

Expression of *Egr3* in mouse gonads and its localization and function in oocytes

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Objective: The early growth response (Egr) family consists of four members (Egr1, Egr2, Egr3, and Egr4) that are zinc finger transcription factors. Among them, Egr3 is involved in transcriptional regulation of target genes during muscle spindle formation and neurite outgrowth. We previously showed that the immunoreactive Egr3 is localized on oocyte spindle and accumulate near the microtubule organizing center during meiosis I in mice. Egr3 was also shown to be localized on spermatocytes. We herein investigated if *Egr3* is expressed in mouse gonads and if *Egr3* blockade results in any defect in oocyte maturation.

Methods: Expression of *Egr3* in mouse gonads was examined by reverse transcription-polymerase chain reaction. Full-length *Egr3* and truncated *Egr3* (Δ Egr3) complementary RNAs (cRNAs) with Xpress tag at N-terminus and DsRed2 at C-terminus, and small interfering RNA (siRNA) targeting *Egr3* were microinjected into mouse oocytes at germinal vesicle stage. Localization of microinjected *Egr3* was examined by confocal live imaging and immunofluorescence staining.

Results: *Egr3* mRNA was detected in mouse ovaries and testes from 1 to 4 week-old mice. An uncharacterized longer transcript containing 5' untranslated region was also detected in 3 and 4 week-old gonads. Microinjected Xpress-Egr3-DsRed2 or Xpress- Δ Egr3-DsRed2 localized to nuclei and chromosomes during meiotic progression. Microinjection of these cRNAs or *Egr3* siRNA in oocytes did not affect meiotic maturation. Immunofluorescence staining of *Egr3* in Xpress- Δ Egr3-DsRed2-injected oocytes showed a positive signal only on meiotic spindle, suggesting that this antibody does not detect endogenous or exogenous *Egr3* in mouse oocytes.

Conclusion: The results show that *Egr3* localizes to chromosomes during meiotic progression and that certain antibodies may not faithfully represent localization of target proteins in oocytes. *Egr3* seems to be dispensable during oocyte maturation in mice.

Keywords: Mouse; Oocytes; Ovary; Testis; Egr3

INTRODUCTION

The early growth response (Egr) family of zinc finger transcription factors regulates critical genetic programs in cellular growth, differentiation, and function [1,2]. There are four Egr members, Egr1, Egr2, Egr3, and Egr4, and each member has specific and overlapping functions in a given physiological setting. Some Egr factors have shown to be involved in reproductive function. Egr1 regulates expression of the luteinizing hormone beta subunit, and *Egr1* deficient mice suffer from female infertility resulting from hormone insufficiency [3]. Egr1 is induced by estrogen in the mouse uterus and expressed in stromal cells surrounding implanting blastocysts, suggesting a role for Egr1 in uterine biology [4]. Egr3 is implicated in neurodevelopment, learning and memory, immune response, and fibrogenic response [5-8], and this list shows functional diversity of this factor. As for *Egr3* deficient mice, neurodevelopmental phenotypes are well described [5].

Whether *Egr3* deficiency leads to other tissue-specific functions is just beginning to unravel with the availability of the *Egr3* floxed mice [9]. In reproduction, expression of *Egr3* in mouse oocytes and spermatocytes was reported by us [2]. In bovine granulosa cells, the administration of mycotoxin was shown to induce *Egr3* expression [10]. Expression of *Egr3* and *Egr4* in the pig ovary is also recently reported [11]. In male mice, *Egr4* exhibits a dynamic pattern of localization in germ cells and gonadal somatic cells depending on the stage of sexual maturity [12].

Previously, we showed that the immunoreactive *Egr3* colocalizes with meiotic spindle and accumulates near cytosolic microtubule organizing centers (MTOCs) in oocytes during meiotic maturation. As a transcription factor, *Egr3* was expected to localize to nucleus in cells and tissues, but localization of the immunoreactive *Egr3* was noted in the cytoplasmic structures in mouse oocytes, early preimplantation embryos, and spermatocytes [2]. Several transcriptional regulators including *Egr1* have been reported to exhibit similar localization on microtubule-associated structures [13-15].

