

Cold Atmospheric Plasma Ameliorates Skin Diseases Involving Reactive Oxygen/Nitrogen Species-Mediated Functions

Si-yue Zhai^{1,2}, Michael G. Kong^{2,3*} and Yu-min Xia^{1*}

¹ Department of Dermatology, The Second Affiliated Hospital of Xi'an Jiaotong University, Xi'an, China, ² Center of Plasma Biomedicine, State Key Laboratory of Electrical Insulation and Power Equipment, Xi'an Jiaotong University, Xi'an, China, ³ School of Electrical Engineering, Xi'an Jiaotong University, Xi'an, China

OPEN ACCESS

Edited by:

Kai Fang, University of California, Los Angeles, United States

Reviewed by:

Chih-Yao Hou, National Kaohsiung University of Science and Technology, Taiwan Lia-Mara Ditu, University of Bucharest, Romania Xiaofeng Dai, Jiangnan University, China

*Correspondence:

Michael G. Kong mglin5g@gmail.com Yu-min Xia xiayumin1202@163.com

Specialty section:

This article was submitted to Inflammation, a section of the journal Frontiers in Immunology

Received: 02 February 2022 Accepted: 09 May 2022 Published: 26 May 2022

Citation:

Zhai S-y, Kong MG and Xia Y-m (2022) Cold Atmospheric Plasma Ameliorates Skin Diseases Involving Reactive Oxygen/Nitrogen Species-Mediated Functions. Front. Immunol. 13:868386. doi: 10.3389/fimmu.2022.868386 Skin diseases are mainly divided into infectious diseases, non-infectious inflammatory diseases, cancers, and wounds. The pathogenesis might include microbial infections, autoimmune responses, aberrant cellular proliferation or differentiation, and the overproduction of inflammatory factors. The traditional therapies for skin diseases, such as oral or topical drugs, have still been unsatisfactory, partly due to systematic side effects and reappearance. Cold atmospheric plasma (CAP), as an innovative and non-invasive therapeutic approach, has demonstrated its safe and effective functions in dermatology. With its generation of reactive oxygen species and reactive nitrogen species, CAP exhibits significant efficacies in inhibiting bacterial, viral, and fungal infections, facilitating wound healing, restraining the proliferation of cancers, and ameliorating psoriatic or vitiligous lesions. This review summarizes recent advances in CAP therapies for various skin diseases and implicates future strategies for increasing effectiveness or broadening clinical indications.

Keywords: cold atmospheric plasma, reactive oxygen species, reactive nitrogen species, skin disease, therapy

INTRODUCTION

It is widely accepted that there are various kinds of skin diseases, including infectious diseases, cancers, wounds, and non-communicable inflammatory skin diseases (ncISDs). The pathogenic factors of these skin disorders include microbial infections, excessive cytokines release, inflammatory cell infiltration, and aberrant cell proliferation or differentiation (1–3). Currently, the comprehensive treatments include systemic or topical medications or in combination with different phototherapies. However, these applications have still been challenged due to systemic side effects, the high recurrence rate of the disease, or the serious impact on daily life or working (4). As an innovative non-invasive application, cold atmospheric plasma (CAP) has exhibited significant efficacy in treating skin diseases such as microbial infections, cutaneous wounds, cancers, and ncISDs (5, 6). It is well clarified that the biological functions of CAP involve ultra-violet radiation, reactive oxygen species, and reactive nitrogen species while exhibiting little damage to the normal cells or tissues (7). CAP may destroy biofilms or even the integrity of cells, degrade cellular proteins and DNA, generate DNA and protein crosslinking, and dysregulate pH value- or calcium-related

1

microenvironments (7–11). In this review, we summarize recent advances in the applications of CAP in treating skin diseases and possible therapeutic mechanisms.

THE CHARACTERISTICS OF CAP

Containing with the positive and negative changed species, plasma is the fourth state of matter, which is different from the common three substances (solid, liquid, and gas) (12). According to the gas temperature of plasma, the plasma generated by different methods can be divided into hot plasm, warm plasma, and cold plasma. Because high temperature can cause thermal damage to organisms, the plasma applied to medical treatment should be considered a kind of plasma at atmospheric pressure with its temperature close to room temperature (13). CAP has been used in medical treatments widely (6). As **Figure 1** described, there are two common devices for CAP generators, dielectric barrier discharges (DBD) and atmospheric pressure plasma jet (APPJ). Surface DBD is consisted of two parallel electrodes sandwiching a ceramic slab to strike a surface discharge plasma at a high voltage in a narrow discharge gap (7). The other is the APPJ, consisting of a quartz capillary equipped with a pin-type electrode at a high voltage generated in an open space (12). In order to improve the efficiency of CAP, helium (He), argon (Ar), nitrogen (N₂), oxygen (O₂), artificial air, and two or more mixtures of these gases can be utilized to generate CAP (14). It is documented that reactive oxygen species (ROS) (15) or reactive nitrogen species (RNS) (16), play an essential role in CAP application in dermatology. Figure 2 has shown the interconverting ROS and RNS that could be classified into short-lived and long-lived species, such as singlet oxygen (¹O₂), superoxide anions (O₂⁻), hydroxyl radicals ([•]OH), hydrogen peroxide (H₂O₂), nitric oxide (NO), peroxynitrite (ONOO⁻), nitrite (HNO₂/NO₂⁻) and nitrate (HNO₃/NO₃⁻). $^{1}O_{2}$ is a highly reactive molecule that participates in apoptosis processes and produces some oxidants and cytotoxic molecules. O₂ has relatively high reactivity and results in oxidative stress in cells to generate hydroxyl radicals. It is displayed that 'OH belongs to extremely reactive oxidizing





FIGURE 2 | The internal structure of surface DBD and APPJ and the transformation of CAP particles. The internal structures of **(A)** surface DBD and **(B)** APPJ were consisted of electrodes, high-voltage probes, quartz tubes, and a dielectric layer. The **(C)** RNS and **(D)** RNS generated under high voltage could be converted into different molecules or ions and finally synthesized into effective particles like ¹O₂, ONOO⁻, NO₂⁻, and H₂O₂.

species, which can damage the lipid peroxidation, protein, membrane and result in cell death. As for ONOO⁻, it reacted with H_2O_2 to generate 1O_2 , which inhibited the catalase activity of tumor cells, resulting in the accumulation of H₂O₂ in the part of the peripheral environment. Then H₂O₂ entered tumor cells through aquaporin on the cell surface and consumed internal glutathione, which could promote the apoptosis of tumor cells via lipid peroxidation (17, 18). Besides, as a long-lived species and second messenger for signaling cascades, H₂O₂ can inhibit the activity of microbes and kill them directly (19). It has been documented that CAP-induced changes in target cells are often closely related to intervention time, voltage, and other factors. For example, CAP could induce cell cycle arrest, senescence, and autophagy at low doses, but high doses of CAP could lead to irreversible cell necrosis (20). Moreover, RNS has been confirmed to regulate cell differentiation, self-renewal, and the balance between quiescence and proliferation (21). The occurrence and development of tumors can be influenced by NO concentration, which can accelerate tumor growth in the low-level concentration but inhibit it at high levels (22). The typical functions of ROS and RNS have been presented in Figure 3.

ROS AND RNS INACTIVATED MICROORGANISMS BY DESTROYING CELL BIOFILMS AND COMPONENTS

It is well displayed that CAP has shown promising effects in inactivating microorganisms (**Figure 4**). CAP can inhibit the proliferation of microorganisms on the surface of medical devices but prohibit the infection of various microorganisms, including *Staphylococcus aureus*, *Streptococcus pyogenes*, *Pseudomonas aeruginosa*, *Yersinia enterocolitica*, MRSA, and MRGN (8, 23, 24). In addition, related studies have reported that CAP has satisfactory efficacy in treating the pathogens of infectious skin diseases, such as *Trichophyton rubrum*, *Mycobacterium canis*, and *Candida albicans* (**Table 1**) (8, 25). In this section, we will explore the inactivation mechanisms of CAP against bacteria, fungi, and viruses, respectively.

