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# *Ex vivo* drug sensitivity profiles of *Plasmodium falciparum* field isolates from Cambodia and Thailand, 2005 to 2010, determined by a histidine-rich protein-2 assay

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## Abstract

**Background:** *In vitro* drug susceptibility assay of *Plasmodium falciparum* field isolates processed “immediate *ex vivo*” (IEV), without culture adaptation, and tested using histidine-rich protein-2 (HRP-2) detection as an assay, is an expedient way to track drug resistance.

**Methods:** From 2005 to 2010, a HRP-2 *in vitro* assay assessed 451 *P. falciparum* field isolates obtained from subjects with malaria in western and northern Cambodia, and eastern Thailand, processed IEV, for 50% inhibitory concentrations (IC<sub>50</sub>) against seven anti-malarial drugs, including artesunate (AS), dihydroartemisinin (DHA), and piperazine.

**Results:** In western Cambodia, from 2006 to 2010, geometric mean (GM) IC<sub>50</sub> values for chloroquine, mefloquine, quinine, AS, DHA, and lumefantrine increased. In northern Cambodia, from 2009–2010, GM IC<sub>50</sub> values for most drugs approximated the highest western Cambodia GM IC<sub>50</sub> values in 2009 or 2010.

**Conclusions:** Western Cambodia is associated with sustained reductions in anti-malarial drug susceptibility, including the artemisinins, with possible emergence, or spread, to northern Cambodia. This potential public health crisis supports continued *in vitro* drug IC<sub>50</sub> monitoring of *P. falciparum* isolates at key locations in the region.

**Keywords:** Cambodia, malaria, *Plasmodium falciparum*, HRP-2, Anti-malarial drugs, Drug resistance

## Background

Since the 1980s, measuring *in vitro* drug 50% inhibitory concentrations (IC<sub>50</sub>) against *P. falciparum* field isolates has been useful in tracking clinical drug susceptibility patterns [1-3]. In Southeast Asia, especially along the Thailand-Cambodia border, changes in drug susceptibility often emerge first, with worldwide implications,

underscoring the region's importance as a sentinel site for anti-malarial drug resistance [3].

In 2003, artesunate (AS) + mefloquine (MQ) was implemented as the first-line artemisinin-combination therapy (ACT) for *falciparum* malaria in Cambodia. By 2005, subjects with *falciparum* malaria along Thai-Cambodia border treated with oral artesunate were showing longer parasite clearance times, suggesting the emergence of reduced susceptibility to artemisinins, as well as to the partner drug [4].

The non-radioisotope histidine-rich protein-2 (HRP-2) ELISA, a relatively sensitive assay, which reliably depicts drug IC<sub>50</sub> values for *P. falciparum* isolates, reduces obstacles for conducting assays in remote settings [5,6].

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HRP-2 assays are expedient, safer and less costly than radioisotope assays. HRP-2 assay, when field deployed proximal to *P. falciparum* collection sites, allows for "immediate *ex vivo*" (IEV) field isolate processing [7]. IEV, by avoiding cryopreservation and culture-adaptation before IC<sub>50</sub> determination, may reduce clonal selection, better preserving parasite subpopulations with variable drug susceptibility profiles, the latter likely present in a smaller proportion than wild-type drug susceptible parasites.

Here, to characterize recent geographical and temporal trends in western and northern Cambodia, and eastern Thailand, including years when reduced artemisinin susceptibility was first described, an HRP-2 assay, with IEV field isolate processing was used to determine IC<sub>50</sub> values of *P. falciparum* field isolates obtained from 2005 to 2010.

## Methods

### Protocol, sites and subjects

All studies were approved by 1 or more ethical review boards which included Cambodian National Ethics Committee for Health Research (NECHR), Ethical Review Committee for Research in Human Subjects, Thailand Ministry of Public Health (MoPH), Walter Reed Army Institute of Research (WRAIR) Institutional Review Board and World Health Organization (WHO) Ethical Review Committee (protocol numbers: WRAIR 973, WRAIR 1327, WRAIR 1296, WRAIR 1396, WRAIR 1576, and WRAIR 1737). All protocols complied with International Conference on Harmonization Good Clinical Practice (ICH-GCP) guidelines. Enrollment centers were located in western and northern Cambodia (Pailin, Battambang, Oddar Meanchey and Preah Vihear Provinces (Figure 1), and Trat, eastern Thailand. All sites were considered low transmission, malaria endemic regions at the time of sample collection [8,9].

Among six studies involved in subject recruitment, summarized in Table 1, subjects of five to 70 years of age with smear-confirmed, uncomplicated *P. falciparum* malaria were eligible to participate. Written informed consent was obtained from adult subjects, or from legal guardians with assent from minors. Subjects who gave a history of anti-malarial drug use within the previous 7–30 days, were pregnant or nursing, or who had signs suggestive of severe malaria, were excluded from participation (Table 1). Treatment following malaria diagnosis was according to national standards of care at the respective times, or involved randomized allocation to an artemisinin containing regimen as part of an interventional clinical trial.

