

Review

The Role of Persistent Organic Pollutants in Obesity: A Review of Laboratory and Epidemiological Studies

Jan Aaseth ^{1,2,*}, Dragana Javorac ³, Aleksandra Buha Djordjevic ³, Zorica Bulat ³, Anatoly V. Skalny ^{4,5}, Irina P. Zaitseva ⁶, Michael Aschner ^{7,8} and Alexey A. Tinkov ^{6,8,*}

- ¹ Research Department, Innlandet Hospital Trust, P.O. Box 104, 2381 Brumunddal, Norway
 - ² Faculty of Health and Social Sciences, Inland Norway University of Applied Sciences, P.O. Box 400, 2418 Elverum, Norway
 - ³ Department of Toxicology “Akademik Danilo Soldatović”, University of Belgrade-Faculty of Pharmacy, Vojvode Stepe 450, 11000 Belgrade, Serbia; dragana.javorac@pharmacy.bg.ac.rs (D.J.); aleksandra.buha@pharmacy.bg.ac.rs (A.B.D.); zorica.bulat@pharmacy.bg.ac.rs (Z.B.)
 - ⁴ World-Class Research Center “Digital Biodesign and Personalized Healthcare”, IM Sechenov First Moscow State Medical University (Sechenov University), 119435 Moscow, Russia; skalny3@microelements.ru
 - ⁵ Department of Bioelementology, KG Razumovsky Moscow State University of Technologies and Management, 109004 Moscow, Russia
 - ⁶ Laboratory of Ecobiomonitoring and Quality Control, Yaroslavl State University, 150003 Yaroslavl, Russia; irisha-zip@yandex.ru
 - ⁷ Department of Molecular Pharmacology, Albert Einstein College of Medicine, Bronx, NY 10461, USA; michael.aschner@einsteinmed.org
 - ⁸ Laboratory of Molecular Dietetics, IM Sechenov First Moscow State Medical University (Sechenov University), 119435 Moscow, Russia
- * Correspondence: jaol-aas@online.no (J.A.); tinkov.a.a@gmail.com (A.A.T.)



Citation: Aaseth, J.; Javorac, D.; Djordjevic, A.B.; Bulat, Z.; Skalny, A.V.; Zaitseva, I.P.; Aschner, M.; Tinkov, A.A. The Role of Persistent Organic Pollutants in Obesity: A Review of Laboratory and Epidemiological Studies. *Toxics* **2022**, *10*, 65. <https://doi.org/10.3390/toxics10020065>

Academic Editor: Mirta Milić

Received: 20 December 2021

Accepted: 30 January 2022

Published: 2 February 2022

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Persistent organic pollutants (POPs) are considered as potential obesogens that may affect adipose tissue development and functioning, thus promoting obesity. However, various POPs may have different mechanisms of action. The objective of the present review is to discuss the key mechanisms linking exposure to POPs to adipose tissue dysfunction and obesity. Laboratory data clearly demonstrate that the mechanisms associated with the interference of exposure to POPs with obesity include: (a) dysregulation of adipogenesis regulators (PPAR γ and C/EBP α); (b) affinity and binding to nuclear receptors; (c) epigenetic effects; and/or (d) proinflammatory activity. Although in vivo data are generally corroborative of the in vitro results, studies in living organisms have shown that the impact of POPs on adipogenesis is affected by biological factors such as sex, age, and period of exposure. Epidemiological data demonstrate a significant association between exposure to POPs and obesity and obesity-associated metabolic disturbances (e.g., type 2 diabetes mellitus and metabolic syndrome), although the existing data are considered insufficient. In conclusion, both laboratory and epidemiological data underline the significant role of POPs as environmental obesogens. However, further studies are required to better characterize both the mechanisms and the dose/concentration-response effects of exposure to POPs in the development of obesity and other metabolic diseases.

Keywords: obesity; dioxin; polybromated diphenylethers; dichlorodiphenyltrichloroethane; polychlorinated biphenyls; polyaromatic hydrocarbons; bisphenol A; phthalates; perfluorinated compounds; adipogenesis

1. Introduction

Obesity is considered as a worldwide epidemic posing significant negative health effects. Specifically, current estimates indicate that more than 1.9 billion adults are overweight, with more than 650 million of them characterized as obese [1]. It has also been extrapolated that the majority of the global adult population will be overweight/obese by

2030 [1]. In addition to adults, the prevalence of obesity in children is also characterized by a drastic increase, reaching up to 5.6% and 7.8% in boys and girls, respectively [2]. Of note, parental obesity is associated with a higher risk and severity of obesity in children [3].

Obesity leads to significant health effects [4] due to association with metabolic disturbances, including insulin resistance and type 2 diabetes mellitus, atherogenic dyslipidemia, and hypertension, altogether clustered to the metabolic syndrome [5] being the major cardiovascular risk factor [6]. Obesity is also associated with other systemic disorders including respiratory [7] and neurological [8] diseases, as well as cancer [9]. Finally, obesity is considered one of the key risk factors for COVID-19 severity and mortality [10].

Beyond the key role of caloric excess in the development of obesity, multiple biological, environmental, and behavioral factors were shown to affect susceptibility to obesity [11]. The potential role of exposure to chemical toxins in expanding the obesity epidemic was proposed in 2002 [12]. Further studies have shown a significant association between exposure to persistent organic pollutants (POPs) and obesity in humans [13], although certain inconsistencies in the epidemiologic data exist [14].

POPs refer to a chemically heterogeneous group of pollutants that are categorized according to their origin to (i) intentionally produced POPs including organochlorine pesticides and industrial chemicals (polychlorinated biphenyls), and (ii) unintentionally produced agents including polyaromatic hydrocarbons, dioxins, and furans [15,16].

POPs are chemically resistant to environmental degradation and hence persist for long periods of time in the environment and can accumulate and pass from one species to the next through the food chain [17]. Some POPs have been banned or have been nominated to be banned from production at the Stockholm Convention due to their persistency in the environment and their bio-accumulative and toxic properties in both animals and humans. Those chemicals which are still on the market are regulated through the Registration, Evaluation, and Authorization of Chemicals (REACH). Nevertheless, POPs are still to be found in both human and animal tissues worldwide [18–20].

These chemicals also accumulate in human tissue with long biological half-lives; they can cross the placenta during gestation. The main route of exposure to these chemicals is through dietary intake, while the environmental exposure pathways, mainly dermal exposure, account for less than 2% of exposure for most of them [21].

Being hydrophobic, brominated and chlorinated POPs accumulate predominantly in lipid rich tissues [22], while perfluorinated compounds have a higher affinity for plasma proteins [23]. Various POPs chemicals are present in children's blood and breast milk, and due to their ability to cross both the placental and the blood brain barriers, can reach the developing nervous system [24]. However, adipose tissue is the one that plays a major role in the storage and toxicokinetics of POPs. La Merrill et al. [25] suggested some additional functions of adipose tissue in context of POPs toxicity. Adipose tissue acts as a unique buffering system by protecting other organs and tissue by sequestering POPs. However, at the same time, this tissue represents a continual source of internal exposure to POPs.

POPs were shown to promote obesogenic effects through endocrine disruption, thus being considered as obesogens [26,27]. Obesogens have been shown to affect adipose tissue development and functioning through interference with key adipocyte metabolic pathways, including the regulation of adipogenesis [28]. Such dysregulation may also be mediated by the epigenetic effects of the pollutants [29]. In addition, alterations in other endocrine tissues, as well as hypothalamic centers, may contribute to energy dyshomeostasis upon exposure to POPs [30]. Moreover, it has been observed that accumulation of POPs in adipocytes promotes inflammation in adipose tissue, thus contributing to the dysfunction of the tissue [31]. However, the mechanisms of the obesogenic effects of various POPs may be quite different [31] and have yet to be fully characterized.

Given the above observations, the objective of the present review is to discuss the key mechanisms linking exposure to POPs to adipose tissue dysfunction and obesity in laboratory studies, as well as to review the epidemiological evidence supporting this association.

2. Perfluorinated Compounds

The role of perfluorooctanoic acid (PFOA) (Figure 1A) and perfluorooctane sulphonate (PFOS) (Figure 1B) in obesity has yet to be fully clarified, although some data derived from epidemiological studies suggest that PFOS and PFOA exposures are associated with overweight and obesity [32]. Higher levels of PFOS and PFOA were found in mothers with obesity and in underweight mothers when compared to those of a normal weight [33]. Moreover, prenatal exposure to PFAS has been linked with obesity, metabolic disorders, and alterations in children's growth [34–36]. In a large multicenter prospective cohort study, the “European Youth Heart Study” performed in young participants (N = 369), scientists found that PFOS and PFOA exposure predicted adiposity at 15 and 21 years of age [37]. A recently published cross-sectional study in US children from 12–18 years of age (N = 2473) showed a dose-dependent association between obesity and PFAS exposure [38]. Nonetheless, the European Food Safety Authority reported in 2020 that there is insufficient data to support a link between PFAS exposure and obesity, thus the explanation for this association requires further research [39].

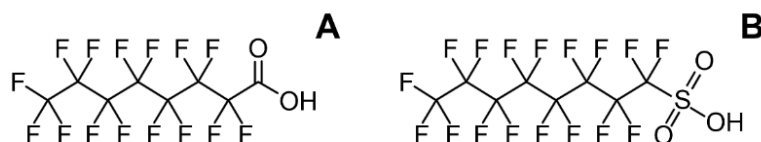


Figure 1. The chemical structures of perfluorooctanoic acid (PFOA) (A) and perfluorooctane sulphonate (PFOS) (B).

In contrast, laboratory data demonstrate that perfluorinated compounds are potent inducers of adipogenesis through interference with peroxisome proliferator-activated receptor gamma (PPAR γ) signaling and other pathways, being most prominent for sulfonated perfluoroalkyl acids, whereas carboxylated agents showed lesser alterations of mouse 3T3-L1 cells gene expression [40]. A similar mechanism was responsible for the PFOA-induced inhibition of human mesenchymal stem cells (MSC) osteogenic differentiation and adipogenesis stimulation [41]. In addition to PPAR γ binding and the associated up-regulation of adipogenesis [42], PFOA was found to increase PPAR γ transcription and the demethylation of PPAR γ promoters during 3T3-L1 preadipocyte differentiation [43]. A study in the nematode *C. elegans* also demonstrated the involvement of PPAR, mitogen activated protein kinase (MAPK), and transforming growth factor beta (TGF β) signaling in the obesogenic effect of PFOA [44]. At the same time, an in vivo study of perinatal PFOA exposure demonstrated a sex-specific response with more profound metabolic alterations in female C57BL/6JxFVB mice [45].

The observed adipogenic effects were accompanied by an increase in insulin-stimulated glucose uptake through the up-regulation of glucose transporter type 4 (GluT4) and insulin receptor substrate 1 (IRS1) expression in murine 3T3-L1 preadipocytes [46]. However, this observation contrasts with findings from an in vivo study demonstrating PFOA-induced insulin resistance in exposed Balb/c mice. These effects are proposed to be mediated by the down-regulation of protein kinase B (Akt) mRNA expression and phosphorylation, as well as increased phosphatase and tensin homolog (PTEN) mRNA expression and protein levels [47].

Similarly, PFOA and PFOS were shown to decrease osteopontin, osteonectin, osteocalcin, and β -catenin expression in human bone marrow-derived mesenchymal stem cells (hBMSCs), thus being indicative of reduced osteogenesis, whereas the expression of adipogenesis-specific marker genes PPAR γ , CCAAT/enhancer-binding protein alpha (C/EBP α), lipoprotein lipase (LPL), and leptin were up-regulated [41]. It has been also demonstrated that in parallel with induction of PPAR γ and C/EBP α expression, PFOS-induced adipogenesis was associated with the activation of the nuclear factor-erythroid factor 2-related factor 2 (Nrf2) pathway in murine 3T3-L1 preadipocytes [46]. In addition to PPAR γ , proadipogenic effects of PFOS may involve induction of activating protein 2

(ap2) [48], as well as PPAR α and PPAR β mRNA expression, affecting stem cell differentiation in hBMSCs [49]. The modulation of DNA methylation may also be considered a potential mechanism of the impact of PFOS on adipogenesis [50].

An adipogenic effect was also demonstrated for certain other perfluorinated compounds. Specifically, perfluorobutanesulfonic acid (PFBS) used as a substituent for perfluorooctanesulfonic acid (PFOA) was demonstrated to be a proadipogenic agent, promoting the differentiation of 3T3-L1 preadipocytes to adipocytes by up-regulating PPAR γ and C/EBP α transcription factors and lipogenic acetyl-CoA carboxylase (ACC) and fatty acid synthase (FAS) [51]. In comparison to PFOS, chlorinated polyfluorinated ether sulfonates (Cl-PFAESs) were shown to be more potent stimulators of 3T3-L1 adipogenesis through the PPAR γ pathway [52]. Perfluorinated alkyl acids (PFAAs) are also capable of inducing adipogenesis in 3T3-L1 cells at human blood-based exposure levels [53].

Altogether, the existing data demonstrate that PFOS and/or PFOA exposure may promote adipogenesis through the up-regulation of PPAR γ and C/EBP α signaling, thus contributing to an increased risk of obesity, although the epidemiological data have yet to confirm this association (Figure 2).

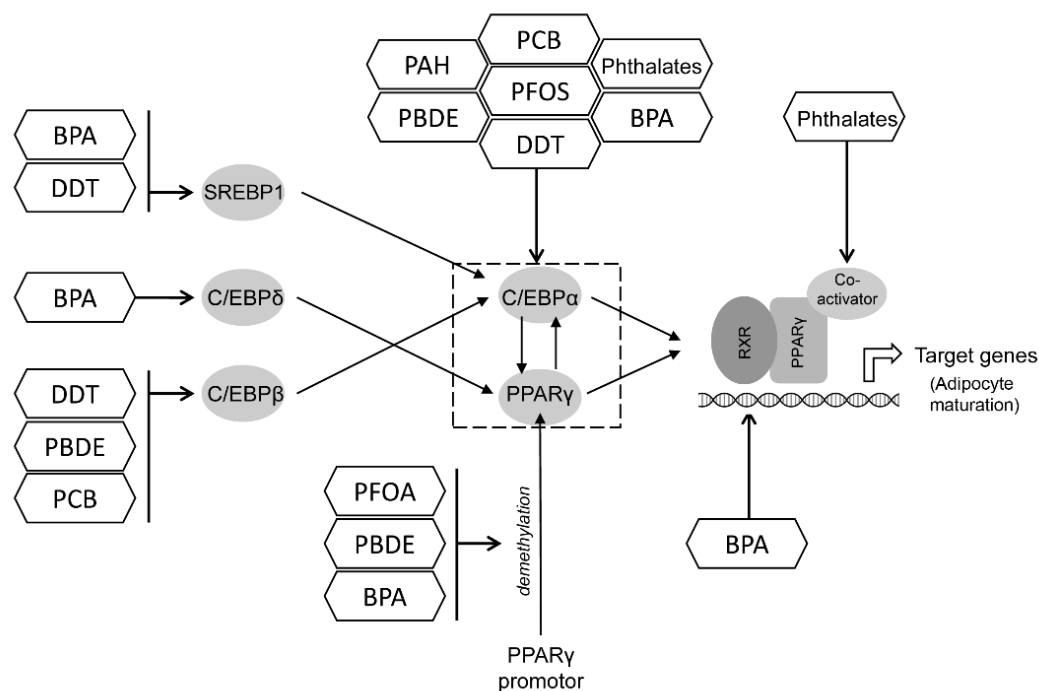


Figure 2. The potential mechanisms underlying the adipogenic effects of POPs in adipocytes. Briefly, POPs were shown to up-regulate PPAR γ and C/EBP α signaling through a variety of mechanisms, including the activation of C/EBP δ and C/EBP β and SREBP1, as well as an increase in PPAR γ promoter demethylation. The activation of PPAR γ results in the up-regulation of adipocyte-specific gene expression and adipocyte maturation. BPA—bisphenol A; PCB—polychlorinated biphenyls; PBDE—polybrominated diphenyl esters; PAH—polyaromatic hydrocarbons; PFOS—perfluorooctane sulphonate; PFOA—perfluorooctanoic acid; DDT—dichlorodiphenyltrichloroethane; DES—diethylstilbestrol; SREBP1—sterol regulatory element-binding protein 1; C/EBP—CCAAT-enhancer-binding proteins; PPAR γ —peroxisome proliferator-activated receptor gamma; RXR—retinoid X receptor.

3. Polybrominated Diphenyl Ethers (PBDE)

Polybrominated diphenyl ethers (PBDEs) (Figure 3A) are man-made chemicals and environmental pollutants used in industry as flame retardants in various commercial goods [54]. PBDEs, due to their lipophilic properties, tend to accumulate in adipose tissue,

possibly changing its function and raising the risk of metabolic diseases. Several studies suggested a link between PBDEs and obesity and metabolic syndrome [55,56].

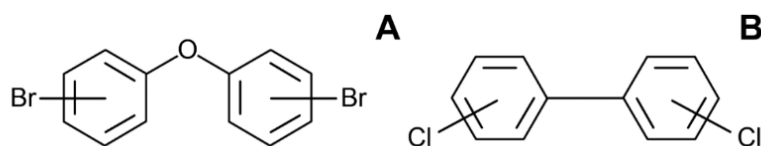


Figure 3. Common formulas of the persistent organic pollutants polybrominated diphenyl ethers (PBDE) (A) and polychlorinated biphenyls (PCB) (B).