Bacteria

Generally speaking, bacterial biofilms have widely existed on the surfaces of various foods, medical devices, and human tissues or organs under pathological conditions. Due to the widespread use



FIGURE 3 | The typical functions of ROS and RNS in CAP. Various kinds of effective particles, including ROS and RNS, are divided into short-lived or long-lived substances, which display different mechanisms in treating skin diseases by CAP.

of antibiotics, opportunistic pathogen infections caused by bacterial biofilms have gradually increased. Once a biofilm is formed, the organism will resist the action of antibiotics, making it difficult for antibiotics to remove bacterial biofilms or kill isolated bacteria on the organism's surface. Due to its antibiofilm properties, CAP has been widely used in sterilization (26, 27). First, CAP can accelerate the death of bacteria via wrecking bacterial biofilms. Theinkom used SMD to treat Enterococcus faecalis to investigate the potential effects of CAP on microbial biofilms with 3.5 kV, 4.0 kHz, and 0.5-1 W/cm². And the results showed that CAP treatment for 1 minute could inactivate bacterial biofilms and cause bacterial death directly. Moreover, a 10-min CAP intervention showed higher inactivating efficiency than a 5-min intervention (28). Utilizing a DBD with helium gas, Govaert treated the biofilms of L. monochromous and S. typhimurium for different minutes to explore the sterilization effects and mechanisms of CAP application by detecting the density of remaining cells (29). Scientists have confirmed the inactivating effects of CAP on bacterial biofilms formed by bacteria, such as S. bacteria, P. aeruginosa, K. pneumoniae, E. faecalis, S. aureus, L. monochromous, and P. fluorescens (30-33). Guo and her teams utilized a 6-min CAP intervention to treat S. aureus to explore the mechanisms of CAP inactivating microbial biofilms. Their results showed that ROS or RNS from CAP could result in biofilm lysis and bacterial death by promoting the oxidative modification of bacterial nucleoproteins, accelerating the filamentous temperature-sensitive Z of *S. aureus* (SaFtsZ) to lose its assembly ability, and disrupting the activity of ATPdependent caseinolytic protease P subunit of *S. aureus* (SaClpP) (34). In conclusion, CAP can expand the ROS/RNS binding area, enhance the oxidative modification of cellular proteins and promote the fusion of microbial biofilms with extracellular matrix components, which triggers the degradation of microbial biofilms and inhibits the proliferation of microorganisms.

Second, the changes in cell composition and microenvironment also affect bacterial inactivation. After DBD treatment, Helmke confirmed that the extracellular environment of bacteria was significantly acidified, and the cell membrane of bacteria gradually collapsed. After a while, the cell membrane of bacteria was destroyed, and its DNA was rapidly broken down (11). The destruction of bacterial DNA is the cause of bacterial inactivation, but the modification of bacterial proteins is considered a significant cause of bacterial inactivation. After DBD treatment, Helmke confirmed that the extracellular environment of bacteria was significantly acidified, and the cell membrane of bacteria gradually collapsed. After a while, the cell membrane of bacteria was



destroyed, and its DNA was rapidly broken down (11). Although the exact type of DNA damage has not been identified, it is well documented that ROS and RNS from CAP can destroy bacterial DNA directly (35, 36). The destruction of bacterial DNA is the cause of bacterial inactivation, but the modification of bacterial proteins is also considered a significant cause of bacterial inactivation. Dezest's study aimed to explore the interactions between active substances from CAP and the activity and proliferation of E. coli. Their results demonstrated that CAP could produce bacterial inactivation through selective carbonylation of proteins (37). Of course, CAP can also produce the interaction of DNA and proteins to inactivate bacteria. Guo, with her faculties, indicated that the CAP could generate DNA and protein crosslinking (DPC) in vitro. They used a DBD device to intervene in E. coli with the gas mixture of helium and artificial air. By analysis of mass spectrometry and hydroperoxide, the results indicated that the formation of DPC caused by ¹O₂ from CAP was the principal mechanism of DNA damage (7). In addition, other studies also documented that the shape of bacteria and the thickness of the bacterial cell wall might also be relevant to the CAP inactivation of bacteria (38-40). Taken together, CAP could inhibit bacterial infection by attacking the bacterial biofilms, destroying the integrity of the cell membrane, and inducing the selective carbonylation protein, DNA degradation, and protein crosslinking. The phenomenon could be explained by the mutual transformation and synergy of the ROS/RNS by CAP.

The inactivation of CAP for the main pathogenic bacteria in the skin has been shown in **Table 2**.

Fungus and Virus

A series of chemical processes based on ROS and RNS have been considered the main reason for the destruction of fungal biofilm (40). Fricke, with his faculties, confirmed that the biofilm of C. albicans could be wrecked by CAP (41). Heinlin et al. used CAP application to assess its efficacy against the dermatophytes such as T. rubrum and M. canis in vitro. The fungal growth was then measured after CAP treatment. The results proved that CAP exhibited efficient effects on the decontamination of T. rubrum and M. canis (42). Ouf et al. used CAP to treat five types of skin fungus- E. floccosum, M. canis, M. gypseum, and T. rubrum. They found that the fungal cell wall had been damaged under the scanning electron microscope, which indicated that CAP exhibited therapeutic effects in inhibiting fungal growth. Furthermore, they also indicated that the CAP could ameliorate symptoms of fungal skin diseases by the generation of NO, which could inhibit the activity of fungal spores via reducing the activity of keratinase and increasing the mycelium permeability (43). Generally speaking, ROS and RNS from CAP could destroy fungal spores and the integrity of cell membrane to inhibit the growth of fungus.

TABLE 1 | The mechanisms of CAP for inactivating pathogens.

Bacteria	Time	Device	Gas	Target
E. faecalis	2019	DBD	0 ₂	Biofilm
L. monochromous	2019	DBD	He	Biofilm
S. typhimurium	2019	DBD	He	Biofilm
S. bacteria	2014	APPJ	Air	Biofilm
P. aeruginosa	2014	APPJ	Air	Biofilm
K. pneumonia	2014	APPJ	Air	Biofilm
S. aureus	2014	APPJ	He	Biofilm
	2012	DBD	Air	Membrane
P. fluorescens	2014	APPJ	He	Biofilm Membrane
	2012	DBD	Air	
S. epidermidis	2011	DBD	Air	Biofilm
				DNA
E. coli	2018	DBD	He + air	Protein
	2017	APPJ	He + O_2	DNA
			-	DPC
Fungus	Time	Device	Gas	Target
C. albicans	2012	APPJ	Argon	Biofilm
			Argon $+ O_2$	
T. rubrum	2013	DBD	Air	Biofilm
M. canis	2015	APPJ	Air	Biofilm
	2013	DBD	Air	Protein
				Spores
E. floccosum	2015	APPJ	Air	Protein
				Spores
M. gypseum	2015	APPJ	Air	Protein
				Spores
Visus	Time	Device	Gas	Target
hAV	2018	DBD	Argon	Protein
			0	DNA
COVID-19	2021	APPJ	He	Ca ²⁺
	2022	APP.I	Argon	ACE2

E. faecalis, Enterococcus faecalis; L. monochromous, Listeria monochromous; S. typhimurium, Salmonella typhimurium; S. bacteria, Salmonella bacteria; P. aeruginosa, Pseudomonas aeruginosa; K. pneumonia, Klebsiella pneumonia; S. aureus, Staphylococcus aureus; P. fluorescens, Pseudomonas fluorescens; S.epidermidis, Staphylococcus epidermidis; E. coli, Escherichia coli; DPC, DNA and protein crosslinking; C. albicans, Candida albicans; T. rubrum, Trichophyton rubrum; M. canis, Microsporum canis; E. floccosum, Epidermophyton floccosum; M. gypseum, Microsporum gypseum; hAV, Human adenovirus; COVID-19, coronavirus disease-19; ACE2, angiotensin-converting enzyme 2; DBD, dielectric barrier discharge; APPJ, atmospheric pressure plasma jet.

TABLE 2 | The inactivation of CAP for the main skin pathogenic microorganisms.

Microorganism	Skin disease	Skin lesion	Device	Target
S. aureus	Impetigo	VesiclesBullousBlisters	APPJDBD	Biofilm
C. albicans	Tinea	ItchyWhite PlaquesErythema	APPJ	Biofilm
T. rubrum	Tinea capitis, Tinea corporis, Tinea manus	ErythemaScalyItchy	DBD	Biofilm
M. canis	Tinea capitis, Tinea corporis, Tinea manus	ErythemaScalyItchy	APPJDBD	Biofilm Protein Spores
M. gypseum	Tinea capitis, Tinea manus	ErythemaScalyItchy	APPJ	Protein Spores

Staphylococcus aureus, S. aureus; Candida albicans, C. albicans; Trichophyton rubrum, T. rubrum; Microsporum canis, M. canis; Microsporum gypseum, M. gypseum; DBD, dielectric barrier discharge; APPJ, atmospheric pressure plasma jet.