### Malaria microscopy

Microscopic examination of Giemsa-stained peripheral blood smears was performed by two microscopists for

each subject (200 high-powered fields [HPF] per reader) to determine parasitaemia, and presence/absence of a second malaria species. Microscopists conducted all reads in the thick smear unless there were > 500 parasites/200 WBCs, in which case they switched to the thin smear. Discrepancies between microscopists were resolved in real-time by a third reader (reviewing an additional 200 HPFs), who determined the final result. Parasite density was calculated as the number of parasites/200 WBC's in the thick smear or the number of parasites/5000 RBC's in the thin smear. A geometric mean (GM) of the two microscopist's results was used as the final parasitaemia for each subject. Time zero peripheral blood specimens from WRAIR 1396, 1576, and 1737 were also confirmed by species-specific real-time PCR (RT-PCR) targeting the 18 s rRNA gene. Specimens with a microscopically or RT-PCR confirmed malaria coinfection at time zero, *Plasmodium vivax* or other malaria species, were excluded from analysis. Subject parasite density ranges at study enrolment are shown in Table 1.

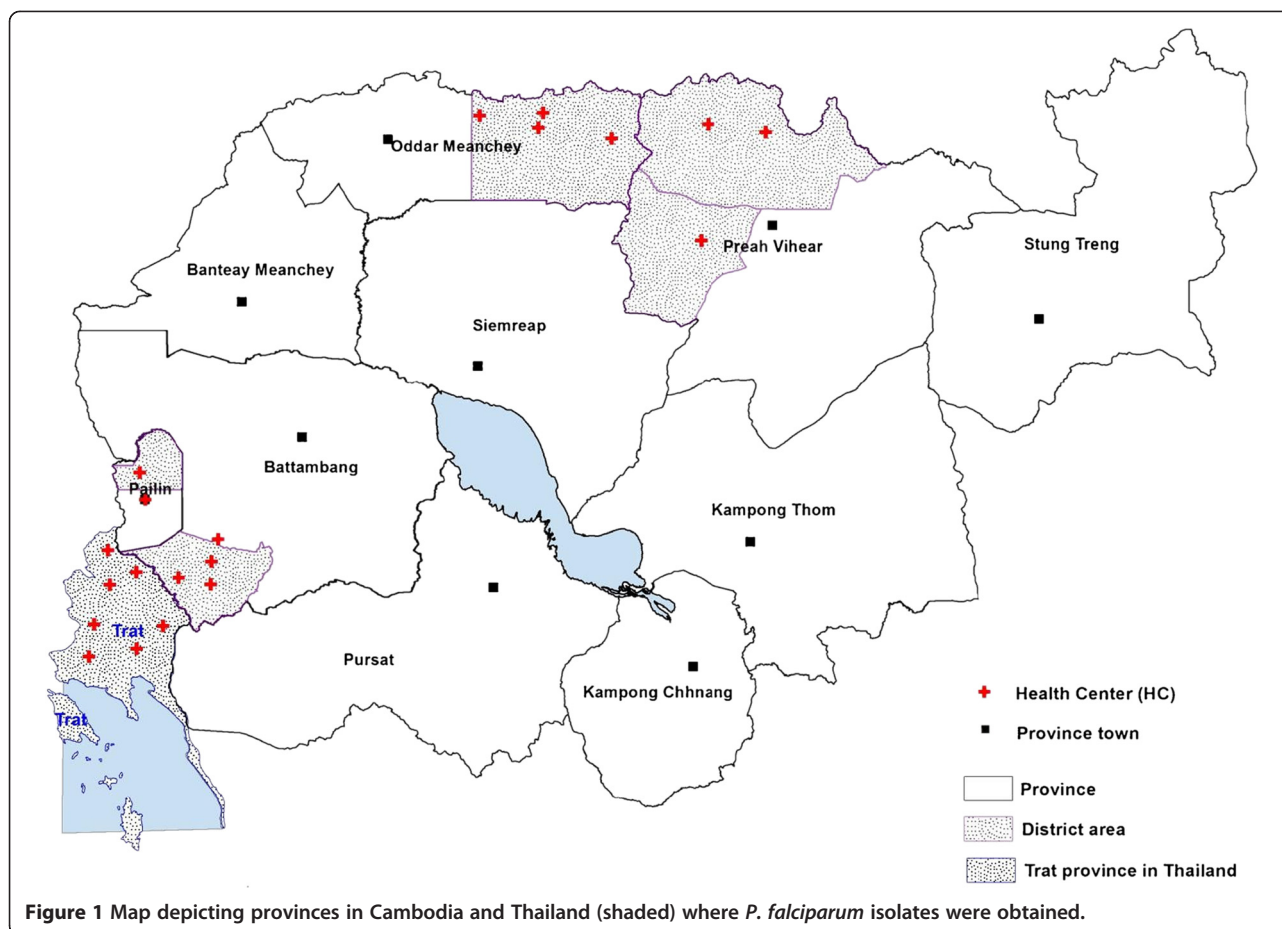
### Sample collection and preparation

Enrolled subjects provided up to 8 ml of venous blood in sodium heparin tubes for the HRP-2 *ex vivo* IC<sub>50</sub> assay. In all cases, *P. falciparum* isolates from submitting study locations were plated directly onto malaria drug coated plates without prior culture adaptation [5]. All applications of falciparum isolates to drug coated plate occurred at field sites or in central laboratories located at Battambang Referral Hospital (BRH) or at Anlong Veng Health Center (AVHC). Samples analyzed at BRH or AVHC were refrigerated during transportation (2 - 8° C) and plated < 6 hours after being drawn from the subject.

