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SPECIALTY SECTION

This article was submitted to Evolutionary and Population Genetics, a section of the journal Frontiers in Genetics

RECEIVED 16 May 2022 ACCEPTED 08 August 2022 PUBLISHED 05 October 2022

CITATION

Xu B, Yang G, Jiao B and Zhu H (2022), Analysis of ancient and modern horse genomes reveals the critical impact of IncRNA-mediated epigenetic regulation on horse domestication. *Front. Genet.* 13:944933. doi: 10.3389/fgene.2022.944933

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Analysis of ancient and modern horse genomes reveals the critical impact of IncRNA-mediated epigenetic regulation on horse domestication

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Background: The domestication of horses has played critical roles in human civilizations. The excavation of ancient horse DNA provides crucial data for studying horse domestication. Studies of horse domestication can shed light on the general mechanisms of animal domestication.

Objective: We wish to explore the gene transcription regulation by long noncoding RNAs (IncRNAs) that influence horse domestication.

Methods: First, we assembled the ancient DNA sequences of multiple horses at different times and the genomes of horses, donkeys, and Przewalski horses. Second, we extracted sequences of lncRNA genes shared in ancient horses and sequences of lncRNA genes and the promoter regions of domestication-critical genes shared in modern horses, modern donkeys, and Przewalski horses to form two sample groups. Third, we used the LongTarget program to predict potential regulatory interactions between these lncRNAs and these domestication-critical genes and analyzed the differences between the regulation in ancient/modern horses and between horses/donkeys/ Przewalski horses. Fourth, we performed functional enrichment analyses of genes that exhibit differences in epigenetic regulation.

Results: First, genes associated with neural crest development and domestication syndrome are important targets of lncRNAs. Second, compared with undomesticated Przewalski horses, more lncRNAs participate in the epigenetic regulation in modern horses and donkeys, suggesting that domestication is linked to more epigenetic regulatory changes. Third, lncRNAs' potential target genes in modern horses are mainly involved in two functional areas: 1) the nervous system, behavior, and cognition, and 2) muscle, body size, cardiac function, and metabolism.

Conclusion: Domestication is linked to substantial epigenetic regulatory changes. Genes associated with neural crest development and

domestication syndrome underwent noticeable lncRNA-mediated epigenetic regulation changes during horse domestication.

KEYWORDS

domestication, IncRNA, epigenetic regulation, neural crest cell, horse

1 Introduction

The domestication of animals and plants has greatly changed human life. For example, pigs and cattle provide food for humans, cats and dogs are companions for people, wheat and rice are the main food crops for humans (Kilian et al., 2010), and horses and camels are important means of human transportation (Vigne, 2011; Zeder, 2012). Researchers study animal domestication mainly using two methods. One method is based on archaeological records, which analyze the changes in animal body structure based on fossils from different periods and locations (Conolly et al., 2011). Animal skeletal changes, age at death, sex ratio, and isotopic characteristics can be used as indicators for analyzing domestication in archaeological studies. However, archaeological data are fragmented, and many characteristic differences may have already existed between domesticated animals and their wild ancestors in the early domestication phase, including coat color, temperament, and reproductive ability, which cannot be discovered through skeletal archaeology. The other method analyzes the genome of modern domesticated animals to obtain temporal and geographic information on domestication and to discover the genetic basis of domestication (Luikart et al., 2001; Vilà et al., 2001; Pedrosa et al., 2005; Naderi et al., 2008), including inferring the initial domestication centers based on the genetic diversity generated by geographical differences. A large amount of phenotypic diversity exists among and within species of modern domesticated animals, and genomic analysis can reveal the influence of specific loci on phenotypes, including coat color, body size, fat content, biological clock, and behavior (Wright, 2015). However, genomic information obtained from living animals can reveal only a small fraction of domestication information. Over the past decade, researchers have gained access to a wealth of genomic information from ancient species using new sequencing techniques. The first extraction of ancient animal DNA, an extinct zebra species, was achieved in the 1980s (Higuchi et al., 1984). Since then, the combined archaeological and genomic analysis have led to a better understanding of the role of location, timing, and artificial selection in the domestication of animals.

Domesticated mammals share common traits. Especially, in the early stage, all domesticated mammals yield reduced fear and reactive aggression. Wilkins et al. (2014) focused on a special group of cells found in embryos, neural crest cells (NCCs), proposed that the genetic changes affecting their development were at the root of vertebrate domestication, and termed these traits the "neural crest/domestication syndrome" (NCDS). NCCderived tissues include the major parts of the jaws and teeth, pigmentation cells, components of the external ears, and cells involved in sympathetic responses docility. Many genes are required for NCC formation and development (which are referred to as neural crest/domestication syndrome-associated genes, or NCDS genes in this study), but their roles in the NCC genetic regulatory network remain poorly known (Simoes-Costa and Bronner, 2015). It is proposed that the neural crest cell contribution to final target tissues could be modified by mutations (Wilkins et al., 2021), but it is questionable whether sufficient mutations can be generated and accumulated in a short period for domestication. Whether the NCDS hypothesis provides a unified explanation for domestication, these genes should be important targets of combined archaeological and genomic analysis for revealing the mechanisms of animal domestication.

The domestication of horses not only revolutionized the way humans migrated but also changed the way human activities and diseases were spread and goods were distributed, allowing for the first real globalization of humankind (Kelekna, 2009). Fossils of horses at all stages of domestication have been excavated and reported (Orlando, 2015), with related studies reporting the time and place of horse domestication (Orlando, 2020; Librado et al., 2021). Early studies of ancient DNA analysis focused on proteincoding genes, and no computational studies on lncRNAs and their potential epigenetic target genes have been reported. However, most protein-coding genes are highly conserved in mammals and would unlikely have changed much in just a few thousands of years. We posit that it should be the regulation of the expression of these genes, but not the sequence of these genes, that undergoes significant changes during domestication. Gene expression is regulated by multiple mechanisms, including transcriptionally by transcription factors and long noncoding RNAs (lncRNAs). Many lncRNAs can bind to DNA sequences by forming RNA: DNA triplexes and recruit epigenomic modification enzymes to DNA binding sites. Thus, transcription factors and their DNA-binding sites and lncRNAs and their DNA-binding sites are two mechanisms controlling gene expression changes. Mammalian lncRNA genes are less conserved, and many are species-specific (Ulitsky, 2016; Sarropoulos et al., 2019), suggesting that they may have a greater effect on gene expression than transcription factors within a short period. Therefore, they should be essential regulators of domestication. An example from dog domestication suggests that diverse phenotypic changes can be produced in a