Initially, research on whether CAP could effectively inactivate human pathogenic virus was mainly based on the studies conducted in different phage models (8). Yasuda et al. studied the effects of CAP treatment on λ phage. They utilized the DBD device to treat λ phage for 20 seconds. Then they found that due to the rapid denaturation of its protein, the virus was inactivated totally. Their study noted that CAP inactivated viral proteins through denaturation or chemical/physical modification (44). Aboubakr et al. supported that the degradation of viral capsid protein might be the primary mechanism by which CAP caused viral inactivation. Their study showed that CAP treatment for 2 min could wreck the capsid protein (45). From the studies mentioned above, the degradation of viral protein could be the responsible mechanism for CAPdependent virus inactivation. Compared with protein degradation, was the mechanism by which CAP inactivated viruses included the destruction of viral DNA or RNA by CAP? Guo utilized CAP to treat water-containing bacteriophages T4, Φ 174, and MS2. According to the analysis of the genetic materials of bacteriophages, CAP could destroy both their nucleic acids and proteins *via* ${}^{1}O_{2}$ (46). Besides the viral protein and DNA/RNA, the unique structure "envelope" should also be considered as the third potential mechanism. CAP is more conducive to suppressing nonenveloped viruses (47). HAV is a kind of non-enveloped dsDNA virus. Zimmermann et al. studied the inactivating effect of CAP on HAV. After CAP treatment for 240 seconds, they found that the virus was inactivated efficiently. Moreover, before infecting CMS-5 cells, HAV, eGFP, and firefly luciferase, were treated by CAP. The results showed that the infectivity and replication of HAV were inhibited directly. They believed that the effect of inactivating HAV was mainly due to the RNS from CAP, which could destroy viral proteins and DNA (48). Virus infection of eukaryotic cells was complicated, including multiple signaling mechanisms inside and outside (49). The COVID-19 virus has spread worldwide in the past two years, and people's health is facing a considerable threat (50). To explore the interaction between CAP and COVID-19 virus, we selected surface discharge plasma to inactivate the COVID-19 pseudovirus. We utilized CAP with 8.36 kV, 23 kHz, and 0.25 W/ cm² to treat deionized water to generate solutions including ROS and RNS. Then a COVID-19 pseudovirus incorporated with SARS-CoV-2S protein was recognized as the virus model. The results showed that the short-lived species from CAP-activated deionized water, such as ONOO⁻ and O₂⁻, could modify the amino acids in proteins, inactivate S protein and prevent the binding between receptor-binding domain and angiotensin-converting enzyme 2 (ACE2) to achieve the disinfection of COVID-19 virus (25). As we all know, developing a new coronavirus vaccine is crucial in helping humans defend against the inflammatory outbreak caused by kinds of viruses. A recent article demonstrated that CAP could promote ROS accumulation in host cells, leading to the lipid peroxidation of the cell membrane, increasing the influx of Ca²⁺, and activating selective autophagy. The phenomenon caused by CAP would accelerate the entry of host cells into G1 and stop at the G2/M stage to increase the nutrients for viral multiplication, which was conducive to the development of new vaccines and clinical treatments (51). Furthermore, although vaccines that treat SARS-CoV-2 have been produced, the continuous mutation of SARS-CoV-2 may reduce the specificity of the vaccines. Dai, with his faculties, explored the effect of CAP on mutant SARS-CoV-2 viruses. Their results confirmed that the interaction of 'OH and other substances produced by CAP or CAP-activated medium (PAM) could trigger the nuclear translocation of the viral receptor ACE2 and inhibit the SARS-CoV-2 virus to invasive host cells (52). Due to the different complexity of virus particles, viruses infect host cells through a series of strategies, resulting in physical interactions between host cells and viruses. The ROS and RNS produced by CAP can promote the inactivation of the viruses by destroying the virus envelope, degrading their DNA or protein, and modifying the protein binding site.

ROS AND RNS ACCELERATED WOUND HEALING VIA CELLS' PROLIFERATION, GROWTH FACTORS' SECRETION, AND VESSELS' GENERATION

Recent studies have demonstrated that CAP can promote wound healing by regulating cell viability, proliferation, migration, and

inflammatory reaction (53–55). As a secondary messenger, H_2O_2 could drive redox signaling pathways and participate in the steady-state, inflammation, proliferation, and remodeling stages of wound healing (56). In addition, ROS and RNS also acted as the mediator of various cellular reactions, affecting cell differentiation, cell apoptosis, cell structure, and the structural integrity of connective tissue (57-60). Shome used a co-culture model to study the effect of CAP on wound healing and paracrine crosstalk between keratinocytes and fibroblasts in vitro. They observed that CAP treatment resulted in the up-regulation of the HIPPO transcription factor YAP in HaCaT and fibroblasts. As downstream effectors of the HIPPO signaling pathway, CTGF and Cyr61 were also up-regulated in their studies (56). In conclusion, their results confirmed that at the beginning of wound healing, CAP activated regeneration signaling pathways and stimulated communication between skin fibroblasts and keratinocytes, which promoted the healing of keratinocyte wounds co-culture model in vitro.

Schmidt et al. utilized the APPJ with Ar working gas to treat female mouse wound models for 20 seconds daily. Over 14 days, compared with the control group, the CAP group significantly accelerated the wound's epithelial regeneration from day three to nine by quantitative analysis. They demonstrated that CAP could stimulate the migration of keratinocytes and fibroblasts in vitro, which resulted in the accelerated closure of gaps in scratch assays. The cause of this effect was related to the downregulation of the gap junctional protein Cx43 mentioned above. Due to the down-regulation of E- cadherin, several integrins, and actin reorganization after CAP application, they confirmed that CAP could induce changes in adhesion junctions and cytoskeleton dynamics to accelerate wound healing (61). Lin utilized a DBD device with helium gas to treat the wound on the mice's skin at 66 kHz and 6.0 kV. They found that galectin-1, as a β-galactose-binding lectin, was involved in many physiological functions to induce myofibroblast activation, which wound secret growth factors and chemoattractants to create new substrates and proteins in the extracellular matrix in wound healing. It was reported that galectin-1 accelerated wound healing by regulating the theneuropilin-1/Smad3/NOX4 pathway and ROS production in myofibroblasts (62). They examined the efficacy of low-temperature plasma on galectin expression in the skin using immunoelectron microscopy and found that the galectins increased quickly after the lowtemperature plasma treatment and accelerated wound healing through the Smad pathway (6). Besides, CAP could increase the generation of fibroblasts and inner wall cells of blood vessels in wounds. The effect resulted from the link to the activity of some proteins, such as the vascular-associated migration cell protein (AAMP). CAP could strongly up-regulate the expression of the gene encoding AAMP, which could drive the RhoA-ROCK and Nod2–NF-κB signaling pathway to accelerate cell movement and promote wound healing (63). In summary, as Figure 5 displayed, CAP promoted wound healing by enhancing the proliferation of keratinocytes, up-regulating the expression of cell growth-related genes, strengthening the release of cytokines, inhibiting wound microbial infection, and inducing the formation of blood vessels.



ROS AND RNS ATTENUATED NCISDS VIA KERATINOCYTES' APOPTOSIS, IMMUNE REGULATION, AND ANTIOXIDANT SYSTEMS' ACCELERATING

According to the lesions and pathological changes of skin diseases, ncISDs include nearly 100 different immune diseases caused by immune cells. For example, Th17 and ILC3 cells can lead to psoriasis characterized by neutrophil infiltration. Th2 cell can lead to eczema characterized by the abnormal infiltration of eosinophils. As for vitiligo, the CD8+T cell is critical, resulting in the high secretion of CXCL10, IFN- γ , and IL-17. These diseases have seriously affected the quality of patients' lives and caused enormous socio-economic costs. Due to the genetic susceptibility of the body and the

influence of the peripheral environments, ncISDs often lead to impaired epithelial function and pathological changes, including immune cell infiltration (64).