### Drugs and plate coating

Seven anti-malarial drugs for IC<sub>50</sub> testing were used: chloroquine diphosphate (CQ), mefloquine hydrochloride (MQ), quinine sulfate hydrate (QN) artesunate (AS), dihydroartemisinin (DHA), lumefantrine (LUM) and piperazine (PPQ). CQ, MQ, QN and PPQ were provided as salts, AS, DHA, and LUM as bases. All drug stocks were prepared using a weight/volume convention with salt or base formulation weights used to calculate nM IC<sub>50</sub> values. Test drugs were provided by Walter Reed Army Institute of Research (Silver Spring, Maryland, USA).

To prepare test drugs, stock drug solutions at 1 mg/mL were prepared in 70% ethanol for DHA, AS, MQ, QN, and CQ or a solution of 100% ethanol : linoleic acid : Tween 80 (1:1:1) for LUM and 0.5% lactic acid in distilled water for PPQ. Further drug dilutions were performed in distilled water for DHA, AS, MQ, QN, CQ,



**Figure 1** Map depicting provinces in Cambodia and Thailand (shaded) where *P. falciparum* isolates were obtained.

and PPQ or 70% ethanol for LUM to the desired starting concentration, followed by serial 3-fold dilutions to generate 7 concentrations for IC<sub>50</sub> testing. The last row of the drug plate contained no drugs. Dried drug coated plates were prepared as described [5,6,10].

The concentration ranges (ng/ml), from highest to lowest, and molecular weights (g/mole) used to convert to nM were, respectively: CQ (2000 to 2.74; 515.92), MQ (200 to 0.274; 414.78), QN (1250 to 1.71; 782.97), AS (20 to 0.027; 384.42), DHA (20 to 0.027; 284.35), LUM (50 to 0.07; 528.95) and PPQ (625 to 0.86; 999.56). The drug solutions were prepared and used within one week. Drug solutions were stored at 2 – 8°C and used within 1 week of preparation.

#### ***Plasmodium falciparum* W2 reference clone and quality control**

W2, a well characterized *P. falciparum* laboratory reference clone, recovered from cryopreserved sample and maintained in continuous culture for ≤ 5 months, served as quality control for drug coating plate lots and to assess assay performance within a previously validated acceptable range. W2 also served as a comparator for *P. falciparum* field isolate IC<sub>50</sub> values. W2 IC<sub>50</sub> assays

against each drug were conducted annually from 2008 to 2010. W2, *in vitro*, is considered CQ “resistant” (CQ-R), and MQ “susceptible” (MQ-S). The W2 GM IC<sub>50</sub> values of CQ, MQ and QN, especially when assessed simultaneously with *P. falciparum* field isolates, provide context for non-radioisotopic IEV assays [11]. W2 has no defined IC<sub>50</sub> value discriminative for AS or DHA “resistance”.

Earlier reports describe drug IC<sub>50</sub> “cut-off” values for “culture-adapted” *P. falciparum* field isolates, once generally considered discriminative for *in vivo* resistance; these include CQ (> 45.5 ng/ml; 85 nM), MQ (> 10 ng/ml; 24 nM), and QN (> 275 ng/ml; 351 nM) [12,13]. These historical cut-off values may provide rough context, primarily for assay reliability, and perhaps for *P. falciparum* field isolate IC<sub>50</sub> trends, including non-radioisotopic IEV assays [11].

For quality control (QC), conducted from 2008 to 2010, a subset of all drug-plate coating lots was tested using the HRP-2 assay with W2 and an IC<sub>50</sub> value was derived for each anti-malarial compound before the lot was sent to the field. Lots that performed within previously validated criteria were cleared to be sent to the field. Out of range lots were destroyed. Drug plating consistency and stability between different lots were

**Table 1 Studies providing *P. falciparum* isolates for HRP-2 IC<sub>50</sub> drug assays**

Study	Years of enrolment	Location		Relevant inclusion criteria
		Province, Country		
1	2005-07	Trat, Thailand <sup>1</sup>		> 19 years old, any asexual <i>P. falciparum</i> parasitaemia
2	2006-07	Trat, Thailand <sup>1</sup>		5-70 years old, any asexual <i>P. falciparum</i> parasitaemia
3	2006-07	Western Cambodia <sup>2</sup> : Battambang		18-65 years old, asexual <i>P. falciparum</i> parasitaemia, > 100 and < 100,000/μL
4	2008-09	Western Cambodia <sup>2</sup> : Battambang		18-65 years old, asexual <i>P. falciparum</i> parasitaemia, > 1000 and < 200,000/μL
5	2009-10	Western Cambodia <sup>2</sup> : Battambang, Pailin Northern Cambodia <sup>3</sup> : Oddar Mean Chey Preah Vihear		≥ 13 years old, any asexual <i>P. falciparum</i> parasitaemia
6	2010-11	Northern Cambodia <sup>3</sup> : Oddar Mean Chey Preah Vihear		18-65 years old, any asexual <i>P. falciparum</i> parasitaemia

<sup>1</sup> Thailand Ministry of Public Health Clinics in Borai, Khaosaming and Muang districts.