In this study, we elaborate further on the expression of *Egr3* in gonads and its role in oocyte maturation. Whereas several pieces of information indicate involvement of *Egr* factors in female reproduction, it is not clear if *Egr3* is required for oocyte maturation in mice. Thus, we herein investigated if functional blockade of *Egr3* hinders meiotic maturation of oocytes by microinjecting dominant-negative *Egr3* complementary RNA (cRNA) and small interfering RNA (siRNA) specific to *Egr3*. The results show that *Egr3* is dispensable for meiotic maturation in mouse oocytes and that some of the commercially available anti-*Egr3* antibodies may not faithfully represent endogenous and exogenous *Egr3* proteins in oocytes.

MATERIALS AND METHODS

Mice

All mice were maintained in accordance with the policies of the Konkuk University Institutional Animal Care and Use Committee (IACUC). This study was approved by the Konkuk University IACUC (approval number KU15109). Mice were sacrificed after administration of avertin to minimize suffering. Four-week-old virgin ICR or BDF female mice were purchased from Orient-Bio (Seongnam, Korea).

Collection and culture of oocytes

Four-to-five-week-old virgin *CrljOri:CD-1* (ICR) female mice were intraperitoneally injected with 7.5 IU of pregnant mare's serum gonadotropin (Sigma-Aldrich, St. Louis, MO, USA) to induce folliculogenesis. Fully grown follicles were punctured 48 h later and cumulus-oocyte complexes were collected. The cumulus cells were then removed by using 300 µg/mL hyaluronidase (Sigma-Aldrich, USA) and germinal vesicle (GV) stage oocytes were retrieved in Quinn's Advantage Medium with HEPES (Sage BioPharma Inc., Bedminster, NJ, USA) containing 10% substitute protein serum and 200 µM 3-isobutyl-1-methylxanthine (IBMX) (I5879; Sigma-Aldrich, USA). Cumulus cells were gently removed from the cumulus-oocyte complexes using a Pasteur glass pipette. The oocytes were incubated in microdrops of M16 medium (M7292; Sigma-Aldrich, USA) containing 200 µM IBMX covered with mineral oil (Sage, USA) at 37°C under 5% CO₂ in atmosphere until microinjection was conducted. After microinjection, these oocytes were placed in drops of M16 medium with 200 µM IBMX for 8 h. For oocyte maturation, these oocytes were placed in IBMX-free M16 medium for 20 h. Rates of GV breakdown (GVBD) and polar body extrusion were monitored.

Reverse transcription-polymerase chain reaction

Reverse transcription-polymerase chain reaction (RT-PCR) was conducted as described previously [2]. Gonads were individually collected from 1, 2, 3, and 4 week-old female and male ICR mice. Each sample was subjected to RNA preparation by using TRI Reagent (Life Technologies, Invitrogen, Carlsbad, CA, USA) following the manufacturer's protocol. Isolated RNA (2 µg) was subjected to reverse transcription (RT) using M-MLV reverse transcriptase (Beams Biotechnology, Seongnam, Korea). PCR was carried out using Prime Taq Premix (Genet Bio, Daejeon, Korea). Primer sequence are given in Table 1. PCR reaction was also performed for the mouse ribosomal protein L7 (*Rpl7*), a housekeeping gene. Annealing temperature is 59°C for all reactions. This experiments were repeated 6 times with independent samples.

Plasmid constructs

Full-length mouse *Egr3* gene (NM_018781.3) was amplified by PCR from the mouse ovary cDNA sample [2]. To generate the dominant negative form of *Egr3* [16], the truncated form of *Egr3* (missing the first 249 amino acids, Δ*Egr3*) was generated by PCR from the full-length mouse *Egr3* cDNA. Full-length and truncated

Table 1. Primers used for reverse transcription-polymerase chain reaction

Gene name	Sequence (5'-3')	Product size (bp)	GenBank accession no.
<i>Egr3</i>	F: CAA TCT GTA CCC CGA GGA GAT	204	NM_018781.3
	R: GCG AAC TTT CCC AAG TAG GTC	Variant: 257	NM_001289925.1
<i>Rpl7</i>	F: TCA ATG GAG TAA GCC CAA AG	246	NM_011291.5
	R: CAA GAG ACC GAG CAA TCA AG		

F, forward; R, reverse.

Egr3 DNAs were inserted into BglII and AgeI site of pDsRed2-N1 vector (Clontech, Mountain View, CA, USA) and cloned into KpnI and XhoI site of pcDNA3.1/Myc-His vector (Invitrogen, USA). For *in vitro* transcription, full-length and truncated *Egr3* construct containing Xpress tag at the N-terminus and DsRed2 at the C-terminus were amplified by PCR and cloned into BamHI and HindIII site of pRSET-A vector (Invitrogen, USA). The constructs are shown in Figure 1A, 1B.