In individuals with obesity, the accumulation of certain PBDEs in human adipose tissues is linked to insulin resistance [57]. In a study conducted of 224 mothers during pregnancy and afterwards, including their children at age 7, it was revealed that the levels of BDE 47 and BDE 153 in maternal blood were positively associated with the body mass index (BMI) of boys. Contrary, in girls, the PBDE levels in maternal blood negatively correlated with BMI [58]. Furthermore, the serum levels of BBDEs were found to be positively associated with the expression of obesity biomarkers in subcutaneous and visceral adipose tissue, such as leptin, adiponectin, tumor necrosis factor α (TNF α), and PPAR γ , implying that exposure to these pollutants may contribute to the development of obesity in humans [59].

In agreement, laboratory data have shown that PBDE exposure potentiates dexamethasone-induced adipogenesis in 3T3-L1 cells with the up-regulation of C/EBP α , PPAR γ , and liver X receptor alpha (LXR α) [60,61]. Specifically, PBDE 99 was shown to up-regulate C/EBP β with subsequent the activation of C/EBP α and PPAR γ along with promotion of mitotic clonal expansion. In addition, the methylation status of PPAR γ promoter was found to be reduced in response to PBDE 99 exposure [62]. Furthermore, a stimulatory effect of BDE-47 on 3T3-L1 adipocyte differentiation was shown to be dependent on *Ppar γ 2* gene induction and *Ppar γ 2* promoter demethylation [63].

However, some other studies indicated that an obesogenic effect of PBDE may be mediated by other distinct mechanisms. Specifically, adipogenesis stimulation by PBDE-47 may be dependent on the activation of purine metabolism, oxidative stress, and mitochondrial respiration [64]. These effects are also associated with a reduction of lipid catabolism through the inhibition of β -oxidation and increased lipid biosynthesis, altogether resulting in increased lipid accumulation [65], as well as the inflammatory infiltration of adipose tissue in C57BL/6J mice [66].

The systemic effects of PBDE may also significantly contribute to its modulation of metabolic risk in obesity. It has been proposed that the obesogenic effect of PBDE exposure may be related to reduced T4 levels with a subsequent decrease in systemic thyroid hormone effects in Wistar rats [67]. In addition, PBDEs also significantly alter the murine gut microbiome, which may, at least partially, mediate the association between PBDE exposure and metabolic syndrome [68].

In corroboration with epidemiological studies demonstrating the association between PBDE exposure and obesity, as well as the accumulation of PBDE in adipose tissue, laboratory data indicate that adipogenic effect of PBDE is mediated by the up-regulation of PPAR γ and C/EBP α signaling (Figure 1) through a variety of mechanisms including epigenetic effects, modulation of oxidative stress, adipose tissue inflammation, thyroid functioning, and gut microbiota.

4. Dichlorodiphenyltrichloroethane (DDT)

Introduced as an insecticide in the 1940s, dichlorodiphenyltrichloroethane (DDT) (Figure 4A) is a persistent organic pollutant and endocrine disrupting chemical. In the last decade, a huge number of studies aimed to evaluate the association of the serum and fat levels of DDT and its metabolite dichlorodiphenyldichloroethylene (DDE) (Figure 4B) with body weight, visceral obesity, or BMI [69,70].

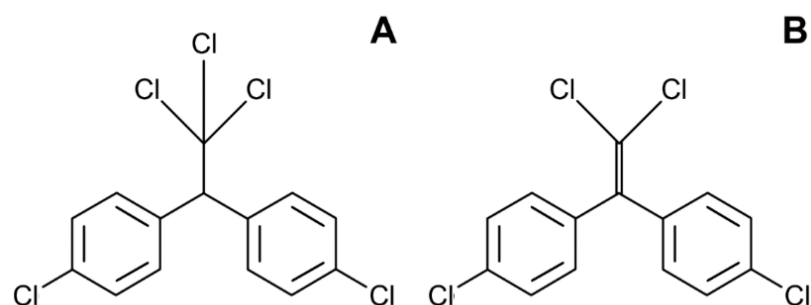


Figure 4. The structures of dichlorodiphenyltrichloroethane (DDT) (A) and its toxic metabolite dichlorodiphenyldichloroethane (DDE) (B).

In a study conducted in Spain on 298 participants, a positive association was found between lipid DDT levels and BMI [71]. A larger study encompassing 775 men and 808 women has shown a positive correlation between BMI and DDT serum levels [72]. Interestingly, a similar trend was observed in children. The positive connection between DDE/DDT prenatal exposure and childhood BMI and growth patterns was determined in the CHAMACOS cohort study, which included 249 participants. The study showed that this correlation was stronger in boys than in girls [73–76]. Certain studies evaluated the association between obesity and prenatal DDT exposure. The Child Health and Development Studies in California demonstrated the effects of in utero exposure to DDT on middle-aged daughters (44–53 years, N = 511). Researchers have found a higher risk for being overweight (26–31%) as estimated from the BMI and waist circumference in the DDT exposed group [77]. In addition, in utero exposure to DDT was found to be associated with higher body weight in the postnatal period during the first and third years of life [78].

Moreover, DDT also plays a role in obesity-associated diseases. In a recent study, Henríquez-Hernández et al. have shown that DDT and/or its metabolites can contribute to obesity development and related diseases as recorded by altered fasting blood glucose and metabolic disorders [79]. Furthermore, adipose tissue DDT levels, a measure of the deposition of this chemical, were found to be positively associated with triglycerides, glycemic parameters, and central obesity in a study involving 100 participants [80].

Corroborating the epidemiological studies, both laboratory in vitro and in vivo studies revealed the potent adipogenic effects of DDT and DDE. Specifically, both substances were shown to promote adipogenesis in 3T3-L1 adipocytes, resulting in increased fatty acid synthase (FAS) and acetyl-CoA carboxylase (ACC) expression as well as lipid accumulation, likely mediated by the induction of C/EBP α and PPAR γ expression. It has also been shown that both DDT and DDE are capable of up-regulating AMP-activated protein kinase- α (AMPK α) expression while reducing its phosphorylation [81]. These findings are in agreement with the observations in two adipocyte lines, 3T3-L1 and 3T3-F442A, that demonstrated a DDT-induced increase in C/EBP α and PPAR γ protein levels, as well as C/EBP α interaction with DNA, absent influence on C/EBP β expression [82].

The adipogenic effect of DDE was shown to be associated with the up-regulation of PPAR γ expression, modulation of lipid metabolism, and a concomitant reduction of pluripotency genes in human adipose derived MSCs [83]. Correspondingly, DDT exposure reduced the self-renewal of human MSCs while increasing its adipogenic differentiation with increased mRNA expression of adipocyte-specific PPAR γ , GluT4, and LPL. It is noteworthy that the effects of DDT were similar to those observed in response to estrogen exposure, while antiestrogen ICI 182780 ameliorated the influence of DDT, both being indicative of the role of estrogen receptor signaling in DDT-induced adipogenesis [84]. In a model of the suboptimal adipogenic differentiation of 3T3-L1 preadipocytes, DDE exposure increased lipid accumulation due to an increased expression of sterol regulatory element-binding protein 1 (SREBP1), fatty acid binding protein (FABP), and fatty acid synthase (FASN) [85].

It has been proposed that the effects of DDT exposure in adipocytes on the offspring from DDT-exposed rats may be significantly mediated by DNA methylation changes affecting adipogenesis regulation and metabolic functioning [86].

Of note, DDE was shown to induce insulin resistance in NIH3T3-L1 adipocytes through induction of adipose tissue endocrine dysfunction more efficiently than DDT [87]. In addition, perinatal DDT exposure in C57BL/6J mice may impair thermogenesis and reduce energy expenditure, thus promoting the dysregulation of lipid and carbohydrate metabolism and increasing susceptibility to metabolic syndrome [88].

Taken together, epidemiological studies provide strong evidence for the association between DDT exposure and the risk of obesity and obesity-related metabolic disturbances leading to metabolic syndrome. Such an obesogenic effect may be mediated by the up-regulation of C/EBP α and PPAR γ signaling and a subsequent shift to adipogenesis (Figure 1), which is, at least in part, mediated by estrogen receptor signaling and epigenetic effects. The reduction of insulin sensitivity and energy expenditure may also contribute to the disturbances of carbohydrate and lipid metabolism upon DDT exposure (Figure 5).

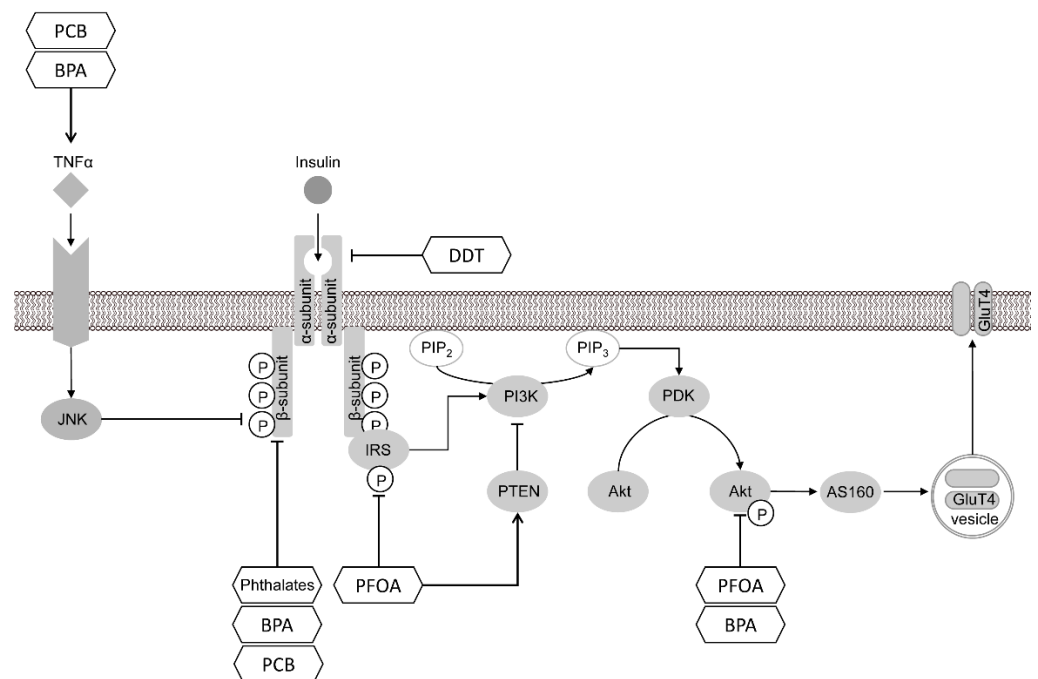


Figure 5. The molecular mechanisms involved in the dysregulation of insulin signaling in adipocytes upon exposure to POPs. POPs were shown to impair insulin signaling in adipocytes through the inhibition of insulin receptors and IRS phosphorylation, thus reducing downstream signaling from insulin receptors. Certain POPs may contribute to altered insulin signaling through the up-regulation of PTEN activity, as well as the inhibition of Akt phosphorylation. The proinflammatory effects of POPs may also contribute to the inhibition of insulin signaling and development of adipocyte insulin resistance. BPA—bisphenol A; PCB—polychlorinated biphenyls; PBDE—polybrominated diphenyl esters; PAH—polyaromatic hydrocarbons; PFOS—perfluorooctane sulphonate; PFOA—perfluorooctanoic acid; DDT—dichlorodiphenyltrichloroethane; DES—diethylstilbestrol; GluT4—glucose transporter type 4; JNK—c-Jun N-terminal kinases; AS160—Akt substrate of 160 kDa; PTEN—phosphatase and tensin homolog; IRS—insulin receptor substrate; PDK—phosphoinositide dependent kinase-1; PI3K—phosphoinositide 3-kinase; PIP3—phosphatidylinositol 3,4,5 trisphosphate; TNF α —tumor necrosis factor alpha.

5. Polychlorinated Biphenyls (PCBs)

Polychlorinated biphenyls (PCBs) (Figure 4B) are persistent organic chemicals that have been used in industry for more than 50 years. Although PCBs were globally banned in the late 1970s, they are still present in the environment. Exposure to PCBs has been

shown to affect endocrine functions and metabolic processes by mimicking natural thyroid, estrogen, and androgen actions [89–91]. Recent published data suggest an association between exposure to PCBs and the development of obesity [92,93]. In the cross-sectional National Health and Nutrition Examination Survey's (NHANES) 1999–2002 study of 721 non-diabetic adults with metabolic syndrome, the blood concentration of PCBs was linearly related to waist circumference [94]. Furthermore, in a longitudinal study with 12,313 participants, individuals with higher PCBs intake levels were at a greater risk of being obese, supporting the strong association between PCB exposure and the occurrence of obesity [95]. A number of studies have shown that the in utero exposure of embryos to PCBs can contribute to the development of metabolic syndrome, dyslipidemia, glucose intolerance, and other obesity-related metabolic disorders in postnatal life [55,96–98]. Maternal exposure to PCBs was associated with more pronounced obesity in girls than in boys, indicating a sex-dependent effect [99]. The overweight incidence in children has been shown to be sex-dependent in a study performed in 344 children, supporting an association of exposure to PCBs and overweight only in the girls' group [97].

Based on findings from laboratory studies, PCBs have been implicated as endocrine-disrupting chemicals that are capable of disturbing lipid metabolism secondary to their accumulation in adipose tissue [100]. Specifically, PCB-118, PCB-153, and especially PCB-23 were shown to accumulate extensively within a lipid droplet of 3T3-L1 and mouse embryonic fibroblast-derived cultured adipocytes, being independent of caveolin 1 [101]. In turn, it has been proposed that molecular lipophilicity might be considered as a key factor in the regulation of PCB accumulation in adipose tissue [102].

PCBs were shown to impair adipogenesis through modulation of key adipogenesis regulators PPAR γ and C/EPBs. Specifically, the adipogenic response to PCB-77 exposure was shown to be biphasic with increased 3T3-L1 adipocyte differentiation and PPAR γ expression at low-dose treatments and inhibition upon high-dose exposure. Moreover, PCB-77 exposure in vivo was shown to induce obesity, dyslipidemia, and atherosclerosis in ApoE $-/-$ mice [103]. In addition, PCB-153 was also shown to induce adipogenesis through PPAR γ -independent mechanisms [104]. Aroclor 1254 (a mixture of chlorinated biphenyls) exposure in C57B6/J mice was shown to induce adipocyte differentiation via the stimulation of PPAR γ signaling and the alteration of redox metabolism due to the inhibition of hepatocyte nuclear factor 1b (HNF1b) [105].

PCB 180 was shown to promote adipogenesis in murine 3T3-L1 preadipocytes and human visceral preadipocytes through the activation of C/EPB β by reducing its SUMOylation and ubiquitination, thus reducing to hyperplastic obesity [106].

In agreement with PCBs' ability to activate nuclear receptors [100], several effects of PCBs in adipose tissue, including insulin resistance, were shown to be largely attributed to the modulation of the aryl hydrocarbon receptor (AhR). Specifically, the administration of coplanar PCBs to C57BL/6 mice was shown to affect glucose homeostasis and insulin signaling through an AhR-mediated increase in adipocyte TNF α expression [107].

At the same time, an AhR-dependent increase in proinflammatory cytokine expression in human preadipocytes was found to be delayed in comparison to the classic AhR-targeted gene CYP1A1 [108]. Concomitantly, another study demonstrated that the PCB-153-induced glucose dyshomeostasis and inflammatory response may be mediated through NF- κ B activation and the up-regulation of downstream proinflammatory cytokine genes in 3T3-L1 cells [109]. In addition to the up-regulation of adipocyte-specific proinflammatory cytokine production, PCB-138 is also capable of increasing 3T3-L1 adipocytes adipocyte resistance to TNF α -induced apoptosis via the up-regulation of surviving expression, thus promoting further lipid droplet enlargement assisted by Fsp27 and perilipin [110].

Another study demonstrated that the PCB-induced dysregulation of glucose metabolism in 3T3-L1 cells is reversed by resveratrol treatment and the resulting up-regulation of Nrf2 signaling, thus being indicative of the role of oxidative stress in PCB-associated insulin resistance [111].

The PCB-126-induced activation of AhR was also shown to be involved in the alteration of adipocyte beiging through the inhibition of UCP1 up-regulation in response to β -adrenergic stimulation, thus affecting energy metabolism in the subcutaneous human normal preadipocyte (NPAD) clone B cell line [112].

In addition, the activation of retinoid X and retinoic acid receptors (RAR), which play a significant role in adipogenesis regulation, may be also considered as a possible mechanism of the PCB adipogenic effect [113].

In vivo studies demonstrated that the hazardous effects of PCB exposure may be diet-dependent and aggravated in obesity. Specifically, in high-fat-fed mice, PCB-153 exposure resulted in a significant increase in body adiposity, hepatic steatosis, circulating leptin, adiponectin, resistin, and tPAI-1 concentrations, as well as the activation of hepatic lipid anabolism, whereas in STD-fed mice, no such effect was observed [114]. It is also notable that male zebrafish were more sensitive to PCB-induced alterations in lipid metabolism when compared to females, as evidenced by a higher expression of genes related to lipid biosynthesis and catabolism [115].

Generally, the existing data demonstrate that both life-long and prenatal PCB exposure may be considered as a risk factor for obesity due to the increased expression of adipogenic regulators PPAR γ and C/EPB α , as well as C/EPB β (Figure 2). In addition, PCB-induced AhR signaling was shown to be responsible for the development of adipose tissue inflammation, insulin resistance (Figure 5), and the inhibition of adipocyte energy expenditure and beiging.