<sup>2</sup> Western Cambodia: Pailin (Pailin Hospital) and Battambang Provinces (Tasanh, Samlot, Chanlong Kuoy and Kampong Lpov health centers);

<sup>3</sup> Northern Cambodia: Oddar Mean Chey (Anlong Veng, Trapang Tav, Trapang Prey, Trapang Prasat and Sraem health centers), and Preah Vihear Provinces.

consistent with previous reports [14]. A subset of each plate-coating lot taken to the field was stored refrigerated (2 - 8°C) for up to 8 weeks, then transported back to the AFRIMS laboratory in Bangkok where QC was performed again with W2, to determine if transport and storage conditions in the field affected IC<sub>50</sub> analysis.

#### HRP-2 IC<sub>50</sub> drug susceptibility assay

An established HRP-2 IC<sub>50</sub> drug susceptibility assay was used to assess each *P. falciparum* field isolate, processed immediate *ex vivo* [5,6]. Falciparum isolates were processed within six hours after phlebotomy, including parasitaemia adjustment, as previously described, then placed onto drug coated plates (0 hours) [5,15]. Falciparum isolates within the range of plate parasitaemia (0.2% - 0.5%) were immediately transferred to the drug coated plates as the 0 hr screening sample; samples with a parasitaemia > 0.5% were diluted to within the plate parasitaemia range by adding 50% haematocrit human O+ red blood cells in 10% serum RPMI 1640 culture media; and samples with a parasitaemia below 0.2% were plated without adjustment [15].

Samples (~200 μl) from untreated wells were collected every 24 hours for 72 hours, with parasite growth rate interpreted as the difference in HRP-2 production

between 0 and 72 hours. Due to the high sensitivity of the HRP-2 assay, dilution of the samples was sometimes required before performing the ELISA to bring the concentration within assay OD range.

#### Statistical analysis of IC<sub>50</sub> values

HRP-2 optical densities (OD) were plotted against drug concentrations and IC<sub>50</sub> values estimated by nonlinear regression analysis using ICEstimator [16]. In *P. falciparum* field isolates which showed measurable growth, ICEstimator rules were used to determine the concentration-response relationship and IC<sub>50</sub>, i.e., ratio of growth without drug to that with maximum drug. For each drug, IC<sub>50</sub> values were expressed as geometric means (GM). Statistical significance within and between groups was determined by nonparametric Kruskal-Wallis or Mann-Whitney tests, as indicated (Graph-Pad Prism 4, GraphPad Software, Inc., La Jolla, California, USA).

## Results

#### HRP-2 IC<sub>50</sub> drug assays

Among 590 *P. falciparum* isolates collected, IC<sub>50</sub> values on 451 (76%) were obtained.

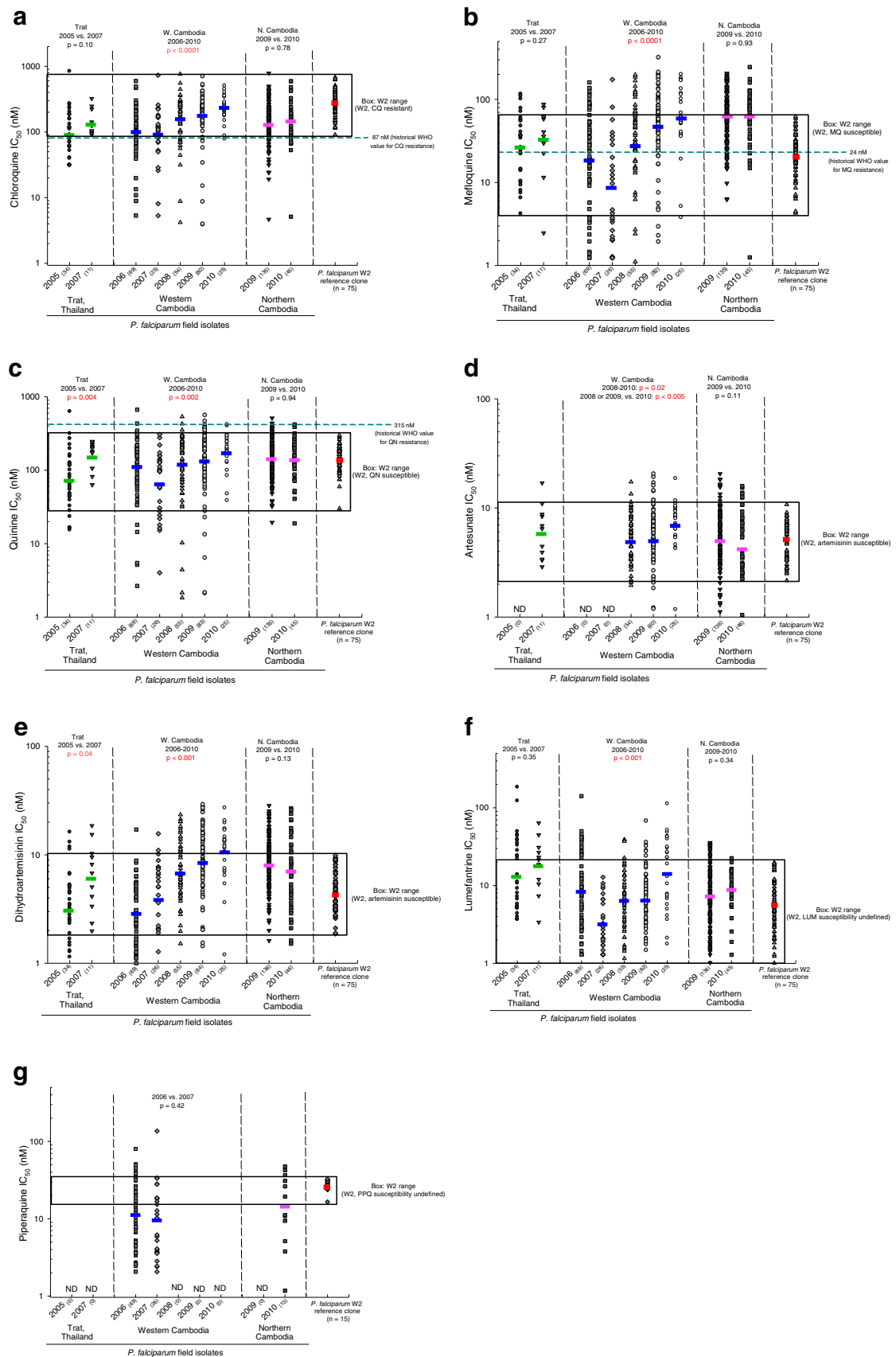
The success rate for all assays was about 75%, including 78% with a parasitaemia > 0.5% and 61% with a parasitaemia ≤ 0.5.