In vitro transcription and microinjection

For live imaging of *Egr3* expression in mouse oocytes, full-length and truncated *Egr3* cRNAs were produced by *in vitro* transcription using mMessage mMachin T7 transcription kit (Ambion, Austin, TX, USA). To visualize microtubule in some experiments, β -tubulin-green fluorescent protein (GFP) cRNA was produced from pRN3- β -tubulin-GFP construct (a gift from Marie-Hélène Verlhac, CIRB, Collège de France, Paris, France) using T3 transcription kit. These RNAs were cleaned and concentrated by using RNeasy MinElute Clean up kit (Qiagen, Valencia, CA, USA).

These constructs were confirmed by *in vitro* translation by using TNT T7 Coupled Reticulocyte Lysate System kit (Promega, Madison, WI, USA). SMARTpool *Egr3* siRNA (#M-046392-01-0005) and non-targeting siRNA (#D-001810-01-05) were purchased from Dharmacon (Lafayette, CO, USA). Knockdown capacity of this siRNA was first confirmed in 293T cells transfected with full-length *Egr3* plasmid. *Egr3* siRNA (25 μ M) was microinjected into GV oocytes and rates of GVBD and polar body extrusion were monitored.

To visualize *Egr3* localization by live imaging, *Egr3*-DsRed2 or truncated *Egr3*-DsRed2 cRNA (0.0034 μ M) was microinjected into GV oocytes. Injection volume was 5 to 10 pL, which is about 1% to 3% of total oocyte volume. Mouse oocytes were placed in drops of culture media containing 200 μ M IBMX for microinjection. Microinjection of cRNA or siRNA was performed using a Nikon (Nikon Corporation, Shinagawa-Ku, Tokyo, Japan) manipulator with picoinjector (Femtojet, Eppendorf, Hamburg, Germany) via pneumatic pressure.