6. Polycyclic Aromatic Hydrocarbons (PAHs)

Polycyclic aromatic hydrocarbons (PAHs) are a group of chemicals with endocrine disruptive potential that are released into the environment through industrial processes, food preparation, and tobacco smoke, with benzo[a]pyrene (BaP) (Figure 6) being one of the most toxic.

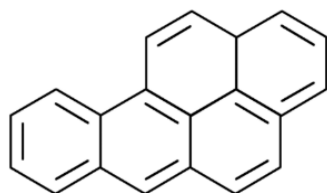


Figure 6. The structural formula of polycyclic aromatic hydrocarbon benzo[a]pyrene (BaP).

In the NANHES 2001–2006, total urinary PAH was analyzed in 3189 participants from 6–19 years of age. The results have shown that urinary PAH concentration was positively correlated with BMI and obesity, with the strongest association in children 6–11 years of age [116]. Another study, conducted in Iran on a smaller number of participants from 6–18 years of age, also showed that urinary monohydroxy PAH was associated with an increased risk of obesity [117]. In addition, a cross-sectional study that used data from NANHES 2003–2008 revealed that simultaneous exposure to PAHs and tobacco smoke synergistically increased the risk of obesity [118]. In other studies, exposure to PAHs was found to be associated with metabolic syndrome, diabetes, and the risk of cardiovascular disease, all health conditions that are strongly connected with obesity [117,119–121].

In laboratory studies, PAHs were shown to have a significant impact on adipose tissue functioning both in vitro and in vivo, although the effect of such influence may be quite different. Specifically, the exposure of BaP in combination with PCBs was shown to down-regulate adipogenesis genes while increasing inflammatory gene expression through the up-regulation of AhR signaling in 3T3-L1 cells [122]. These findings corroborate the observation on BaP-induced inhibition of human MSC-derived adipogenesis mediated by AhR activation [123]. Specifically, BaP was shown to down-regulate AhR protein expression and increase AhR translocation to the nucleus, as well as to inhibit PPAR γ , resulting in the down-regulation of adipogenesis in canine adipose-derived MSCs [124].

In contrast to *in vitro* investigations, *in vivo* studies demonstrated the profound obesogenic effect of PAH exposure. Specifically, prenatal PAH exposure to BALB/cByJ mice through the maternal inhalation of PAH mixtures was shown to increase offspring adiposity through an increase in PPAR γ and C/EBP α expression, at least partially due to a decrease in PPAR γ promoter DNA methylation [125]. It has been also demonstrated that the adipogenic effect of prenatal BaP exposure in mice may be glutathione-dependent [126]. *In utero* rat exposure to another PAH, 2-aminoanthracene (2AA), was also shown to induce adipogenesis in offspring [127]. BaP was shown to accumulate in murine adipose tissue with a subsequent inhibition of adrenaline-induced lipolysis [128], thus promoting weight gain in C57Bl/6J mice [129].

Prenatal phenanthrene exposure in mice was also shown to impair adipokine levels through the alteration of adiponectin and resistin gene DNA methylation, thus promoting glucose intolerance [130]. Another adipose tissue-targeted mechanism contributing to insulin resistance upon PAH exposure may involve an increase in insulin receptor substrate 2 methylation [131].

It is also notable that obesity may promote 7, 12-dimethylbenz [a] anthracene-induced carcinogenesis both in NIH/3T3 fibroblasts and exposed C57BL/6J female mice [132].

Therefore, the existing findings revealed a significant association between PAH exposure, obesity, and obesity-associated metabolic disturbances. These observations are consistent with *in vivo* laboratory studies indicating increased adipogenesis through a decrease in PPAR γ promoter DNA methylation (Figure 1), lipid overaccumulation due to the inhibition of lipolysis, and insulin resistance (Figure 2). In contrast, *in vitro* data demonstrate that PAH-induced AhR signaling may result in the inhibition of adipogenesis via the down-regulation of PPAR γ . Such an inconsistency may be mediated by the differences in PAH doses used in *in vivo* and *in vitro* studies as well as by the mechanisms of PAH metabolism in the organism, thus raising a question on the estimation of conditions for consistency between *in vivo* and *in vitro* models.

7. Dioxin

The relation between exposure to 2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD) (Figure 7), the most prominent member of dioxin and other dioxin-like compounds, to obesity and metabolic effects in general has been examined in several epidemiologic studies producing conflicting data.

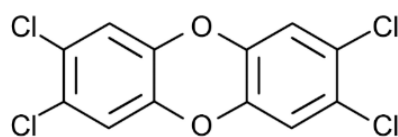


Figure 7. The chemical structure of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD).

The Seveso Women's Health Study (SWHS), a cohort study of the health of the women relying on the data obtained after a chemical explosion on 10 July 1976 in Seveso, Italy, prompted Warner et al. [133] to examine the relation of serum TCDD to diabetes, metabolic syndrome, and obesity more than 30 years later. The study enrolled 981 women who were aged from newborns to 40 at the time of the explosion and who were residents of the most contaminated areas. However, the authors did not find association between a 10-fold increase in serum TCDD and obesity. An increased prevalence of metabolic syndrome was observed; however, only among women who were ≤ 12 years of age at the time of the explosion [133].

In a cross-sectional descriptive study aiming to investigate the link between abdominal obesity and concomitant exposure to serum dioxins (PCDD/Fs), seventeen 2,3,7,8-substituted PCDD/Fs congeners were measured [134]. The authors hypothesized that the subjects with the highest serum PCDD/Fs levels adjusted as TEQDF-1998 and abdominal obesity levels had elevated the chances of developing insulin resistance (IR). In a cross-

sectional study evaluating the associations of body burden levels of dioxins and related compounds with the prevalence of metabolic syndrome among the 1374 participants from the general population in Japan, it was found that the body burden levels of dioxins and related compounds (particularly, DL-PCBs) were associated with metabolic syndrome [135]. Furthermore, the same study revealed that high blood pressure, elevated triglycerides, and glucose intolerance were also connected to the levels of these pollutants. In a study investigating 1234 nondiabetic persons living near a deserted pentachlorophenol factory, the association between exposure to dioxins and insulin resistance was investigated [136]. The study proved a positive association between serum dioxins and the prevalence of insulin resistance. The different results obtained by the studies can be partly explained by the fact that the chemical concentration in the blood may be lower in obese subjects due to dilution, while the actual cumulative exposure may be higher due to an extended half-life [137].

The role of this class of chemicals in obesity remains to be further elucidated, especially when considering the hydrocarbon receptor (AhR) as the main mediator of dioxins' toxicity. AhR has been implicated in the regulation of energy metabolism and is currently being investigated as a potential therapeutic target for obesity [138]. Specifically, 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) exposure was shown to inhibit adipogenesis in 3T3-L1 cells through the AhR-dependent down-regulation of PPAR γ and C/EBP α at an early stage of differentiation, whereas the dioxin-induced inhibition of glucose uptake was not AhR-dependent [139]. In addition, the inhibitory effect of TCDD on adipogenesis in C3H10T1/2 cells was shown to be aggravated by the epidermal growth factor (EGF) and the fibroblast growth factor (FGF) that were shown to potentiate AhR activation and the subsequent down-regulation of PPAR γ expression [140]. Inhibition of PPAR γ by dioxin may be at least partially mediated by the down-regulation of PPAR γ coactivator 1 alpha production in murine 3T3-L1 adipocytes [141]. ERK activation was also considered as the potential mechanism of the inhibitory effect of TCDD on PPAR γ 1 expression in C3H10T1/2 cells [142]. In addition, the dioxin-induced inhibition of both isoforms p30 and p42 of C/EBP α may be characterized as an additional mechanism of adipogenesis inhibition in the 3T3-L1 preadipocyte cell line [143].

In addition to the inhibition of PPAR γ and C/EBP α in the adipose tissue of exposed guinea pigs, dioxin was also shown to down-regulate sterol regulatory element binding protein (SREBP) 1 and 2, a key regulator of lipid anabolism, as well as its downstream genes, acetyl-CoA carboxylase and HMG-CoA synthase [144]. However, an earlier study demonstrated that TCDD exposure is also capable of inhibiting adipose tissue lipoprotein lipase activity, thus decreasing lipid catabolism [145].

In contrast to *in vitro* data, *in vivo* studies have demonstrated a role of TCDD in obesity promotion in laboratory rodents. Specifically, in female C57BL/6J mice fed HFD, TCDD exposure resulted in increased adiposity and reduced adipose tissue triglyceride lipase mRNA expression, as well as elevated liver triglyceride content along with stearoyl-coA desaturase-1 mRNA expression, altogether being associated with higher AhR and androgen receptor mRNA expression [146]. Correspondingly, in another study, TCDD exposure was shown to promote obesity-associated non-alcoholic fatty liver disease in C57BL/6J mice fed a high-fat diet [147].

Moreover, transient low-dose TCDD exposure was shown to induce post-exposure weight gain and glucose intolerance in C57BL/6 mice [148]. However, TCDD-induced hyperglycemia was also shown to be sex-specific, being characteristic only for female C57BL/6 mice [149].

Taken together, the existing data on the role of dioxin exposure in obesity are inconsistent. Despite certain positive findings, other epidemiological studies did not reveal an association between dioxin exposure and obesity. The above-mentioned laboratory data indicate that dioxin-induced AhR signaling may be responsible for the inhibition of adipogenesis *in vitro*, whereas *in vivo* studies demonstrate that dioxin exposure may promote excessive adiposity and obesity-associated metabolic disturbances in laboratory animals.

Hypothetically, the AhR-dependent down-regulation of adipogenesis upon dioxin exposure may result in adipose tissue dysfunction and increased ectopic lipid accumulation, thus promoting additional metabolic risk, although this suggestion requires further elaboration.

8. Bisphenol A (BPA)

Bisphenol A (BPA) (Figure 8A) is a synthetic organic chemical with endocrine disrupting potential that is used with other chemicals in the production of various plastic products and epoxy resins [150].

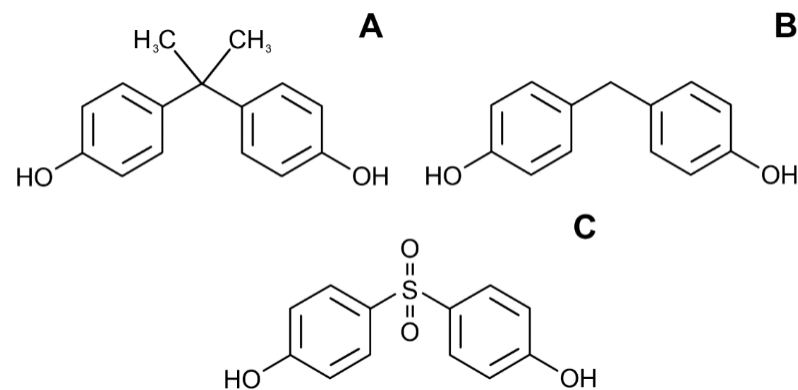


Figure 8. The structure of bisphenol A (A) and related compounds bisphenol F (B) and A (C).

Several recent studies have demonstrated the link between BPA and obesity development. In 296 women of reproductive age, BPA urinary levels were positively correlated with BMI and waist circumference [151]. In the recently published and previously mentioned Korean National Environmental Health Survey (KoNEHS) 2015–2017, higher BPA urinary levels showed a significantly higher risk of developing diabetes mellitus and obesity [152]. In the Korean cross-sectional study ($n = 702$), urinary BPA concentration was associated with abdominal obesity in women but not in men, with the strongest association determined in postmenopausal women, indicating the roles of gender and women's menopausal status [153]. The data processed from the U.S. NANHES (2003–2006) study on 2747 adult subjects showed that participants in the highest BPA urinary quartiles had a higher obesity incidence when compared to the lowest BPA quartile, and generally higher BPA levels were associated with higher body weight and central obesity [154]. Growing evidence suggests that BPA can interfere with physiological metabolism and promote obesity in children and adolescents [155]. In a study investigating 1326 children and adolescents, it was observed that in male participants, BPA urinary levels were not linked to obesity. However, in females, this association was found, especially among girls entering the pubertal stage (BPA > 2 mg/L), and their risk of being overweight was doubled when compared to those with lower urine BPA levels [156]. A large study in Chinese school children (more than 2000 participants from middle and high school) demonstrated that urine BPA levels were positively associated with BMI [157]. The limitation of the most presented studies is their cross-sectional design, so there is a need for longitudinal studies to further scrutinize the link between BPA and obesity development.

Recent studies demonstrate that BPA exposure may significantly affect adipogenesis through the modulation of key adipogenesis regulators (PPAR γ and C/EBPs) through interference with receptor signaling. Specifically, BPA exposure was shown to up-regulate adipogenesis in human adipose stromal/stem cells due to the activation of PPAR γ and C/EBP α in an estrogen receptor-dependent manner [158]. It has also been shown that the adipogenic effect of BPA may be mediated through its interference with glucocorticoid signaling. Specifically, BPA exposure was shown to stimulate PPAR- γ mRNA expression in human visceral (pre)adipocytes by increasing mRNA expression and the activity of 11 β -HSD1, an enzyme involved in cortisol formation [159]. In addition to the earlier reported role of PPAR γ and C/EBP α , the adipogenic effect of BPA exposure in 3T3-L1 cells may be

also mediated by the increase in transcriptional activity of the glucocorticoid receptor and C/EBP δ [160]. At the same time, another study demonstrated that BPA-induced adipogenesis in human preadipocytes is dependent on estrogen receptor signaling, rather than the glucocorticoid pathway [161]. It is also notable that the impact of BPA on PPAR γ signaling during adipogenesis in committed 3T3L1 and uncommitted NIH3T3 preadipocytes may be mediated by the BPA-induced reduction of preadipocyte peroxisome proliferator-activated receptor gamma (PPAR γ) promoter methylation [162]. The activation of the PI3K/Akt pathway is also considered as the potential mechanism of BPA proadipogenic activity in 3T3-L1 cells [163].

In addition to the modulation of adipocyte proliferation and differentiation, BPA exposure promoted lipid accumulation, proinflammatory cytokine production, and reduced insulin sensitivity in mature 3T3-L1 adipocytes [164]. Moreover, BPA-induced insulin resistance in adipocytes was shown to be independent of adipogenesis, being associated with reduced insulin-induced Akt phosphorylation and increased proinflammatory cytokine mRNA levels and being indicative of the role of BPA-induced inflammation in insulin resistance [165]. Correspondingly, the BPA-induced alteration of insulin receptor phosphorylation and signaling in adipocytes derived from subcutaneous adipose tissue and differentiated 3T3-L1 cells was shown to be JNK-dependent [166]. Moreover, environmentally relevant doses of BPA were also shown to decrease adiponectin production by human adipose tissue [167], which may significantly contribute to insulin resistance.

Being in agreement with *in vitro* studies, *in vivo* experiments also demonstrated the impact of BPA exposure on adipogenesis. Specifically, exposure to low doses, but not high doses, of BPA induced a significant increase in adipose tissue mass with an elevation in both adipocyte size and volume, as well as circulating leptin levels and insulin resistance [168]. Perinatal BPA exposure in Sprague-Dawley rats was shown to increase white adipose tissue mass through inducing adipocyte hypertrophy due to the up-regulation of PPAR- γ , C/EBP- α , SREBP-1C, LPL, FAS, and stearoyl-CoA desaturase 1 (SCD-1), although this effect was observed only in females [169].

At the same time, *in vivo* studies revealed the significant sex-specific effects of BPA on adiposity [170]. Specifically, BPA exposure was shown to induce an increase in body adiposity and adipose tissue inflammation in STD-fed female C57BL/6J mice, whereas no effect was observed in HFD-fed and/or male mice [171]. In turn, another study demonstrated a significant BPA-induced increase in the body adiposity in male Sprague-Dawley rats [172]. Given this inconsistency and the observation of sex-specific effects, it is proposed that BPA is not considered as a specific obesogen in laboratory rodents (C57BL/6JxFVB mice), although it programs for metabolic dysregulation [173].

Several studies have evaluated the impact of BPA derivatives BPA-glucuronide (BPA-G) [174] and bisphenol A diglycidyl ether (BPA-DGE) [175] on adipogenesis. Specifically, BPA-glucuronide (BPA-G), the main BPA metabolite, was also shown to induce adipogenesis in human and 3T3L1 murine preadipocytes through a mechanism involving estrogen receptor activation without the direct estrogenic activity of the compound [174]. Bisphenol A diglycidyl ether also possessed a more profound adipogenic effect as compared to BPA by inducing adipogenesis both in mesenchymal stromal stem cells and 3T3-L1 preadipocytes, whereas BPA was capable of inducing an adipogenic response only in the latter [175].

A comparative analysis of various bisphenol species using the preadipocytic 3T3-L1 cell line demonstrated that bisphenol F and especially bisphenol S possess a more profound adipogenic effect as compared to BPA, as demonstrated by higher PPAR γ and C/EBP α protein expression [176]. A similar PPAR γ -mediated effect was observed in primary human preadipocytes [177]. Both BPA and BPS were shown to activate PPAR γ in murine preadipocytes through targeting the PPAR γ response element, although the mechanism may be quite different [178]. Correspondingly, BPA and BPS induced distinct transcriptional patterns in differentiating human primary preadipocytes that may underlie the differences in adipogenic effects of these compounds. While BPA was shown to affect mainly the pathways involved in liver X receptor/retinoid X receptor (LXR/RXR) activation, hepatic

fibrosis, cholestasis, and atherosclerosis signaling, BPS exposure perturbed adenosine monophosphate-activated protein kinase (AMPK) signaling, cholesterol biosynthesis, and adipogenesis pathways, as well as LXR/RXR and PPAR α /RXR α activation [179]. In agreement with in vitro observations, in vivo prenatal BPS exposure was shown to result in white adipose tissue hypertrophy and the up-regulation of PPAR γ gene expression in HFD-fed mice, being indicative of the role of BPS in increasing susceptibility to dietary obesity [180].

