

# Human $\gamma\delta$ T lymphocytes for immunotherapeutic strategies against cancer

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## Abstract

$\gamma\delta$  T lymphocytes are a numerically small subset of T cells with potent cytotoxic activity against a variety of tumor cells. Human  $\gamma\delta$  T cells expressing the V $\gamma$ 9V $\delta$ 2 T cell antigen receptor recognize endogenous pyrophosphate molecules that are overproduced in transformed cells. Moreover, the intracellular accumulation of such pyrophosphates is strongly enhanced by aminobisphosphonates used in the treatment of osteoporosis and bone metastasis in certain cancer patients. A new concept of cancer immunotherapy is based on the endogenous activation of  $\gamma\delta$  T cells with aminobisphosphonates plus low-dose interleukin-2.

## Introduction and context

Approximately 1-5% of peripheral blood T cells express the  $\gamma\delta$  T-cell receptor instead of the conventional  $\alpha\beta$  T-cell receptor [1]. The  $\alpha\beta$  versus  $\gamma\delta$  T-cell lineage commitment during intrathymic T-cell development seems to be controlled by the signal strength provided to the  $\gamma\delta$  T-cell receptor [2]. In healthy donors, most blood  $\gamma\delta$  T cells carry a specific T-cell receptor composed of V $\gamma$ 9 and V $\delta$ 2 elements. In addition to effector functions shared with  $\alpha\beta$  T cells, the V $\gamma$ 9V $\delta$ 2 T cells can acquire professional antigen-presenting capacity characteristic of dendritic cells [3]. In contrast to  $\alpha\beta$  T cells, V $\gamma$ 9V $\delta$ 2 T cells do not see processed antigenic peptides presented by major histocompatibility complex molecules, but rather recognize small phosphorylated non-peptide molecules ('phosphoantigens') produced by many microorganisms but also by transformed eukaryotic cells [4,5]. While microbial phosphoantigens are active at pico- to nanomolar concentrations, micromolar concentrations of the eukaryotic phosphoantigen isopentenyl pyrophosphate (IPP) are required for  $\gamma\delta$  T-cell activation. Such high concentrations are not achieved in the mevalonate pathway of isoprenoid synthesis used in non-transformed cells. Interestingly, human V $\gamma$ 9V $\delta$ 2 T cells can kill a broad variety of epithelial tumor and

leukemia/lymphoma cells [6,7]. The sensitivity of tumor cells to  $\gamma\delta$  T-cell-mediated killing is increased upon treatment of tumor cells with aminobisphosphonates (N-BPs), drugs that are used in clinical practice for the treatment of osteoporosis and bone metastasis in cancer patients [8]. N-BPs inhibit the IPP-processing enzyme farnesyl diphosphate synthase (FPPS), thereby leading to an accumulation of IPP, which is then sensed by the  $\gamma\delta$  T cells [9].  $\gamma\delta$  T cells are poor producers of interleukin-2 (IL-2), which is required for expansion of  $\gamma\delta$  T cells. Therefore, attempts to activate tumor-reactive  $\gamma\delta$  T cells endogenously by treating patients with N-BPs must take into consideration an appropriate supply of IL-2. Alternative strategies consider the adoptive transfer of *in vitro* expanded tumor-reactive  $\gamma\delta$  T cells [10-14].

## Recent advances

The critical role of FPPS in the control of intracellular IPP levels, and thus of the sensitivity of tumor cells toward  $\gamma\delta$  T-cell killing, has been recently demonstrated using short hairpin RNA-mediated knock-down of FPPS [15]. Knock-down of FPPS caused tumor cells, which otherwise were not recognized by  $\gamma\delta$  T cells, to be susceptible to  $\gamma\delta$  T-cell killing [15]. Therefore, V $\gamma$ 9V $\delta$ 2 T cells recognize and kill tumor cells on the basis of the unbalanced isoprenoid