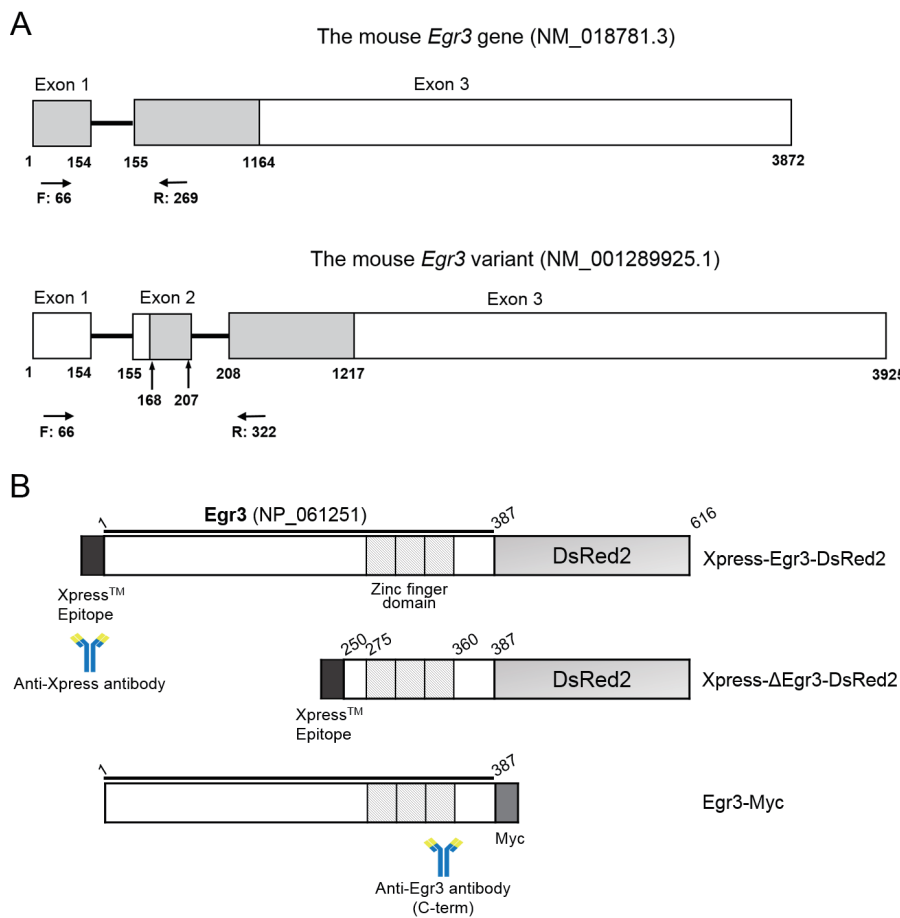


Figure 1. Diagrams depicting the structure of *Egr3* gene and constructs used in this study. (A) The mouse *Egr3* gene and the variant detected by RT-PCR. Coding regions are shown as gray shaded areas. Locations of forward (F) and reverse (R) primers are shown as arrows. The variant has another exon (exon 2) in the middle thus the PCR product is 53 bp longer. The predicted size of translated product is shorter because it uses a downstream start codon at 168. Gray box, translated region; while box, untranslated region; black line, intron. (B) Fusion constructs used in this study. Full-length and truncated (Δ Egr3 without the first 249 amino acids) were cloned into pRSET vector containing Xpress epitope at the N-terminus. At the C-terminus, DsRed2 fluorescence tag was cloned in frame for live imaging. The locations of epitopes for the anti-Xpress and anti-Egr3 antibodies are shown. *Egr3*, early growth response protein 3; RT-PCR, reverse transcription-polymerase chain reaction.

Immunofluorescence staining

The oocytes injected with *Egr3* siRNA or Xpress- Δ *Egr3*-DsRed2 RNA were fixed in 3.7% formaldehyde in phosphate-buffered saline (PBS) and permeabilized in 0.25% triton X-100 in PBS. They were blocked and incubated with 4 μ g/mL antibody or 2 μ g/mL antibody of specific primary antibodies. Primary antibodies are the rabbit polyclonal anti-*Egr3* antibody (sc-191; Santa Cruz Biotechnology, CA, USA) and the mouse anti-Xpress antibody (P/N 46-0528; Invitrogen, USA). Primary antibodies were probed with chick anti-rabbit IgG-Alexa Fluor 488 antibody and donkey anti-mouse IgG-Alexa Fluor 488 antibody (1:250; Invitrogen, USA). DNA was counter-stained with TO-PRO-3-iodide (1:500; Invitrogen, USA). Rabbit IgG was used at the same concentration as a mock control in each experiment. Images were obtained using the Olympus Fluoview FV1000 spectral confocal microscope (Tokyo, Japan) and analyzed using the software Fluoview version 1.5, a platform associated with the confocal microscope.

Protein extraction and western blotting

293T cells were co-transfected with pcDNA3.1/Myc-*Egr3* plasmid and *Egr3* siRNA. Cells transfected with empty plasmid or non-targeting siRNA served as negative controls. Cell lysates were prepared in RIPA buffer (50 mM Tris [pH 7.5], 150 mM NaCl, 1% Nonidet P-40, 0.5% Na-deoxycholate, 0.1% SDS, 1 mM dithiothreitol (DTT), 1 mM phenylmethylsulfonyl fluoride (PMSF), 1X Protease Inhibitor [Roche, Indianapolis, IN, USA]) and centrifuged at 13,000 rpm. The supernatant was subjected to bicinchoninic acid (BCA) assays (Thermo Scientific, Rockford, IL, USA) for quantitation. Fifty micrograms of protein were loaded per lane and separated in 10% SDS-polyacrylamide gel

electrophoresis (PAGE) gels. Western blotting was performed as previously described [2]. Anti- α -tubulin (Sigma-Aldrich, USA) and anti-*Egr3* (sc-191; Santa Cruz Biotechnology, USA) antibodies were used at 1:2,000 and 1:500, respectively. Anti-Myc antibody (#631206; Clontech, USA) was used at 1:2,000. Horseradish peroxidase (HRP)-conjugated goat anti-rabbit and goat anti-mouse antibodies (GeneDEPOT, Barker, TX, USA) were used at 1:10,000. Super Signal West Femto ECL reagent (Thermo Scientific, USA) was used. Chemiluminescence signal was detected by LAS3000 (FUJIFILM, Tokyo, Japan).

RESULTS

Expression of *Egr3* in ovaries and testes in mice

To examine if *Egr3* is expressed in mouse gonads at different stages of sexual maturation, we collected ovaries and testes from 1, 2, 3, and 4 week-old mice. The location of primers is depicted in Figure 1A. RT-PCR analysis showed that *Egr3* mRNA is expressed in ovaries and testes in all groups. We also found that an uncharacterized longer transcript of *Egr3* (NM_001289925.1) was present in 3 and 4-week-old gonads (Figure 2). Whether this form of *Egr3* is functional or expressed in other tissues is not known.