As a kind of complex hyperproliferative skin disease, psoriasis is an autoimmune disease. The typical clinical features are characterized by erythema, plaques, and scaly (65). In addition, excessive proliferation of keratinocytes, inflammation, infiltration, and pathological angiogenesis has been regarded as the typical pathological changes of psoriasis (66, 67). Our team used a DBD device with 0.06 W/cm² to treat keratinocytes and evaluated the initial treatment effect of CAP on psoriasis by observing the death of keratinocytes and the release of related cytokines. We placed a petri dish containing HaCaT cells 10 mm below the surface DBD device. Then we explored the therapeutic effects of CAP on psoriasis by detecting the concentrations of ROS and RNS, observing the morphology, the viability, the apoptosis of keratinocytes, and the secretion of different cytokines, which played an essential role in the pathogenesis in psoriasis. From the results, we confirmed that CAP could suppress psoriasis. The main mechanisms of the results were exhibited: producing keratinocyte apoptosis, inhibiting excessive proliferation of keratinocyte and T lymphocytes, up-regulating the expression of IFN-y and the mRNA of VEGF, increasing IL-6, IL-8, TNF- α , and VEGF release and decreasing the content of IL-12 (68). Lu et al. used cell and animal experiments to confirm CAP's therapeutic function in psoriasis. They chose an APPJ device that flushed into helium and oxygen working gas with 15 kV and 1 kHz. Firstly, the cell model of psoriasis-like inflammation was simulated by adding LPS or TNF- α into HaCaT keratinocytes and then using a CAP-treated medium to cultivate HaCaT cells. Secondly, APPJ was directly applied to imiquimod-induced psoriasis-like dermatitis in mice. They observed that CAP increased ROS in HaCaT keratinocytes and caused keratinocyte apoptosis. Moreover, it was interesting that the IMQ-induced psoriasiform dermatitis gradually ameliorated lesions that displayed weakened epidermal proliferation and performed a thinner epidermal layer after APPJ treatment (69). Lee et al. indicated that CAP treatment might suppress psoriasislike lesions in mice. Compared with the mice without CAP treatment, increasing epithelial thickness and the expression of pro-inflammatory molecules were inhibited in the skin of psoriasis-like mice treated with CAP. It has been well accepted that the expression of PD-L1 was relevant to the excessive activation of T cells. To their surprise, CAP could inhibit excessive activation of T cells via increasing PD-L1 expression in HaCaT cells (70). ROS and RNS from CAP could inhibit the excessive proliferation of keratinocytes and promote their apoptosis but restrain the proliferation of T lymphocytes and enhance the expression of PD-L1 protein, which was negatively correlated with the T lymphocytes' proliferation. Moreover, ROS and RNS could also directly regulate the release of inflammatory factors such as IL-6, IL-12, and VEGF in lesions to achieve the effects of alleviating psoriasis.

Vitiligo is the most frequent cause of depigmentation globally (71). Its standard treatment methods make vitiligo treatment challenging, owing to side effects, low cure, and high recurrence rates (72). Our team recently conducted experiments on CAP to treat vitiligo in vitro and in vivo. We used APPJ and DBD generating devices to treat vitiligo-like mice and patients with active focal vitiligo up to one month. With 8 kV and 9 kHz, CAP showed satisfactory efficiency in attenuating vitiligo. We used CAP-activated hydrogel and APPJ device to observe the interaction between CAP and vitiligo. From the vitiligo-like mouse, we documented that CAP could ameliorate vitiligo lesions and accelerate the distribution of follicular melanin. Furthermore, we confirmed that CAP could inhibit the infiltration of immune cells such as CD8⁺T cells, CD3⁺ cells, and CD11c⁺ cells, decrease the secretion of immune cytokines, and strengthen the responsibility of anti-oxidative systems. Moreover, we also observed the same changes in active focal vitiligo patients in the clinical trial without any side effects.

In conclusion, we thought that CAP could inhibit the excessive activation of immune cells, restrain the release of inflammatory chemokines, induce the expression of Nrf2 and weaken the inducible nitric oxide synthase activity to attenuate vitiligo (73). From these studies, we considered that CAP ameliorated vitiligo by producing the generation of follicular melanocytes, inhibiting the excessive infiltration of immune cells, restraining the release of inflammatory cytokines, and regulating the activity of oxidative or anti-oxidative systems (Figure 6). As we all know, ncISDs are chronic and T-cell-associated inflammatory skin diseases. For psoriasis, with the characteristics of thickening and scaling, its pathogenesis was considered the proliferation of keratinocytes, infiltration of inflammatory or immune cells, and form of vessels. As for vitiligo, it involved oxidative stress pathways, immune inflammation, and the destruction of melanocytes. It could be known from the above documents that CAP attenuated ncISDs by promoting the apoptosis of keratinocytes, inhibiting the proliferation of keratinocytes, restraining excessive immune responses, regulating inflammatory cytokines, and reinforcing antioxidant systems.

ROS AND RNS INHIBITED CANCER *VIA* PRODUCING CELL APOPTOSIS, INCREASING CYTOKINES RELEASE, AND ACTIVATING THE DIFFERENT PATHWAYS

Recent studies have suggested that researchers have concluded that the primary mechanism of CAP's anti-cancer treatment might be related to the generation of ROS and RNS (74, 75). Free radicals, especially ROS and RNS, had been reported as the common mediator of apoptosis. ROS, RNS, and charged particles have been determined to be significant contributors to plasmainduced cell death (76). The excessive generation of ROS and RNS might damage cellular components, including DNA, proteins, and lipid membranes. However, most apoptosis pathways observed in CAP-treated cancer cells mainly depend on the injury of significant protein and DNA or the damage to the cell microenvironment.

Melanoma is considered as a highly invasive cancer that is highly resistant to most traditional chemotherapy and radiation therapies (77). Previous studies had provided several possible mechanisms by which CAP inactivated cancer cells, including ROS and RNS, charged particles, heat, pressure gradients, and changes in electrostatic and electromagnetic fields. In Keidar's study, CAP had been translated into in vivo models of tumor treatment. From the results, they found that the mid-sized tumors in nude mice ameliorated after 2-min CAP treatment without any thermal damage (78). Chernets et al. utilized CAP application to treat melanoma in a mouse model, and the results confirmed that CAP could prohibit the activity and proliferation of melanoma. Their study showed that ROS, RNS, and its radicals from CAP created a chemistry response for eliminating subdermal B16 melanoma. They also indicated that the heat from CAP was insufficient to prohibit melanoma



proliferation without the presence of reactive species (79). Ishaq et al. showed that CAP caused melanoma cells to overexpression TP73, which will lead to growth arrest and apoptosis (80). Ardnt et al. discovered that CAP treatment for 2 seconds could lead to irreversible cell cycle arrest, induction of phosphor-H2AX, and DNA damage (81, 82). In addition, Schneider et al. analyzed intracellular Ca2+ indicator fura-2 AM to analyze changes in cytoplasmic Ca²⁺ levels after CAP treatment of malignant melanoma cells. The results showed that CAP caused the acidification of water and solution, and the influx of Ca²⁺ in melanoma cells induced by CAP was enhanced in an acidic environment. Besides, NO induced by CAP was dose- and pH-dependent, and CAP-treated solutions could cause protein nitrification in cells under acidic conditions. Therefore, they believed that the decrease in pH caused by CAP treatment inhibited tumor cells' proliferation and growth. The possible mechanism was that the synergistic effect of ROS, RNS, and acidic conditions produced by CAP affected the intracellular Ca²⁺ level of melanoma cells. Since the microenvironment of tumors was usually acidic, further acidification may be one reason for the specific anti-cancer effect of CAP (10). These results might indicate that CAP shared some similarities in their mechanism of action, possibly by causing ROS-dependent DNA or protein damage in melanoma. Of course, acidifying the microenvironment of tumor cells and enhancing the influx of Ca²⁺ to tumor cells could also cause the death of tumors.