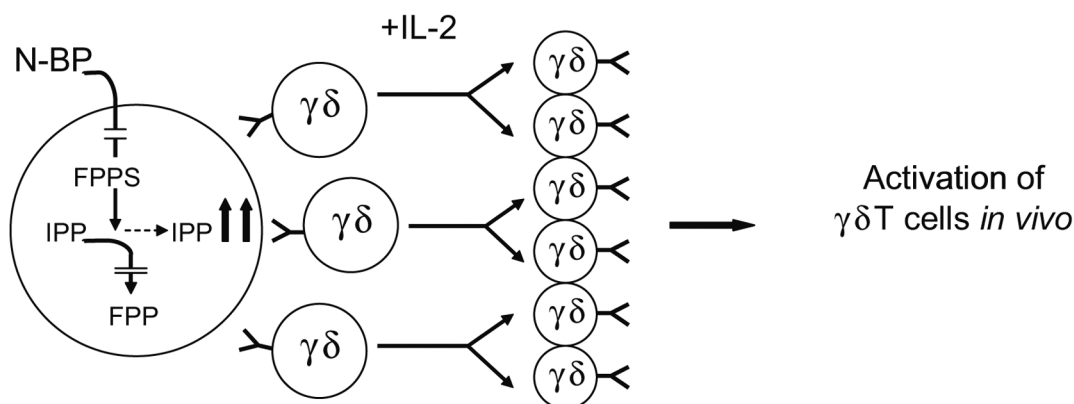
metabolic pathway in transformed cells, a pathway that is stable in non-malignant cells.

The discovery that N-BPs activate  $\gamma\delta$  T cells by inhibiting FPPS, thereby leading to accumulation of IPP, has paved the way for proof-of-principle studies to activate  $\gamma\delta$  T cells in patients with advanced cancer. In a phase I clinical trial, Dieli and colleagues [16] treated patients with hormone-refractory prostate cancer with a standard application of the N-BP zoledronate (4 mg intravenous infusion every 21 days) either with or without additional low-dose ( $6 \times 10^5$  IU) subcutaneous application of IL-2. Various parameters, including subset analysis of  $\gamma\delta$  T cells, and serum levels of prostate-specific antigen and cytokines, were monitored over time. Although the two cohorts comprised only a few patients, statistically significant

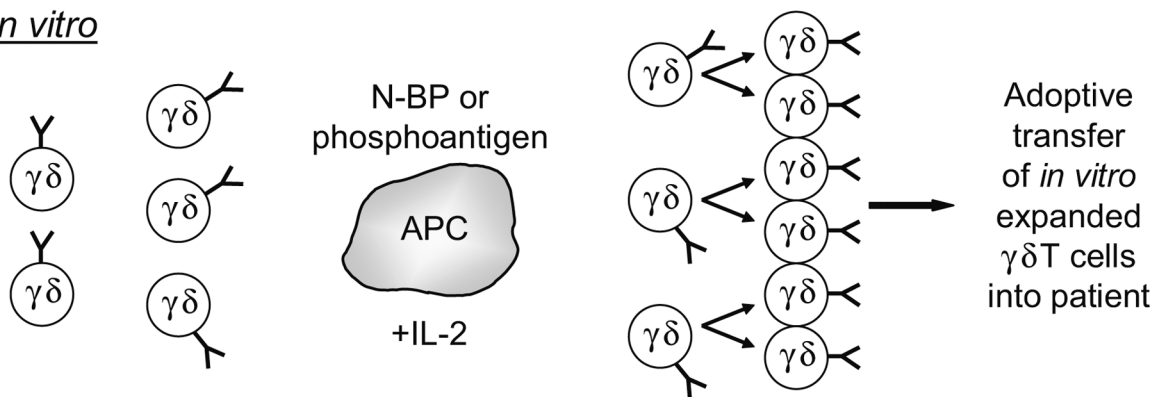
effects of zoledronate plus IL-2 on the mobilization and effector cell maturation of  $\gamma\delta$  T cells were recorded. Very importantly, the two cohorts showed distinct clinical outcomes, with clinical responses seen in six of nine patients treated with zoledronate plus IL-2 but only in one of nine patients treated with zoledronate alone. Interestingly, a correlation between favorable outcome at 12 months and  $\gamma\delta$  T-cell numbers or functional status (or both) was observed [16]. Similarly, Wilhelm and colleagues had previously shown that the combined application of N-BP plus low-dose IL-2 can induce objective tumor responses in patients with lymphoid malignancies [17]. Together, these studies support the view that application of N-BPs plus IL-2 is safe, induces *in vivo* activation/maturation of  $\gamma\delta$  T cells, and may have beneficial effects in advanced cancer (Figure 1a).