| Sample | Age (BP or BCE) | Genome/GFF version | Assembly level | Source |
|------------------------|-----------------|--------------------|----------------|-------------------------|
| CGG10022 | 42,012 (BCE) | _ | Shotgun reads | Schubert et al. (2014b) |
| Botai_2_5500 | 5,500 | _ | BAM(2.0) | Gaunitz et al. (2018) |
| Borly4_PAVH8_4978 | 4,071 | _ | BAM(2.0) | Gaunitz et al. (2018) |
| 838-64_NB46 | 4,978 | _ | Shotgun reads | Librado et al. (2017) |
| UushgiinUvur_Mon84_312 | 3,123 | _ | BAM (2.0) | Gaunitz et al. (2018) |
| BER09_I | 2,300 | _ | Shotgun reads | Librado et al. (2017) |
| I-K3_Arz2 | 2,760 | _ | Shotgun reads | Librado et al. (2017) |
| Yenikapi_Tur172_1695 | 1,695 | _ | BAM (2.0) | Gaunitz et al. (2018) |
| Modern horse | | EquCab3.0 | Genome | NCBI |
| Modern donkey | | ASM303372v1 | Scaffold | NCBI |
| Przewalskii horse | | GCA_000696695.1 | Scaffold | NCBI |

TABLE 1 Sample information.

Data download links or accession numbers: CGG10022: https://www.ebi.ac.uk/ena/browser/view/PRJEB7537; Botai_2_5500: https://www.ebi.ac.uk/ena/browser/view/PRJEB22390; BER09_I: https://www.ebi.ac.uk/ena/browser/view/PRJEB19970; Modern horses: https://www.ncbi.nlm.nih.gov/assembly/GCF_002863925.1/; Modern donkey: https://www.ncbi.nlm.nih.gov/assembly/GCF_002863925.1/; Przewalskii horse: https://www.ncbi.nlm.nih.gov/assembly/GCF_00286925.1/

short time without much change in the genome (Larson et al., 2012; Botigué et al., 2017; Pendleton et al., 2018)

In this study, we used two groups of published data to analyze the effects of lncRNAs on horse domestication. One group contains the DNA sequences from ancient and modern horses, and the other group contains the DNA sequences of modern horses, modern donkeys (a closely related species to horses), and Przewalski horses. The Przewalski horse was once assumed to be the only surviving wild horse (Schubert et al., 2014b), but the sequencing of its genome and subsequent genome analysis in 2018 revealed that it is actually a descendant of the ancient Botai horse. In other words, the Przewalski horses are semi-domesticated or undomesticated horses (Gaunitz et al., 2018); thus, it provides important information on horse domestication when combined with ancient horse DNA. The promoter regions of genes are the most important binding sites of lncRNAs and are critical for gene expression. To make data analyses less influenced by ancient DNA's relatively poor sequencing quality, we only used promoter regions to predict lncRNAs' binding sites and analyze lncRNAmediated gene expression regulation. Our results support the hypothesis that lncRNAs and NCDS genes significantly influenced the domestication of horses. The results also indicate that domestication is associated with an increase in epigenetic regulation.

2 Methods

2.1 Samples and candidate genes

We examined the DNA sequencing data of 8 ancient horses (Table 1). Ancient DNA sequencing results with relatively good quality were obtained from three ancient horses in three typical



the ancient horse Botai_2_5500.

periods: a 40,000-year-old wild horse (CGG10022), a 5,500-yearold Botai horse (Botai_2_5500), and a 2300-year-old ancient horse (BER09_I). CGG10022 was undomesticated, and BER09_I was largely (but not completely) domesticated. These ancient genomes, together with the genome of modern horses, form a



chronological data group (called the "CGG/Botai/BER/modern horse group"). Moreover, the genomes of modern horses, modern donkeys, and Przewalski horses form an interspecific data group (called the "modern horse/modern donkey/ Przewalski horse group") (Table 1; Figure 1).

Researchers identified 125 genes with positive selection signals related to horse domestication (Supplementary Table S1) (Schubert et al., 2014b). These genes can be divided into two groups. One group of genes is involved in muscle and limb development, joint connectivity, and the cardiac system; the other is involved in cognitive functions, including social behavior, learning ability, fear response, and conformation. These genes are considered essential common genes involved in domestication. Librado et al. (2017) found that the NCDS genes (Supplementary Table S2), 15 of which have been reported in the relevant literature (Wilkins et al., 2014), underlie the phenotypic differences between wild and domesticated horses. We examined the intersection of the 125 positively selected genes (Schubert et al., 2014b) in the modern horse/modern donkey/ Przewalski horse group and the intersection of the 15 NCDS genes (Wilkins et al., 2014) in the modern horse/modern donkey/

Przewalski horse group, respectively. 73 out of the 125 positive selection-related genes and 10 out of the 15 NCDS genes were identified in all three species (Supplementary Table S3).

2.2 Extracting ancient DNA sequences of long noncoding RNAs and promoter regions of genes

Since promoter regions are the most important regions for lncRNA-mediated epigenetic regulation, we only analyzed the promoter regions of candidate genes. In the CGG/Botai/BER/ Modern horse group, the DNA sequencing results of CGG10022 and BER09_I were shotgun reads, so we processed all data with the following steps (Figure 2). 1) All data of ancient horse genomes were processed using the ancient DNA sequence processing pipeline PALEOMIX (Version 1.2.13.2) (Schubert et al., 2014a). 2) Then, lncRNA genes and promoter regions of candidate genes were identified. 3) Ancient DNA reads were aligned to the modern horse reference genome EquCab3.0 to generate VCF files. 4) The location of each gene's transcription start site (TSS) was determined according to the EquCab3.0 gene annotation file. 5) The FASTA sequence of the ancient DNA was generated using the *vcf_to_fasta* command of PALEOMIX. 6) Sequences -3,500 bp-+1500 bp upstream/downstream of the TSS of the candidate genes were obtained as the promoter regions. 7) The exon sequences of each lncRNA were obtained and assembled into lncRNA sequences.

The DNA sequencing results of Botai_2_5500 have a pairwise sequence alignment with the modern horse genome EquCab2.0. However, no lncRNA genes are annotated in EquCab2.0. We used the *liftover* tool in the UCSC Genome Browser (http://genome.UCSC.edu) to convert the positions of exons in EquCab3.0 to those in EquCab2.0 and then used PALEOMIX's *vcf_to_fasta* to generate the FASTA sequences and obtain the lncRNA sequences. The promoter regions of the candidate genes were similarly obtained, which ranged from -3,500 bp-+1500 bp upstream/downstream of the TSS.

In the modern horse/modern donkey/Przewalski horse group, promoter regions (-3,500 bp-+1500 bp upstream/ downstream of the TSS) and lncRNA sequences were extracted based on genome positions in the downloaded genome sequence files in FASTA format and gene annotation files.

