



# Complete Genome Sequence of Megaplasmid-Bearing *Streptococcus salivarius* Strain LAB813, Isolated from the Dental Plaque of a Caries-Free Child

Siew-Ging Gong,<sup>a</sup> Yuki Chan,<sup>b</sup>  Céline M. Lévesque<sup>a,b</sup>

<sup>a</sup>Faculty of Dentistry, University of Toronto, Toronto, Ontario, Canada

<sup>b</sup>Faculty of Dentistry, The University of Hong Kong, Hong Kong SAR, China

**ABSTRACT** *Streptococcus salivarius* strain LAB813 was isolated from the dental plaque biofilm of a caries-free child with healthy oral tissues. We report here the complete genome sequence of *S. salivarius* strain LAB813. This genome consists of a chromosome of 2.2 Mb and a megaplasmid, pSAL813, of 183 kb.

*Streptococcus salivarius* is a predominant member of the oral microbiome that persists throughout the human life. It is not known to initiate infections in healthy or immunocompetent individuals. In fact, several reports have indicated that *S. salivarius* plays a positive role in oral and digestive tract ecology. *S. salivarius* may exert its positive impact through effects on the stability of the microbiome, bacterial interference, and/or host interaction (1). *S. salivarius* colonizes the buccal epithelium and is a resident of the tongue dorsum. *S. salivarius* is also able to coaggregate with various oral colonizers (2). Some strains of *S. salivarius* display antimicrobial activity against virulent streptococci (e.g., group A streptococcus [GAS] and group B streptococcus [GBS]) and therefore contribute to the maintenance of oral, pharyngeal, and intestinal health (3). *S. salivarius* has thus emerged as an important source of safe and efficacious probiotics capable of fostering more balanced, health-associated oral microbiota (4). In a screening of *S. salivarius* strains with probiotic potential, our group isolated strain LAB813 from the supragingival plaque collected from the facial and lingual smooth surfaces of the primary maxillary incisors of a caries-free child aged 5 years and 2 months (University of Toronto REB protocol reference number 32740). The plaque sample was plated on mitis-salivarius-bacitracin agar supplemented with 20% sucrose using a spiral plater. Isolate LAB813 was verified as *S. salivarius* by PCR (5) and 16S rRNA gene sequencing (6). Here, we present the complete genome sequence of this strain.

The LAB813 strain was cultivated in a 50-ml volume of brain heart infusion broth at 37°C in air with 5% CO<sub>2</sub> for 18 h without agitation. Total genomic DNA was extracted using an in-house protocol. Briefly, LAB813 cells were lysed with lysozyme (50 mg/ml at 37°C for 1 h), and proteins were digested with proteinase K (20 mg/ml at 37°C for 15 min) and precipitated with ice-cold potassium acetate buffer and ice-cold isopropanol. The DNA was fished out using a glass pipette and treated with RNase A (10 mg/ml at 37°C for 1 h). DNA was quantified using the Quant-iT PicoGreen double-stranded DNA (dsDNA) assay kit (Thermo Fisher). Whole-genome sequencing was performed using the Pacific Biosciences sequencing technology. The DNA library was prepared following the Pacific Biosciences 20-kb template preparation using the BluePippin size selection system protocol. Qualified genomic DNA was fragmented using the Covaris g-TUBE device and then end repaired to prepare SMRTbell DNA template libraries (with a fragment size of 15 kb to 50 kb) selected using a BluePippin system. Sequencing was performed in a Pacific Biosciences RS II sequencer using the MagBead OneCellPerWell (OCPW) protocol at the Génome Québec Innovation Centre

**Citation** Gong S-G, Chan Y, Lévesque CM. 2019. Complete genome sequence of megaplasmid-bearing *Streptococcus salivarius* strain LAB813, isolated from the dental plaque of a caries-free child. Microbiol Resour Announc 8:e01092-19. <https://doi.org/10.1128/MRA.01092-19>.

**Editor** Julie C. Dunning Hotopp, University of Maryland School of Medicine

**Copyright** © 2019 Gong et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Céline M. Lévesque, [celine.levesque@dentistry.utoronto.ca](mailto:celine.levesque@dentistry.utoronto.ca).

