [SBE - mEq/L] = 0.9287 × (HCO₃⁻ - 24.4 + 14.83 × (pH - 7.4))

Statistical analysis

Normality was assessed by the Shapiro-Wilk test. Variables are shown as mean ± standard deviation if normally distributed and median and the 25th and 75th percentiles if otherwise. Percent variations of PaCO₂ at the end of the 50-min observations in each sweep gas flow were normally distributed and compared with the paired t test. The R

free source statistical package and comprehensive-R archive network (CRAN)-specific libraries were used to build the graphics and analyze the data [15].

Results

During the entire experiment, arterial oxygen saturation (SatO₂), pH, and standard base excess varied across the combinations of sweep gas and blood flow; however, arterial O₂ and CO₂ contents did not vary significantly. There were slight variations in hemoglobin and electrolytes. Table 1 shows all variables measured at the end of each 50-min period of observation

Variations of PaCO₂ over time with an ECMO blood flow of 1500 mL/min are shown in Fig. 2. As expected, PaCO₂ increased with the reduction of sweep gas flow from 3 to 1.5 L/min; however, an elevation of sweep gas flow from 1.5 to 10 L/min was associated with a slight fall of PaCO₂ to levels similar to those observed with 3 L/min (the PaCO₂ with sweep = 3.0 L/min was 66 [55,78] mmHg, and the PaCO₂ with sweep = 10.0 L/min was 61 [48,92] mmHg, $P = 0.814$). In all sweep gas flows tested, the PaCO₂ variation was not statistically different at the end of 50 min; Furthermore, a percent mean difference of PaCO₂ between 45 and 50 min was $2 \pm 3 \%$ ($P = 0.124$, paired t test), $-1 \pm 3 \%$ ($P = 0.168$, paired t test), and $4 \pm 7 \%$ ($P = 0.338$, paired t test) with the sweep gas flow set at 3.0, 1.5, and 10.0 L/min, respectively. However, after fitting the model of PaCO₂ equilibrium, we observed that with a blood flow of 1500 mL/min, the time for ascendant PaCO₂ stabilization phase had a time constant as high as 30 min, therefore an expected time to equilibrium of around 90 min. It is interesting to note that, differently from expected, after reducing the sweep gas flow from 3.0 to 1.5 L/min, the time constant fell from 33 ± 3 to 17 ± 4 min.

In Fig. 3, the kinetics of PaCO₂ with an ECMO blood flow of 3500 mL/min is shown. Unlike the previous condition, the increase of sweep gas flow up to 12 L/min was associated with a progressive reduction in the PaCO₂. The percent mean difference of the PaCO₂ between 45 and 50 min was $0 \pm 2 \%$ ($P = 0.393$, paired t test), $1 \pm 1 \%$ ($P = 0.098$, paired t test), $-2 \pm 5 \%$ ($P = 0.357$, paired t test), and $2 \pm 0.4 \%$ ($P = 0.100$, paired t test) with the sweep gas flow set at 2.0, 3.5, 7.0, and 12.0 L/min, respectively.

Both ECMO blood flow and sweep gas flow affected the time to equilibrium of PaCO₂ (Fig. 4). Within the studied range of sweep gas flow and blood flow, the time constant values varied from 6 to 33 min.

Discussion

In all conditions tested, carbon dioxide had a long time to equilibrium after changes in ECMO settings. With an ECMO blood flow of 1500 mL/min, increases in sweep gas flow beyond 3.0 L/min did not significantly affect PaCO₂. With an ECMO blood flow of 3500 mL/min, all increments of sweep gas flow up to 12 L/min resulted in additional reductions of PaCO₂. CO₂ equilibrium was faster with higher blood and sweep gas flows.