Figure 1. Approaches to immunotherapy with  $\gamma\delta$  T cells

**(a) *in vivo***



**(b) *in vitro***



(a) N-BPs inhibit farnesyl diphosphate synthase (FPPS), thus preventing processing of isopentenyl pyrophosphate (IPP) to farnesyl diphosphate (FPP). This leads to accumulation of IPP which then activates V $\gamma$ 9V $\delta$ 2 T cells. V $\gamma$ 9V $\delta$ 2 T cells require exogenous interleukin (IL)-2 for cellular expansion. The combined application of N-BP plus IL-2 leads to *in vivo* activation of  $\gamma\delta$  T cells. (b) Alternatively,  $\gamma\delta$  T cells can be activated *in vitro* with N-BP or synthetic phosphoantigens in the presence of antigen-presenting cells (APC) and can be subsequently expanded to large cell numbers by an exogenous supply of IL-2 for subsequent adoptive transfer into cancer patients. The cell preparation can be performed under GMP (Good Manufacturing Practice) conditions.

In a case study reported by Laggner *et al.* [18], regression of lung and bone metastases was observed in a patient with advanced stage melanoma upon systemic treatment with zoledronate and localized radiotherapy. Although  $\gamma\delta$  T-cell subsets were analyzed, it is difficult to ascertain a substantial role of  $\gamma\delta$  T-cell activation in the resolution of metastases in this single case, particularly since IL-2 was omitted in the treatment of this patient [18].

In addition to their  $\gamma\delta$  T-cell activating properties, N-BPs also exhibit direct anti-tumor activities by both inhibiting proliferation and inducing apoptosis in tumor cells [19]. While zoledronate seems to be the most potent  $\gamma\delta$  T-cell-activating substance among the N-BPs licensed for clinical application [9], derivatives of zoledronate with further improved  $\gamma\delta$  T-cell-stimulating capacity and enhanced direct anti-tumor activity are under development [20]. Such modified N-BPs might also exert improved *in vivo* activation of  $\gamma\delta$  T cells when given to patients together with IL-2.

An alternative and not mutually exclusive  $\gamma\delta$  T-cell-based immunotherapeutic strategy is the adoptive transfer of *in vitro* expanded V $\gamma$ 9V $\delta$ 2 T cells from tumor patients (Figure 1b). Recently, efficient protocols for the large-scale *in vitro* expansion of V $\gamma$ 9V $\delta$ 2 T cells based on stimulation with synthetic phosphoantigens [11,13,14] or zoledronate [21] have been established. First results indicate that the repetitive adoptive transfer of *in vitro* expanded  $\gamma\delta$  T cells is well tolerated and may induce anti-tumor responses in patients with solid tumors, including renal cell carcinoma [13,14] and myeloma [12].

### Implications for clinical practice

The protocol developed by Dieli *et al.* [16] for the *in vivo* activation of  $\gamma\delta$  T cells based on zoledronate plus low-dose IL-2 application is ready to be explored in larger clinical trials and in other tumor entities with poor prognosis, for example, pancreatic ductal adenocarcinoma where it might be combined with standard regimens such as gemcitabine. It is conceivable that this protocol can be further improved, for instance, by combination with tumor-targeting monoclonal antibodies. Along this line, it has been shown that B-cell lymphoma or breast tumor cell killing by Fc $\gamma$  receptor-expressing  $\gamma\delta$  T cells is enhanced in the presence of targeting antibodies rituximab (CD20) or trastuzumab (HER2/neu), respectively [22]. Moreover, a  $\gamma\delta$  T-cell-stimulating synthetic phosphoantigen was found to enhance the depletion of CD20<sup>+</sup> B cells by rituximab in a non-human primate model *in vivo*, pointing to the possible use of phosphoantigen plus anti-CD20 antibodies in the treatment of CD20<sup>+</sup> leukemias and lymphomas [23]. Furthermore, cytokines promoting

homeostatic proliferation and survival of T cells, such as IL-15 [24], or cytokines potentiating the cytolytic activity and pro-inflammatory response, such as IL-21 [25], might be combined with IL-2 or used instead of IL-2. This could be considered both for *in vivo* application together with N-BPs and for optimization of *in vitro* expansion of  $\gamma\delta$  T cells. In addition, future study protocols might include the combination of *in vivo* activation of  $\gamma\delta$  T cells (by N-BP or phosphoantigen plus IL-2) followed by the adoptive transfer of *in vitro* expanded  $\gamma\delta$  T cells. Finally, it should be stressed that  $\gamma\delta$  T-cell-based immunotherapy is not expected to replace established therapeutic protocols. Rather, it might offer additional benefit to the patient, for instance, in combination with conventional chemotherapy [26].