Non-melanoma skin tumor primarily includes basal cell and squamous cell carcinoma. Basal cell carcinoma (BCC), most common in the head and neck region, is the most common form of skin tumor and 3 to 5 times more common than squamous cell carcinoma. Squamous cell carcinoma mainly occurs in the head and neck region and is more common in older men (83). Recently, Wang studied the effects and the related mechanisms of CAP-activated medium on cutaneous squamous carcinoma cells (SCC). They selected A431 epidermoid carcinoma cell lines and a CAP-activated PBS medium to incubate the A431 cells for 2 hours. Their results showed that the CAP-activated PBS medium could increase the intracellular ROS levels and inhibit the proliferation of A431 cells in a dose/time-dependent manner. At the same time, the CAPactivated PBS medium could lead to the apoptosis of A431 cells (84). Yang's study aimed to explore the effects and related mechanisms of two CAP-activated solutions on TE354T cell and HaCaT keratinocyte cell lines. PAS was prepared by CAP irradiation of DMEM and PBS. Then they treated TE354T cells with PAS *in vitro* and evaluated the viability and apoptosis rate of TE354T cells and HaCaT cells. Western blotting and RNA sequencing were also detected. The results showed that CAPactivated PAS solution strengthened the apoptosis signaling in BCC, and the mechanisms were related to the activation of the different signaling pathways (85).

Besides the skin tumors mentioned above, CAP has shown remarkable effects on other kinds of tumors, such as the triplenegative breast cancers through hsa_circRNA_0040462 (86-88), glioblastoma multiforme (89), lung cancers (90), and pancreatic ductal adenocarcinoma (91). Recent studies confirmed that invivopen, a new source of CAP for cancer therapy, could ameliorate triple-negative breast cancer successfully. They found that CAP or PAM could inhibit the proliferation of triple-negative breast cancer, which showed a more significant inactivation on mesenchymal breast cancer than epithelialderived breast cancer. Surprisingly, the above results also applied to the CAP inactivation of mesenchymal-derived bladder cancers (92, 93). In addition, tumor recurrence is often related to the rapid proliferation of remaining tumor cells in the body. In order to solve the threat of residual tumor cells, our team developed a new type of therapeutic plasma-activated biogel that could be implanted into mice to inhibit the proliferation of residual tumor cells. With 10 kHz and 7.5 kV, CAP could transform the solution into the plasma-activated biogel containing ROS with its temperature rising to 35°C. Then the plasma-activated biogel was placed in the tissue where the tumor cells were removed, and the tumor growth on the skin of the

right-back of the mouse was monitored three times a week. The results showed that compared with other groups, the mouse in the plasma-activated biogel group was ameliorated entirely within two weeks. Moreover, the survival rate was more than six weeks, and no tumor recurrence had been seen. Through experiments, it could be known that the effect substances, such as H_2O_2 , NO_2^- , NO_3^- , $ONOO^-$ and O_2^- , could be released by plasma-activated biogel would inhibit the growth and recurrence of residual tumor tissues without any side effects (94). All in all, CAP could promote tumor cell apoptosis by activating the cell culture medium, acting on tumor cells, and directly inhibiting the proliferation of tumors. It could be seen from the above findings that the increase in the concentration of ROS and RNS in tumor cells was the key to the treatment of tumors by CAP. On the one hand, ROS and RNS could damage the DNA of tumor cells and the proteins such as tumor protein 73. On the other hand, the acidification of the tumor's microenvironment and the increased concentration of Ca²⁺ in the microenvironment were also crucial in prohibiting tumor cells by CAP. In addition, the MAPK/JNK or NF-KB signaling pathway was also the main mechanism for CAP to inhibit the proliferation of tumors.

CONCLUSION

CAP has shown its apparent effects on different skin diseases such as infectious diseases, skin tumors, psoriasis, vitiligo, and wound healing. Figure 7 has described the main mechanisms of CAP application on skin diseases. In terms of inhibiting microbial infections, with ROS and RNS, CAP destroyed biofilms or the cell membranes of microorganisms and damaged the DNA, RNA, or protein of microbes in a time or dose-dependent manner directly or indirectly; In wound healing, CAP could promote the proliferation of keratinocytes, enhance the expression of genes related to cell growth, and enhance the release of inflammatory factors. As for the ncISDs, CAP could promote the apoptosis of keratinocytes, inhibit the proliferation of keratinocytes, and suppress the excessive activation of T lymphocytes, which played a crucial role in psoriasis. CAP could also ameliorate the occurrence of vitiligo by reducing the excessive activation of the immune response and enhancing the ability of the antioxidant system. Finally, CAP could activate the apoptosis of cells, restrain the acidification of the cells' microenvironment, increase intracellular Ca²⁺, and damage DNA to ameliorate melanoma. Last but not least, for the other tumor such as glioblastoma multiforme,



lung cancers, and pancreatic ductal adenocarcinoma, ROS, and RNS could activate the apoptosis of the tumor and accelerate the release of the necrosis factors related to MAPK or NF- κ B signaling pathways to inhibit tumors. As a new type of non-invasive physical tool, CAP has shown satisfactory results in medicine. It is believed that CAP will be applied to treat more skin diseases in the future and apply more therapeutic options for the treatment of skin diseases.

AUTHOR CONTRIBUTIONS

Y-mX and MK designed the manuscript. S-yZ wrote the manuscript. S-yZ and Y-mX created the figures. Y-mX and

REFERENCES

- Lugović-Mihić L, Ćesić D, Vuković P, Novak Bilić G, Šitum M, Špoljar S. Melanoma Development: Current Knowledge on Melanoma Pathogenesis. Acta Dermatovenerol Croat (2019) 27:163–8.
- Rendon A, Schäkel K. Psoriasis Pathogenesis and Treatment. Int J Mol Sci (2019) 23:1475. doi: 10.3390/ijms20061475
- Wang P, Huang B, Horng H, Yeh C, Chen Y. Wound Healing. J Chin Med Assoc (2018) 81:94–101. doi: 10.1016/j.jcma.2017.11.002
- Mruwat R, Cohen Y, Yedgar S. Phospholipase A₂ Inhibition as Potential Therapy for Inflammatory Skin Diseases. *Immunotherapy* (2013) 5:315–7. doi: 10.2217/imt.13.18
- Wang B, Liu D, Zhang Z, Li Q, Wang X, Kong MG. A New Surface Discharge Source: Plasma Characteristic and Delivery of Reactive Species. *IEEE Trans Plasma Sci* (2016) 44:1–7. doi: 10.1109/TPS.2016.0620565
- Akimoto Y, Ikehara S, Yamaguchi T, Kim J, Kawakami H, Shimizu N, et al. Galectin Expression in Healing Wounded Skin Treated With Low-Temperature Plasma: Comparison With Treatment by Electronical Coagulation. Arch Biochem Biophys (2016) 1:86–94. doi: 10.1016/ j.abb.2016.01.012
- Guo L, Zhao Y, Liu D, Liu Z, Chen C, Xu R, et al. Cold Atmospheric-Pressure Plasma Induces DNA-Protein Crosslinks Through Protein Oxidation. *Free Radic Res* (2018) 52:783–98. doi: 10.1080/10715762
- Weiss M, Daeschlein G, Kramer A, Burchardt M, Brucker S, Wallwiener D, et al. Virucide Properties of Cold Atmospheric Plasma for Future Clinical Applications. J Med Virol (2017) 89:952–9. doi: 10.1002/jmv.24701
- Brun P, Pathak S, Castagliuolo I, Palù G, Zuin M, Cavazzana R, et al. Helium Generated Cold Plasma Finely Regulates Activation of Human Fibroblast-Like Primary Cells. *PloS One* (2014) 9:e104397. doi: 10.1371/ journal.pone.0104397
- Schneider C, Gebhardt L, Arndt S, Karrer S, Zimmermann JL, Fischer MJM, et al. Acidification is an Essential Process of Cold Atmospheric Plasma and Promotes the Anti-Cancer Effect on Malignant Melanoma Cells. *Cancers* (*Basel*) (2019) 11:671. doi: 10.3390/cancers11050671
- 11. Helmke A, Hoffmeister D, Berge F, Emmert S, Laspe P, Mertens N, et al. Physical and Microbiological Characterisation of *Staphylococcus Epidermidis* Inactivation by Dielectric Barrier Discharge Plasma. *Plasma Process Polym* (2011) 8:278–86. doi: 10.1002/ppap201000168
- Friedman PC. Cold Atmospheric Pressure (Physical) Plasma in Dermatology: Where are We Today? Int J Dermatol (2020) 59:1171–84. doi: 10.1111/ ijd.15110
- von Woedtke T, Metelmann HR, Weltmann KD. Clinical Plasma Medicine: State and Perspectives of *In Vivo* Application of Cold Atmospheric Plasma. *Contrib Plasma Physics* (2014) 54:104–17. doi: 10.1002/ctpp201300168
- Nguyen DB, Lee WG. Effects of Ambient Gas on Cold Atmospheric Plasma Discharge in the Decomposition of Trifluoromethane. *RSC Advances* (2016) 32:26505–13. doi: 10.1039/C6RA01485B
- Bernhardt T, Semmler ML, Schäfer M, Bekeschus S, Emmert S, Boeckmann L. Plasma Medicine: Applications of Cold Atmospheric Pressure Plasma in Dermatology. Oxid Med Cell Longev (2019) 3:3873928. doi: 10.1155/2019/ 3873928

MK revised the manuscript. All authors contributed to the article and approved the submitted version.