Certain studies also demonstrated that bisphenol AF may also promote adipogenesis and induce proinflammatory signaling pathways in murine 3T3L1 preadipocytes [181].

Generally, BPA exposure may be considered a risk factor for human obesity as evidenced by epidemiological studies. The promotion of PPAR γ and C/EBP α -dependent adipogenesis upon exposure to BPA may be mediated by glucocorticoid and estrogen receptor signaling, also involving the up-regulation of C/EBP δ (Figure 2). The epigenetic effects of BPA as well as the induction of adipose tissue inflammation may also contribute to obesity pathogenesis and obesity-associated insulin resistance (Figure 5). Involvement of the estrogen receptor may also underlie certain sex-specific obesogenic effects of BPA. It is also notable that other bisphenol (S, F, AF) species may possess obesogenic effects through similar mechanisms.

9. Phthalates

Phthalates (Figure 9), diesters of 1,2-benzendicarboxylic acid, are a group of chemicals used as additives in plastics. These chemicals are listed as endocrine disrupting chemicals [182]. Many epidemiological studies have examined the association between phthalates (measured as metabolites in urine) and body weight and obesity [183–187]. Results from a cross sectional study conducted on 242 participants (6–18 years of age) have shown an association between urinary phthalate metabolites and obesity, triglyceride, and blood pressure [183]. Hatch et al. used data from the US National Health and Nutrition Examination Surveys (NANHES) and revealed a number of different positive associations between phthalate exposure and BMI and waist circumference, with the most coherent results in males from 20–59 years old [184]. A study investigating 128 newborns demonstrated the positive association between urine di(2-ethylhexyl)phthalate (DEHP) levels and an increase in body weight at the 3rd month of life [185]. In the Korean National Environmental Health Survey (KoNEHS) conducted between 2015 and 2017 with a sample size of 3782 subjects, DEHP and benzylbutylphthalate (BzBP) urine levels were found to be associated with obesity [152]. A case–control study performed in Iran ($n = 320$) showed a positive association between phthalates metabolites and the BMI of participants [186]. Contrary to this, a Korean cross-sectional study of 702 participants did not find a significant association between the urinary levels of six phthalates metabolites and general and/or abdominal obesity in both genders [153]. And lastly, a negative association between phthalates metabolites and BMI was reported in a study investigating these parameters in American children [187]. It is evident that there are controversial data describing the connection between exposure to phthalates and obesity. It should be highlighted that most of the studies are of cross-sectional design. Hence, there is a need for large prospective studies which would confirm or infirm the existence of this connection.

The existing data demonstrate the adipogenic potential of phthalate exposure due to the up-regulation of PPAR γ [188]. Specifically, the role of PPAR γ activation in the adipogenic response in 3T3L1 cells was demonstrated for MEHP and DEHP [189], as well as other phthalates, including monobenzyl phthalate (MBzP) and mono-sec-butyl phthalate (MBuP) [190], benzyl butyl phthalate (BBP) [191,192], and diisononyl phthalate (DINP) [193]. It is also notable that the DEHP-induced up-regulation of PPAR γ also requires the activation of Med1 and PGC-1 α coregulators in 3T3L1 cells [189].

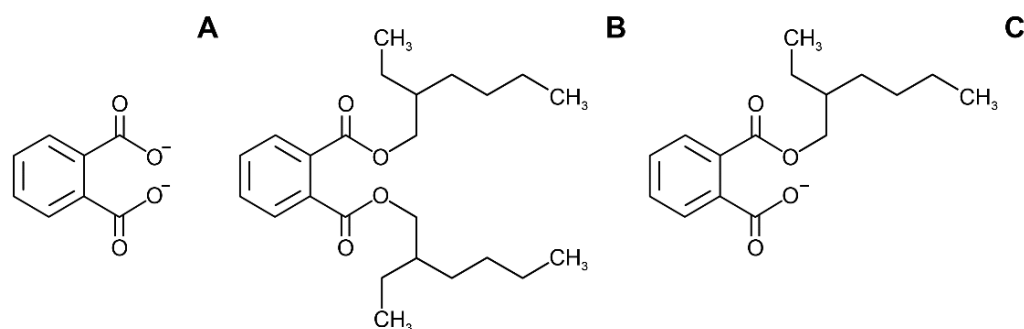


Figure 9. The common structure of phthalates (A), as well as the most common representatives di(2-ethylhexyl)phthalate (DEHP) (B) and its metabolite mono(2-ethylhexyl)phthalate (MEHP) (C).

Correspondingly, certain studies demonstrate that phthalate exposure to pluripotent bone marrow stromal cells may promote adipogenesis at the expense of osteoblastogenesis [194] and Leydig cell differentiation [195] through the up-regulation of PPAR γ and C/EBP α expression.

Phthalate-induced adipogenesis in 3T3L1 cells may be also dependent on the activation of the glucocorticoid receptor [196]. In agreement, molecular docking analysis demonstrated that dicyclohexyl phthalate (DCHP) and mono-cyclohexyl phthalate (MCHP) bind GR active sites with binding affinities close to that of dexamethasone [197].

In agreement with the phthalate-induced modulation of adipogenesis, exposure to this compound was also shown to affect adipokine secretion. Specifically, a DEHP-induced increase in leptin and FABP4 mRNA expression was associated with adiponectin down-regulation [198]. In contrast, another study revealed a significant inhibition in adipokine production in 3T3-L1 adipocytes upon MEHP exposure [199].

In parallel with the promotion of 3T3-L1 murine preadipocyte differentiation to adipocytes, MEHP exposure also increased the expression of genes implicated in lipid uptake, biosynthesis, and accumulation [200]. In addition, MEHP-induced lipid accumulation in 3T3-L1 adipocytes may be associated with the up-regulation of the Notch pathway as evidenced by increased Notch-1 and Jagged-2 expression [201]. The activation of the TYK2/STAT-3 pathway may be also considered as another candidate mechanism promoting 3T3-L1 adipocyte differentiation and lipid accumulation [202].

In contrast, one of the studies demonstrated that DEHP exposure may induce white adipocyte browning, as evidenced by the increased expression of *PPARG*, *ADRB1*, *ADRB3*, *PPARGC1a*, and *UCP1* [203]. MEHP accumulation in adipocytes was also shown to reduce adipocyte size through a significant increase in lipolysis, glucose uptake, glycolysis, mitochondrial respiration, and mitochondrial biogenesis in 3T3-L1 cells [204].

The results of in vivo studies generally correspond to in vitro data, demonstrating the impact of phthalate exposure on body adiposity and obesity-associated metabolic disturbances. Specifically, i.p. DEHP injection to C57BL/6J mice resulted in a significant increase in adipose tissue weight, as well as circulating total cholesterol, glucose, and triglyceride levels [205]. A similar effect was observed following in utero phthalate exposure. In utero MEHP exposure significantly increased body weight and adipose tissue mass along with the development of hyperglycemia and dyslipidemia in mouse offspring [206]. In agreement, in utero exposure to DEHP in C57BL/6J mice was shown to increase body adiposity in the offspring accompanied by the elevation of circulating leptin, insulin, lipid, and glucose levels [207], as well as blood pressure through the dysregulation of AT1R signaling and eNOS activation [208], thus promoting metabolic syndrome.

The observed phthalate-induced insulin resistance in male Wistar rats may be associated with a decrease in adipocyte glucose uptake, resulting in hyperglycemia and insulin resistance in rats through the down-regulation of insulin receptor and IRS-1 mRNA expression, as well as the inhibition of Akt phosphorylation [209]. Moreover, the protective role

of ascorbic acid and α -tocopherol demonstrates the role of oxidative stress in the alteration of adipose tissue insulin signaling [209].

BBP exposure in C57BL/6 mice was also shown to potentiate the adipogenic effect of HFD through an increase in liver and adipose tissue mass, also inducing insulin resistance, although the effect was observed only in moderate, but not low or high, doses [210].

DEHP exposure was also shown to induce rat adipose tissue infiltration with macrophages with the subsequent secretion of $\text{TNF}\alpha$ and IL-1b, which promoted adipose tissue dysfunction and altered lipid metabolism [211]. In agreement, in utero DEHP exposure in Sprague–Dawley rats induced both adipose tissue and systemic inflammation in parallel with increasing preadipocyte differentiation [212]. It is also proposed that $\text{PPAR}\gamma$ may be at least partially responsible for the proinflammatory response in differentiated murine adipocytes in parallel with the adipogenic effects [213].

Additional mechanisms linking phthalate exposure and obesity pathogenesis were also proposed. Specifically, the epigenetic effects of phthalate exposure were shown to contribute to phthalate-induced adipogenesis in MSCs and high-fat fed rats [214,215], although this effect in rats is expected to be sex-specific [216]. In addition, the most recent study also demonstrated that altered gut microbiota with a decrease of the Firmicutes-to-Bacteroidetes ratio may be associated with DEHP-induced obesity in mice [217].

Despite the contradictory epidemiological data, laboratory findings demonstrate that exposure to phthalates promoted adipogenesis through the up-regulation of $\text{C}/\text{EBP}\alpha$ and $\text{PPAR}\gamma$ signaling (Figure 2), as well as lipid accumulation in adipocytes due to the activation of lipid biosynthesis. In addition, phthalate-induced adipokine dysregulation, adipose tissue inflammation, and epigenetic effects may also contribute to obesity and obesity-associated metabolic disturbances.

10. Diethylstilbestrol (DES)

Diethylstilbestrol (DES) (Figure 10) is a synthetic drug with estrogen activity that was used as a therapy for the prevention of miscarriage and other pregnancy complications during the period from 1940–1970 and was recognized as a potential obesogenic in animal studies [218,219]. However, there are only a few epidemiological studies that examined the obesogenic potential of DES in in utero-exposed children. A large prospective study, covering three cohorts, has shown that DES-exposed women had a slightly greater weight than unexposed controls [220]. Another study performed using data from the Collaborative Perinatal Project ($n = 34,419$), found strong a positive association between prenatal exposure to DES at 3–4 months and 4–5 months of pregnancy and obesity in children at 7 years of age [221].

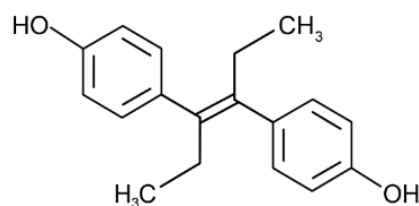


Figure 10. The structural formula of diethylstilbestrol (DES).

DES exposure was shown to induce adipogenesis via the estrogen receptor-mediated activation of $\text{PPAR}\gamma$ and $\text{C}/\text{EBP}\alpha$ expression in 3T3-L1 preadipocytes with the subsequent up-regulation of target genes including aP2 , FAS , and LPL . A similar effect was observed in vivo [218]. In contrast, in mature adipocytes, DES exposure resulted in a significant decrease in PPAR expression due to the down-regulation of $\text{ER}\alpha$ and $\text{ER}\beta$ [222].

Certain studies demonstrated that DES may also affect other mechanisms implicated in obesity pathogenesis. Specifically, DES was capable of inhibiting mitochondrial respiration and glycolysis as well as activating ERK in 3T3-L1 adipocytes in an ER-dependent

manner [223]. In addition, DES exposure was shown to induce WAT inflammation in obese mice [224].

Taken together, epidemiological data on the potential role of DES as an environmental obesogen are insufficient, although laboratory findings indicate that adipose tissue could be considered as a potential target for this pollutant.

11. POPs Mixtures and Obesity

In view of the recent rate of environmental pollution, humankind is more frequently exposed to a mixture of various chemicals rather than to one particular pollutant [225]. Therefore, an investigation of the impact of various POPs mixtures on obesity and adipogenesis is of particular interest.

Data from the Center for Health Assessment of Mothers and Children of Salinas (CHAMACOS) study demonstrated that obesity is directly associated with serum dichloro-diphenyltrichloroethane, hexachlorocyclohexane, and PBDE-47 concentrations, whereas PBDE-153 was characterized by an inverse association [93]. Another study also revealed an association between obesity and urinary mono (carboxyoctyl) phthalate, BPA, and BPS levels in the NHANES 2013–2014 cohort [226]. Although these findings clearly demonstrate that exposure to POPs mixtures may be associated with anthropometric markers of obesity, the particular effect of multiple pollutant co-exposure is unclear.

Only single studies demonstrated the impact of POPs mixtures on obesity and adipogenesis mechanisms. Specifically, the most recent study demonstrated that a mixture of POPs in human exposure-relevant doses promoted adipogenesis in 3T3-L1 cells, inducing higher lipid accumulation as compared to single chemicals [53]. It has been also demonstrated that female zebrafish exposed to natural mixture of POPs are characterized by significantly increased body weight, which may be associated with the modulation of PPAR γ and C/EBP α signaling or with other endocrine disturbances [227]. Further *in vivo* and *in vitro* studies aimed at the investigation of the interactive effects of POPs on adipogenesis upon co-exposure are required [228].

12. A Summary of Key Targets for POPs Obesogenic Effects

The reviewed studies demonstrated that the obesogenic effects of POPs may be mediated by their impact on adipogenic transcription regulators, namely PPAR γ and C/EBPs, as well as nuclear receptors including ER and GR. The role of these mechanisms in adipogenesis and adipose tissue functioning will be briefly reviewed.

12.1. PPAR and C/EBP

The above reviewed studies clearly indicate that PPAR γ may be considered as a target for all discussed POPs. PPAR γ is a key regulator of adipogenesis that activates preadipocyte differentiation and the expression of adipocyte-specific genes involved in lipid metabolism and accumulation, insulin sensitivity, and thermogenesis [229]. Correspondingly, targeted adipose tissue PPAR γ knockout results in reduced adipogenesis [230]. It is also notable that PPAR γ is also essential for the survival of mature adipocytes [231]. At the same time, whole-body PPAR γ knockout was shown to be lethal due to placental defects [232].

PPAR γ expression is regulated by a wide spectrum of transcription factors, with the C/EBP family playing a key role [233]. C/EBP β and C/EBP δ are induced at early steps of adipogenesis, subsequently up-regulating PPAR γ expression [234]. PPAR γ activity is integrated with another transcription factor, C/EBP α , and both transcriptions potentiate the expression of each other [234]. However, certain studies demonstrated that C/EBP α is unable to induce adipogenesis without PPAR γ , which is considered as the proximal regulator of adipogenesis [235].

PPAR activation is also regulated by epigenetic modifications including chromatin remodeling and histone acetylation and methylation [236]. Specifically, the demethylation of PPAR γ promoter during adipogenesis is associated with its activation [237]; this was also observed upon exposure to POPs.

PPAR activation is also known to be involved in the regulation of insulin sensitivity through the up-regulation of GluT4, PI3K, and IRS1/2 [238]. However, upon PPAR γ overexpression, increased glucose and fatty acid uptake, in parallel with increased adipogenesis, may promote weight gain, resulting in visceral obesity [229]. In view of the data demonstrating the interference of POPs with insulin signaling through the inhibition of insulin receptor IRS1/2 and Akt phosphorylation, as well as PTEN activation (Figure 5), the insulin-sensitizing effect of PPAR γ activation may be abrogated by exposure to POPs.

Therefore, aberrant PPAR γ activation upon exposure to POPs may be considered as one of the key mechanisms mediating the obesogenic effects of POPs, although the particular mode of POP-PPAR interaction is still to be further explored [239].

12.2. Estrogen Receptor (ER)

Certain POPs, including DDT, PCB, BPA, and DES, were shown to mediate their adipogenic effects through targeting estrogen receptors. These observations correspond to the earlier reported estrogenic activity of PCBs and DES.

ER α and ER β are known to be involved in the regulation of adipose tissue functioning as well as adipocyte differentiation, although the effects are receptor-specific [240].

ER β was shown to inhibit PPAR γ transcription and activity resulting in reduced adipogenesis [241], which may be at least partially mediated by the competition between PPAR γ and ER β for coactivator binding [242]. Correspondingly, selective ER β agonists were shown to inhibit basal and PGC-1-coactivated PPAR- γ transactivation [243]. In vivo, ER β activation significantly reduced both visceral and subcutaneous adipose tissue mass in HFD-fed mice [244]. In turn, ER β inhibition is associated with the activation of PPAR γ signaling and subsequent adipogenesis [245].

At the same time, ER α is believed to have a predominant role in the regulation of adipogenesis when compared to ER β . ER α activation upon estradiol treatment was shown to increase 3T3-L1 adipocyte differentiation through C/EBP α signaling, along with increased GluT4 expression and improved insulin sensitivity [246]. Correspondingly, ER α knockdown is associated with adipocyte hypertrophy [247].

ER α expression in white adipocytes is also associated with reduced visceral adiposity through the modulation of mitochondrial dynamics due to the up-regulation of Polg1 (mtDNA polymerase γ -subunit) [248]. Correspondingly, ER α activation is also associated with adipocyte beiging through the up-regulation of AMPK, adipose tissue triglyceride lipase (ATGL), and UCP-1 expression [249].