### Abbreviations

FPPS, farnesyl diphosphate synthase; IL, interleukin; IPP, isopentenyl pyrophosphate; N-BP, aminobisphosphonate.

### Competing interests

The author declares that he has no competing interests.

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### References

- Hayday A:  $\gamma\delta$  T cells: a right time and a right place for a conserved third way of protection. *Annu Rev Immunol* 2000, **18**:975-1026.
  - Hayes SM, Love PE: TCR signal strength influences  $\alpha\beta/\gamma\delta$  lineage fate. *Immunity* 2005, **22**:583-93.
- F1000 Factor 3.0 Recommended  
Evaluated by Harald von Boehmer 02 Jun 2005
- Brandes M, Willimann K, Moser B: Professional antigen-presentation function by human  $\gamma\delta$  T cells. *Science* 2005, **309**:264-8.
- F1000 Factor 8.4 Exceptional  
Evaluated by Pedro Romero 29 Jun 2005, Dan Conrad 21 Jul 2005, Stephen Jameson 09 Aug 2005, Peter Van Endert 16 Aug 2005
- Morita CT, Jin C, Sarikonda G, Wang H: Nonpeptide antigens, presentation mechanisms, and immunologic memory of human V $\gamma$ 2V $\delta$ 2 T cells: discriminating friend from foe through the recognition of prenylpyrophosphate antigens. *Immunol Rev* 2007, **215**:59-76.
  - Kabelitz D: Small molecules for the activation of human  $\gamma\delta$  T cell responses against infection. *Recent Pat Anti-Infect Drug Discov* 2008, **3**:1-9.
  - Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M: Stimulation of  $\gamma\delta$  T cells by aminobisphosphonates and induction of antiplasma cell activity in multiple myeloma. *Blood* 2000, **96**:384-92.
  - Wrobel P, Shojaei H, Schitteck B, Gieseler F, Wollenberg B, Kalthoff H, Kabelitz D, Wesch D: Lysis of a broad range of epithelial tumour cells by human  $\gamma\delta$  T cells: Involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. *Scand J Immunol* 2007, **66**:320-8.