Unlike oxygen, which has a fast time to equilibrium and can be monitored by pulse oximetry at the bedside [16], CO₂ requires serial arterial blood gases measurements. Differently from our initial expectative, we found long time constants for CO₂ equilibrium with a blood flow of 1500 mL/min, suggesting that late (>50 min) assessments of arterial blood gases are usually necessary to capture the full impact of new ECMO

Table 1 Physiological variables after 50 min of sweep or blood flow modifications

Variable	Blood flow = 1500 mL/min			Blood flow = 3500 mL/min				P value*
	Sweep flow 3.0 L/min	Sweep flow 1.5 L/min	Sweep flow 10 L/min	Sweep flow 2.0 L/min	Sweep flow 3.5 L/min	Sweep flow 7.0 L/min	Sweep flow 12 L/min	
pH	7.21 [7.17,7.32]	7.03 [7.01,7.16]	7.26 [7.19,7.38]	7.08 [7.04,7.22]	7.16 [7.14,7.29]	7.31 [7.27,7.40]	7.43 [7.37,7.49]	<0.001
SBE (mmol/L)	0.9 [0.0,1.4]	-5.2 [-9.1,-1.2]	0.3 [-2.2,2.5]	-0.3 [-1.7,0.4]	1.2 [-1.2,2.0]	4.3 [2.9,4.9]	4.8 [4.7,6.3]	<0.001
SatO ₂ (%)	87 [81,98]	85 [75,96]	97 [80,99]	99 [91,100]	99 [91,100]	99 [96,99]	99 [98,100]	0.003
CaO ₂ (mL O ₂ /100 mL of blood)	17 [14,17]	16 [13,18]	17 [14,18]	15 [14,18]	17 [15,18]	15 [15,19]	15 [14,18]	0.764
CaCO ₂ (mL CO ₂ /100 mL of blood)	58 [57,64]	65 [54,66]	64 [58,66]	67 [63,74]	64 [61,64]	65 [63,65]	63 [63,69]	0.179
Lactate (mmol/L)	1.6 [1.3,2.2]	2.7 [1.7,3.6]	1.9 [1.7,4.3]	2.6 [1.1,3.4]	1.6 [1.6,3.2]	1.0 [1.0,1.8]	1.7 [0.8,2.1]	0.031
Hemoglobin (g/dL)	13 [12,14]	14 [13,14]	13 [12,13]	12 [11,13]	12 [12,13]	12 [11,13]	11 [10,13]	0.003
Chloride (mmol/L)	104 [103,104]	103 [103,104]	103 [103,104]	103 [103,104]	103 [103,104]	103 [102,105]	103 [103,104]	0.880
Sodium (mmol/L)	140 [138,143]	140 [139,142]	140 [137,141]	141 [138,141]	139 [138,142]	137 [137,139]	139 [136,139]	0.007
Calcium (mmol/L)	1.4 [1.4,1.4]	1.4 [1.3,1.4]	1.3 [1.3,1.3]	1.4 [1.3,1.4]	1.3 [1.3,1.4]	1.3 [1.3,1.4]	1.3 [1.3,1.3]	0.007
Glucose (mg/dL)	112 [103,131]	129 [116,214]	135 [115,158]	137 [124,138]	116 [112,127]	126 [121,128]	119 [118,121]	0.007
Pulmonary shunt (%)	47 [26,54]	60 [33,67]	45 [29,66]	67 [55,72]	54 [47,55]	56 [31,59]	50 [44,56]	0.115
Temperature (°C)	37.2 [36.8,38.5]	37.5 [36.6,38.4]	37.5 [36.4,38.2]	38.0 [37.0,38.1]	37.7 [36.9,38.2]	37.5 [37.0,38.2]	37.5 [37.0,38.2]	0.534
O ₂ transfer (mL/min)	116 [108,134]	115 [103,128]	133 [98,190]	246 [211,246]	201 [197,222]	206 [200,217]	207 [181,237]	<0.001
CO ₂ transfer (mL/min)	209 [204,283]	185 [178,204]	319 [275,377]	255 [223,275]	259 [238,320]	365 [294,386]	383 [296,400]	<0.001
Rotations (RPM)	2075 [1930,2175]	2075 [1980,2175]	2075 [1980,2175]	3485 [3380,3780]	3485 [3380,3780]	3485 [3380,3725]	3485 [3380,3725]	<0.001
CO (L/min)	6.5 [6.4,6.9]	6.3 [4.0,6.7]	7.7 [3.6,9.4]	7.3 [4.3,9.3]	9.0 [4.4,9.1]	7.6 [4.8,8.2]	6.3 [4.8,7.9]	0.834
Heart rate (bpm)	134 [124,145]	133 [95,137]	135 [123,147]	128 [124,149]	137 [131,160]	121 [121,153]	128 [123,161]	0.867
ABPm (mmHg)	126 [106,126]	106 [100,125]	101 [91,112]	98 [92,105]	103 [94,114]	113 [112,122]	122 [119,123]	0.154