2.3 Filtering of the long noncoding RNA genes and promoter regions of candidate genes

The DNA sequences of CGG/Botai/BER ancient horses have considerable "N"s. We chose only lncRNA genes with "N" < 10% to filter out unreliable lncRNA genes. Upon this criterion, we identified 65 lncRNA genes in the intersection between CGG10022, Botai_2_5500, BER09_I, and modern horses (Supplementary Table S4). Upon the criterion "N" < 30%, we identified 22,341 promoter regions in the three ancient horses (CGG/Botai/BER ancient horses).

The genomes of the modern horse/donkey/Przewalski horse contain many lncRNA genes, but what lncRNAs function in the nucleus and in the cytoplasm are unknown. A lncRNA capable of regulating gene expression in the nucleus may bind to duplex DNA sequences by forming triplexes and recruit epigenomic modification enzymes to binding sites. Each triplex comprises triplex-forming oligonucleotides (TFO) in the lncRNA and a triplex-targeting site (TTS) in the DNA sequence. Overlapping TFOs indicate a DNA binding domain (DBD) in the lncRNA, and overlapping TTSs indicate a DNA binding site (DBS) in the DNA sequence. We first used the Triplexator program (which is very fast) to predict lncRNAs' TTSs in promoter regions genomewide (Buske et al., 2012). We assumed that if a lncRNA had > 2,000 TTSs, the lncRNA likely regulate gene expression in the nucleus. This empirical cutoff helped us choose the proper lncRNAs. A total of 4,562 lncRNA genes were annotated in the horse genome, 987 of which had > 2,000 TTSs. In addition, if multiple DBDs were predicted in a lncRNA, only the DBSs that correspond to the best DBDs (which have the most DBSs) were analyzed. Compared with the horse genome, only 900 lncRNA genes were annotated in the donkey genome. Due to the lack of annotated lncRNA genes in donkeys and Przewalski horses, we used the BLAST program to search the 987 horse lncRNAs against the modern donkey and Przewalski horse genomes (Johnson et al., 2008). The search identified 378 homologous lncRNA genes in modern horses, modern donkeys, and Przewalski horses (Supplementary Table S5).

2.4 Predicting DBSs in the promoter regions of genes

In all, 65 lncRNA genes and 22,341 promoter regions were obtained from the genomes of the CGG/Botai/BER/Modern horse group, and 378 lncRNA genes and the promoter regions in the 83 key domestication-related genes were obtained from the genomes of the modern horse/modern donkey/Przewalski horse group. We used the LongTarget program we developed previously (Lin et al., 2019) to predict the DBSs of these lncRNAs in these promoter regions, which helped us find which genes might be the targets of these lncRNAs. In total, 4 (samples) × 65 (lncRNAs) × 22,341 (promoter regions) (Supplementary Material S1-S4) and 3 (samples) × 378 (lncRNAs) × 83 (promoter regions) predictions were performed (Supplementary Material S5-S7). The default LongTarget parameters were used (*Identity* \geq 60%, which means > 60% RNA: DNA bases are paired, and Minimal *triplex* \geq 50, which means DBSs \geq 50 bp).

We computed the binding affinity for each pair of lncRNA and DBS to measure the strength of lncRNA binding. Binding affinity was defined as the product of DBS length and the averaged Identity values of all triplexes at the site. Thus, a binding affinity \geq 60 means DBS \geq 100 bp if the Identity = 60%. In the modern horse/modern donkey/Przewalski horse group, only DBSs with affinity > 60 were analyzed.

To check whether the sequencing quality of ancient horses influences DBS prediction, we examined DBSs' mean length in the three ancient horses and in the three modern species. The results indicate that DBSs have comparable lengths in the two groups (Table 2).

2.5 Calculating sequence differences in the long noncodingRNAs in the CGG/ Botai/BER/Modern horse group

To estimate lncRNAs' sequence differences in the CGG/ Botai/BER/Modern horse group, we first performed multiple sequence alignment using Clustalw2 (Larkin et al., 2007).

| Species | LncRNA number | DBS number | Mean DBS length | Mean binding affinity |
|-------------------|---------------|------------|-----------------|-----------------------|
| CGG10022 | 65 | 70,445 | 140 | 89 |
| Botai_2_5500 | 65 | 49,598 | 120 | 77 |
| BER09_I | 65 | 54,283 | 130 | 82 |
| Modern horse | 376 | 28,825 | 157 | 101 |
| Modern donkey | 376 | 22,537 | 156 | 100 |
| Przewalski horses | 319 | 11,007 | 136 | 87 |

TABLE 2 LncRNAs' number and DBSs' number, mean length, and mean affinity value in samples in the two groups.

DBSs, in ancient horses were predicted in 22,341 promoter regions, and DBSs, in modern horses, modern donkeys, and Przewalski horses were predicted in the promoter regions of 378 genes.

Then, we used the Phylip program (http://evolution.genetics. washington.edu/phylip.html) to calculate the sequence differences.

2.6 Identification of the species specificity of DBSs

To examine whether lncRNAs' target genes are conserved, we used each DBS in the modern horse genome as the query sequence to search for orthologous sequences in the genomes of modern donkeys and Przewalski horses. The sequence search was performed using the BLAST program, and the 100,000 bp region upstream/downstream of the TSS of the corresponding gene was the target region. If no orthologous sequence was obtained for a DBS, the DBS was assumed to be modern horse specific.

2.7 Analysis of the functional enrichment of target genes

The functions of genes in horses and donkeys are poorly annotated because published studies remain very limited. We determined the functions of horse and donkey protein-coding genes based on the GeneCards Human Gene database (https:// www.genecards.org/), assuming that these protein-coding genes have the same functions in humans, horses, and donkeys.

We performed functional enrichment analysis for the epigenetic target genes of lncRNAs in both the CGG/Botai/ BER/Modern horse group and the modern horse/modern donkey/Przewalski horse group using the g: Profiler program (https://biit.cs.ut.ee/gprofiler/gost) (Raudvere et al., 2019). For the modern horse/modern donkey/Przewalski horse group, functional enrichment analysis was applied to the genes shared by the three species and the genes specific to modern horses, respectively (g:Profiler parameters: Organism = Equus caballus, Ordered query = No, Statistical domain scope = All known genes, Statistical threshold = Benjamini–Hochberg FDR, User threshold = 0.05, 2< term size < 2,000).