Based on the above mentioned studies, it is considered that physiologically high Era expression is required for adipogenesis and normal adipose tissue functioning, whereas its Era down-regulation affects adipogenesis and promotes adipocyte hypertrophy together with ectopic lipid accumulation [250].

At the same time, certain studies demonstrated that Era signaling may also repress adipogenesis through the up-regulation of AKT with the subsequent recruitment of co-repressors GATA3 and β -catenin/TCF4 complex to the PPAR γ promoter, ultimately leading to reduced PPAR γ activity. Moreover, AKT activation was also shown to facilitate ER α nuclear translocation and localization at the PPAR γ gene promoter through the inhibition of trimethylation of lysine 27 of histone H3 (H3K27me3) and increased wingless-integrated (Wnt) 1 and 10b expression [251]. Correspondingly, the role of H3K27me2 and H3K27me3 in Era induced a decrease in PPAR γ and C/EBP α expression [252].

In agreement with the distinct effects of ERs in adipocytes, it has been demonstrated that the ER α /ER β ratio is associated with obesity and leptin production, with higher BMI values in cases with a shift to the prevalence of ER β [253].

The specific patterns of estrogen receptor signaling may mediate the observed sexual dimorphism in the obesogenic response to POPs. Specifically, it has been demonstrated that 17 β -Estradiol is capable of inducing ER α and ER β mRNA expression in female subcutaneous adipocytes, whereas in male cells, only ER α was up-regulated [246]. In addition,

Er α deficiency in adipose tissue was shown to induce adipocyte hypertrophy as well as fibrosis and inflammation, being more profound in males [247].

Given the significant role of estrogens in the regulation of adipose tissue functioning, the difference in estrogen production and the distinct patterns of ERs distribution in men and women may mediate the observed sex-specific effects of exposure to POPs [254].

12.3. Glucocorticoid Receptor

The activation of glucocorticoid receptor signaling by its ligand, dexamethasone, is known to be involved in the activation of adipogenesis through the up-regulation of proadipogenic transcription factors [255]. Specifically, GR activation was shown to be involved in the up-regulation of C/EBP β [256,257] and C/EBP δ [258] expression with the subsequent induction of C/EBP α . It has been also demonstrated that GR-induced adipogenesis may be mediated by KLF-15-induced PPAR γ activation [259].

At the same time, certain studies demonstrated that GR signaling is not required for *in vivo* adipogenesis [260]. Correspondingly, another study demonstrated that the activation of GR signaling by dexamethasone accelerates adipocyte differentiation through the up-regulation of C/EBP α , C/EBP β , C/EBP δ , KLF5, KLF9, and PPAR γ expression, being yet dispensable for adipogenesis, as demonstrated in GR-deficient preadipocytes that were capable of further differentiation [261]. Being in line with these observations, an *in vivo* study demonstrated that targeted adipocyte GR deletion does not reduce adipose tissue weight and distribution, although it does promote adipose tissue inflammation upon high-fat feeding [262].

At the same time, the particular effect of GR signaling in POPs-associated obesity is unclear, especially in view of the earlier demonstrated predominant role of ER activation upon exposure to POPs [161].

13. Conclusions

The existing epidemiological data demonstrate a significant association between lifetime and prenatal exposure to POPs and obesity and obesity-associated metabolic disturbances (e.g., type 2 diabetes mellitus and metabolic syndrome), although the existing data are rather insufficient and sometimes contradictory.

Laboratory *in vitro* data have clearly and consistently demonstrated the potential mechanisms associated with the interference of exposure to POPs with obesity, including: (i) the effects on adipogenesis regulators (PPAR γ , C/EBP α); (ii) the endocrine-disrupting effects and binding to nuclear receptors (ER, GR); (iii) the epigenetic effects; (iv) proinflammatory activity; and (v) the induction of insulin resistance. The involvement of estrogen-like effects of certain POPs and ER signaling may also underlie the sex-specific differences in POPs' adipogenic effects. Although *in vivo* data are generally corroborative of the *in vitro* results, studies in living organisms demonstrated that the impact of POPs on adipogenesis is largely affected by biological factors including sex, age, and period of exposure.

Therefore, both laboratory and epidemiological data underline the significant role of POPs as environmental obesogens. However, further studies are required to better characterize both the mechanisms and the dose-response effects of exposure to POPs in the etiology of obesity and other metabolic diseases.

Author Contributions: Conceptualization, J.A., A.B.D. and A.A.T.; methodology, A.B.D. and A.A.T.; formal analysis, D.J. and Z.B.; investigation, D.J., A.B.D., Z.B., A.V.S., I.P.Z. and A.A.T.; writing—original draft preparation, D.J., A.B.D., Z.B., I.P.Z. and A.A.T.; writing—review and editing, J.A., A.V.S. and M.A.; visualization, I.P.Z. and A.A.T.; supervision, J.A., M.A. and A.V.S. All authors have read and agreed to the published version of the manuscript.

Funding: The study was supported by the Ministry of Science and Higher Education of the Russian Federation within the framework of state support for the creation and development of World-Class Research Centers “Digital biodesign and personalized healthcare” 075-15-2020-926.

Institutional Review Board Statement: Not applicable.

Acknowledgments: This work of DJ, ABD, and ZB was partly funded by the Ministry of Education, Science and Technological Development of the Republic of Serbia (No: 451-03-9/2021-14/200161).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Haththotuwa, R.N.; Wijeyaratne, C.N.; Senarath, U. Worldwide Epidemic of Obesity. In *Obesity*; Elsevier: Amsterdam, The Netherlands, 2013; pp. 3–11, ISBN 9780124160453.
2. Blüher, M. Obesity: Global Epidemiology and Pathogenesis. *Nat. Rev. Endocrinol.* **2019**, *15*, 288–298. [[CrossRef](#)]
3. Martínez-Villanueva, J.; González-Leal, R.; Argente, J.; Martos-Moreno, G.Á. La Obesidad Parental Se Asocia Con La Gravedad de La Obesidad Infantil y de Sus Comorbilidades. *An. Pediatr.* **2019**, *90*, 224–231. [[CrossRef](#)]
4. The GBD 2015 Obesity Collaborators. Health Effects of Overweight and Obesity in 195 Countries over 25 Years. *N. Engl. J. Med.* **2017**, *377*, 13–27. [[CrossRef](#)]
5. Maury, E.; Brichard, S.M. Adipokine Dysregulation, Adipose Tissue Inflammation and Metabolic Syndrome. *Mol. Cell. Endocrinol.* **2010**, *314*, 1–16. [[CrossRef](#)]
6. Mottillo, S.; Filion, K.B.; Genest, J.; Joseph, L.; Pilote, L.; Poirier, P.; Rinfret, S.; Schiffrin, E.L.; Eisenberg, M.J. The Metabolic Syndrome and Cardiovascular Risk. *J. Am. Coll. Cardiol.* **2010**, *56*, 1113–1132. [[CrossRef](#)]
7. Zammit, C.; Liddicoat, H.; Moonsie, I.; Makker, H. Obesity and Respiratory Diseases. *Am. J. Clin. Hypn.* **2011**, *53*, 335–343. [[CrossRef](#)]
8. O'Brien, P.D.; Hinder, L.M.; Callaghan, B.C.; Feldman, E.L. Neurological Consequences of Obesity. *Lancet Neurol.* **2017**, *16*, 465–477. [[CrossRef](#)]
9. Avgerinos, K.I.; Spyrou, N.; Mantzoros, C.S.; Dalamaga, M. Obesity and Cancer Risk: Emerging Biological Mechanisms and Perspectives. *Metabolism* **2019**, *92*, 121–135. [[CrossRef](#)]
10. Dietz, W.; Santos-Burgoa, C. Obesity and Its Implications for COVID-19 Mortality. *Obesity* **2020**, *28*, 1005. [[CrossRef](#)]
11. Kadouh, H.C.; Acosta, A. Current Paradigms in the Etiology of Obesity. *Tech. Gastrointest. Endosc.* **2017**, *19*, 2–11. [[CrossRef](#)]
12. Baillie-Hamilton, P.F. Chemical Toxins: A Hypothesis to Explain the Global Obesity Epidemic. *J. Altern. Complement. Med.* **2002**, *8*, 185–192. [[CrossRef](#)]
13. Wang, Y.; Hollis-Hansen, K.; Ren, X.; Qiu, Y.; Qu, W. Do Environmental Pollutants Increase Obesity Risk in Humans? *Obes. Rev.* **2016**, *17*, 1179–1197. [[CrossRef](#)]
14. Hatch, E.E.; Nelson, J.W.; Stahlhut, R.W.; Webster, T.F. Association of Endocrine Disruptors and Obesity: Perspectives from Epidemiological Studies. *Int. J. Androl.* **2010**, *33*, 324–332. [[CrossRef](#)]
15. Ashraf, M.A. Persistent Organic Pollutants (POPs): A Global Issue, a Global Challenge. *Environ. Sci. Pollut. Res.* **2017**, *24*, 4223–4227. [[CrossRef](#)]
16. Gaur, N.; Narasimhulu, K.; PydiSetty, Y. Recent Advances in the Bio-Remediation of Persistent Organic Pollutants and Its Effect on Environment. *J. Clean. Prod.* **2018**, *198*, 1602–1631. [[CrossRef](#)]
17. World Health Organisation. *Food Safety: Persistent Organic Pollutants (POPs)*; World Health Organisation: Geneva, Switzerland, 2020.
18. González, N.; Domingo, J. Concentrations of Persistent Organic Pollutants in Blood of the Spanish Population: Temporal Trend. *Arch. Farm.* **2021**, *71*, 455–479. [[CrossRef](#)]
19. Eskenazi, B.; Chevrier, J.; Rosas, L.G.; Anderson, H.A.; Bornman, M.S.; Bouwman, H.; Chen, A.; Cohn, B.A.; de Jager, C.; Henshel, D.S.; et al. The Pine River Statement: Human Health Consequences of DDT Use. *Environ. Health Perspect* **2009**, *117*, 1359–1367. [[CrossRef](#)]
20. Panieri, E.; Buha-Dorđević, A.; Saso, L. Endocrine Disruption by PFAS: A Major Concern Associated with Legacy and Replacement Substances. *Arch. Farm.* **2021**, *71*, 429–454. [[CrossRef](#)]
21. Linares, V.; Perelló, G.; Nadal, M.; Gómez-Catalán, J.; Llobet, J.M.; Domingo, J.L. Environmental versus Dietary Exposure to POPs and Metals: A Probabilistic Assessment of Human Health Risks. *J. Environ. Monit.* **2010**, *12*, 681–688. [[CrossRef](#)]
22. Jones, K.C.; de Voogt, P. Persistent Organic Pollutants (POPs): State of the Science. *Environ. Pollut.* **1999**, *100*, 209–221. [[CrossRef](#)]
23. Kärman, A.; van Bavel, B.; Järnberg, U.; Hardell, L.; Lindström, G. Perfluorinated Chemicals in Relation to Other Persistent Organic Pollutants in Human Blood. *Chemosphere* **2006**, *64*, 1582–1591. [[CrossRef](#)] [[PubMed](#)]
24. Davidsen, N.; Lauvås, A.J.; Myhre, O.; Ropstad, E.; Carpi, D.; Gyves, E.M.; Berntsen, H.F.; Dirven, H.; Paulsen, R.E.; Bal-Price, A.; et al. Exposure to Human Relevant Mixtures of Halogenated Persistent Organic Pollutants (POPs) Alters Neurodevelopmental Processes in Human Neural Stem Cells Undergoing Differentiation. *Reprod. Toxicol.* **2021**, *100*, 17–34. [[CrossRef](#)]
25. La Merrill, M.; Emond, C.; Kim, M.J.; Antignac, J.-P.; Le Bizec, B.; Clément, K.; Birnbaum, L.S.; Barouki, R. Toxicological Function of Adipose Tissue: Focus on Persistent Organic Pollutants. *Environ. Health Perspect.* **2013**, *121*, 162–169. [[CrossRef](#)]
26. Darbre, P.D. Endocrine Disruptors and Obesity. *Curr. Obes. Rep.* **2017**, *6*, 18–27. [[CrossRef](#)]
27. Nappi, F.; Barrea, L.; Di Somma, C.; Savanelli, M.; Muscogiuri, G.; Orio, F.; Savastano, S. Endocrine Aspects of Environmental “Obesogen” Pollutants. *Int. J. Environ. Res. Public Health* **2016**, *13*, 765. [[CrossRef](#)]
28. Heindel, J.J.; Blumberg, B. Environmental Obesogens: Mechanisms and Controversies. *Annu. Rev. Pharmacol. Toxicol.* **2019**, *59*, 89–106. [[CrossRef](#)]

29. Stel, J.; Legler, J. The Role of Epigenetics in the Latent Effects of Early Life Exposure to Obesogenic Endocrine Disrupting Chemicals. *Endocrinology* **2015**, *156*, 3466–3472. [[CrossRef](#)]
30. Grün, F.; Blumberg, B. Endocrine Disrupters as Obesogens. *Mol. Cell. Endocrinol.* **2009**, *304*, 19–29. [[CrossRef](#)]
31. Griffin, M.D.; Pereira, S.R.; DeBari, M.K.; Abbott, R.D. Mechanisms of Action, Chemical Characteristics, and Model Systems of Obesogens. *BMC Biomed. Eng.* **2020**, *2*, 6. [[CrossRef](#)]
32. Inoue, K.; Goto, A.; Sugiyama, T.; Ramlau-Hansen, C.H.; Liew, Z. The Confounder-Mediator Dilemma: Should We Control for Obesity to Estimate the Effect of Perfluoroalkyl Substances on Health Outcomes? *Toxics* **2020**, *8*, 125. [[CrossRef](#)]
33. Apelberg, B.J.; Goldman, L.R.; Calafat, A.M.; Herbstman, J.B.; Kuklenyik, Z.; Heidler, J.; Needham, L.L.; Halden, R.U.; Witter, F.R. Determinants of Fetal Exposure to Polyfluoroalkyl Compounds in Baltimore, Maryland. *Environ. Sci. Technol.* **2007**, *41*, 3891–3897. [[CrossRef](#)]
34. Lauritzen, H.B.; Larose, T.L.; Øien, T.; Sandanger, T.M.; Odland, J.O.; Van De Bor, M.; Jacobsen, G.W. Prenatal Exposure to Persistent Organic Pollutants and Child Overweight/Obesity at 5-Year Follow-up: A Prospective Cohort Study. *Environ. Health Glob. Access Sci. Source* **2018**, *17*, 9. [[CrossRef](#)]
35. Braun, J.M. Early-Life Exposure to EDCs: Role in Childhood Obesity and Neurodevelopment. *Nat. Rev. Endocrinol.* **2017**, *13*, 161–173. [[CrossRef](#)]
36. Halldorsson, T.I.; Rytter, D.; Haug, L.S.; Bech, B.H.; Danielsen, I.; Becher, G.; Henriksen, T.B.; Olsen, S.F. Prenatal Exposure to Perfluorooctanoate and Risk of Overweight at 20 Years of Age: A Prospective Cohort Study. *Environ. Health Perspect.* **2012**, *120*, 668–673. [[CrossRef](#)]
37. Domazet, S.L.; Grøntved, A.; Timmermann, A.G.; Nielsen, F.; Jensen, T.K. Longitudinal Associations of Exposure to Perfluoroalkylated Substances in Childhood and Adolescence and Indicators of Adiposity and Glucose Metabolism 6 and 12 Years Later: The European Youth Heart Study. *Diabetes Care* **2016**, *39*, 1745–1751. [[CrossRef](#)]
38. Geiger, S.D.; Yao, P.; Vaughn, M.G.; Qian, Z. PFAS Exposure and Overweight/Obesity among Children in a Nationally Representative Sample. *Chemosphere* **2021**, *268*, 128852. [[CrossRef](#)]
39. Schrenk, D.; Bignami, M.; Bodin, L.; Chipman, J.K.; del Mazo, J.; Grasl-Kraupp, B.; Hogstrand, C.; Hoogenboom, L.; Leblanc, J.C.; Nebbia, C.S.; et al. Risk to Human Health Related to the Presence of Perfluoroalkyl Substances in Food. *EFSA J.* **2020**, *18*, e06223. [[CrossRef](#)]
40. Watkins, A.M.; Wood, C.R.; Lin, M.T.; Abbott, B.D. The Effects of Perfluorinated Chemicals on Adipocyte Differentiation in Vitro. *Mol. Cell. Endocrinol.* **2015**, *400*, 90–101. [[CrossRef](#)]
41. Liu, W.; Qin, H.; Pan, Y.; Luo, F.; Zhang, Z. Low Concentrations of Perfluorooctane Sulfonate Repress Osteogenic and Enhance Adipogenic Differentiation of Human Mesenchymal Stem Cells. *Toxicol. Appl. Pharmacol.* **2019**, *367*, 82–91. [[CrossRef](#)]
42. Yamamoto, J.; Yamane, T.; Oishi, Y.; Kobayashi-Hattori, K. Perfluorooctanoic Acid Binds to Peroxisome Proliferator-Activated Receptor γ and Promotes Adipocyte Differentiation in 3T3-L1 Adipocytes. *Biosci. Biotechnol. Biochem.* **2015**, *79*, 636–639. [[CrossRef](#)]
43. Ma, Y.; Yang, J.; Wan, Y.; Peng, Y.; Ding, S.; Li, Y.; Xu, B.; Chen, X.; Xia, W.; Ke, Y.; et al. Low-Level Perfluorooctanoic Acid Enhances 3 T3-L1 Preadipocyte Differentiation via Altering Peroxisome Proliferator Activated Receptor Gamma Expression and Its Promoter DNA Methylation. *J. Appl. Toxicol.* **2018**, *38*, 398–407. [[CrossRef](#)]
44. Li, Z.; Yu, Z.; Gao, P.; Yin, D. Multigenerational Effects of Perfluorooctanoic Acid on Lipid Metabolism of *Caenorhabditis Elegans* and Its Potential Mechanism. *Sci. Total Environ.* **2020**, *703*, 134762. [[CrossRef](#)]
45. Van Esterik, J.C.J.; Sales, L.B.; Dollé, M.E.T.; Håkansson, H.; Herlin, M.; Legler, J.; van der Ven, L.T.M. Programming of Metabolic Effects in C57BL/6JxFVB Mice by in Utero and Lactational Exposure to Perfluorooctanoic Acid. *Arch. Toxicol.* **2016**, *90*, 701–715. [[CrossRef](#)]
46. Xu, J.; Shimpi, P.; Armstrong, L.; Salter, D.; Slitt, A.L. PFOS Induces Adipogenesis and Glucose Uptake in Association with Activation of Nrf2 Signaling Pathway. *Toxicol. Appl. Pharmacol.* **2016**, *290*, 21–30. [[CrossRef](#)]
47. Du, G.; Sun, J.; Zhang, Y. Perfluorooctanoic Acid Impaired Glucose Homeostasis through Affecting Adipose AKT Pathway. *Cytotechnology* **2018**, *70*, 479–487. [[CrossRef](#)]
48. Gao, Y.; Guo, X.; Wang, S.; Chen, F.; Ren, X.; Xiao, H.; Wang, L. Perfluorooctane Sulfonate Enhances MRNA Expression of PPAR γ and Ap2 in Human Mesenchymal Stem Cells Monitored by Long-Retained Intracellular Nanosensor. *Environ. Pollut.* **2020**, *263*, 114571. [[CrossRef](#)]
49. Lu, Z.; Yifan, P.; Hui, Q.; Jiangyu, Z.; Wei, L. Interference of Perfluorooctane Sulfonate (PFOS) on PPARs Subtypes and Differentiation Potential in Human Bone Marrow Mesenchymal Stem Cells. *Asian J. Ecotoxicol.* **2021**, *2*, 151–157. [[CrossRef](#)]
50. Van den Dungen, M.W.; Murk, A.J.; Kok, D.E.; Steegenga, W.T. Persistent Organic Pollutants Alter DNA Methylation during Human Adipocyte Differentiation. *Toxicol. In Vitro* **2017**, *40*, 79–87. [[CrossRef](#)]
51. Qi, W.; Clark, J.M.; Timme-Laragy, A.R.; Park, Y. Perfluorobutanesulfonic Acid (PFBS) Potentiates Adipogenesis of 3T3-L1 Adipocytes. *Food Chem. Toxicol.* **2018**, *120*, 340–345. [[CrossRef](#)]
52. Li, C.H.; Ren, X.M.; Ruan, T.; Cao, L.Y.; Xin, Y.; Guo, L.H.; Jiang, G. Chlorinated Polyfluorinated Ether Sulfonates Exhibit Higher Activity toward Peroxisome Proliferator-Activated Receptors Signaling Pathways than Perfluorooctanesulfonate. *Environ. Sci. Technol.* **2018**, *52*, 3232–3239. [[CrossRef](#)]
53. Xie, Y.; Berntsen, H.F.; Zimmer, K.E.; Ropstad, E.; Verhaegen, S.; Connolly, L. Lipogenic Potency of Individual Perfluorinated Alkyl Acids (PFAAs) and Persistent Organic Pollutant (POP) Mixtures at Human Blood-Based Exposure Levels on Adipogenesis in 3T3-L1 Cells. *Expo. Health* **2021**. [[CrossRef](#)]