Figure 2 (a-g) shows individual and geometric mean (GM) IC<sub>50</sub> values for IEV *P. falciparum* field isolates, by location and year, and W2 *P. falciparum* reference clone (CQ-R, MQ-S, QN-S, artemisinin-S) IC<sub>50</sub> values against each drug. In western Cambodia, assessed annually from 2006 to 2010, geometric mean GM IC<sub>50</sub> values for CQ, MQ, QN, AS, DHA, and LUM increased (p < 0.05). In northern Cambodia, assessed in 2009 and 2010, GM IC<sub>50</sub> values for most drugs approximated the highest western Cambodia GM IC<sub>50</sub> values in 2009 or 2010. In Thailand, assessed in 2005 and 2007, GM IC<sub>50</sub> values increased for QN and DHA (p < 0.05).

## Discussion

Among *P. falciparum* field isolates obtained in western Cambodia from 2005 to 2010, steady increases were observed for GM IC<sub>50</sub> values measured by a HRP-2 *in vitro* assay against a range of anti-malarial drugs, including AS and DHA. Moreover, in northern Cambodia, assessed in 2009 and 2010, most GM IC<sub>50</sub> values approximated those in western Cambodia, during the same period. This supports the notion that western Cambodia is associated with sustained and likely progressive reductions in anti-malarial drug susceptibility, with possible spread to northern Cambodia.

As the same HRP-2 IEV assay was used for all *P. falciparum* field isolates, increasing GM IC<sub>50</sub> values for AS and DHA may reflect possible emerging artemisinin



**Figure 2** (See legend on next page.)



(See figure on previous page.)

**Figure 2 (a-g): *In vitro* HRP-2, *P. falciparum* field isolate IC<sub>50</sub> values (nM), determined by HRP-2 against 7 drugs shown as a scatter plots, with geometric mean (GM; solid color bars), by year and region.** Sample numbers, in parentheses, are next to each year. P values at top of graphs determined by Mann-Whitney (2 groups) or Kruskal-Wallis. (> 2 groups) tests; p < 0.05 in red. *P. falciparum* W2 reference clone values for each drug shown at far right (GM, solid red bars); boxes denote ranges. Dashed horizontal lines (green) on CQ, MQ and QN graphs represent approximate historical. IC<sub>50</sub> values, denoting clinical "resistance". nM = nanomolar. ND = not done.

resistance, first described in subjects in western Cambodia in 2005 [7,17,18]. Statistically significant increases in GM IC<sub>50</sub> values for DHA in Trat Province, Thailand, adjacent to the Cambodian border, from 2005 to 2007, coincided with declining efficacy of AS + MQ in Thailand [19]. These observations may reflect a > 30 year history of artemisinin use in this region.

In western Cambodia, a steady increase in GM IC<sub>50</sub> values for CQ from 2006 to 2010 was observed. This may reflect continued drug pressure due to its use for *P. vivax* blood stage treatment, as well as widespread unregulated availability. This is in contrast to the situation in parts of Africa, where the introduction of ACTs and withdrawal of CQ use has led to a reduction in mean IC<sub>50</sub> values from those recorded when CQ was the first-line treatment for *P. falciparum* infections [20,21].

QN showed no remarkable trends in GM IC<sub>50</sub> values, paralleling continued effectiveness of this 2nd line agent. For LUM, not widely used in Cambodia, increasing GM IC<sub>50</sub> values in western Cambodia from 2006–2010, in parallel with trends in other drugs, possibly mirrored increases in MQ IC<sub>50</sub>s (discussed below). For PPQ, the clinical relevance of our preliminary *in vitro* IC<sub>50</sub> data remains unclear. Of note, 15% of subjects in two recent malaria treatment trials in Cambodia had detectable pre-treatment PPQ levels, suggesting increasing drug pressure [22]. Continued *in vivo* and *ex vivo* monitoring of PPQ, the recently adopted first-line partner drug for DHA in Cambodia, is of paramount importance.