ACKNOWLEDGMENTS

We thank helpful discussions with Dr. Ding-xin Liu (State Key Laboratory of Electrical Insulation and Power Equipment, Xi'an Jiaotong University). And we also thank the financial support by the National Natural Science Foundation of China (No. 82173445) and the Innovation Capability Support Plan of Shaanxi Province (No.2019TD-034).

- Schmidt A, Wende K, Bekeschus S, Bundscherer L, Barton A, Ottmüller K, et al. Non-Thermal Plasma Treatment is Associated With Changes in Transcriptome of Human Epithelial Skin Cells. *Free Radic Res* (2013) 47:577–92. doi: 10.3109/10715762.2013.804623
- Lukes P, Dolezalova E, Sisrova I, Clupek M. Aqueous-Phase Chemistry and Bactericidal Effects From an Air Discharge Plasma in Contact With Water: Evidence for the Formation of Peroxynitrite Through a Pseudo-Second-Order Post-Discharge Reaction of H₂O₂ and HNO₂. *Plasma Sourc Sci Technol* (2014) 23:015019. doi: 10.1088/0963-0252/23/1/015019
- Bauer G, Sersenová D, Graves DB, Machala Z. Cold Atmospheric Plasma and Plasma-Activated Medium Trigger RONS-Based Tumor Cell Apoptosis. Sci Rep (2019) 9:14210. doi: 10.1038/s41598-019-50291-0
- Mitra S, Nguyen LN, Akter M, Park G, Choi EH, Kaushik NK. Impact of ROS Generated by Chemical, Physical, and Plasma Techniques on Cancer Attenuation. *Cancers (Basel)* (2019) 11:1030. doi: 10.3390/cancers11071030
- Dai X, Zhang Z, Zhang J, Ostrikov K. Dosing: The Key to Precision Plasma Oncology. *Processes Polymers* (2020) 10:e1900178. doi: 10.1002/ppap.201900178
- Vlaski-Lafarge M, Ivanovic Z. Reliability of ROS and RNS Detection in Hematopoietic Stem Cells–Potential Issues With Probes and Target Cell Population. J Cell Sci (2015) 128:3849–60. doi: 10.1242/jcs.171496
- Xia J, Zeng W, Xia Y, Wang B, Xu D, Liu D, et al. Cold Atmospheric Plasma Induces Apoptosis of Melanoma Cells via Sestrin2-Mediated Nitric Oxide Synthase Signaling. J Biophotonics (2019) 12:e201800046. doi: 10.1002/ jbio.201800046
- Daeschlein G, Scholz S, von Woedtke T, Niggemeier M, Kindel E, Brandenburg R, et al. *In Vitro* Killing of Clinical Fungal Strains by Low-Temperature Atmospheric Pressure Plasma Jet. *IEEE Trans Plasma Sci* (2011) 39:815–21. doi: 10.1109/tps.2010.2063441
- Daeschlein G, Napp M, von Podewils S, Scholz S, Arnold A, Emmert S, et al. Antimicrobial Efficacy of a Historical High-Frequency Plasma Apparatus in Comparison With 2 Modern, Cold Atmospheric Pressure Plasma Devices. Surg Innov (2015) 22:394–400. doi: 10.1177/1553350615573584
- Guo L, Yao Z, Yang L, Zhang H, Qi Y, Gou L, et al. Plasma-Activated Water: An Alternative Disinfectant for S Protein Inactivation to Prevent SARS-CoV-2 Infection. *Chem Eng J* (2012) 421:127742. doi: 10.1016/j.cej.2020.127742
- Ali A, Kim YH, Lee JY, Lee SH, Uhm HS, Cho G, et al. Inactivation of Propionibacterium Acnes and its Biofilm by non-Thermal Plasma. *Curr Appl Physics* (2014) 14:S142–8. doi: 10.1016/j.cap.2013.12.034
- Del Pozo JL. Biofilm-Related Disease. Expert Rev Anti Infect Ther (2018) 16:51–65. doi: 10.1080/14787210.2018.1417036
- Theinkom F, Singer L, Cieplik F, Cantzler S, Weilemann H, Cantzler M, et al. Antibacterial Efficacy of Cold Atmospheric Plasma Against Enterococcus Faecalis Planktonic Cultures and Biofilms *In Vitro. PloS One* (2019). 14: e0223925. doi: 10.1371/journal.pone.0223925
- Govaert M, Smet C, Walsh JL, van Impe JFM. Dual-Species Model Biofilm Consisting of Listeria Monocytogenes and Salmonella Typhimurium: Development and Inactivation With Cold Atmospheric Plasma (CAP). Front Microbiol (2019) 7:2524. doi: 10.3389/fmicb.2019.02524
- Niemira BA, Boyd G, Sites J. Cold Plasma Rapid Decontamination of Food Contact Surfaces Contaminated With Salmonella Biofilms. J Food Sci (2014) 79:M917–922. doi: 10.1111/1750-3841.12379