54. Ćurčić, M.; Janković, S.; Jačević, V.; Stanković, S.; Vučinić, S.; Durgo, K.; Bulat, Z.; Antonijević, B. Combined Effects of Cadmium and Decabrominated Diphenyl Ether on Thyroid Hormones in Rats. *Arhiv za Higijenu Rada i Toksikologiju* **2012**, *63*, 255–262. [[CrossRef](#)]
55. Yang, C.; Kong, A.P.S.; Cai, Z.; Chung, A.C.K. Persistent Organic Pollutants as Risk Factors for Obesity and Diabetes. *Curr. Diabetes Rep.* **2017**, *17*, 132. [[CrossRef](#)]
56. Llm, J.S.; Lee, D.H.; Jacobs, D.R. Association of Brominated Flame Retardants with Diabetes and Metabolic Syndrome in the U.S. Population, 2003–2004. *Diabetes Care* **2008**, *31*, 1802–1807. [[CrossRef](#)]
57. Helaleh, M.; Diboun, I.; Al-Tamimi, N.; Al-Sulaiti, H.; Al-Emadi, M.; Madani, A.; Mazloum, N.A.; Latiff, A.; Elrayess, M.A. Association of Polybrominated Diphenyl Ethers in Two Fat Compartments with Increased Risk of Insulin Resistance in Obese Individuals. *Chemosphere* **2018**, *209*, 268–276. [[CrossRef](#)]
58. Erkin-Cakmak, A.; Harley, K.G.; Chevrier, J.; Bradman, A.; Kogut, K.; Huen, K.; Eskenazi, B. In Utero and Childhood Polybrominated Diphenyl Ether Exposures and Body Mass at Age 7 Years: The CHAMACOS Study. *Environ. Health Perspect.* **2015**, *123*, 636–642. [[CrossRef](#)]
59. Pereira-Fernandes, A.; Dirinck, E.; Dirtu, A.C.; Malarvannan, G.; Covaci, A.; Van Gaal, L.; Vanparys, C.; Jorens, P.G.; Blust, R. Expression of Obesity Markers and Persistent Organic Pollutants Levels in Adipose Tissue of Obese Patients: Reinforcing the Obesogen Hypothesis? *PLoS ONE* **2014**, *9*, 84816. [[CrossRef](#)]
60. Tung, E.W.Y.; Boudreau, A.; Wade, M.G.; Atlas, E. Induction of Adipocyte Differentiation by Polybrominated Diphenyl Ethers (PBDEs) in 3T3-L1 Cells. *PLoS ONE* **2014**, *9*, e94583. [[CrossRef](#)]
61. Armstrong, L.E.; Akinbo, S.; Slitt, A.L. 2,2',4,4',5-Pentabromodiphenyl Ether Induces Lipid Accumulation throughout Differentiation in 3T3-L1 and Human Preadipocytes in Vitro. *J. Biochem. Mol. Toxicol.* **2020**, *34*, e22485. [[CrossRef](#)]
62. Wen, Q.; Xie, X.; Zhao, C.; Ren, Q.; Zhang, X.; Wei, D.; Emanuelli, B.; Du, Y. The Brominated Flame Retardant PBDE 99 Promotes Adipogenesis via Regulating Mitotic Clonal Expansion and PPAR γ Expression. *Sci. Total Environ.* **2019**, *670*, 67–77. [[CrossRef](#)]
63. Kamstra, J.H.; Hruba, E.; Blumberg, B.; Janesick, A.; Mandrup, S.; Hamers, T.; Legler, J. Transcriptional and Epigenetic Mechanisms Underlying Enhanced in Vitro Adipocyte Differentiation by the Brominated Flame Retardant BDE-47. *Environ. Sci. Technol.* **2014**, *48*, 4110–4119. [[CrossRef](#)]
64. Yang, C.; Wong, C.M.; Wei, J.; Chung, A.C.K.; Cai, Z. The Brominated Flame Retardant BDE 47 Upregulates Purine Metabolism and Mitochondrial Respiration to Promote Adipocyte Differentiation. *Sci. Total Environ.* **2018**, *644*, 1312–1322. [[CrossRef](#)]
65. Yang, C.; Zhu, L.; Kang, Q.; Lee, H.K.; Li, D.; Chung, A.C.K.; Cai, Z. Chronic Exposure to Tetrabromodiphenyl Ether (BDE-47) Aggravates Hepatic Steatosis and Liver Fibrosis in Diet-Induced Obese Mice. *J. Hazard. Mater.* **2019**, *378*, 120766. [[CrossRef](#)]
66. Yang, C.; Wei, J.; Cao, G.; Cai, Z. Lipid Metabolism Dysfunction and Toxicity of BDE-47 Exposure in White Adipose Tissue Revealed by the Integration of Lipidomics and Metabolomics. *Sci. Total Environ.* **2022**, *806*, 150350. [[CrossRef](#)]
67. Allgood, E.L.; Carey, G. The Effect of Diet and Polybrominated Diphenyl Ether Exposure on Adipocyte and Whole Body Metabolism in Male Wistar Rats. *FASEB J.* **2009**, *23*, 505.3. [[CrossRef](#)]
68. Scoville, D.K.; Li, C.Y.; Wang, D.; Dempsey, J.L.; Raftery, D.; Mani, S.; Gu, H.; Cui, J.Y. Polybrominated Diphenyl Ethers and Gut Microbiome Modulate Metabolic Syndrome-Related Aqueous Metabolites in Mice. *Drug Metab. Dispos.* **2019**, *47*, 928–940. [[CrossRef](#)]
69. Cano-Sancho, G.; Salmon, A.G.; La Merrill, M.A. Association between Exposure to p,p'-DDT and Its Metabolite p,p'-DDE with Obesity: Integrated Systematic Review and Meta-Analysis. *Environ. Health Perspect.* **2017**, *125*, 096002. [[CrossRef](#)]
70. González-Casanova, J.E.; Pertuz-Cruz, S.L.; Caicedo-Ortega, N.H.; Rojas-Gomez, D.M. Adipogenesis Regulation and Endocrine Disruptors: Emerging Insights in Obesity. *BioMed Res. Int.* **2020**, *2020*, 7453786. [[CrossRef](#)]
71. Arrebola, J.P.; Ocaña-Riola, R.; Arrebola-Moreno, A.L.; Fernández-Rodríguez, M.; Martín-Olmedo, P.; Fernández, M.F.; Olea, N. Associations of Accumulated Exposure to Persistent Organic Pollutants with Serum Lipids and Obesity in an Adult Cohort from Southern Spain. *Environ. Pollut.* **2014**, *195*, 9–15. [[CrossRef](#)]
72. Dhooge, W.; Den Hond, E.; Koppen, G.; Bruckers, L.; Nelen, V.; Van De Mierop, E.; Bilau, M.; Croes, K.; Baeyens, W.; Schoeters, G.; et al. Internal Exposure to Pollutants and Body Size in Flemish Adolescents and Adults: Associations and Dose-Response Relationships. *Environ. Int.* **2010**, *36*, 330–337. [[CrossRef](#)]
73. Heggeseth, B.; Harley, K.; Warner, M.; Jewell, N.; Eskenazi, B. Detecting Associations between Early-Life DDT Exposures and Childhood Growth Patterns: A Novel Statistical Approach. *PLoS ONE* **2015**, *10*, e0131443. [[CrossRef](#)]
74. Warner, M.; Schall, R.A.; Harley, K.G.; Bradman, A.; Barr, D.; Eskenazi, B. In Utero DDT and DDE Exposure and Obesity Status of 7-Year-Old Mexican-American Children in the CHAMACOS Cohort. *Environ. Health Perspect.* **2013**, *121*, 631–636. [[CrossRef](#)]
75. Warner, M.; Wesselink, A.; Harley, K.G.; Bradman, A.; Kogut, K.; Eskenazi, B. Prenatal Exposure to Dichlorodiphenyl-trichloroethane and Obesity at 9 Years of Age in the CHAMACOS Study Cohort. *Am. J. Epidemiol.* **2014**, *179*, 1312–1322. [[CrossRef](#)]
76. Warner, M.; Ye, M.; Harley, K.; Kogut, K.; Bradman, A.; Eskenazi, B. Prenatal DDT Exposure and Child Adiposity at Age 12: The CHAMACOS Study. *Environ. Res.* **2017**, *159*, 606–612. [[CrossRef](#)]
77. La Merrill, M.A.; Krigbaum, N.Y.; Cirillo, P.M.; Cohn, B.A. Association between Maternal Exposure to the Pesticide Dichlorodiphenyltrichloroethane (DDT) and Risk of Obesity in Middle Age. *Int. J. Obes.* **2020**, *44*, 1723–1732. [[CrossRef](#)]

78. Verhulst, S.L.; Nelen, V.; Hond, E.D.; Koppen, G.; Beunckens, C.; Vael, C.; Schoeters, G.; Desager, K. Intrauterine Exposure to Environmental Pollutants and Body Mass Index during the First 3 Years of Life. *Environ. Health Perspect.* **2009**, *117*, 122–126. [[CrossRef](#)]
79. Henríquez-Hernández, L.A.; Luzardo, O.P.; Valerón, P.F.; Zumbado, M.; Serra-Majem, L.; Camacho, M.; González-Antuña, A.; Boada, L.D. Persistent Organic Pollutants and Risk of Diabetes and Obesity on Healthy Adults: Results from a Cross-Sectional Study in Spain. *Sci. Total Environ.* **2017**, *607–608*, 1096–1102. [[CrossRef](#)]
80. Tawar, N.; Banerjee, B.; Mishra, B.; Sharma, T.; Tyagi, S.; Madhu, S.; Agarwal, V.; Gupta, S. Adipose Tissue Levels of DDT as Risk Factor for Obesity and Type 2 Diabetes Mellitus. *Indian J. Endocrinol. Metab.* **2021**, *25*, 160–165. [[CrossRef](#)]
81. Kim, J.; Sun, Q.; Yue, Y.; Yoon, K.S.; Whang, K.-Y.; Marshall Clark, J.; Park, Y. 4,4'-Dichlorodiphenyltrichloroethane (DDT) and 4,4'-Dichlorodiphenyldichloroethylene (DDE) Promote Adipogenesis in 3T3-L1 Adipocyte Cell Culture. *Pestic. Biochem. Physiol.* **2016**, *131*, 40–45. [[CrossRef](#)]
82. Moreno-Aliaga, M.J.; Matsumura, F. Effects of 1,1,1-Trichloro-2,2-Bis(p-Chlorophenyl)-Ethane (p,p'-DDT) on 3T3-L1 and 3T3-F442A Adipocyte Differentiation. *Biochem. Pharmacol.* **2002**, *63*, 997–1007. [[CrossRef](#)]
83. Pesta, M.; Cedikova, M.; Dvorak, P.; Dvorakova, J.; Kulda, V.; Srbecka, K.; Muller, L.; Bouchalova, V.; Kralickova, M.; Babuska, V.; et al. Trends in Gene Expression Changes during Adipogenesis in Human Adipose Derived Mesenchymal Stem Cells under Dichlorodiphenyldichloroethylene Exposure. *Mol. Cell. Toxicol.* **2018**, *14*, 369–379. [[CrossRef](#)]
84. Strong, A.L.; Shi, Z.; Strong, M.J.; Miller, D.F.B.; Rusch, D.B.; Buechlein, A.M.; Flemington, E.K.; McLachlan, J.A.; Nephew, K.P.; Burrow, M.E.; et al. Effects of the Endocrine-Disrupting Chemical DDT on Self-Renewal and Differentiation of Human Mesenchymal Stem Cells. *Environ. Health Perspect.* **2015**, *123*, 42–48. [[CrossRef](#)] [[PubMed](#)]
85. Mangum, L.H.; Howell, G.E.; Chambers, J.E. Exposure to p,p'-DDE Enhances Differentiation of 3T3-L1 Preadipocytes in a Model of Sub-Optimal Differentiation. *Toxicol. Lett.* **2015**, *238*, 65–71. [[CrossRef](#)]
86. King, S.E.; Nilsson, E.; Beck, D.; Skinner, M.K. Adipocyte Epigenetic Alterations and Potential Therapeutic Targets in Trans-generationally Inherited Lean and Obese Phenotypes Following Ancestral Exposures. *Adipocyte* **2019**, *8*, 362–378. [[CrossRef](#)] [[PubMed](#)]
87. Howell, G.; Mangum, L. Exposure to Bioaccumulative Organochlorine Compounds Alters Adipogenesis, Fatty Acid Uptake, and Adipokine Production in NIH3T3-L1 Cells. *Toxicol. In Vitro* **2011**, *25*, 394–402. [[CrossRef](#)] [[PubMed](#)]
88. La Merrill, M.; Karey, E.; Moshier, E.; Lindtner, C.; La Frano, M.R.; Newman, J.W.; Buettner, C. Perinatal Exposure of Mice to the Pesticide DDT Impairs Energy Expenditure and Metabolism in Adult Female Offspring. *PLoS ONE* **2014**, *9*, 103337. [[CrossRef](#)] [[PubMed](#)]
89. Ghosh, S.; Murinova, L.; Trnovec, T.; Loffredo, C.; Washington, K.; Mitra, P.; Dutta, S. Biomarkers Linking PCB Exposure and Obesity. *Curr. Pharm. Biotechnol.* **2014**, *15*, 1058–1068. [[CrossRef](#)]
90. Buha Djordjevic, A.; Antonijevic, E.; Curcic, M.; Milovanovic, V.; Antonijevic, B. Endocrine-Disrupting Mechanisms of Polychlorinated Biphenyls. *Curr. Opin. Toxicol.* **2020**, *19*, 42–49. [[CrossRef](#)]
91. Buha, A.; Antonijević, B.; Bulat, Z.; Jačević, V.; Milovanović, V.; Matović, V. The Impact of Prolonged Cadmium Exposure and Co-Exposure with Polychlorinated Biphenyls on Thyroid Function in Rats. *Toxicol. Lett.* **2013**, *221*, 83–90. [[CrossRef](#)]
92. Domazet, S.L.; Grøntved, A.; Jensen, T.K.; Wedderkopp, N.; Andersen, L.B. Higher Circulating Plasma Polychlorinated Biphenyls (PCBs) in Fit and Lean Children: The European Youth Heart Study. *Environ. Int.* **2020**, *136*, 105481. [[CrossRef](#)]
93. Warner, M.; Rauch, S.; Coker, E.S.; Harley, K.; Kogut, K.; Sjödin, A.; Eskenazi, B. Obesity in Relation to Serum Persistent Organic Pollutant Concentrations in CHAMACOS Women. *Environ. Epidemiol.* **2018**, *2*, e032. [[CrossRef](#)]
94. Lee, D.H.; Lee, I.K.; Porta, M.; Steffes, M.; Jacobs, D.R. Relationship between Serum Concentrations of Persistent Organic Pollutants and the Prevalence of Metabolic Syndrome among Non-Diabetic Adults: Results from the National Health and Nutrition Examination Survey 1999–2002. *Diabetologia* **2007**, *50*, 1841–1851. [[CrossRef](#)]
95. Donat-Vargas, C.; Gea, A.; Sayon-Orea, C.; Carlos, S.; Martinez-Gonzalez, M.A.; Bes-Rastrollo, M. Association between Dietary Intakes of PCBs and the Risk of Obesity: The SUN Project. *J. Epidemiol. Community Health* **2014**, *68*, 834–841. [[CrossRef](#)]
96. Lee, D.H.; Lind, L.; Jacobs, D.R.; Salihovic, S.; van Bavel, B.; Lind, P.M. Associations of Persistent Organic Pollutants with Abdominal Obesity in the Elderly: The Prospective Investigation of the Vasculature in Uppsala Seniors (PIVUS) Study. *Environ. Int.* **2012**, *40*, 170–178. [[CrossRef](#)]
97. Valvi, D.; Mendez, M.A.; Martinez, D.; Grimalt, J.O.; Torrent, M.; Sunyer, J.; Vrijheid, M. Prenatal Concentrations of Polychlorinated Biphenyls, DDE, and DDT and Overweight in Children: A Prospective Birth Cohort Study. *Environ. Health Perspect.* **2012**, *120*, 451–457. [[CrossRef](#)] [[PubMed](#)]
98. Lee, D.H.; Steffes, M.W.; Sjödin, A.; Jones, R.S.; Needham, L.L.; Jacobs, D.R. Low Dose Organochlorine Pesticides and Polychlorinated Biphenyls Predict Obesity, Dyslipidemia, and Insulin Resistance among People Free of Diabetes. *PLoS ONE* **2011**, *6*, e15977. [[CrossRef](#)] [[PubMed](#)]
99. Hertz-Picciotto, I.; Charles, M.J.; James, R.A.; Keller, J.A.; Willman, E.; Teplin, S. In Utero Polychlorinated Biphenyl Exposures in Relation to Fetal and Early Childhood Growth. *Epidemiology* **2005**, *16*, 648–656. [[CrossRef](#)]
100. Hassan, W.; Ahmed, H.; Murtaza, G.; Umar, M.I.; Iqbal, F.M. *Role of Polychlorinated Biphenyls as EDCs in Metabolic Disorders*; Springer: Cham, Switzerland, 2021; pp. 161–174.