3 Results

3.1 Domestication is associated with an increase in epigenetic regulation

We used the genome data of the three ancient horses to analyze the regulatory function of lncRNAs on gene expression in ancient horses and used the genome data of modern horses, modern donkeys, and Przewalski horses to analyze the regulatory function of lncRNAs in modern horses and related species (Table 1). Given that most protein-coding genes have unlikely evolved much in the short domestication period, we postulated that gene expression linked to horse domestication has been more subject to epigenetic regulation. In the CGG/Botai/BER/ Modern horse group, 65 lncRNA genes and 22,341 promoter regions of genes were analyzed. We first examined whether lncRNA sequences show clear divergence across ancient and modern horses. Agreeing with that predicted DBSs in ancient and modern horses have a comparable length (Table 2), the computed sequence differences indicated that these lncRNA sequences have limited variation (especially in ancient horses) (Table 3), from which we infer that it is their DBSs that may have played important roles in the changes of domestication-related epigenetic regulation.

Then, we predicted lncRNAs' DBSs, assuming that genes at DBSs were lncRNAs' target genes. For the CGG/Botai/BER/ Modern horse group, 65 lncRNAs had DBSs in the promoter regions of 2,988 genes (Supplementary Table S6). For example, the lncRNA LOC106783351 had 1104 DBSs in 644 proteincoding genes and 358 noncoding genes, and the lncRNA LOC11773204 had 1492 DBSs in 657 protein-coding genes and 327 noncoding genes. The target genes with high DBS binding affinity were mainly immune and nervous systemrelated. The functional enrichment analysis indicates that target genes are mainly enriched in immune, metabolism, and response to external stimulus-related GO terms (Figure 3A) and metabolism and neuroactive ligand-receptor interaction pathways (Figure 3B). Notably, more DBSs were predicted in the undomesticated CGG10022 (than in the semi-domesticated Botai_2_5500 and BER09_I), but in modern species, more DBSs TABLE 3 Distances between the 65 lncRNAs in the CGG/Botai/BER ancient horse group.

| Sample | CGG10022 | Botai_2_5500 | BER09_I | Modern horse |
|--------------|----------|--------------|----------|--------------|
| CGG10022 | 0 | 0.000285 | 0.000417 | 0.000926 |
| Botai_2_5500 | 0.000285 | 0 | 0.000288 | 0.000547 |
| BER09_I | 0.000417 | 0.000288 | 0 | 0.000561 |
| Modern horse | 0.000926 | 0.000547 | 0.000561 | 0 |

| | GO:BP | | stats | |
|---|--|--|--|--------------------------------|
| | Term name | Term ID | Padj | 0 ^{_log10(padj)} ≤16 |
| | intracellular signal transduction | GO:0035556 | 2.726×10 ⁻¹⁶ | |
| | immune system process | GO:0002376 | 1.433×10 ⁻¹³ | |
| | regulation of localization | GO:0032879 | 1.444×10 ⁻¹² | |
| | regulation of catalytic activity | GO:0050790 | 2.958×10-12 | |
| | regulation of intracellular signal transduction | GO:1902531 | 1.150×10 ⁻¹¹ | |
| | response to external stimulus | GO:0009605 | 3.638×10 ⁻¹¹ | |
| | positive regulation of response to stimulus | GO:0048584 | 1.038×10 ⁻¹⁰ | |
| | regulation of multicellular organismal process | GO:0051239 | 1.078×10-10 | |
| | positive regulation of molecular function | GO:0044093 | 2.306×10 ⁻¹⁰ | |
| | organic substance transport | GO:0071702 | 2.560×10-10 | |
| | response to biotic stimulus | GO:0009607 | 3.595×10-10 | |
| | movement of cell or subcellular component | GO:0006928 | 1.258×10 ⁻⁹ | in the second |
| | biological process involved in interspecies interaction b | GO:0044419 | 1.281×10 ⁻⁹ | |
| | regulation of immune system process | GO:0002682 | 2.097×10 ⁻⁹ | |
| | immune response | GO:0006955 | 3.255×10 ⁻⁹ | |
| | regulation of immune response | GO:0050776 | 1.394×10 ⁻⁸ | |
| В | KEGG | | stats | |
| | Term name | Term ID | Padj | 0 ^{-log} 10(padj) ≤16 |
| | Amoebiasis | KEGG:05146 | 4.706×10 ⁻⁴ | |
| | Staphylococcus aureus infection | KEGG:05150 | 4.706×10 ⁻⁴ | |
| | Metabolic pathways | KEGG:01100 | 6.185×10 ⁻⁴ | |
| | Endocytosis | KEGG:04144 | 6.185×10 ⁻⁴ | |
| | Neuroactive ligand-receptor interaction | KEGG:04080 | 8.464×10 ⁻⁴ | |
| | Complement and coagulation cascades | KEGG:04610 | 1.049×10 ⁻³ | |
| | Pertussis | KEGG:05133 | 2.962×10 ⁻³ | |
| | Regulation of actin cytoskeleton | KEGG:04810 | 4.527×10 ⁻³ | |
| | Calcium signaling pathway | KEGG:04020 | 1.072×10 ⁻² | |
| | Cytokine-cytokine receptor interaction | KEGG:04060 | 1.072×10 ⁻² | |
| | | KE00:02012 | 1.386×10 ⁻² | |
| | Nucleocytoplasmic transport | KEGG-03013 | | |
| | Nucleocytoplasmic transport Phosphatidylinositol signaling system | KEGG:03013 | 1.907×10 ⁻² | |
| | Nucleocytoplasmic transport Phosphatidylinositol signaling system Viral protein interaction with cytokine and cytokine rece | KEGG:04070 KEGG:04061 | 1.907×10 ⁻² 3.109×10 ⁻² | |
| | Nucleocytoplasmic transport Phosphatidylinositol signaling system Viral protein interaction with cytokine and cytokine rece Inflammatory mediator regulation of TRP channels | KEGG:04070 KEGG:04061 KEGG:04750 | 1.907×10 ⁻² 3.109×10 ⁻² 3.654×10 ⁻² | |
| | Nucleocytoplasmic transport Phosphatidylinositol signaling system Viral protein interaction with cytokine and cytokine rece Inflammatory mediator regulation of TRP channels Cell adhesion molecules | KEGG:04070 KEGG:04061 KEGG:04750 KEGG:04514 | 1.907×10 ⁻² 3.109×10 ⁻² 3.654×10 ⁻² 3.654×10 ⁻² | |

FIGURE 3

Target genes of the 65 lncRNAs in the CGG/Botai/BER/modern horses are enriched in immune, metabolism, and neuroactive ligand-receptor interaction related GO terms (A) and KEGG pathways (B).

were predicted in the domesticated horses (Table 2). This is confirmed when we examined DBSs in the NCDS genes. This result could indicate a substantial turnover of DBSs during domestication.