Microinjection of full-length and truncated Xpress-*Egr3*-DsRed2 does not hinder oocyte maturation

We previously showed that the immunoreactive *Egr3* is specifically localized near MTOCs and on spindle during meiotic maturation of oocytes [2]. We examined the subcellular localization of *Egr3* with live imaging after microinjecting full-length *Egr3* cRNA. This construct has an Xpress tag at N-terminus and

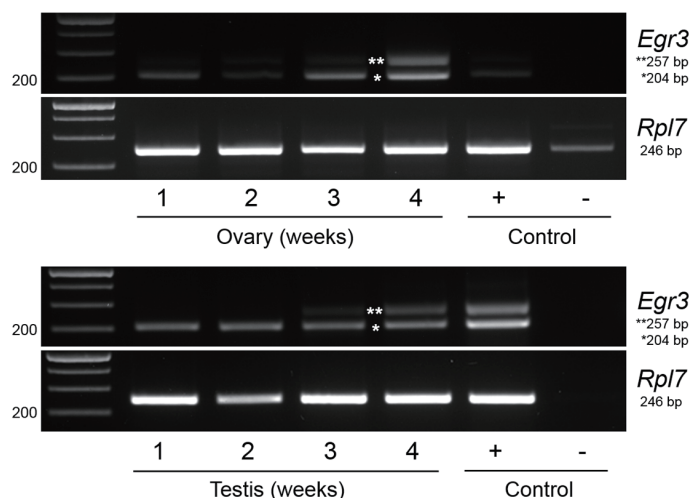


Figure 2. mRNA expression of the *Egr3* gene in mouse ovaries and testes from 1 to 4 week-old mice. (A) Mouse ovaries from 1 to 4 week-old mice were subjected to RT-PCR. RNA from 4 week-old mouse testis was used as a positive control (+), and RNA from 4 week-old ovary without reverse transcription (RT) was used as a negative control (-). Expression of a housekeeping gene, *Rpl7*, was also examined. (B) Mouse testes from 1 to 4 week-old mice were subjected to RT-PCR. RNA from 4 week-old mouse ovary was used as a positive control (+), and RNA from 4 week-old testis without RT was used as a negative control (-). *, *Egr3*; **, a longer variant (NM_001289925.1). *Egr3*, early growth response protein 3; RT-PCR, reverse transcription-polymerase chain reaction; *Rpl7*, ribosomal protein L7.

DsRed2 at C-terminus (Figure 1B). We also microinjected truncated Egr3 RNA (Xpress-ΔEgr3-DsRed2) lacking the first 249 amino acids, which is known as a dominant-negative form of Egr3 [16] (Figure 1B). As shown in Figure 3 by confocal live imaging, products of both Xpress-Egr3-DsRed2 and Xpress-ΔEgr3-DsRed2 cRNAs localize to nucleus and chromosomes during meiotic maturation. We then monitored if microinjection of these Egr3 RNAs affect meiotic maturation rate in oocytes. Microinjection of either construct did not influence the rate of meiotic maturation (Table 2).