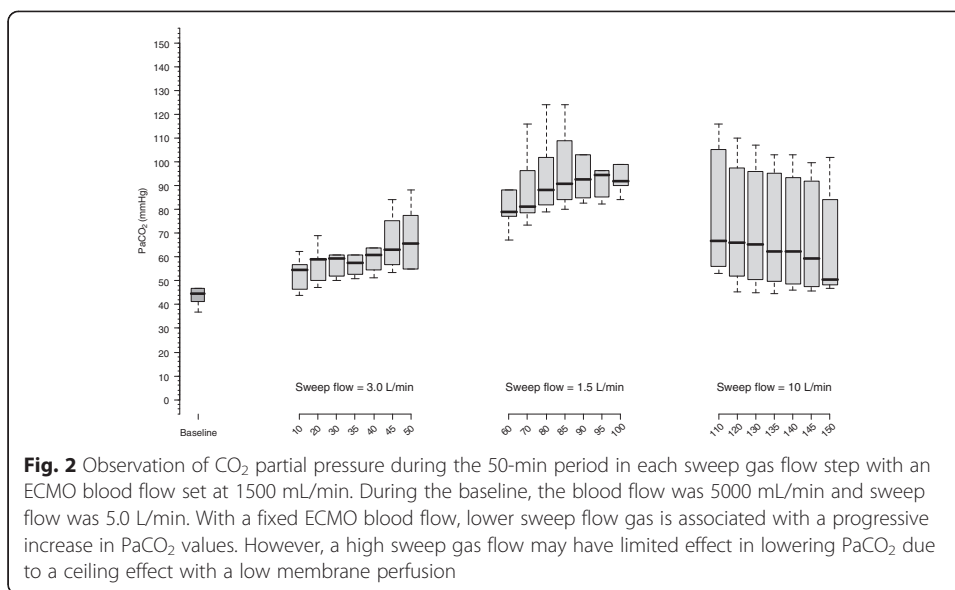
Table 1 Physiological variables after 50 min of sweep or blood flow modifications (Continued)

PAPm (mmHg)	54 [41,59]	46 [42,47]	36 [32,46]	47 [34,48]	40 [31,47]	42 [35,45]	39 [33,44]	0.248
CVP (mmHg)	6 [5,13]	6 [4,16]	5 [4,13]	5 [3,15]	5 [4,14]	5 [4,11]	5 [4,11]	0.299
PAOP (mmHg)	11 [10,12]	8 [8,11]	8 [7,11]	7 [5,9]	8 [7,8]	8 [6,16]	8 [7,17]	0.715

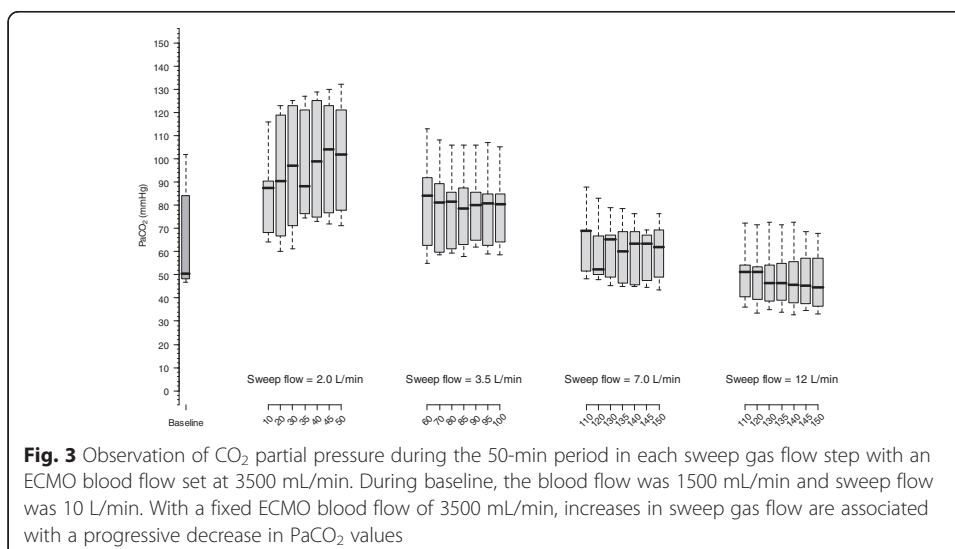
Post hoc analyses were not done due to the high spectrum of comparisons (varying through the blood flow and sweep gas flows domains)