In western Cambodia, MQ GM IC<sub>50</sub> values showed a steady increase from 2006 to 2010, and comparably high GM IC<sub>50</sub> values were observed in northern Cambodia in 2009–2010; all were well above the proposed WHO historical cut-off indicative of clinical MQ monotherapy "resistance", paralleling reduced clinical efficacy of MQ in this region [23,24]. The cause of reduced MQ sensitivity (and possibly LUM) is generally accepted to be related to increased *Pfmdr1* copy number, so the association of increased MQ GM IC<sub>50</sub>s with a recent description of reduced *Pfmdr1* copy numbers in western Cambodia from 2005 to 2007 is unclear [25]. The WHO 2010 Global Report suggests this molecular event was due to a switch in treatment policy from MQ-AS to DHA-PPQ [4]; if so, a continued rise in MQ mean IC<sub>50</sub> values to 2010 is potentially concerning. As before, unregulated availability of antimalarial medications during

that period may have been a contributing factor. Notably, MQ-AS still appears to retain high efficacy despite reduced *in vitro* sensitivity to both drugs, shown most recently in a trial comparing mefloquine-artesunate with pyronaridine-artesunate [26].

For northern Cambodia, little is known about IC<sub>50</sub> malaria drug susceptibility [27]. Relatively high GM IC<sub>50</sub> values in northern Cambodia in 2009 and 2010 are supported by antecedent GM IC<sub>50</sub> values in eastern Thailand and western Cambodia [27-29]. In Thailand, GM IC<sub>50</sub> values for CQ, MQ, QN and LUM increased from 2005 to 2007, an increase mirrored in nearby western Cambodia, starting in 2006. QN GM IC<sub>50</sub> values remained below the WHO historic cut-off of 315 nM denoting "resistance", whereas CQ and MQ GM IC<sub>50</sub> values rose above WHO historic cut-off values, and DHA showed marked increases in both countries. These trends followed drug policies during the survey period, with AS-MQ the first-line regimen for *P. falciparum*, and CQ as the first-line agent for blood stage *P. vivax* in both Thailand and Cambodia.

In western Cambodia, measured annually 2006–2010, statistically significant, relatively steady increases in GM IC<sub>50</sub> values for CQ, MQ, QN, AS, DHA and LUM were noted. However, GM IC<sub>50</sub> values dropped in 2007, versus 2006, but then steadily increased from 2008 to 2010. It is unclear why GM IC<sub>50</sub> decreases occurred in 2007 only. Review of data from 2007 showed a smaller sample size (n = 26) compared to other years, although the location of sampled sites was unchanged. Interestingly Lim *et al.* observed a similar dip in IC<sub>50</sub> values for a range of antimalarial drugs in 2006, and attributed the observation to sampling bias since most samples that year were collected from a single, new field site; the data was included in the report for completeness and so that future comparisons could be made [27]. This perhaps illustrates the importance considering factors such as sampling bias when interpreting antimalarial drug IC<sub>50</sub> surveys.

HRP-2 assays with IEV isolate processing conducted at field laboratories, which are simpler techniques than radioisotopic assays and culture-adaptation, may also reduce clonal selection and better preserve sub-populations of susceptible and potentially drug resistant parasites [11]. In this survey about 75% of HRP-2 IEV assays were successful, and W2 clone IC<sub>50</sub> values for

CQ, MQ, QN and AS were within expected ranges, providing context for falciparum isolate IC<sub>50</sub> values. This bodes well for field based HRP-2 IEV assays. Moreover, GM IC<sub>50</sub> values reported here generally paralleled other published data from eastern Thailand and Cambodia, which assayed *P. falciparum* isolates by <sup>3</sup>H-hypoxanthine uptake [28,30]. For example, a comparison of western Cambodia GM IC<sub>50</sub> ranges from 2001 to 2007 [27], with these results were, respectively (nM): CQ (131–237 vs. 96–242), MQ (12–57 vs. 7–58), QN (94–302 vs. 53–163), and AS (0.6–1.8 vs. 5.1–6.8). The higher AS GM IC<sub>50</sub> range reported here could reflect continuing emergence of artemisinin resistance, paralleling GM IC<sub>50</sub> increases for AS and DHA from 2008 to 2010. Alternatively, it might reflect methodological differences, which affect drug or parasite behaviour, illustrating the importance of inter-assay consistency.

Trends in IC<sub>50</sub> values, coupled with molecular marker assays, help track emergence and spread of anti-malarial drug resistance. For the artemisinins, although associations between validated resistance markers, treatment outcome and IC<sub>50</sub> susceptibility remain elusive [18,31], in part because the clinical definition of artemisinin “resistance” is unclear; the converse is not true for partner drugs, such as MQ and CQ, which have established thresholds of resistance and validated molecular markers. Nonetheless, determining AS and DHA GM IC<sub>50</sub> values in a substantial number of *P. falciparum* isolates from 2005 to 2010 establishes a baseline which may be useful for future comparisons if evidence of reduced artemisinin susceptibility grows, and especially if the methodology remains consistent.

## Conclusion

Increases in GM IC<sub>50</sub> values for a range of important anti-malarial drugs in a region of emerging drug resistance underscores the importance of harmonizing methodologies that allow for accurate comparison between geographical locations, and over time, so that potentially important trends in drug susceptibility can be identified.