Two results were obtained from the modern horse/donkey/ Przewalski horse group. First, of the 378 lncRNAs shared in the three species, 376 in modern horses and donkeys but 319 in Przewalski horses have DBSs, indicating that more lncRNAs have functions in the domesticated horse and donkey. The 376 lncRNAs have 28,825 and 22,537 DBSs in 83 positively selected or NCDS genes in modern horses and modern donkeys, but the 319 lncRNAs have only 11,007 DBSs in the 83 genes in Przewalski horses (Tables 2, 4). Second, to confirm that domesticated horses acquired specific epigenetic regulation, we used the BLAST program to perform six pairwise sequence searches to examine how many DBSs obtained in a species are

TABLE 4 Orthologues of DBSs in different species.

| No hits |
|----------------|
| 13,756 (47.7%) |
| 7,781 (27.0% |
| 8,546 (37.9%) |
| 3,949 (17.5%) |
| 1,013 (9.2%) |
| 1,591 (14.5%) |
| |



FIGURE 4

The Binding affinity of DBSs in species and the number of DBSs in different NCDS genes. (A) LncRNAs' mean binding affinity at DBSs in modern horses, modern donkeys, and Przewalski horses (indicated by H, D, and P, respectively). The label "H > D > P 44%" indicates that the mean binding affinity of 44% of the lncRNAs has the largest values in modern horses but the smallest values in Przewalski horses. Similarly, the label "D > H > P 29%" indicates that the mean binding affinity of 29% of the lncRNAs has the largest values in modern donkeys but the smallest values in Przewalski horses. (B) The regulatory relationship between the 319 lncRNAs and the 10 NCDS genes in modern horses, modern donkeys, and Przewalski horses. Species in the inner layer correspond to the 10 NCDS genes in the second layer, and NCDS genes in the second layer correspond to lncRNA numbers in the outer layer.

also obtained in the other two species. Since DBS sequences are quite long, these BLAST searches basically yielded unique highscoring hits. The results indicate that modern horses share many DBSs with donkeys but not with Przewalski horses (Table 4). That is, many DBSs in modern horses and donkeys do not have homologous sequences in Przewalski horses. However, most DBSs in Przewalski horses have homologous sequences in modern horses and donkeys. These results indicate an increase in epigenetic regulatory relationships in modern horses and donkeys and suggest that the increased regulatory relationships may have greatly influenced the domestication of horses and donkeys.

3.2 Epigenetic regulation is stronger in domesticated horses

To further confirm that domestication is associated with epigenetic regulation, we analyzed the binding affinity of the DBSs in the modern horses/modern donkeys/Przewalski horses group. Excluding lncRNAs with very few DBSs, we calculated the average binding affinity of all DBSs for the 319 lncRNAs. In all, 140 lncRNAs (44%) showed the binding affinity pattern of "modern horse > modern donkey > Przewalski horse", and 93 lncRNAs (29%) showed the binding affinity pattern of "modern donkeys > modern



horses > Przewalski horses" (Supplementary Tables S7, S8) (Figure 4A). This result implies that a majority of the lncRNAs show stronger epigenetic regulation in domesticated animals (horses and donkeys) than in undomesticated animals (Przewalski horses). When examining detailed lncRNAs, it is clear that the average binding affinity of lncRNAs in horses and donkeys was higher than that in Przewalski horses (Figure 5).

3.3 Regulation of genes associated with neural crest/domestication syndrome

In the modern horse/modern donkey/Przewalski horse group, we further analyzed the potential regulatory relationships between the 10 NCDS genes and the 376 lncRNAs. Of note, each NCDS gene (except CHD7) has DBSs of > 60 lncRNAs, and more lncRNAs have DBSs in NCDS genes in modern horses and modern donkeys than in Przewalski horses (the total DBSs of lncRNAs in the 10 genes are 1,940, 1,895, and 1,155 in the modern horses, donkeys, and Przewalski horses) (Figure 4B). Also, 9 out of the 10 NCDS genes have fewer regulatory lncRNAs in Przewalski horses than in modern horses and modern donkeys. These results indicate that the increased epigenetic regulation in domesticated animals is enriched at least in NCDS genes, and this enrichment lends a support for the "neural crest/ domestication syndrome" hypothesis.

The NCDS genes *FGF8*, *KIT*, *PAX3*, *RET*, and *SOX9* were predicted to be regulated by > 50% of the lncRNAs in both modern horses and donkeys. *FGF8* is related to the cardiovascular system and craniofacial phenotypes. *KIT* is related to pigmentation and integument phenotypes. *PAX3* is related to pigmentation and nervous system phenotypes. *RET* is related to renal/urinary system and digestive/ alimentary phenotypes. *SOX9* is related to skeleton, growth/ size/body region and limb/digit/tail phenotypes. The intensive regulations of these genes (some of which may have been