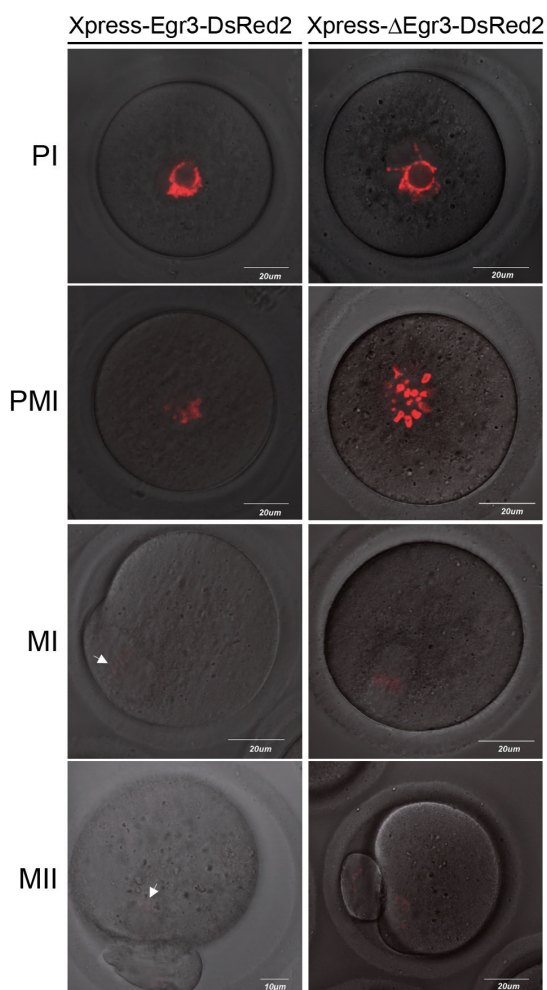


Figure 3. Microinjection of truncated Egr3 cRNA does not hinder oocyte maturation. Xpress-Egr3-DsRed2 or Xpress-ΔEgr3-DsRed2 cRNA (0.0034 μM) was microinjected into GV oocytes in the presence of 200 μM IBMX and oocytes were kept in M16 containing IBMX for 8 h. They were then washed and cultured in IBMX-free M16 medium for 20 h. Localization of microinjected constructs were monitored by confocal live imaging examining the expression of red fluorescent protein, DsRed2. During oocytes maturation, both Xpress-Egr3-DsRed2 or Xpress-ΔEgr3-DsRed2 proteins are localized in nuclei and chromosomes. Expression of Xpress-Egr3-DsRed2 protein is lower than that of Xpress-ΔEgr3-DsRed2, as translation efficiency of Xpress-Egr3-DsRed2 is lower due to its length (about 750 bp longer than Xpress-ΔEgr3-DsRed2). White arrows indicate chromosomes. PI, prophase I; PMI, prometaphase I; MI, metaphase I; MII, metaphase II; Egr3, early growth response protein 3; GV, germinal vesicle; IBMX, 3-isobutyl-1-methylxanthine.

Table 2. Meiotic maturation rate of oocytes injected with Xpress-Egr3-DsRed2 or Xpress-ΔEgr3-DsRed2 cRNA¹⁾

Group	Total oocytes	Oocytes maturation rate (%)	
		GVBD oocytes (%)	MII oocytes (%)
Xpress-Egr3-DsRed2	85	84 (98.82)	51 (60.00)
Xpress-ΔEgr3-DsRed2	78	74 (94.87)	52 (66.66)
Water injection	40	40 (100)	29 (72.50)
No injection	68	65 (95.59)	54 (79.41)

GVBD, germinal vesicle breakdown; MII, metaphase II; IBMX, 3-isobutyl-1-methylxanthine.

¹⁾ Microinjection was performed in the presence of IBMX and oocytes were incubated in M16 medium containing 200 μM IBMX for 8 h. They were then cultured in IBMX-free M16 medium for 20 h. GVBD was monitored after 3 to 5 h of culture and polar body extrusion (MII) after 20 h from the beginning of IBMX-free culture.

Meiotic maturation is not hindered by Egr3 siRNA in oocytes

We next tested if microinjection of Egr3 siRNA has any influence on meiotic maturation. The specificity of Egr3 siRNA was first confirmed by western blotting in 293T cells transfected with Egr3-Myc plasmid (Figure 4A). However, microinjection of Egr3 siRNA to GV oocytes did not affect GVBD or polar body formation (Table 3). Spindle forms normally in metaphase I (MI) oocytes injected with Egr3 siRNA, as evidenced by normal distribution of tubulin-GFP (Figure 4B).

Egr3 localization: potential artefact in immunolocalization

The observation that the microinjected Xpress-ΔEgr3-DsRed2 constructs are only confined to nuclei and chromosomes contradicts our previous observation that immunoreactive Egr3 localized to MTOCs and microtubules [2]. Thus, we side-by-side compared localization of Egr3 in microinjected oocytes by confocal live imaging, and anti-Xpress and anti-Egr3 immunofluorescence staining in fixed oocytes. As shown in Figure 5, DsRed2 signal is clearly confined to chromosomes. Anti-Xpress antibody detected a diffuse pattern of signal on or near metaphase chromosomes. This pattern is less clear than DsRed2 live imaging, but some signal overlaps with DNA. In contrast, anti-Egr3 antibody shows positive signal only on meiotic spindle (Figure 5). This anti-Egr3 antibody does not detect exogenous Xpress-ΔEgr3-DsRed2, because the signal is negative on chromosomes. This

Table 3. Meiotic maturation rate of oocytes injected with Egr3 siRNA or non-targeting siRNA¹⁾

Group	Total oocytes	Oocytes maturation rate (%)	
		GVBD oocytes (%)	MII oocytes (%)
Egr3 siRNA	167	164 (98.20)	119 (71.26)
Non-targeting siRNA	155	146 (94.19)	116 (74.84)
No injection	63	62 (98.41)	45 (71.42)

GVBD, germinal vesicle breakdown; MII, metaphase II; IBMX, 3-isobutyl-1-methylxanthine.