## Competing interests

The authors declare that they have no competing interests.

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## Authors' contributions

Study designs, oversight, data interpretation, manuscript preparation: CL, ST, DS (Dr Socheat), YS, HN, DB, KS, WR, WS, MF, SC (Dr. Suwanna), AT, DS (Dr Saunders), DW Subject recruitment, interactions: DS (Dr Sea), YS, SC (Ms. Chann), NB, SS (Ms Sabaithip) Conducted experiments, summarized data: ST, KS, WR, SC, KY, SS (Ms Siratchana), PC. All authors read and approved the final manuscript.

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## References

- Desjardins RE, Canfield CJ, Haynes JD, Chulay JD: **Quantitative assessment of antimalarial activity *in vitro* by a semiautomated microdilution technique.** *Antimicrob Agents Chemother* 1979, **16**:710–718.
- Webster HK, Boudreau EF, Pavanand K, Yongvanitchit K, Pang LW: **Antimalarial drug susceptibility testing of *Plasmodium falciparum* in Thailand using a microdilution radioisotope method.** *Am J Trop Med Hyg* 1985, **34**:228–235.
- Wongsrichanalai C, Pickard AL, Wernsdorfer WH, Meshnick SR: **Epidemiology of drug-resistant malaria.** *Lancet Infect Dis* 2002, **2**:209–218.
- World Health Organization: *Global report on antimalarial efficacy and drug resistance: 2000–2010.* Geneva: WHO; 2010.
- Noedl H, Attlmayr B, Wernsdorfer WH, Kollaritsch H, Miller RS: **A histidine-rich protein 2-based malaria drug sensitivity assay for field use.** *Am J Trop Med Hyg* 2004, **71**:711–714.
- Noedl H, Bronnert J, Yingyuen K, Attlmayr B, Kollaritsch H, Fukuda M: **Simple histidine-rich protein 2 double-site sandwich enzyme-linked immunosorbent assay for use in malaria drug sensitivity testing.** *Antimicrob Agents Chemother* 2005, **49**:3575–3577.
- Noedl H, Se Y, Schaecher K, Smith BL, Socheat D, Fukuda MM: **Evidence of artemisinin-resistant malaria in western Cambodia.** *NEJM* 2008, **359**:2619–2620.
- Cui W: **WHO urges the phasing out of artemisinin based monotherapy for malaria to reduce resistance.** *BMJ* 2011, **342**:d2793.
- Incardona S, Vong S, Chiv L, Lim P, Nhem S, Sem R, Khim N, Doung S, Mercereau-Puijalon O, Fandeur T: **Large-scale malaria survey in Cambodia: novel insights on species distribution and risk factors.** *Malar J* 2007, **6**:37.
- Noedl H, Teja-Isavadharm P, Miller RS: **Nonisotopic, semiautomated *plasmodium falciparum* bioassay for measurement of antimalarial drug levels in serum or plasma.** *Antimicrob Agents Chemother* 2004, **48**:4485–4487.
- Akala HM, Eyase FL, Cheruiyot AR, Omondi AA, Ogutu BR, Waters NC, Johnson JD, Polhemus ME, Schnabel DC, Walsh DS: **Anti-malarial drug sensitivity profile of western Kenya *Plasmodium falciparum* field isolates determined by a SYBR Green I *in vitro* assay, and molecular analysis.** *Amer J Trop Med Hyg* 2011, **85**:34–41.
- Mbaisi A, Liyala P, Eyase F, Achilla R, Akala H, Wangui J, Mwangi J, Osuna F, Alam U, Smoak BL, Davis JM, Kyle DE, Coldren RL, Mason C, Waters NC: **Drug susceptibility and genetic evaluation of *Plasmodium falciparum* isolates obtained in four distinct geographical regions of Kenya.** *Antimicrob Agents Chemother* 2004, **48**:3598–3601.
- World Health Organization: *Mark III in vitro micro-test for the assessment of the response of *Plasmodium falciparum* to chloroquine, mefloquine, quinine, amodiaquine, sulfadoxine/pyrimethamine, and artemisinin.*; CTD/MAL/9720 Rev 2 2001.