**Received** 9 September 2019

**Accepted** 18 September 2019

**Published** 10 October 2019

and Canadian Centre for Computational Genomics (McGill University, Québec, Canada). The Pacific Biosciences sequencing using 1 single-molecule real-time (SMRT) cell generated a total of 52,186 raw subreads with an average length of 14,073 bp, with an  $N_{50}$  value of 24,132 bp. Genome assembly was done using the Hierarchical Genome Assembly Process (HGAP) workflow with default settings (7). The assembled genome had 350× genome coverage. The assembly contained both a complete chromosome of 2,242,557 bp, with a G+C content of 40.1%, and a megaplasmid named pSAL813 of 183,700 bp, with a G+C content of 34.9%. Gene prediction and annotation were performed using RAST (8) and BLASTp (9). A total of 2,101 and 174 protein-coding genes (CDSs), 75 and 0 tRNAs, and 5 and 0 rRNAs were annotated in the chromosome and megaplasmid pSAL813, respectively. The genomic information was analyzed to predict putative bacteriocin gene clusters through the BAGEL4 (10) Web server with default search options. The pipeline predicted a novel multi-peptide lantibiotic locus on pSAL813 that is highly similar to the *pld* locus that drives the production of the broad-spectrum bacteriocin pneumolancidin in *Streptococcus pneumoniae* (11). The megaplasmid also encodes the chromosomal PezTA toxin-antitoxin system found in pneumococci (12). We also identified a chromosomal locus encoding three major fimbrial subunits belonging to the serine-rich repeat proteins family in streptococcal and staphylococcal species and that comprised many repeats of a motif of four amino acid residues (13); we also identified numerous genes encoding enzymes responsible for the glycosylation of the subunits and their transport through the accessory SecA2/Y2 system.

**Data availability.** The complete genome sequence has been deposited in GenBank under the accession numbers CP040803 (megaplasmid pSAL813) and CP040804 (chromosome). Raw sequencing reads were deposited in the NCBI Sequence Read Archive (SRA) under accession number SRR9298671 and BioProject number PRJNA546007.

## ACKNOWLEDGMENTS

We thank Lindsay Jackson and Abdelahhad Barbour for technical assistance in the lab.

This research was supported by Canadian Institutes of Health Research (CIHR) grant CMA-151711. C.M.L. is a recipient of a Canada Research Chair in Oral Microbial Genetics.

## REFERENCES

- Delorme C, Abraham AL, Renault P, Guédon E. 2015. Genomics of *Streptococcus salivarius*, a major human commensal. *Infect Genet Evol* 33:381–392. <https://doi.org/10.1016/j.meegid.2014.10.001>.
- Nobbs AH, Lamont RJ, Jenkinson HF. 2009. *Streptococcus* adherence and colonization. *Microbiol Mol Biol Rev* 73:407–450. <https://doi.org/10.1128/MMBR.00014-09>.
- Hols P, Ledesma-Garcia L, Gabant P, Mignolet J. 2019. Mobilization of microbiota commensals and their bacteriocins for therapeutics. *Trends Microbiol* 27:690–702. <https://doi.org/10.1016/j.tim.2019.03.007>.
- Wescombe PA, Hale JD, Heng NC, Tagg JR. 2012. Developing oral probiotics from *Streptococcus salivarius*. *Future Microbiol* 7:1355–1371. <https://doi.org/10.2217/fmb.12.113>.
- Nakanishi H, Kido A, Ohmori T, Takada A, Hara M, Adachi N, Saito K. 2009. A novel method for the identification of saliva by detecting oral streptococci using PCR. *Forensic Sci Int* 183:20–23. <https://doi.org/10.1016/j.forsciint.2008.10.003>.
- Edwards U, Rogall T, Blöcker H, Emde M, Böttger EC. 1989. Isolation and direct complete nucleotide determination of entire genes. Characterization of a gene coding for 16S ribosomal RNA. *Nucleic Acids Res* 17:7843–7853. <https://doi.org/10.1093/nar/17.19.7843>.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 25:3389–3402. <https://doi.org/10.1093/nar/25.17.3389>.
- van Heel AJ, de Song A, Song C, Viel JH, Kok J, Kuipers OP. 2018. BAGEL4: a user-friendly Web server to thoroughly mine RIPPs and bacteriocins. *Nucleic Acids Res* 46:W278–W281. <https://doi.org/10.1093/nar/gky383>.
- Maricic N, Anderson ES, Opipari AE, Yu EA, Dawid S. 2016. Characterization of a multi-peptide lantibiotic locus in *Streptococcus pneumoniae*. *mBio* 7:e01656-15. <https://doi.org/10.1128/mBio.01656-15>.
- Chan WT, Moreno-Córdoba I, Yeo CC, Espinosa M. 2012. Toxin-antitoxin genes of the Gram-positive pathogen *Streptococcus pneumoniae*: so few and yet so many. *Microbiol Mol Biol Rev* 76:773–791. <https://doi.org/10.1128/MMBR.00030-12>.
- Lévesque C, Vadeboncoeur C, Chandad F, Frenette M. 2001. *Streptococcus salivarius* fimbriae are composed of a glycoprotein containing a repeated motif assembled into a filamentous nondissociable structure. *J Bacteriol* 183:2724–2732. <https://doi.org/10.1128/JB.183.9.2724-2732.2001>.