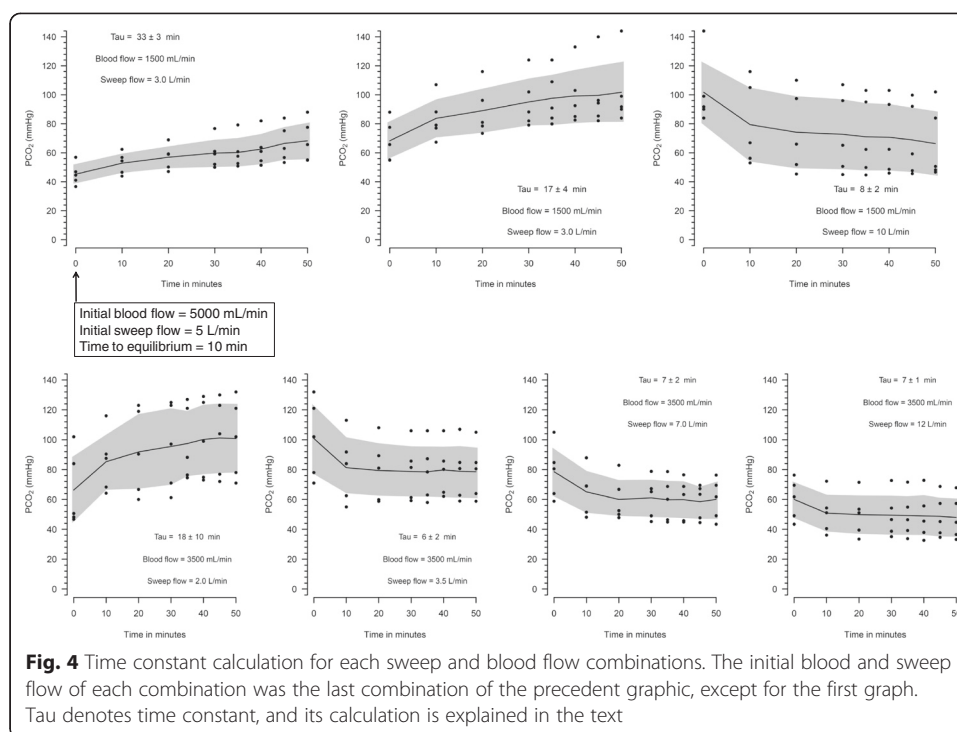
SBE standard base excess, *CaO₂* arterial content of oxygen, *CaCO₂* arterial content of CO₂, *CO* cardiac output, *ABPm* mean arterial blood pressure, *PAPm* mean pulmonary arterial blood pressure, *CVP* central venous pressure, *PAOP* pulmonary artery occlusion pressure

*The *P* values were obtained through the Friedman's test



settings in low ECMO blood flow states. Furthermore, this finding is also explained by the large volume of distribution of CO₂ [17], also stored as bicarbonate, bound to hemoglobin, and dissolved in the peripheral tissues [14, 18, 19]. The plasma dissolved fraction (PaCO₂) actually accounts for less than 5 % of the total body storage of CO₂. Therefore, small variations in PaCO₂ represent large total CO₂ variations [20]. Because the changes in CO₂ elimination are small in relation to its volume of distribution, the time to equilibrium is expected to be long. Moreover, after the elevation of effective ventilation, the reduction of the PaCO₂ will depend mainly of the effective ventilation and volume of CO₂ distribution; and after the reduction of effective ventilation, the elevation of PaCO₂ will depend of the volume of distribution and of the CO₂ production. The last depends on other variables such as temperature, sedation, inflammatory status, etc. Additionally, the interchange among the different molecular forms of CO₂ depends on other modulators, such as temperature, local pH, carbonic anhydrase activity, and





others [19, 21]. Thus, the time to PaCO₂ equilibrium is a complex function derived from the combination of many variables. In this study, a 50-min period was sufficient to achieve the PaCO₂ equilibrium in all sweep gas flow tested only with a blood flow of 3500 mL/min, otherwise in using 1500 mL/minute of blood flow, it may take as long as 90 min to reach the equilibrium.