¹⁾ Microinjection was performed in the presence of IBMX and oocytes were incubated in M16 medium containing 200 μM IBMX for 8 h. They were then cultured in IBMX-free M16 medium for 20 h. GVBD was monitored after 3 to 5 h of culture and polar body extrusion (MII) after 20 h from the beginning of IBMX-free culture.

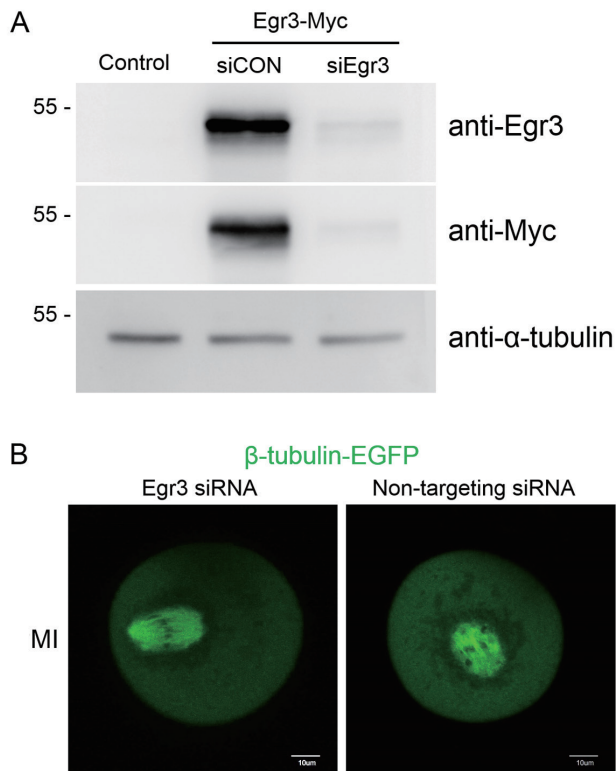


Figure 4. Microinjection of *Egr3* siRNA does not hinder oocyte maturation. (A) Western blot analysis of Egr3 in 293T cell co-transfected with pcDNA3.1/myc-his-Egr3 (Egr3-Myc) and *Egr3* siRNA (siEgr3) or non-targeting siRNA (siCON). Empty plasmid served as a negative control (Control). These cells were subjected to western blot analysis with anti-Egr3 antibody. Fifty micrograms of cell lysates were run on 10% SDS-PAGE gels. (B) Microinjection of *Egr3* siRNA in GV oocytes does not interfere with meiotic maturation. Confocal live image of meiotic spindle after co-injection of 0.5 μ g/ μ L β -tubulin-EGFP cRNA and 25 μ M *Egr3* siRNA or non-targeting siRNA. After microinjection, the oocytes were incubated in M16 medium supplemented with 200 μ M IBMX for 8 h, then they were washed and cultured in IBMX-free M16 medium for 6 h. Spindle forms normally in both groups. MI, metaphase I; *Egr3*, early growth response protein 3; SDS-PAGE, SDS-polyacrylamide gel electrophoresis; GV, germinal vesicle; EGFP, enhanced green fluorescent protein; IBMX, 3-isobutyl-1-methylxanthine.

result suggests that spindle-positive signal detected by anti-Egr3 antibody seems to be nonspecific.

DISCUSSION

In this study, we show that mouse gonads express *Egr3* mRNA, both at immature and sexually maturing stages of age. Herein we present experiments to test functionality of Egr3 in mouse oocytes, providing evidence that Egr3 is dispensable for meiotic maturation in mouse oocytes.

In RT-PCR analysis, a bigger, uncharacterized *Egr3* isoform was noted in 3 and 4-week ovaries. This variant, NM_001289925.1, has another exon inserted between two original exons of the mouse *Egr3* gene (Figure 1A). This form using a downstream start codon is expected to produce a shorter product with distinct N-terminus. Several isoforms of Egr3 have been reported in brain and they seem to have different transcriptional activity [17,18]. Interestingly, the appearance of this variant is associated with sexual maturation of the ovary. Whether this particular variant in mouse gonads is expressed in brain or other tissues and functionally active is currently unknown.