| GO:BP | | stats | |
|---|--|--|---|
| Term name | Term ID | Padj | 0 ^{-log10(padj)} ≤1 |
| stem cell development | GO:0048864 | 7.892×10 ⁻⁵ | |
| mesenchyme development | GO:0060485 | 7.892×10 ⁻⁵ | |
| tube development | GO:0035295 | 1.313×10 ⁻⁴ | |
| stem cell differentiation | GO:0048863 | 2.288×10 ⁻⁴ | |
| primary sex determination | GO:0007538 | 4.584×10 ⁻⁴ | |
| primary sex determination, germ-line | GO:0007542 | 4.584×10 ⁻⁴ | |
| male germ-line sex determination | GO:0019100 | 4.584×10 ⁻⁴ | |
| germ-line sex determination | GO:0018992 | 4.584×10 ⁻⁴ | |
| epithelial tube formation | GO:0072175 | 5.125×10 ⁻⁴ | |
| nephron development | GO:0072006 | 5.125×10 ⁻⁴ | |
| tube formation | GO:0035148 | 7.230×10 ⁻⁴ | |
| morphogenesis of embryonic epithelium | GO:0016331 | 7.230×10 ⁻⁴ | |
| neural crest cell development | GO:0014032 | 9.157×10 ⁻⁴ | |
| regulation of developmental process | GO:0050793 | 9.845×10 ⁻⁴ | |
| regulation of kidney development | 60.0090183 | 9845×10-4 | |
| regulation of kidney development | 00.0030103 | 0.040410 | |
| negative regulation of lipid metabolic process | GO:0045833 | 1.101×10 ⁻³ | |
| GO:BP Term name | GO:0045833 | 1.101×10 ⁻³ stats | -log10(padi) |
| GO:BP Term name myoloid progenitor cell differentiation | GO:0045833 | 1.101×10 ⁻³ stats Padj | 0log10(padj)1 |
| GO:BP Term name myeloid progenitor cell differentiation locomation | G0:0045833 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ | 0 ^{_log10(padj)} ≤10 |
| GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation | G0:0045833 G0:0045833 G0:0002318 G0:0040011 G0:0007548 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ | 0 -log10(Padj) 0 |
| regulation of Namey development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation peuronepesis | GO:0045833 GO:0045833 GO:0002318 GO:0040011 GO:0007548 GO:0022008 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ | 0log10(padj) 0 |
| regulation of Namey development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development | GO:0045833 GO:0045833 GO:0045833 GO:0045833 GO:0002318 GO:0007548 GO:0007548 GO:0022008 | 1.101×10 ⁻³ stats padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ | |
| Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurops | GO:0045833 GO:0045833 GO:0002318 GO:0040011 GO:0007548 GO:0022008 GO:0022008 GO:0007417 GO:0048699 | 1.101×10 ⁻³ stats padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ | 0log10(Padj)1 |
| regative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development | GO:0045833 GO:0045833 Term ID GO:0002318 GO:0040011 GO:0007548 GO:0022008 GO:0048699 GO:0048699 GO:0048468 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.338×10 ⁻⁴ | 0log10(Padj)≤10 |
| regative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development bead development | G0:0045833 G0:0045833 G0:0045833 G0:0022318 G0:0022008 G0:007417 G0:0048699 G0:0048699 G0:0048468 G0:0060322 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.007×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.338×10 ⁻⁴ | o ^{_l} og10(padj) _≤ 10 |
| regative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development regulation of cell migration | GO:0045833 GO:0045833 GO:0045833 GO:0022318 GO:0022008 GO:007417 GO:0048699 GO:0048468 GO:0060322 GO:0060322 GO:0030334 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.338×10 ⁻⁴ 3.723×10 ⁻⁴ 3.754×10 ⁻⁴ | 0 ^{-log} 10 (Padj) _≤ 10 |
| regulation of Namey development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development regulation of cell migration nervous system development | GO:0045833 GO:0045833 Term ID GO:002318 GO:0022008 GO:007548 GO:007548 GO:007548 GO:007548 GO:007417 GO:0048699 GO:0048468 GO:0060322 GO:0030334 GO:0030334 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.007×10 ⁻⁴ 3.077×10 ⁻⁴ 3.338×10 ⁻⁴ 3.723×10 ⁻⁴ 3.754×10 ⁻⁴ 4.627×10 ⁻⁴ | 0 ^{-log} 10 (Padj) _ ≤10 |
| regulation of Namey development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development regulation of cell migration nervous system development cell migration | GO:0045833 GO:0045833 GO:0045833 GO:0002318 GO:0022008 GO:007548 GO:007548 GO:007417 GO:0048699 GO:0048699 GO:0048468 GO:0060322 GO:0030334 GO:0007399 GO:0016477 | 1.101×10 ⁻³ stats 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.007×10 ⁻⁴ 3.077×10 ⁻⁴ 3.338×10 ⁻⁴ 3.723×10 ⁻⁴ 3.754×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ | 0 ^{-log} 10 (Padj) _ ≤10 |
| regulation of kinely development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development regulation of cell migration nervous system development cell migration regulation of cell motility | GO:0045833 GO:0045833 GO:0045833 GO:0002318 GO:0007548 GO:0007548 GO:0007548 GO:0007417 GO:0048699 GO:0048699 GO:0048468 GO:0060322 GO:0030344 GO:0007399 GO:0016477 GO:2000145 | 1.101×10 ⁻³ stats 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.338×10 ⁻⁴ 3.754×10 ⁻⁴ 3.754×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ | 0 -log10(Padj) ≤10 |
| regulation of kinely development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development regulation of cell migration nervous system development cell migration regulation of cell motility movement of cell or subcellular component | GO:0045833 GO:0045833 GO:0045833 GO:0045833 GO:0002318 GO:0007548 GO:0007548 GO:0007548 GO:0007548 GO:0007548 GO:0007548 GO:0007548 GO:00048699 GO:0048699 GO:0048699 GO:0048468 GO:0060322 GO:003334 GO:0007399 GO:0016477 GO:2000145 GO:0006928 | 1.101×10 ⁻³ stats 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.38×10 ⁻⁴ 3.754×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ | 0 -log10(Padj) ≤10 |
| regulation of kinely development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development regulation of cell migration nervous system development cell migration regulation of cell motility movement of cell or subcellular component positive regulation of developmental process | GO:0045833 GO:0045833 GO:0045833 GO:002218 GO:0022008 GO:007548 GO:0022008 GO:0048699 GO:0048699 GO:0048699 GO:0048699 GO:0048468 GO:0060322 GO:0030334 GO:0007399 GO:0016477 GO:2000145 GO:0006928 GO:006928 GO:0051094 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.38×10 ⁻⁴ 3.754×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ | 0 ^{-log10(Padj)} ≤10 |
| regulation of kinely development negative regulation of lipid metabolic process GO:BP Term name myeloid progenitor cell differentiation locomotion sex differentiation neurogenesis central nervous system development generation of neurons cell development head development nervous system development cell migration nervous system development cell migration regulation of cell motility movement of cell or subcellular component positive regulation of developmental process | GO:0045833 GO:0045833 GO:0045833 GO:002218 GO:0022008 GO:007548 GO:0022008 GO:0048699 GO:0048699 GO:0048699 GO:0048468 GO:0060322 GO:0030334 GO:0007399 GO:0016477 GO:2000145 GO:0006928 GO:0051094 GO:0040012 | 1.101×10 ⁻³ stats Padj 9.654×10 ⁻⁵ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.009×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.077×10 ⁻⁴ 3.754×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ 4.627×10 ⁻⁴ | |

FIGURE 6

GO analysis of genes containing DBSs for the 376 lncRNAs shared by modern horses, modern donkeys, and Przewalski horses. (A) The enriched GO terms of genes in horses with DBSs that have hits (counterparts) in modern donkeys and Przewalski horses. (B) The enriched GO terms of genes in horses with DBSs that do not have hits (counterparts) in modern donkeys and Przewalski horses (see Table 4).

acquired recently) may help explain the following phenotypic changes: 1) changes in coat color (Orlando, 2020): in ancient times, a spotted coat was very common, but by the Middle Ages it was replaced by a uniform chestnut coat color from red to brown; 2) rapid dental evolution (MacFadden and Ceding, 1994): as the food source changed, shifting from young leaves to herbs, the teeth became more worn, and in turn, the crowns were raised, and the teeth became larger; 3) changes in the craniofacial area: as the teeth changed, the upper and lower jaws of the teeth also became larger and longer, creating the typical long face; and 4) changes in the toes: this was an important change in the evolutionary process of the horse (Solounias et al., 2018); eohippus had three toes on the hind limbs and four toes on the forelimbs, while the modern horse now walks with a single toe, which greatly enhances its running ability.