- Modic M, McLeod NP, Sutton JM, Walsh JL. Cold Atmospheric Pressure Plasma Elimination of Clinically Important Single- and Mixed-Species Biofilms. Int J Antimicrob Agents (2017) 49:375–8. doi: 10.1016/ j.ijantimicag.2016.11.022
- Patange A, Boehm D, Ziuzina D, Cullen PJ, Gilmore B, Bourke P. High Voltage Atmospheric Cold Air Plasma Control of Bacterial Biofilms on Fresh Produce. Int J Food Microbiol (2019) 16:137–45. doi: 10.1016/ j.ijfoodmicro.2019.01.005
- Fahmide F, Ehsani P, Atyabi SM. Time-Dependent Behavior of the Staphylococcus Aureus Biofilm Following Exposure to Cold Atmospheric Pressure Plasma. Iran J Basic Med Sci (2021) 24:744–51. doi: 10.22038/ Ijbms.2021.52541.11866
- 34. Guo L, Yang L, Qi Y, Niyazi G, Huang L, Gou L, et al. Cold Atmospheric-Pressure Plasma Caused Protein Damage in Methicillin-Resistant Cells in Biofilms. *Microorganisms* (2021) 9:1072. doi: 10.3390/ microorganisms9051072
- 35. Colagar AH, Memariani H, Sohbatzadeh F, Omran AV. Nonthermal Atmospheric Argon Plasma Jet Effects on Escherichia Coli Biomacromolecules. *Appl Biochem Biotechnol* (2013) 171:1617-29. doi: 10.1007/s12010-013-0430-9
- Han X, Cantrell WA, Escobara EE, Ptasinskab S. Plasmid DNA Damage Induced by Helium Atmospheric Pressure Plasma Jet. *Eur Phys J D* (2014) 68:7. doi: 10.1140/epjd/e2014-40753-y
- Dezest M, Bulteau AL, Quinton D, Chavatte L, Bechec ML, Cambus JP, et al. Oxidative Modification and Electrochemical Inactivation of Upon Cold Atmospheric Pressure Plasma Exposure. *PloS One* (2017) 12:e0173618. doi: 10.1371/journal.pone.0173618
- Hähnel M, von Woedtke T, Weltmann KD. Influence of the Air Humidity on the Reduction of Bacillus Spores in a Defined Environment at Atmospheric Pressure Using a Dielectric Barrier Surface Discharge. *Plasma Process Polym* (2010) 7:244–9. doi: 10.1002/ppap.200900076
- Cahill OJ, Claro T, O'Connor N, Cafolla AA, Stevens NT, Daniels S, et al. Cold Air Plasma to Decontaminate Inanimate Surfaces of the Hospital Environment. Appl Environ Microbiol (2014) 80:2004–10. doi: 10.1128/ AEM.03480-13
- 40. Joshi SG, Cooper M, Yost A, Paff M, Ercan UK, Fridman G, et al. Nonthermal Dielectric-Barrier Discharge Plasma-Induced Inactivation Involves Oxidative DNA Damage and Membrane Lipid Peroxidation in Escherichia Coli. Antimicrob Agents Chemother (2011) 55:1053–62. doi: 10.1128/AAC.01002-10
- Fricke K, Koban I, Tresp H, Jablonowski L, Schroder K, Kramer A, et al. Atmospheric Pressure Plasma: A High- Performance Tool for the Efficient Removal of Biofilms. *PloS One* (2012) 7:e42539. doi: 10.1371/ journal.pone.0042539
- Heinlin J, Maisch T, Zimmermann JL, Shimizu T, Holzmann T, Simon M, et al. Contact-Free Inactivation of Trichophyton Rubrum and Microsporum Canis by Cold Atmospheric Plasma Treatment. *Future Microbiol* (2013) 8:1097–106. doi: 10.2217/fmb.13.86
- Ouf SA, El-Adly AA, Mohamed AH. Inhibitory Effect of Silver Nanoparticles Mediated by Atmospheric Pressure Air Cold Plasma Jet Against Dermatophyte Fungi. J Med Microbiol (2015) 64:1151-61. doi: 10.1099/ jmm.0.000133
- 44. Yasuda H, Miura T, Kurita H, Takashima K, Mizuno A. Biological Evaluation of DNA Damage in Bacteriophages Inactivated by Atmospheric Pressure Cold Plasma. *Plasma Process Polym* (2010) 7:301–8. doi: 10.1002/ppap.200900088
- Aboubakr HA, Mor SK, Higgins LA, Armien A, Youssef MM, Bruggenman PJ, et al. Cold Argon-Oxygen Plasma Species Oxidize and Disintegrate Capsid Protein of Feline Calicivirus. *PloS One* (2018) 13:e0194618. doi: 10.1371/ journal.pone.0194618
- 46. Guo L, Xu R, Gou L, Liu Z, Zhao Y, Liu D, et al. Mechanism of Virus Inactivation by Cold Atmospheric-Pressure Plasma and Plasma-Activated Water. Appl Environ Microbiol (2018) 84:e00726-18. doi: 10.1128/ AEM.00726-18
- Brun P, Vono M, Venier P, Tarricone E, Deligianni V, Martines E, et al. Disinfection of Ocular Cells and Tissues by Atmospheric-Pressure Cold Plasma. *PloS One* (2012) 7:e33245. doi: 10.1371/journal.pone.0033245
- Zimmermann JL, Dumler K, Shimizu T, Morfill GE, Wolf A, Boxhammer V, et al. Effects of Cold Atmospheric Plasmas on Adenoviruses in Solution. J Phys D Appl Phys (2011) 44:505201. doi: 10.1088/0022-3727/44/50/505201

- Galdiero S, Falanga A, Vitiello M, Grieco P, Caraglia M, Morelli G, et al. Exploitation of Viral Properties for Intracellular Delivery. J Pept Sci (2014) 20:468–78. doi: 10.1002/psc.2649
- Xiang Y, Li W, Zhang Q, Jin Y, Rao W, Zeng L, et al. Timely Research Papers About COVID-19 in China. *Lancet* (2020) 395:684–5. doi: 10.1016/S0140-6736(20)30375-5
- Miao Y, Han P, Hua D, Zhou R, Guan Z, Lv Q, et al. Cold Atmospheric Plasma Increases IBRV Titer in MDBK Cells by Orchestrating the Host Cell Network. *Virulence* (2021) 1:679–89. doi: 10.1080/21505594.2021.1883933
- Wang P, Zhou R, Zhou R, Li W, Weerasinghe J, Chen S, et al. Cold Atmospheric Plasma for Preventing Infection of Viruses That Use ACE2 for Entry. *Theranostics* (2022) 6:2811–32. doi: 10.7150/thno.70098
- 53. Wende K, Strassenburg S, Haertel B, Harms M, Holtz S, Barton A, et al. Atmospheric Pressure Plasma Jet Treatment Evokes Transient Oxidative Stress in HaCaT Keratinocytes and Influences Cell Physiology. *Cell Biol Int* (2014) 38:412–25. doi: 10.1002/cbin.10200
- Bekeschus S, Schmidt A, Bethge L, Masur K, von Woedtke T, Hasse S, et al. Stimulation of Human THP-1 Monocytes in Response to Cold Physical Plasma. Oxid Med Cell Longev (2016) 2016:5910695. doi: 10.1155/2016/5910695
- 55. Arndt S, Landthaler M, Zimmermann JL, Unger P, Wacker E, Shimizu T. Effects of Cold Atmospheric Plasma (CAP) on ß-Defensins, Inflammatory Cytokines, and Apoptosis-Related Molecules in Keratinocytes *In Vitro* and *In Vivo. PloS One* (2015) 10:e0120041. doi: 10.1371/journal.pone.0120041
- Shome D, von Woedtke T, Riedel K, Masur K. The HIPPO Transducer YAP and its Targets CTGF and Cyr61 Drive a Paracrine Signalling in Cold Atmospheric Plasma-Mediated Wound Healing. Oxid Med Cell Longev (2020) 13:4910280. doi: 10.1155/2020/4910280
- Balzer J, Heuer K, Demir E, Hoffmanns MA, Baldus S, Fuchs PC, et al. Non-Thermal Dielectric Barrier Discharge (DBD) Effects on Proliferation and Differentiation of Human Fibroblasts are Primary Mediated by Hydrogen Peroxide. *PloS One* (2015) 10:e0144968. doi: 10.1371/journal.pone.0144968
- Bekeschus S, von Woedtke T, Kramer A, Weltmann KD, Masur K. Cold Physical Plasma Treatment Alters Redox Balance in Human Immune Cells. *Plasma Med* (2013) 3:267–78. doi: 10.1615/plasmamed.2014011972
- Schmidt A, Bekeschus S, von Woedtke T, Hasse S. Cell Migration and Adhesion of a Human Melanoma Cell Line is Decreased by Cold Plasma Treatment. *Clin Plasma Med* (2015) 3:24–31. doi: 10.1016/j.cpme.2015.05.003
- 60. Arndt S, Unger P, Wacker E, Shimizu T, Heinlin J, Li Y, et al. Cold Atmospheric Plasma (CAP) Changes Gene Expression of Key Molecules of the Wound Healing Machinery and Improves Wound Healing *In Vitro* and *In Vivo*. *PloS One* (2013) 8:e79325. doi: 10.1371/journal.pone.0079325
- Schmidt A, Bekeschus S, Wende K, Vollmar B, Woedtkevon T. A Cold Plasma Jet Accelerates Wound Healing in a Murine Model of Full-Thickness Skin Wounds. *Exp Dermatol* (2017) 26:156–62. doi: 10.1111/exd.13156
- 62. Lin Y, Chen J, Wu M, Hsieh I, Liang C, Hsu C, et al. Galectin-1 Accelerates Wound Healing by Regulating the Neuropilin-1/Smad3/NOX4 Pathway and ROS Production in Myofibroblasts. J Invest Dermatol (2015) 135:258–68. doi: 10.1038/jid.2014.288
- 63. Daeschlein G, von Woedtke T, Kindel E, Brandenburg R, Weltmann KD, Jünger M. Antibacterial Activity of an Atmospheric Pressure Plasma Jet Against Relevant Wound Pathogens *In Vitro* on a Simulated Wound Environment. *Plasma Processes Polymers* (2010) 7:224–30. doi: 10.1002/ppap.200900059
- Eyerich K, Eyerich S. Immune Response Patterns in non-Communicable Inflammatory Skin Diseases. J Eur Acad Dermatol Venereol (2018) 32:692– 703. doi: 10.1111/jdv.14673
- Michalek IM, Loring B, John SM. A Systematic Review of Worldwide Epidemiology of Psoriasis. J Eur Acad Dermatol Venereol (2017) 31:205–12. doi: 10.1111/jdv.13854
- Ogawa E, Sato Y, Minagawa A, Okuyama R. Pathogenesis of Psoriasis and Development of Treatment. J Dermatol (2018) 45:264–72. doi: 10.3390/ ijms20061475
- Nussbaum L, Chen YL, Ogg GS. Role of Regulatory T Cells in Psoriasis Pathogenesis and Treatment. Br J Dermatol (2021) 184:14–24. doi: 10.1111/bjd.19380
- Zhong S, Dong Y, Liu D, Xu D, Xiao S, Chen H, et al. Surface Air Plasma-Induced Cell Death and Cytokine Release of Human Keratinocytes in the Context of Psoriasis. *Br J Dermatol* (2016) 174:542–52. doi: 10.1111/bjd.14236
- 69. Lu G, Duan J, Zhang S, Liu X, Poorun D, Liu X. Cold Atmospheric Plasma Ameliorates Imiquimod-Induced Psoriasiform Dermatitis in Mice by