101. Bourez, S.; Le Lay, S.; Van den Daelen, C.; Louis, C.; Larondelle, Y.; Thomé, J.-P.; Schneider, Y.-J.; Dugail, I.; Debier, C. Accumulation of Polychlorinated Biphenyls in Adipocytes: Selective Targeting to Lipid Droplets and Role of Caveolin-1. *PLoS ONE* **2012**, *7*, e31834. [[CrossRef](#)]
102. Bourez, S.; Van den Daelen, C.; Le Lay, S.; Poupaert, J.; Larondelle, Y.; Thomé, J.-P.; Schneider, Y.-J.; Dugail, I.; Debier, C. The Dynamics of Accumulation of PCBs in Cultured Adipocytes Vary with the Cell Lipid Content and the Lipophilicity of the Congener. *Toxicol. Lett.* **2013**, *216*, 40–46. [[CrossRef](#)] [[PubMed](#)]
103. Arsenescu, V.; Arsenescu, R.I.; King, V.; Swanson, H.; Cassis, L.A. Polychlorinated Biphenyl-77 Induces Adipocyte Differentiation and Proinflammatory Adipokines and Promotes Obesity and Atherosclerosis. *Environ. Health Perspect.* **2008**, *116*, 761–768. [[CrossRef](#)] [[PubMed](#)]
104. Taxvig, C.; Dreisig, K.; Boberg, J.; Nellemann, C.; Schelde, A.B.; Pedersen, D.; Boergesen, M.; Mandrup, S.; Vinggaard, A.M. Differential Effects of Environmental Chemicals and Food Contaminants on Adipogenesis, Biomarker Release and PPAR γ Activation. *Mol. Cell. Endocrinol.* **2012**, *361*, 106–115. [[CrossRef](#)] [[PubMed](#)]
105. Su, S.; Wu, G.; Cheng, X.; Fan, J.; Peng, J.; Su, H.; Xu, Z.; Cao, M.; Long, Z.; Hao, Y.; et al. Oleanolic Acid Attenuates PCBs-Induced Adiposity and Insulin Resistance via HNF1b-Mediated Regulation of Redox and PPAR γ Signaling. *Free Radic. Biol. Med.* **2018**, *124*, 122–134. [[CrossRef](#)] [[PubMed](#)]
106. Yu, C.; Wen, Q.; Ren, Q.; Du, Y.; Xie, X. Polychlorinated Biphenyl Congener 180 (PCB 180) Regulates Mitotic Clonal Expansion and Enhances Adipogenesis through Modulation of C/EBP β SUMOylation in Preadipocytes. *Food Chem. Toxicol.* **2021**, *152*, 112205. [[CrossRef](#)]
107. Baker, N.A.; Karounos, M.; English, V.; Fang, J.; Wei, Y.; Stromberg, A.; Sunkara, M.; Morris, A.J.; Swanson, H.I.; Cassis, L.A. Coplanar Polychlorinated Biphenyls Impair Glucose Homeostasis in Lean C57BL/6 Mice and Mitigate Beneficial Effects of Weight Loss on Glucose Homeostasis in Obese Mice. *Environ. Health Perspect.* **2013**, *121*, 105–110. [[CrossRef](#)]
108. Gourronc, F.A.; Robertson, L.W.; Klingelutz, A.J. A Delayed Proinflammatory Response of Human Preadipocytes to PCB126 Is Dependent on the Aryl Hydrocarbon Receptor. *Environ. Sci. Pollut. Res.* **2018**, *25*, 16481–16492. [[CrossRef](#)]
109. Wu, H.; Yu, W.; Meng, F.; Mi, J.; Peng, J.; Liu, J.; Zhang, X.; Hai, C.; Wang, X. Polychlorinated Biphenyls-153 Induces Metabolic Dysfunction through Activation of ROS/NF-KB Signaling via Downregulation of HNF1b. *Redox Biol.* **2017**, *12*, 300–310. [[CrossRef](#)]
110. Kim, Y.A.; Kim, H.Y.; Oh, Y.J.; Kwon, W.Y.; Lee, M.H.; Bae, J.Y.; Woo, M.S.; Kim, J.M.; Yoo, Y.H. Polychlorinated Biphenyl 138 Exposure-Mediated Lipid Droplet Enlargement Endows Adipocytes with Resistance to TNF- α -Induced Cell Death. *Toxicol. Lett.* **2018**, *292*, 55–62. [[CrossRef](#)]
111. Baker, N.A.; English, V.; Sunkara, M.; Morris, A.J.; Pearson, K.J.; Cassis, L.A. Resveratrol Protects against Polychlorinated Biphenyl-Mediated Impairment of Glucose Homeostasis in Adipocytes. *J. Nutr. Biochem.* **2013**, *24*, 2168–2174. [[CrossRef](#)] [[PubMed](#)]
112. Gourronc, F.A.; Perdew, G.H.; Robertson, L.W.; Klingelutz, A.J. PCB126 Blocks the Thermogenic Beiging Response of Adipocytes. *Environ. Sci. Pollut. Res.* **2020**, *27*, 8897–8904. [[CrossRef](#)]
113. Kamata, R.; Nakajima, D.; Shiraishi, F. Measurement of the Agonistic Activities of Monohydroxylated Polychlorinated Biphenyls at the Retinoid X and Retinoic Acid Receptors Using Recombinant Yeast Cells. *Toxicol. In Vitro* **2019**, *57*, 9–17. [[CrossRef](#)]
114. Wahlang, B.; Falkner, K.C.; Gregory, B.; Ansert, D.; Young, D.; Conklin, D.J.; Bhatnagar, A.; McClain, C.J.; Cave, M. Polychlorinated Biphenyl 153 Is a Diet-Dependent Obesogen That Worsens Nonalcoholic Fatty Liver Disease in Male C57BL6/J Mice. *J. Nutr. Biochem.* **2013**, *24*, 1587–1595. [[CrossRef](#)] [[PubMed](#)]
115. Li, D.L.; Huang, Y.J.; Gao, S.; Chen, L.Q.; Zhang, M.L.; Du, Z.Y. Sex-Specific Alterations of Lipid Metabolism in Zebrafish Exposed to Polychlorinated Biphenyls. *Chemosphere* **2019**, *221*, 768–777. [[CrossRef](#)] [[PubMed](#)]
116. Scinicariello, F.; Buser, M.C. Urinary Polycyclic Aromatic Hydrocarbons and Childhood Obesity: NHANES (2001–2006). *Environ. Health Perspect.* **2014**, *122*, 299–303. [[CrossRef](#)] [[PubMed](#)]
117. Poursafa, P.; Dadvand, P.; Amin, M.M.; Hajizadeh, Y.; Ebrahimpour, K.; Mansourian, M.; Pourzamani, H.; Sunyer, J.; Kelishadi, R. Association of Polycyclic Aromatic Hydrocarbons with Cardiometabolic Risk Factors and Obesity in Children. *Environ. Int.* **2018**, *118*, 203–210. [[CrossRef](#)]
118. Kim, H.W.; Kam, S.; Lee, D.H. Synergistic Interaction between Polycyclic Aromatic Hydrocarbons and Environmental Tobacco Smoke on the Risk of Obesity in Children and Adolescents: The U.S. National Health and Nutrition Examination Survey 2003–2008. *Environ. Res.* **2014**, *135*, 354–360. [[CrossRef](#)]
119. Yin, W.; Hou, J.; Xu, T.; Cheng, J.; Li, P.; Wang, L.; Zhang, Y.; Wang, X.; Hu, C.; Huang, C.; et al. Obesity Mediated the Association of Exposure to Polycyclic Aromatic Hydrocarbon with Risk of Cardiovascular Events. *Sci. Total Environ.* **2018**, *616–617*, 841–854. [[CrossRef](#)]
120. Hou, J.; Sun, H.; Huang, X.; Zhou, Y.; Zhang, Y.; Yin, W.; Xu, T.; Cheng, J.; Chen, W.; Yuan, J. Exposure to Polycyclic Aromatic Hydrocarbons and Central Obesity Enhanced Risk for Diabetes among Individuals with Poor Lung Function. *Chemosphere* **2017**, *185*, 1136–1143. [[CrossRef](#)]
121. Li, K.; Yin, R.; Wang, Y.; Zhao, D. Associations between Exposure to Polycyclic Aromatic Hydrocarbons and Metabolic Syndrome in U.S. Adolescents: Cross-Sectional Results from the National Health and Nutrition Examination Survey (2003–2016) Data. *Environ. Res.* **2021**, *202*, 111747. [[CrossRef](#)]

122. May, P.; Bremond, P.; Sauzet, C.; Piccerelle, P.; Grimaldi, F.; Champion, S.; Villard, P.-H. In Vitro Cocktail Effects of PCB-DL (PCB118) and Bulky PCB (PCB153) with BaP on Adipogenesis and on Expression of Genes Involved in the Establishment of a Pro-Inflammatory State. *Int. J. Mol. Sci.* **2018**, *19*, 841. [[CrossRef](#)]
123. Podechard, N.; Fardel, O.; Corolleur, M.; Bernard, M.; Lecureur, V. Inhibition of Human Mesenchymal Stem Cell-Derived Adipogenesis by the Environmental Contaminant Benzo(a)Pyrene. *Toxicol. In Vitro* **2009**, *23*, 1139–1144. [[CrossRef](#)]
124. Rathore, K.; Cekanova, M. Effects of Environmental Carcinogen Benzo(a)Pyrene on Canine Adipose-Derived Mesenchymal Stem Cells. *Res. Vet. Sci.* **2015**, *103*, 34–43. [[CrossRef](#)] [[PubMed](#)]
125. Yan, Z.; Zhang, H.; Maher, C.; Arteaga-Solis, E.; Champagne, F.A.; Wu, L.; McDonald, J.D.; Yan, B.; Schwartz, G.J.; Miller, R.L. Prenatal Polycyclic Aromatic Hydrocarbon, Adiposity, Peroxisome Proliferator-Activated Receptor (PPAR) γ Methylation in Offspring, Grand-Offspring Mice. *PLoS ONE* **2014**, *9*, e110706. [[CrossRef](#)] [[PubMed](#)]
126. Ortiz, L.; Nakamura, B.; Li, X.; Blumberg, B.; Luderer, U. In Utero Exposure to Benzo[a]Pyrene Increases Adiposity and Causes Hepatic Steatosis in Female Mice, and Glutathione Deficiency Is Protective. *Toxicol. Lett.* **2013**, *223*, 260–267. [[CrossRef](#)]
127. Gato, W.E.; Hunter, D.A.; Whitby, S.L.; Mays, C.A.; Yau, W. Investigating Susceptibility to Diabetes Using Features of the Adipose Tissue in Response to In Utero Polycyclic Aromatic Hydrocarbons Exposure. *Diabetes Metab. J.* **2016**, *40*, 494. [[CrossRef](#)]
128. Irigaray, P.; Lacomme, S.; Mejean, L.; Belpomme, D. Ex Vivo Study of Incorporation into Adipocytes and Lipolysis-Inhibition Effect of Polycyclic Aromatic Hydrocarbons. *Toxicol. Lett.* **2009**, *187*, 35–39. [[CrossRef](#)] [[PubMed](#)]
129. Irigaray, P.; Ogier, V.; Jacquenet, S.; Notet, V.; Sibille, P.; Mejean, L.; Bihain, B.E.; Yen, F.T. Benzo[a]Pyrene Impairs Beta-Adrenergic Stimulation of Adipose Tissue Lipolysis and Causes Weight Gain in Mice. A Novel Molecular Mechanism of Toxicity for a Common Food Pollutant. *FEBS J.* **2006**, *273*, 1362–1372. [[CrossRef](#)]
130. Guo, J.; Huang, J.; Wang, Q.; Fang, L.; Zhang, S.; Li, B.; Lv, L.; Chen, M.; Wang, C. Maternal Exposure to Phenanthrene during Gestation Disturbs Glucose Homeostasis in Adult Mouse Offspring. *Chemosphere* **2021**, *270*, 128635. [[CrossRef](#)]
131. Kim, Y.H.; Lee, Y.S.; Lee, D.H.; Kim, D.S. Polycyclic Aromatic Hydrocarbons Are Associated with Insulin Receptor Substrate 2 Methylation in Adipose Tissues of Korean Women. *Environ. Res.* **2016**, *150*, 47–51. [[CrossRef](#)]
132. Hsieh, C.C.; Peng, S.H.; Chou, M.J. Obesity Enhances Carcinogen 7, 12-Dimethylbenz [a] Anthracene -Induced Tumorigenesis in Vitro and in Vivo. *Food Chem. Toxicol.* **2017**, *110*, 156–164. [[CrossRef](#)]
133. Warner, M.; Mocarelli, P.; Brambilla, P.; Wesselink, A.; Samuels, S.; Signorini, S.; Eskenazi, B. Diabetes, Metabolic Syndrome, and Obesity in Relation to Serum Dioxin Concentrations: The Seveso Women’s Health Study. *Environ. Health Perspect.* **2013**, *121*, 906–911. [[CrossRef](#)]
134. Chang, J.-W.; Chen, H.-L.; Su, H.-J.; Lee, C.-C. Abdominal Obesity and Insulin Resistance in People Exposed to Moderate-to-High Levels of Dioxin. *PLoS ONE* **2016**, *11*, e0145818. [[CrossRef](#)] [[PubMed](#)]
135. Uemura, H.; Arisawa, K.; Hiyoshi, M.; Kitayama, A.; Takami, H.; Sewachika, F.; Dakeshita, S.; Nii, K.; Satoh, H.; Sumiyoshi, Y.; et al. Prevalence of Metabolic Syndrome Associated with Body Burden Levels of Dioxin and Related Compounds among Japan’s General Population. *Environ. Health Perspect.* **2009**, *117*, 568–573. [[CrossRef](#)] [[PubMed](#)]
136. Chang, J.W.; Chen, H.L.; Su, H.J.; Liao, P.C.; Guo, H.R.; Lee, C.C. Dioxin Exposure and Insulin Resistance in Taiwanese Living near a Highly Contaminated Area. *Epidemiology* **2010**, *21*, 56–61. [[CrossRef](#)] [[PubMed](#)]
137. La Merrill, M.; Birnbaum, L.S. Childhood Obesity and Environmental Chemicals. *Mt. Sinai J. Med. J. Transl. Personal. Med.* **2011**, *78*, 22–48. [[CrossRef](#)] [[PubMed](#)]
138. Girer, N.G.; Tomlinson, C.R.; Elferink, C.J. The Aryl Hydrocarbon Receptor in Energy Balance: The Road from Dioxin-Induced Wasting Syndrome to Combating Obesity with AHR Ligands. *Int. J. Mol. Sci.* **2021**, *22*, 49. [[CrossRef](#)]
139. Hsu, H.F.; Tsou, T.C.; Chao, H.R.; Kuo, Y.T.; Tsai, F.Y.; Yeh, S.C. Effects of 2,3,7,8-Tetrachlorodibenzo-p-Dioxin on Adipogenic Differentiation and Insulin-Induced Glucose Uptake in 3T3-L1 Cells. *J. Hazard. Mater.* **2010**, *182*, 649–655. [[CrossRef](#)]
140. Hanlon, P.R.; Cimafranca, M.A.; Liu, X.; Cho, Y.C.; Jefcoate, C.R. Microarray Analysis of Early Adipogenesis in C3H10T1/2 Cells: Cooperative Inhibitory Effects of Growth Factors and 2,3,7,8-Tetrachlorodibenzo-p-Dioxin. *Toxicol. Appl. Pharmacol.* **2005**, *207*, 39–58. [[CrossRef](#)]
141. Choi, E.M.; Suh, K.S.; Jung, W.W.; Park, S.Y.; Chin, S.O.; Rhee, S.Y.; Kim Pak, Y.; Chon, S. Glabridin Attenuates Antiadipogenic Activity Induced by 2,3,7,8-Tetrachlorodibenzo-p-Dioxin in Murine 3T3-L1 Adipocytes. *J. Appl. Toxicol.* **2018**, *38*, 1426–1436. [[CrossRef](#)]
142. Hanlon, P.R.; Ganem, L.G.; Cho, Y.C.; Yamamoto, M.; Jefcoate, C.R. AhR- and ERK-Dependent Pathways Function Synergistically to Mediate 2,3,7,8-Tetrachlorodibenzo-p-Dioxin Suppression of Peroxisome Proliferator-Activated Receptor- γ 1 Expression and Subsequent Adipocyte Differentiation. *Toxicol. Appl. Pharmacol.* **2003**, *189*, 11–27. [[CrossRef](#)]
143. Liu, P.C.C.; Phillips, M.A.; Matsumura, F. Alteration by 2,3,7,8-Tetrachlorodibenzo-p-Dioxin of CCAAT/Enhancer Binding Protein Correlates with Suppression of Adipocyte Differentiation in 3T3-L1 Cells. *Mol. Pharmacol.* **1996**, *49*, 989–997.
144. Nishiumi, S.; Yabushita, Y.; Furuyashiki, T.; Fukuda, I.; Ashida, H. Involvement of SREBPs in 2,3,7,8-Tetrachlorodibenzo-p-Dioxin-Induced Disruption of Lipid Metabolism in Male Guinea Pig. *Toxicol. Appl. Pharmacol.* **2008**, *229*, 281–289. [[CrossRef](#)]
145. Brewster, D.W. Fumio Matsumura Reduction of Adipose Tissue Lipoprotein Lipase Activity as a Result of in Vivo Administration of 2,3,7,8-Tetrachlorodibenzo-p-Dioxin to the Guinea Pig. *Biochem. Pharmacol.* **1988**, *37*, 2247–2253. [[CrossRef](#)]
146. Brulport, A.; Le Corre, L.; Chagnon, M.C. Chronic Exposure of 2,3,7,8-Tetrachlorodibenzo-p-Dioxin (TCDD) Induces an Obesogenic Effect in C57BL/6J Mice Fed a High Fat Diet. *Toxicology* **2017**, *390*, 43–52. [[CrossRef](#)]