3.4 Modern horses have undergone specific evolution of epigenetic regulation in the nervous system

The 376 lncRNAs (shared in modern horses, modern donkeys, and Przewalski horses) have more DBSs in the 83 positively selected or NCDS genes in horses than in donkeys and Przewalski horses and many of the DBSs in horses are absent in donkeys and Przewalski horses (Table 4); this prompted us to examine whether the target genes with DBSs in the three species and the target genes with DBSs only in the modern horses are enriched in different functions (Supplementary Tables S9, S10). The results show that the target genes with DBSs in the three species are enriched in GO terms such as "stem cell development" and "tube development," and "primary sex determination," and the target genes with DBSs specifically in the modern horses are enriched in GO terms such as "locomotion,"

"neurogenesis," "central nervous system development," and "generation of neurons" (Figure 6). These GO terms reflect the unique traits of domesticated horses. The results suggest that domesticated horses have more evolved nervous system and running ability compared with donkeys and the undomesticated Przewalski horses, which is consistent with the consensus that domestication changes animal cognitive and behavioral characteristics.

4 Discussion

Since a short domestication period, with very limited change in the genome, can make an animal (e.g., dogs, cats, horses) produce many breeds with extremely different phenotypes, it is of great interest to uncover the gene expression regulation mechanisms involved in domestication. The process of animal domestication probably goes through two stages (Zeder, 2015; Pendleton et al., 2018). The first stage is the process of habituation to humans, which produces fear of humans and a reduction in reactive aggression against humans, thereby improving docility. This stage may involve physiological changes that are eventually fixed genetically. The second stage is a longer "breed formation" stage. The two stages are thought to involve distinct genetic changes (Fitak et al., 2020).

The domestication of different animals has commonalities and differences. Some researchers have proposed the NCDS genes as a group of genes reflecting domestication commonalities (Wilkins et al., 2014). However, the neural crest cell hypothesis is not without debate (Johnsson et al., 2021; Wilkins et al., 2021). In addition, exactly what changes these genes undergo in their gene expression regulation remains unclear. In the Equus genus, horses and donkeys have been domesticated, but Przewalski horses remain undomesticated. Thus, the genomes of Przewalski horses and ancient horses provide valuable data for examining the potential regulation of the NCDS genes and the neural crest cell hypothesis.

Computational and comparative genomic analyses are an important complement to and provide valuable clues for experimental studies on animal domestication. This study generates novel results suggesting that changes in the epigenetic regulation of domestication-related genes are an important aspect of horse domestication and that domestication probably leads to more and stronger epigenetic regulation. For horses, the regulation changes are enriched in nervous system development-related genes, which should be common during domestication, and exercise-related genes, which reflect human-specific needs for horse domestication. The results and conclusions from horse domestication may likely apply to the domestication of other animals.

It is worth noting that this study only analyzed highaffinity DBSs of lncRNAs. On the one hand, this made the analysis results quite reliable, but on the other hand, many DBSs with low affinity may be missed, which may also be associated with domestication. LncRNAs and target genes with low-affinity DBSs may reflect more recent, weaker epigenetic regulation. More in-depth analyses of domestication mechanisms can be performed with the identification of more and better ancient DNA sequences of horses and other animals. The methods of this study can be used to analyze the domestication of other animals.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author.

Author contributions

HZ, contributed to the conception and design of the study. BX and GY performed the study. BJ helped data analysis. BX and HZ wrote the manuscript.

Funding

This work was supported by the Department of Science and Technology of Guangdong Province (2020A1515010803) the National Natural Science Foundation of China (31771456).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene. 2022.944933/full#supplementary-material

References

Botigué, L. R., Song, S., Scheu, A., Gopalan, S., Pendleton, A. L., Oetjens, M., et al. (2017). Ancient European dog genomes reveal continuity since the early neolithic. *Nat. Commun.* 8, 16082. doi:10.1038/ncomms16082

Buske, F. A., Bauer, D. C., Mattick, J. S., and Bailey, T. L. (2012). Triplexator: detecting nucleic acid triple helices in genomic and transcriptomic data. *Genome Res.* 22, 1372–1381. doi:10.1101/gr.130237.111

Conolly, J., Colledge, S., Dobney, K., Vigne, J.-D., Peters, J., Stopp, B., et al. (2011). Meta-analysis of zooarchaeological data from SW Asia and SE Europe provides insight into the origins and spread of animal husbandry. *J. Archaeol. Sci.* 38, 538–545. doi:10.1016/j.jas.2010.10.008

Fitak, R. R., Mohandesan, E., Corander, J., Yadamsuren, A., Chuluunbat, B., Abdelhadi, O., et al. (2020). Genomic signatures of domestication in Old World camels. *Commun. Biol.* 3, 316. doi:10.1038/s42003-020-1039-5

Gaunitz, C., Fages, A., Hanghøj, K., Albrechtsen, A., Khan, N., Schubert, M., et al. (2018). Ancient genomes revisit the ancestry of domestic and Przewalski's horses. *Science* 360, 111–114. doi:10.1126/science.aao3297

Higuchi, R., Bowman, B., Freiberger, M., Ryder, O. A., and Wilson, A. C. (1984). DNA sequences from the quagga, an extinct member of the horse family. *Nature* 312, 282–284. doi:10.1038/312282a0

Johnson, M., Zaretskaya, I., Raytselis, Y., Merezhuk, Y., McGinnis, S., and Madden, T. L. (2008). NCBI BLAST: a better web interface. *Nucleic Acids Res.* 36, W5-W9. doi:10.1093/nar/gkn201

Johnsson, M., Henriksen, R., and Wright, D. (2021). The neural crest cell hypothesis: no unified explanation for domestication. *Genetics* 219, iyab097. doi:10.1093/genetics/iyab097

Kilian, B., Martin, W., and Salamini, F. (2010). "Genetic diversity, evolution and domestication of wheat and barley in the Fertile Crescent," in *Evolution in action*. Editor M. Glaubrecht (Berlin, Heidelberg: Springer), 137–166. doi:10.1007/978-3-642-12425-9_8

Kelekna, P. (2009). The horse in human history (Cambridge, New York: Cambridge University Press).