We kept the animals apnoeic to restrict the elimination of CO₂ to the ECMO system. With the subjects ventilating, even with a constant minute alveolar volume, changes in alveolar elimination of CO₂ are important, because alveolar elimination depends on both alveolar CO₂ concentration and alveolar minute ventilation [11]. We reasoned that the apnoeic model would better reflect the isolated effects of changes in ECMO settings. In Table 1, it is possible to note that CO₂ transfer and production varied importantly between study steps and, more importantly, with an ECMO blood flow of 1500 mL/min. We attribute this finding mainly to the absence of CO₂ stabilization throughout the phase 1 study period, instead of marked change on CO₂ production between steps. Moreover, a variation on cardiac output and unstable animal conditions may have contributed to this finding.

At low ECMO blood flow (1500 mL/min), we found that CO₂ elimination had a ceiling effect at sweep gas flow of 3.0 L/min with higher gas flows exerting no significant impact on PaCO₂. This finding suggests that post-membrane CO₂ was already very low with the sweep gas flow of 3.0 L/min, and that CO₂ transfer was thus perfusion limited. This implies that, due to its high diffusibility, a very low CO₂ partial pressure (its dissolved fraction) can be reached early during the passage of blood through the ECMO membrane, limiting CO₂ elimination independently of the sweep gas flow. This finding is of great interest in situations in which ECMO is used mainly for hypercapnia or in low flow devices, such as ECCO₂R removal technologies.

Our study has several limitations: (1) The low number of animals could lead to type-II errors, attenuating the validity of our results. However, such a limitation would not affect our positive findings. (2) We did not study animals with multi-organ failure syndrome, a condition likely closer to a clinical scenario. However, many positive associations will be intuitively the same. (3) As presented in Table 1, we observed a progressive decrease in hemoglobin concentration throughout the experiment. Considering that hemoglobin acts as a CO₂ reservoir, this may have influenced the results. (4) We did the analysis with a static 50-min period to gas equilibrium. With low ECMO blood flows, it was not enough for PaCO₂ to reach the equilibrium and, therefore, definitive time to equilibrium for such situation remains unknown. Moreover, it is possible that late inter-compartment distributions may occur, affecting our results.

Conclusions

In conclusion, our results suggest that 50 min was enough to reach the equilibrium of PaCO₂ after ECMO initiation or after changes in blood and sweep gas flow with an ECMO blood flow of 3500 mL/min. Longer periods are necessary to reach the equilibrium with an ECMO blood flow of 1500 mL/min. This may help bedside decisions and facilitate daily management of patients during ECMO support.

Abbreviations

CO₂: carbon dioxide; ECCO2R: extracorporeal carbon dioxide removal; ECMO: extracorporeal membrane oxygenation; O₂: oxygen; PaCO₂: arterial carbon dioxide partial pressure; PaO₂: arterial oxygen partial pressure; PEEP: positive end expiratory pressure; SatO₂: arterial oxygen saturation.

Competing interests

The authors received a donation of PLS systems from Maquet Cardiopulmonary of Brazil to do experimental research and for patient support. The authors declare that they have no competing interests.

Authors' contributions

PVM and MP performed the statistical analysis, interpreted of data, and drafted the manuscript. PVM and MP contributed similarly to the study. ATM, DPS, NF, EVSB, ASH, GPPS, and LCPA contributed to the conception of the work, interpretation of data, and drafting of the manuscript. ELVC contributed to the conception of the work, performed the statistical analysis, interpreted the data, and did the final critical review of the manuscript. All authors read and approved the final manuscript.