Egr3 without the N-terminal half of the protein is known as a dominant-negative form which blocks activity of Egr factors [16]. Commercially available siRNA specific to *Egr3* efficiently downregulates translation of Egr3 (Figure 4A). Microinjection of either dominant-negative Egr3 or *Egr3* siRNA did not influence the rates of GVBD and PB extrusion, suggesting that Egr3 is functionally dispensable during oocyte maturation (Tables 2, 3). During the course of this study, we noticed that Xpress-Egr3-DsRed2 and Xpress- Δ Egr3-DsRed2 proteins are confined only on nuclei and chromosomes after microinjection when observed by live imaging (Figure 3). This is in discrepancy with our previous observation that anti-Egr3 antibody produced signal near MTOC and on spindle of maturing oocytes. In our previous

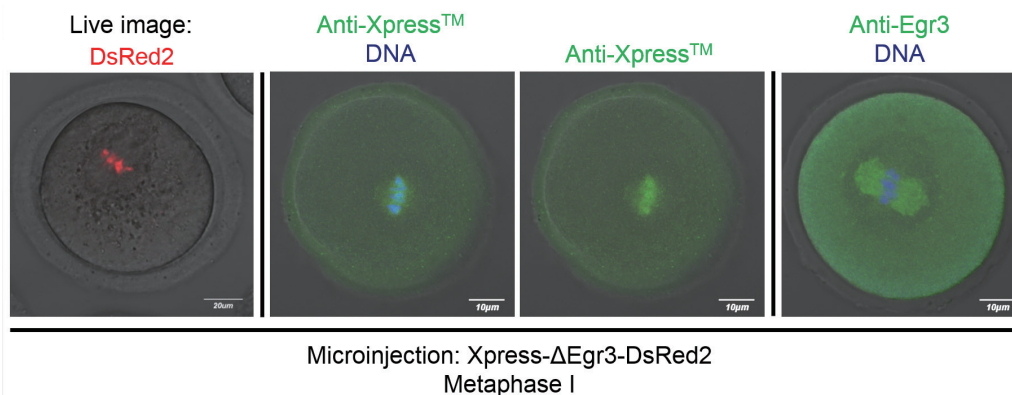


Figure 5. Various methods of validating Egr3 localization in mouse oocytes and potential artefacts. Confocal live imaging and immunofluorescence staining with anti-Xpress and anti-Egr3 antibodies were compared in MI oocytes injected with Xpress- Δ Egr3-DsRed2 cRNA. GV oocytes were microinjected with Xpress- Δ Egr3-DsRed2 cRNA under IBMX and incubated for 8 h in M16 medium supplemented with IBMX. For oocyte maturation, these oocytes were placed in IBMX-free M16 medium for 8 h. At MI stage, oocytes were either subjected to confocal live imaging or immunofluorescence staining with indicated antibodies. *Egr3*, early growth response protein 3; MI, metaphase I; GV, germinal vesicle; IBMX, 3-isobutyl-1-methylxanthine.

study, we had tested five commercially available anti-Egr3 antibodies and two of them had produced the observed pattern of localization in oocytes [2] (Figure 5). Both antibodies, detecting C-terminal region of Egr3 protein, detect Egr3 in western blotting and recognize exogenous Egr3 in nuclei of PC12 or MCF7 cells. Interestingly, one of these antibodies also detect immunoreactive Egr3 in growing neurites of PC12 cells (data not shown). Thus, certain anti-Egr3 antibody recognizes microtubule-based subcellular structures both in PC12 and mouse oocytes; neurite and meiotic spindle. The similarity between these structure is evident, i.e., they both utilize microtubule. However, our side-by-side comparison in Figure 5 shows that anti-Egr3 antibody produces a distinct pattern of subcellular localization compared to live imaging and anti-Xpress immunofluorescence staining. Anti-Xpress antibody detecting Xpress tag on the fusion construct (Figure 1B) produced a signal much closer to Egr3-DsRed2 live imaging (Figure 5), although the pattern was more diffused than that of DsRed2 live imaging. While we vigorously validated the specificity of antibodies in our previous investigation [2], it is possible that this anti-Egr3 antibody might produce artefactual pattern of localization in oocytes. Antibodies are often the most common culprits of problems in biological experiments due to non-specific binding to other proteins [19]. Whether this particular anti-Egr3 antibody detects other target(s) or processed form of Egr3 protein is not clear and it requires further validation.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

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