Mediating Ntiproliferative Effects. *Free Radic Res* (2019) 53:269-80. doi: 10.1080/10715762.2018.1564920

- Lee YS, Lee MH, Kim HJ, Won HR, Kim CH. Non-Thermal Atmospheric Plasma Ameliorates Imiquimod-Induced Psoriasis-Like Skin Inflammation in Mice Through Inhibition of Immune Responses and Up-Regulation of PD-L1 Expression. *Sci Rep* (2017) 7:15564. doi: 10.1038/s41598-017-15725-7
- Ezzedine K, Eleftheriadou V, Whitton M, van Geel N. Vitiligo. Lancet (2015) 386:74–84. doi: 10.1016/S0140-6736(14)60763-7
- Speeckaert R, van Geel N. Vitiligo: An Update on Pathophysiology and Treatment Options. Am J Clin Dermatol (2017) 6:733–44. doi: 10.1007/ s40257-017-0298-5
- Zhai S, Xu M, Li Q, Guo K, Chen H, Kong MG, et al. Successful Treatment of Vitiligo With Cold Atmospheric Plasma–Activated Hydrogel. J Invest Dermatol (2021) 11:2710–9. doi: 10.1016/j.jid.2021.04.019
- Dai X, Bazaka K, Thompson EW, Ostrikov KK. Cold Atmospheric Plasma: A Promising Controller of Cancer Cell States. *Cancers (Basel)* (2020) 11:3360. doi: 10.3390/cancers12113360
- Dai X, Bazaka K, Richard DJ, Thompson ERW, Ostrikov KK. The Emerging Role of Gas Plasma in Oncotherapy. *Trends Biotechnol* (2018) 11:1183–98. doi: 10.1016/j.tibtech.2018.06.010
- 76. Lin A, Chernets N, Han J, Alicea Y, Dobrynin D, Fridman G, et al. Non-Equilibrium Dielectric Barrier Discharge Treatment of Mesenchymal Stem Cells: Charges and Reactive Oxygen Species Play the Major Role in Cell Death. *Plasma Processes Polymers* (2015) 12:1117–27. doi: 10.1002/ppap.201400232
- 77. Hoek KS, Goding CR. Cancer Stem Cells Versus Phenotype-Switching in Melanoma. Pigment Cell Melanoma Res (2010) 23:746. doi: 10.1111/j.1755-148X.2010.00757
- Keidar M, Walk R, Shashurin A, Srinivasan P, Sandler A, Dasgupta S, et al. Cold Plasma Selectivity and the Possibility of a Paradigm Shift in Cancer Therapy. Br J Cancer (2011) 105:1295–301. doi: 10.1038/bjc.2011.386
- Chernets N, Kurpad DS, Alexeev V, Rodrigues DB, Freeman TA. Reaction Chemistry Generated by Nanosecond Pulsed Dielectric Barrier Discharge Treatment is Responsible for the Tumor Eradication in the B16 Melanoma Mouse Model. *Plasma Process Polym* (2015) 12:1400–9. doi: 10.1002/ ppap.201500140
- Ishaq M, Bazaka K, Ostrikov K. Intracellular Effects of Atmospheric-Pressure Plasmas on Melanoma Cancer Cells. *Phys Plasmas* (2015) 22:122003. doi: 10.1021/acsami.0c06500
- Arndt S, Wacker E, Li Y, Shimizu T, Thomas HM, Morfill GE, et al. Cold Atmospheric Plasma, a New Strategy to Induce Senescence in Melanoma Cells. *Exp Dermatol* (2013) 22:284–9. doi: 10.1111/exd.12127
- Kalghatgi S, Kelly CM, Cerchar E, Torabi B, Alekseev O, Fridman A, et al. Effects of non-Thermal Plasma on Mammalian Cells. *PloS One* (2011) 6: e16270. doi: 10.1371/journal.pone.0016270
- Losquadro WD. Anatomy of the Skin and the Pathogenesis of Nonmelanoma Skin Cancer. Facial Plast Surg Clin North Am (2017) 25:283–9. doi: 10.1016/ j.fsc.2017.03.001
- Wang L, Yang X, Yang C, Gao J, Zhao Y, Cheng C, et al. The Inhibition Effect of Cold Atmospheric Plasma-Activated Media in Cutaneous Squamous Carcinoma Cells. *Future Oncol* (2019) 15:495–505. doi: 10.2217/fon-2018-0419
- Yang X, Yang C, Wang L, Cao Z, Wang Y, Cheng C, et al. Inhibition of Basal Cell Carcinoma Cells by Cold Atmospheric Plasma-Activated Solution and

Differential Gene Expression Analysis. Int J Oncol (2020) 56:1262–73. doi: 10.3892/ijo.2020.5009

- Xiang L, Xu X, Zhang S, Cai D, Dai X. Cold Atmospheric Plasma Conveys Selectivity on Triple Negative Breast Cancer Cells Both *In Vitro* and *In Vivo. Free Radic Biol Med* (2018) 124:205–13. doi: 10.1016/j.freeradbiomed.2018.06.001
- Xu X, Dai X, Xiang L, Cai D, Xiao S, Ostrikov K. Quantitative Assessment of Cold Atmospheric Plasma Anti-Cancer Efficacy in Triple-Negative Breast Cancers. Processes Polymers (2018) 8:e1800052. doi: 10.1002/ppap.201800052
- Tian Y, Zhang Z, Zhang Z, Dai X. Hsa_circRNA_0040462: A Sensor of Cells' Response to CAP Treatment With Double-Edged Roles on Breast Cancer Malignancy. *Int J Med Sci* (2022) 4:640–50. doi: 10.7150/ijms.66940
- Almeida ND, Klein AL, Hogan EA, Terhaar SJ, Kedda J, Uppal P, et al. Cold Atmospheric Plasma as an Adjunct to Immunotherapy for Glioblastoma Multiforme. World Neurosurg (2019) 130:369–76. doi: 10.1016/ j.wneu.2019.06.209
- 90. Li W, Yu H, Ding D, Chen Z, Wang Y, Wang S, et al. Cold Atmospheric Plasma and Iron Oxide-Based Magnetic Nanoparticles for Synergetic Lung Cancer Therapy. *Free Radic Biol Med* (2019) 130:71–81. doi: 10.1016/ j.freeradbiomed.2018.10.429
- 91. van Loenhout J, Flieswasser T, Boullosa LF, Waele JD, van Audenaerde J, Marcq E, et al. Cold Atmospheric Plasma-Treated Pbs Eliminates Immunosuppressive Pancreatic Stellate Cells and Induces Immunogenic Cell Death of Pancreatic Cancer Cells. *Cancers (Basel)* (2019) 11:1597. doi: 10.3390/cancers11101597
- Zhou X, Cai D, Xiao S, Ning M, Zhou R, Zhang S, et al. pen: A Novel Plasma Source for Cancer Treatment. J Cancer (2020) 8:2273–82. doi: 10.7150/ jca.38613
- 93. Wang P, Zhou R, Thomas P, Zhao L, Zhou R, Mandal S, et al. Epithelial-To-Mesenchymal Transition Enhances Cancer Cell Sensitivity to Cytotoxic Effects of Cold Atmospheric Plasmas in Breast and Bladder Cancer Systems. *Cancers (Basel)* (2021) 12:2889. doi: 10.3390/cancers13122889
- 94. Zhang H, Xu S, Zhang J, Wang Z, Liu D, Guo L, et al. Plasma-Activated Thermosensitive Biogel as an Exogenous ROS Carrier for Post-Surgical Treatment of Cancer. *Biomaterials* (2021) 276:121057. doi: 10.1016/ j.biomaterials.2021.121057

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zhai, Kong and Xia. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.