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References

1. Australia and New Zealand Extracorporeal Membrane Oxygenation (ANZ ECMO) Influenza Investigators, Davies A, Jones D, Bailey M, Beca J, Bellomo R, Blackwell N, Forrest P, Gattas D, Granger E, Herkes R, Jackson A, McGuinness S, Nair P, Pellegrino V, Pettit V, Plunkett B, Pye R, Torzillo P, Webb S, Wilson M, Ziegenfuss M (2009) Extracorporeal membrane oxygenation for 2009 influenza A(H1N1) acute respiratory distress syndrome. *JAMA* 302(17):1888–1895
2. Peek GJ, Mugford M, Tiruvoipati R, Wilson A, Allen E, Thalany MM et al (2009) Efficacy and economic assessment of conventional ventilatory support versus extracorporeal membrane oxygenation for severe adult respiratory failure (CESAR): a multicentre randomised controlled trial. *Lancet* 374(9698):1351–1363
3. Peek GJ, Moore HM, Moore N, Sosnowski AW, Firmin RK (1997) Extracorporeal membrane oxygenation for adult respiratory failure. *Chest* 112(3):759–764
4. Linden V, Palmer K, Reinhard J, Westman R, Ehren H, Granholm T et al (2000) High survival in adult patients with acute respiratory distress syndrome treated by extracorporeal membrane oxygenation, minimal sedation, and pressure supported ventilation. *Intensive Care Med* 26(11):1630–1637
5. Frenckner B, Palmer P, Linden V (2002) Extracorporeal respiratory support and minimally invasive ventilation in severe ARDS. *Minerva Anesthesiol* 68(5):381–386
6. Pham T, Combes A, Rozé H, Chevret S, Mercat A, Roch A et al (2012) Extracorporeal membrane oxygenation for pandemic influenza A(H1N1) induced acute respiratory distress syndrome. A cohort study and propensity-matched analysis. *Am J Respir Crit Care Med* 187(3):276–285
7. Noah MA, Peek GJ, Finney SJ, Griffiths MJ, Harrison DA, Grieve R et al (2011) Referral to an extracorporeal membrane oxygenation center and mortality among patients with severe 2009 influenza A(H1N1). *JAMA* 306(15):1659–1668
8. Bonacchi M, Harmelin G, Peris A, Sani G (2011) A novel strategy to improve systemic oxygenation in venovenous extracorporeal membrane oxygenation: the “χ”-configuration”. *J Thorac Cardiovasc Surg* 142(5):1197–1204
9. Messaï E, Bouguerra A, Harmelin G, Di Lascio G, Cianchi G, Bonacchi M (2013) A new formula for determining arterial oxygen saturation during venovenous extracorporeal oxygenation. *Intensive Care Med* 39(2):327–334

10. Sidebotham D, McGeorge A, McGuinness S, Edwards M, Willcox T, Beca J (2010) Extracorporeal membrane oxygenation for treating severe cardiac and respiratory failure in adults: part 2-technical considerations. *J Cardiothorac Vasc Anesth* 24(1):164–172
11. Park M, Costa EL, Maciel AT, Silva DP, Friedrich N, Barbosa EV et al (2013) Determinants of oxygen and carbon dioxide transfer during extracorporeal membrane oxygenation in an experimental model of multiple organ dysfunction syndrome. *PLoS One* 8(1):e54954
12. Sidebotham D (2011) Troubleshooting adult ECMO. *J Extra Corpor Technol* 43(1):27–32
13. D'Angelo E, Tavola M, Milic-Emili J (2000) Volume and time dependence of respiratory system mechanics in normal anaesthetized paralysed humans. *Eur Respir J* 16(4):665–672
14. Douglas AR, Jones NL, Reed JW (1988) Calculation of whole blood CO₂ content. *J Appl Physiol* 65(1):473–477
15. Team RDC. R: A language and environment for statistical computing. R Foundation for Statistical Computing. 2009
16. Schmidt M, Tachon G, Devilliers C, Muller G, Hekimian G, Bréchet N et al (2013) Blood oxygenation and decarboxylation determinants during venovenous ECMO for respiratory failure in adults. *Intensive Care Med* 39:838–846
17. Cherniack NS, Longobardo GS (1970) Oxygen and carbon dioxide gas stores of the body. *Physiol Rev* 50(2):196–243
18. Henry RP, Swenson ER (2000) The distribution and physiological significance of carbonic anhydrase in vertebrate gas exchange organs. *Respir Physiol* 121(1):1–12
19. Meldrum NU, Roughton FJ (1933) The state of carbon dioxide in blood. *J Physiol* 80(2):143–170
20. Anderson CT, Breen PH (2000) Carbon dioxide kinetics and capnography during critical care. *Crit Care* 4(4):207–215
21. Meldrum NU, Roughton FJ (1933) Carbonic anhydrase. Its preparation and properties. *J Physiol* 80(2):113–142

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