14. Teja-Isavadharm P, Siriyanon D, Siripokasupkul R, Apinan R, Chanarat N, Lim A, Wannaying S, Saunders D, Fukuda MM, Miller RS, Weina PJ, Melendez V: **A simplified liquid chromatography-mass spectrometry assay for artesunate and dihydroartemisinin, its metabolite, in human plasma.** *Molecules* 2010, **15**:8747–8768.
15. Bethell D, Se Y, Lon C, Socheat D, Saunders D, Teja-Isavadharm P, Khemawoot P, Darapiseth S, Lin J, Sriwichai S, Kuntawungin W, Surasri S, Lee SJ, Sarim S, Tyner S, Smith B, Fukuda MM: **Dose-dependent risk of neutropenia after 7-day courses of artesunate monotherapy in Cambodian patients with acute *Plasmodium falciparum* malaria.** *Clin Infect Dis* 2010, **51**:e105–114.
16. ICEstimator (version 1.2), . Available from: <http://www.antimalarial-icestimator.net/MethodIntro.htm>.
17. Dondorp AM, Yeung S, White L, Nguon C, Day NP, Socheat D, von Seidlein L: **Artemisinin resistance: current status and scenarios for containment.** *Nat Rev Microbiol* 2010, **8**:272–280.
18. Bethell D, Se Y, Lon C, Tyner S, Saunders D, Sriwichai S, Darapiseth S, Teja-Isavadharm P, Khemawoot P, Schaecher K, Ruttvisutinunt W, Lin J, Kuntawungin W, Gosi P, Timmermans A, Smith B, Socheat D, Fukuda MM: **Artesunate dose escalation for the treatment of uncomplicated malaria in a region of reported artemisinin resistance: a randomized clinical trial.** *PLoS One* 2011, **6**:e19283.
19. Vijaykadge S, Rojanawatsirivej C, Cholpol S, Phoungmanee D, Nakavej A, Wongsrichanalai C: **In vivo sensitivity monitoring of mefloquine monotherapy and artesunate-mefloquine combinations for the treatment of uncomplicated falciparum malaria in Thailand in 2003.** *Trop Med Int Health* 2006, **11**:211–219.
20. Fall B, Diawara S, Sow K, Baret E, Diatta B, Fall KB, Mbaye PS, Fall F, Dieme Y, Rogier C, Wade B, Bercion R, Pradines B: **Ex vivo susceptibility of *Plasmodium falciparum* isolates from Dakar, Senegal, to seven standard anti-malarial drugs.** *Malar J* 2011, **10**:310.
21. Ndiaye D, Patel V, Demas A, LeRoux M, Ndir O, Mboup S, Clardy J, Lakshmanan V, Daily JP, Wirth DF: **A non-radioactive DAPI-based high-throughput in vitro assay to assess *Plasmodium falciparum* responsiveness to antimalarials—increased sensitivity of *P. falciparum* to chloroquine in Senegal.** *Am J Trop Med Hyg* 2010, **82**:228–230.
22. Hodel EM, Genton B, Zanolari B, Mercier T, Duong S, Beck HP, Olliaro P, Decosterd LA, Arie F: **Residual antimalarial concentrations before treatment in patients with malaria from Cambodia: indication of drug pressure.** *J Infect Dis* 2010, **202**:1088–1094.
23. Basco LK: *Field application of in vitro assays sensitivity of human malaria parasites antimalarial drugs.*: World Health Organization; 2007.
24. Wongsrichanalai C, Meshnick SR: **Declining artesunate-mefloquine efficacy against falciparum malaria on the Cambodia-Thailand border.** *Emerg Infect Dis* 2008, **14**:716–719.
25. Imwong M, Dondorp AM, Nosten F, Yi P, Mungthin M, Hanchana S, Das D, Phyto AP, Lwin KM, Pukrittayakamee S, Lee SJ, Saisung S, Koecharoen K, Nguon C, Day NP, Socheat D, White NJ: **Exploring the contribution of candidate genes to artemisinin resistance in *Plasmodium falciparum*.** *Antimicrob Agents Chemother* 2010, **54**:2886–2892.
26. Rueangweeraayut R, Phyto AP, Uthaisin C, Poravuth Y, Binh TQ, Tinto H, Penali LK, Valecha N, Tien NT, Abdulla S, Borghini-Fuhrer I, Duparc S, Shin CS, Fleckenstein L: **Pyronaridine-artesunate versus mefloquine plus artesunate for malaria.** *N Engl J Med* 2012, **366**:1298–1309.
27. Lim P, Wongsrichanalai C, Chim P, Khim N, Kim S, Chy S, Sem R, Nhem S, Yi P, Duong S, Bouth DM, Genton B, Beck HP, Gobert JG, Rogers WO, Coppee JY, Fandeur T, Mercereau-Pujjalon O, Ringwald P, Le Bras J, Arie F: **Decreased in vitro susceptibility of *Plasmodium falciparum* isolates to artesunate, mefloquine, chloroquine, and quinine in Cambodia from 2001 to 2007.** *Antimicrob Agents Chemother* 2010, **54**:2135–2142.
28. Chaijaroenkul W, Bangchang KN, Mungthin M, Ward SA: **In vitro antimalarial drug susceptibility in Thai border areas from 1998–2003.** *Malar J* 2005, **4**:37.
29. Suwandittakul N, Chaijaroenkul W, Harnyuttanakorn P, Mungthin M, Na Bangchang K: **Drug resistance and in vitro susceptibility of *Plasmodium falciparum* in Thailand during 1988–2003.** *Korean J Parasitol* 2009, **47**:139–144.
30. Chaijaroenkul W, Wisedpanichkij R, Na-Bangchang K: **Monitoring of in vitro susceptibilities and molecular markers of resistance of *Plasmodium falciparum* isolates from Thai-Myanmar border to chloroquine, quinine, mefloquine and artesunate.** *Acta Trop* 2010, **113**:190–194.
31. Dondorp AM, Nosten F, Yi P, Das D, Phyto AP, Tarning J, Lwin KM, Arie F, Hanpithakpong W, Lee SJ, Ringwald P, Silamut K, Imwong M, Chotivanich K, Lim P, Herdman T, An SS, Yeung S, Singhasivanon P, Day NP, Lindegardh N, Socheat D, White NJ: **Artemisinin resistance in *Plasmodium falciparum* malaria.** *N Engl J Med* 2009, **361**:455–467.

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