8. Gober HJ, Kistowska M, Angman L, Jenö P, Mori L, de Libero G: **Human T cell receptor  $\gamma\delta$  T cells recognize endogenous mevalonate metabolites in tumor cells.** *J Exp Med* 2003, **197**:163-8.
9. Thompson K, Roelofs AJ, Jauhainen M, Mönkkönen H, Mönkkönen J, Rogers MJ: **Activation of  $\gamma\delta$  T cells by bisphosphonates.** *Adv Exp Med Biol* 2010, **658**:11-20.
10. Kabelitz D, Wesch D, He W: **Perspectives of  $\gamma\delta$  T cells in tumor immunology.** *Cancer Res* 2007, **67**:5-8.
11. Salot S, Bercegeay S, Dreno B, Saiagh S, Scaglione V, Bonnafous C, Sicard H: **Large scale expansion of V $\gamma$ 9V $\delta$ 2 T lymphocytes from human peripheral blood mononuclear cells after a positive selection using MACS 'TCR  $\gamma/\delta$ ' T cell isolation kit'. *J Immunol Methods* 2009, **347**:12-8.**
12. Abe Y, Muto M, Nieda M, Nakagawa Y, Nicol A, Kaneko T, Goto S, Yokokawa K, Suzuki K: **Clinical and immunological evaluation of zoledronate-activated V $\gamma$ 9 $\gamma\delta$  T-cell-based immunotherapy for patients with multiple myeloma.** *Exp Hematol* 2009, **37**:956-68.
13. Kobayashi H, Tanaka Y, Yagi J, Osaka Y, Nakazawa H, Uchiyama T, Minato N, Toma H: **Safety profile and anti-tumor effects of adoptive immunotherapy using gamma-delta T cells against advanced renal cell carcinoma: a pilot study.** *Cancer Immunol Immunother* 2007, **56**:469-76.
14. Bennouna J, Bompas E, Neidhardt EM, Rolland F, Philip I, Galéa C, Salot S, Saiagh S, Audrain M, Rimbert M, Lafaye-de Micheaux S, Tiollier J, Négrier S: **Phase-I study of Innacell  $\gamma\delta$ ™, an autologous cell-therapy product highly enriched in  $\gamma\delta$ 2 T lymphocytes, in combination with IL-2, in patients with metastatic renal cell carcinoma.** *Cancer Immunol Immunother* 2008, **57**:1599-609.
15. Li J, Herold MJ, Kimmerl B, Müller I, Rincon-Orozco B, Kunzmann V, Herrmann T: **Reduced expression of the mevalonate pathway enzyme farnesyl pyrophosphate synthase unveils recognition of tumor cells by V $\gamma$ 9V $\delta$ 2 T cells.** *J Immunol* 2009, **182**:8118-24.
16. Dieli F, Vermijlen D, Fulfarò F, Caccamo N, Meraviglia S, Cicero G, Roberts A, Buccheri S, D'Asaro M, Gebbia N, Salerno A, Eberl M, Hayday A: **Targeting human  $\gamma\delta$  T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer.** *Cancer Res* 2007, **67**:7450-7.
17. Wilhelm M, Kunzmann V, Eckstein S, Reimer P, Weissinger F, Ruediger T, Tony HP:  **$\gamma\delta$  T cells for immune therapy of patients with lymphoid malignancy.** *Blood* 2003, **102**:200-6.
18. Laggner U, Lopez JS, Perera G, Warbey VS, Sita-Lumsden A, O'Doherty MJ, Hayday A, Harries M, Nestle FO: **Regression of melanoma metastases following treatment with the n-bisphosphonate zoledronate and localized radiotherapy.** *Clin Immunol* 2009, **131**:367-73.
19. Clezardin P, Ebertino FH, Fournier PG: **Bisphosphonates and cancer-induced bone disease: beyond their anti-resorptive activity.** *Cancer Res* 2005, **65**:4971-4.
20. Simoni D, Gebbia N, Invidiata FP, Eleopra M, Marchetti P, Rondanin R, Baruchello R, Provera S, Marchioro C, Tolomeo M, Marinelli L, Limongelli V, Novellino E, Kwaasi A, Dunford J, Buccheri S, Caccamo N, Dieli F: **Design, synthesis, and biological evaluation of novel aminobisphosphonates possessing an in vivo anti-tumor activity through a  $\gamma\delta$ -T lymphocytes-mediated activation mechanism.** *J Med Chem* 2008, **51**:6800-7.
21. Kondo M, Sakuta K, Noguchi A, Ariyoshi N, Sato K, Sato S, Sao K, Hosoi A, Nakajima J, Yoshida Y, Shiraishi K, Nakagawa K, Kakimi K: **Zoledronate facilitates large-scale ex vivo expansion of functional  $\gamma\delta$  T cells from cancer patients for use in adoptive immunotherapy.** *Cytotherapy* 2008, **10**:842-56.
22. Tokuyama H, Hagi T, Mattarollo SR, Morley J, Wang Q, Fai-So H, Moriyasu F, Nieda M, Nicol AJ: **V $\gamma$ 9V $\delta$ 2 T cell cytotoxicity against tumor cells is enhanced by monoclonal antibody drugs - rituximab and trastuzumab.** *Int J Cancer* 2008, **122**:2526-34.
23. Gertner-Dardenne J, Bonnafous C, Bezombes C, Capietto AH, Scaglione V, Ingoure S, Cendron D, Gross E, Lepage JF, Quillet-Mary A, Ysebaert L, Laurent G, Sicard H, Fournié JJ: **Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies.** *Blood* 2009, **113**:4875-84.
24. Boyman O, Létourneau S, Krieg C, Sprent J: **Homeostatic proliferation and survival of naïve and memory T cells.** *Eur J Immunol* 2009, **39**:2088-94.
25. Thedrez A, Harly C, Morice A, Salot S, Bonneville M, Scotet S: **IL-21-mediated potentiation of antitumor cytolytic and proinflammatory responses of human V $\gamma$ 9V $\delta$ 2 T cells for adoptive immunotherapy.** *J Immunol* 2009, **182**:3423-31.
26. Mattarollo SR, Kenna T, Nieda M, Nicol AJ: **Chemotherapy and zoledronate sensitize solid tumour cells to V $\gamma$ 9V $\delta$ 2 T cell cytotoxicity.** *Cancer Immunol Immunother* 2007, **56**:1285-97.

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