Larkin, M. A., Blackshields, G., Brown, N. P., Chenna, R., McGettigan, P. A., McWilliam, H., et al. (2007). Clustal W and clustal X version 2.0. *Bioinformatics* 23, 2947–2948. doi:10.1093/bioinformatics/btm404

Larson, G., Karlsson, E. K., Perri, A., Webster, M. T., Ho, S. Y., Peters, J., et al. (2012). Rethinking dog domestication by integrating genetics, archeology, and biogeography. *Proc. Natl. Acad. Sci. U. S. A.* 109, 8878–8883. doi:10.1073/pnas. 1203005109

Librado, P., Gamba, C., Gaunitz, C., Der Sarkissian, C., Pruvost, M., Albrechtsen, A., et al. (2017). Ancient genomic changes associated with domestication of the horse. *Science* 356, 442–445. doi:10.1126/science.aam5298

Librado, P., Khan, N., Fages, A., Kusliy, M. A., Suchan, T., Tonasso-Calvière, L., et al. (2021). The origins and spread of domestic horses from the Western Eurasian steppes. *Nature* 598, 634–640. doi:10.1038/s41586-021-04018-9

Lin, J., Wen, Y., He, S., Yang, X., Zhang, H., and Zhu, H. (2019). Pipelines for cross-species and genome-wide prediction of long noncoding RNA binding. *Nat. Protoc.* 14, 795–818. doi:10.1038/s41596-018-0115-5

Luikart, G., Gielly, L., Excoffier, L., Vigne, J.-D., Bouvet, J., and Taberlet, P. (2001). Multiple maternal origins and weak phylogeographic structure in domestic goats. *Proc. Natl. Acad. Sci. U. S. A.* 98, 5927–5932. doi:10.1073/pnas.091591198

MacFadden, B. J., and Ceding, T. E. (1994). Fossil horses, carbon isotopes and global change. *Trends Ecol. Evol.* 9, 481–486. doi:10.1016/0169-5347(94) 90313-1

Miller, C. D., and Lambert, D. M. (2013). Towards a million-year-old genome. *Nature* 499, 34-35.

Naderi, S., Rezaei, H.-R., Pompanon, F., Blum, M. G., Negrini, R., Naghash, H.-R., et al. (2008). The goat domestication process inferred from large-scale mitochondrial DNA analysis of wild and domestic individuals. *Proc. Natl. Acad. Sci. U. S. A.* 105, 17659–17664. doi:10.1073/pnas.0804782105

Orlando, L. (2015). Equids. Curr. Biol. 25, R973–R978. doi:10.1016/j.cub.2015. 09.005

Orlando, L. (2020). Ancient genomes reveal unexpected horse domestication and management dynamics. *BioEssays* 42, 1900164. doi:10.1002/bies.201900164

Orlando, L., Ginolhac, A., Zhang, G., Froese, D., Albrechtsen, A., Stiller, M., et al. (2013). Recalibrating Equus evolution using the genome sequence of an early Middle Pleistocene horse. *Nature* 499, 74–78. doi:10.1038/nature12323

Pedrosa, S., Uzun, M., Arranz, J.-J., Gutiérrez-Gil, B., San Primitivo, F., and Bayón, Y. (2005). Evidence of three maternal lineages in Near Eastern sheep supporting multiple domestication events. *Proc. Biol. Sci.* 272, 2211–2217. doi:10.1098/rspb.2005.3204

Pendleton, A. L., Shen, F., Taravella, A. M., Emery, S., Veeramah, K. R., Boyko, A. R., et al. (2018). Comparison of village dog and wolf genomes highlights the role of the neural crest in dog domestication. *BMC Biol.* 16, 64. doi:10.1186/s12915-018-0535-2

Raudvere, U., Kolberg, L., Kuzmin, I., Arak, T., Adler, P., Peterson, H., et al. (2019). g: Profiler: a web server for functional enrichment analysis and conversions of gene lists (2019 update). *Nucleic Acids Res.* 47, W191–W198. doi:10.1093/nar/gkz369

Sarropoulos, I., Marin, R., Cardoso-Moreira, M., and Kaessmann, H. (2019). Developmental dynamics of lncRNAs across mammalian organs and species. *Nature* 571, 510–514. doi:10.1038/s41586-019-1341-x

Schubert, M., Ermini, L., Sarkissian, C. D., Jónsson, H., Ginolhac, A., Schaefer, R., et al. (2014a). Characterization of ancient and modern genomes by SNP detection and phylogenomic and metagenomic analysis using PALEOMIX. *Nat. Protoc.* 9, 1056–1082. doi:10.1038/nprot.2014.063

Schubert, M., Jónsson, H., Chang, D., Der Sarkissian, C., Ermini, L., Ginolhac, A., et al. (2014b). Prehistoric genomes reveal the genetic foundation and cost of horse domestication. *Proc. Natl. Acad. Sci. U. S. A.* 111, E5661–E5669. doi:10.1073/pnas. 1416991111

Simoes-Costa, M., and Bronner, M. E. (2015). Establishing neural crest identity: a gene regulatory recipe. *Development* 142, 242–257. doi:10.1242/dev.105445

Solounias, N., Danowitz, M., Stachtiaris, E., Khurana, A., Araim, M., Sayegh, M., et al. (2018). The evolution and anatomy of the horse manus with an emphasis on digit reduction. *R. Soc. Open Sci.* 5, 171782. doi:10.1098/rsos. 171782

Ulitsky, I. (2016). Evolution to the rescue: using comparative genomics to understand long non-coding RNAs. *Nat. Rev. Genet.* 17, 601–614. doi:10.1038/nrg.2016.85

Vigne, J.-D. (2011). The origins of animal domestication and husbandry: a major change in the history of humanity and the biosphere. C. R. Biol. 334, 171–181. doi:10.1016/j.crvi.2010.12.009

Vilà, C., Leonard, J. A., Gotherstrom, A., Marklund, S., Sandberg, K., Lidén, K., et al. (2001). Widespread origins of domestic horse lineages. *Science* 291, 474–477. doi:10.1126/science.291.5503.474

Wilkins, A. S., Wrangham, R., and Fitch, W. T. (2021). The neural crest/ domestication syndrome hypothesis, explained: reply to Johnsson, Henriksen, and Wright. *Genetics* 219, iyab098. doi:10.1093/genetics/iyab098

Wilkins, A. S., Wrangham, R. W., and Fitch, W. T. (2014). The "domestication syndrome" in mammals: a unified explanation based on neural crest cell behavior and genetics. *Genetics* 197, 795–808. doi:10.1534/genetics.114.165423

Wright, D. (2015). Article commentary: the genetic architecture of domestication in animals. *Bioinform. Biol. Insights* 9, 11–20. doi:10.4137/BBI.S28902

Zeder, M. A. (2015). Core questions in domestication research. Proc. Natl. Acad. Sci. U. S. A. 112, 3191–3198. doi:10.1073/pnas.1501711112

Zeder, M. A. (2012). The domestication of animals. J. Anthropol. Res. 68, 161–190. doi:10.3998/jar.0521004.0068.201