147. Duval, C.; Teixeira-Clerc, F.; Leblanc, A.F.; Touch, S.; Emond, C.; Guerre-Millo, M.; Lotersztajn, S.; Barouki, R.; Aggerbeck, M.; Coumou, X. Chronic Exposure to Low Doses of Dioxin Promotes Liver Fibrosis Development in the C57BL/6J Diet-Induced Obesity Mouse Model. *Environ. Health Perspect.* **2017**, *125*, 428–436. [[CrossRef](#)] [[PubMed](#)]
148. Hoyeck, M.P.; Merhi, R.C.; Blair, H.L.; Spencer, C.D.; Payant, M.A.; Martin Alfonso, D.I.; Zhang, M.; Matteo, G.; Chee, M.J.; Bruin, J.E. Female Mice Exposed to Low Doses of Dioxin during Pregnancy and Lactation Have Increased Susceptibility to Diet-Induced Obesity and Diabetes. *Mol. Metab.* **2020**, *42*, 101104. [[CrossRef](#)]
149. Matteo, G.; Hoyeck, M.P.; Blair, H.L.; Zebarth, J.; Rick, K.R.C.; Williams, A.; Gagné, R.; Buick, J.K.; Yauk, C.L.; Bruin, J.E. Prolonged Low-Dose Dioxin Exposure Impairs Metabolic Adaptability to High-Fat Diet Feeding in Female but Not Male Mice. *Endocrinology* **2021**, *162*, bqab050. [[CrossRef](#)]
150. Baralić, K.; Živancevic, K.; Bozic, D.; Jennen, D.; Buha Djordjevic, A.; Antonijevic Miljakovic, E.; Đukic-Cosic, D. Potential Genomic Biomarkers of Obesity and Its Comorbidities for Phthalates and Bisphenol A Mixture: In Silico Toxicogenomic Approach. *Biocell* **2022**, *46*, 519–533. [[CrossRef](#)]
151. Hong, S.H.; Sung, Y.A.; Hong, Y.S.; Ha, E.; Jeong, K.; Chung, H.; Lee, H. Urinary Bisphenol A Is Associated with Insulin Resistance and Obesity in Reproductive-Aged Women. *Clin. Endocrinol.* **2017**, *86*, 506–512. [[CrossRef](#)] [[PubMed](#)]
152. Lee, I.; Park, Y.J.; Kim, M.J.; Kim, S.; Choi, S.; Park, J.; Cho, Y.H.; Hong, S.; Yoo, J.; Park, H.; et al. Associations of Urinary Concentrations of Phthalate Metabolites, Bisphenol A, and Parabens with Obesity and Diabetes Mellitus in a Korean Adult Population: Korean National Environmental Health Survey (KoNEHS) 2015–2017. *Environ. Int.* **2021**, *146*, 106227. [[CrossRef](#)] [[PubMed](#)]
153. Lim, J.-e.; Choi, B.K.; Jee, S.H. Urinary Bisphenol A, Phthalate Metabolites, and Obesity: Do Gender and Menopausal Status Matter? *Environ. Sci. Pollut. Res.* **2020**, *27*, 34300–34310. [[CrossRef](#)]
154. Carwile, J.L.; Michels, K.B. Urinary Bisphenol A and Obesity: NHANES 2003–2006. *Environ. Res.* **2011**, *111*, 825–830. [[CrossRef](#)]
155. Amin, M.M.; Ebrahim, K.; Hashemi, M.; Shoshtari-Yeganeh, B.; Rafiei, N.; Mansourian, M.; Kelishadi, R. Association of Exposure to Bisphenol A with Obesity and Cardiometabolic Risk Factors in Children and Adolescents. *Int. J. Environ. Health Res.* **2019**, *29*, 94–106. [[CrossRef](#)]
156. Li, D.K.; Miao, M.; Zhou, Z.J.; Wu, C.; Shi, H.; Liu, X.; Wang, S.; Yuan, W. Urine Bisphenol-A Level in Relation to Obesity and Overweight in School-Age Children. *PLoS ONE* **2013**, *8*, 11–13. [[CrossRef](#)]
157. Wang, H.X.; Zhou, Y.; Tang, C.X.; Wu, J.G.; Chen, Y.; Jiang, Q.W. Association between Bisphenol A Exposure and Body Mass Index in Chinese School Children: A Cross-Sectional Study. *Environ. Health Glob. Access Sci. Source* **2012**, *11*, 79. [[CrossRef](#)]
158. Ohlstein, J.F.; Strong, A.L.; McLachlan, J.A.; Gimble, J.M.; Burrow, M.E.; Bunnell, B.A. Bisphenol a Enhances Adipogenic Differentiation of Human Adipose Stromal/Stem Cells. *J. Mol. Endocrinol.* **2014**, *53*, 345–353. [[CrossRef](#)] [[PubMed](#)]
159. Wang, J.; Sun, B.; Hou, M.; Pan, X.; Li, X. The Environmental Obesogen Bisphenol A Promotes Adipogenesis by Increasing the Amount of 11 β -Hydroxysteroid Dehydrogenase Type 1 in the Adipose Tissue of Children. *Int. J. Obes.* **2013**, *37*, 999–1005. [[CrossRef](#)] [[PubMed](#)]
160. Atlas, E.; Pope, L.; Wade, M.G.; Kawata, A.; Boudreau, A.; Boucher, J.G. Bisphenol A Increases AP2 Expression in 3T3L1 by Enhancing the Transcriptional Activity of Nuclear Receptors at the Promoter. *Adipocyte* **2014**, *3*, 170–179. [[CrossRef](#)] [[PubMed](#)]
161. Boucher, J.G.; Boudreau, A.; Atlas, E. Bisphenol a Induces Differentiation of Human Preadipocytes in the Absence of Glucocorticoid and Is Inhibited by an Estrogen-Receptor Antagonist. *Nutr. Diabetes* **2014**, *4*, e102. [[CrossRef](#)]
162. Longo, M.; Zatterale, F.; Naderi, J.; Nigro, C.; Oriente, F.; Formisano, P.; Miele, C.; Beguinot, F. Low-Dose Bisphenol-a Promotes Epigenetic Changes at Ppary Promoter in Adipose Precursor Cells. *Nutrients* **2020**, *12*, 3498. [[CrossRef](#)]
163. Masuno, H.; Iwanami, J.; Kidani, T.; Sakayama, K.; Honda, K. Bisphenol A Accelerates Terminal Differentiation of 3T3-L1 Cells into Adipocytes through the Phosphatidylinositol 3-Kinase Pathway. *Toxicol. Sci.* **2005**, *84*, 319–327. [[CrossRef](#)]
164. Ariemma, F.; D’Esposito, V.; Liguoro, D.; Oriente, F.; Cabaro, S.; Liotti, A.; Cimmino, I.; Longo, M.; Beguinot, F.; Formisano, P.; et al. Low-Dose Bisphenol-A Impairs Adipogenesis and Generates Dysfunctional 3T3-L1 Adipocytes. *PLoS ONE* **2016**, *11*, e0150762. [[CrossRef](#)] [[PubMed](#)]
165. De Filippis, E.; Li, T.; Rosen, E.D. Exposure of Adipocytes to Bisphenol-A in Vitro Interferes with Insulin Action without Enhancing Adipogenesis. *PLoS ONE* **2018**, *13*, e0201122. [[CrossRef](#)]
166. Valentino, R.; D’Esposito, V.; Passaretti, F.; Liotti, A.; Cabaro, S.; Longo, M.; Perruolo, G.; Oriente, F.; Beguinot, F.; Formisano, P. Bisphenol-A Impairs Insulin Action and up-Regulates Inflammatory Pathways in Human Subcutaneous Adipocytes and 3T3-L1 Cells. *PLoS ONE* **2013**, *8*, e82099. [[CrossRef](#)]
167. Hugo, E.R.; Brandebourg, T.D.; Woo, J.G.; Loftus, J.; Alexander, J.W.; Ben-Jonathan, N. Bisphenol A at Environmentally Relevant Doses Inhibits Adiponectin Release from Human Adipose Tissue Explants and Adipocytes. *Environ. Health Perspect.* **2008**, *116*, 1642–1647. [[CrossRef](#)] [[PubMed](#)]
168. Angle, B.M.; Do, R.P.; Ponzi, D.; Stahlhut, R.W.; Drury, B.E.; Nagel, S.C.; Welshons, W.V.; Besch-Williford, C.L.; Palanza, P.; Parmigiani, S.; et al. Metabolic Disruption in Male Mice Due to Fetal Exposure to Low but Not High Doses of Bisphenol A (BPA): Evidence for Effects on Body Weight, Food Intake, Adipocytes, Leptin, Adiponectin, Insulin and Glucose Regulation. *Reprod. Toxicol.* **2013**, *42*, 256–268. [[CrossRef](#)] [[PubMed](#)]
169. Somm, E.; Schwitzgebel, V.M.; Toulotte, A.; Cederroth, C.R.; Combescure, C.; Nef, S.; Aubert, M.L.; Hüppi, P.S. Perinatal Exposure to Bisphenol A Alters Early Adipogenesis in the Rat. *Environ. Health Perspect.* **2009**, *117*, 1549–1555. [[CrossRef](#)]

170. Lejonklou, M.H.; Dunder, L.; Bladin, E.; Pettersson, V.; Rönn, M.; Lind, L.; Waldén, T.B.; Lind, P.M. Effects of Low-Dose Developmental Bisphenol A Exposure on Metabolic Parameters and Gene Expression in Male and Female Fischer 344 Rat Offspring. *Environ. Health Perspect.* **2017**, *125*, 6. [[CrossRef](#)]
171. Yang, M.; Chen, M.; Wang, J.; Xu, M.; Sun, J.; Ding, L.; Lv, X.; Ma, Q.; Bi, Y.; Liu, R.; et al. Bisphenol a Promotes Adiposity and Inflammation in a Nonmonotonic Dose-Response Way in 5-Week-Old Male and Female C57BL/6J Mice Fed a Low-Calorie Diet. *Endocrinology* **2016**, *157*, 2333–2345. [[CrossRef](#)] [[PubMed](#)]
172. Desai, M.; Ferrini, M.G.; Jellyman, J.K.; Han, G.; Michael, G.; Angeles, L.; Drew, C.R.; Angeles, L.; Angeles, L. In Vivo and In Vitro Bisphenol A Exposure Effects on Adiposity. *J. Dev. Orig. Health Dis.* **2019**, *9*, 678–687. [[CrossRef](#)] [[PubMed](#)]
173. Van Esterik, J.C.J.; Dollé, M.E.T.; Lamoree, M.H.; van Leeuwen, S.P.J.; Hamers, T.; Legler, J.; Van der Ven, L.T.M. Programming of Metabolic Effects in C57BL/6JxFVB Mice by Exposure to Bisphenol A during Gestation and Lactation. *Toxicology* **2014**, *321*, 40–52. [[CrossRef](#)]
174. Boucher, J.G.; Boudreau, A.; Ahmed, S.; Atlas, E. In Vitro Effects of Bisphenol A β -D-Glucuronide (BPA-G) on Adipogenesis in Human and Murine Preadipocytes. *Environ. Health Perspect.* **2015**, *123*, 1287–1293. [[CrossRef](#)] [[PubMed](#)]
175. Chamorro-García, R.; Kirchner, S.; Li, X.; Janesick, A.; Casey, S.C.; Chow, C.; Blumberg, B. Bisphenol A Diglycidyl Ether Induces Adipogenic Differentiation of Multipotent Stromal Stem Cells through a Peroxisome Proliferator-Activated Receptor Gamma-Independent Mechanism. *Environ. Health Perspect.* **2012**, *120*, 984–989. [[CrossRef](#)]
176. Martínez, M.; Blanco, J.; Rovira, J.; Kumar, V.; Domingo, J.L.; Schuhmacher, M. Bisphenol A Analogues (BPS and BPF) Present a Greater Obesogenic Capacity in 3T3-L1 Cell Line. *Food Chem. Toxicol.* **2020**, *140*, 111298. [[CrossRef](#)] [[PubMed](#)]
177. Boucher, J.G.; Ahmed, S.; Atlas, E. Bisphenol S Induces Adipogenesis in Primary Human Preadipocytes from Female Donors. *Endocrinology* **2016**, *157*, 1397–1407. [[CrossRef](#)]
178. Ahmed, S.; Atlas, E. Bisphenol S- and Bisphenol A-Induced Adipogenesis of Murine Preadipocytes Occurs through Direct Peroxisome Proliferator-Activated Receptor Gamma Activation. *Int. J. Obes.* **2016**, *40*, 1566–1573. [[CrossRef](#)]
179. Boucher, J.G.; Gagné, R.; Rowan-Carroll, A.; Boudreau, A.; Yauk, C.L.; Atlas, E. Bisphenol A and Bisphenol S Induce Distinct Transcriptional Profiles in Differentiating Human Primary Preadipocytes. *PLoS ONE* **2016**, *11*, e0163318. [[CrossRef](#)]
180. Ahn, Y.A.; Baek, H.; Choi, M.; Park, J.; Son, S.J.; Seo, H.J.; Jung, J.; Seong, J.K.; Lee, J.; Kim, S. Adipogenic Effects of Prenatal Exposure to Bisphenol S (BPS) in Adult F1 Male Mice. *Sci. Total Environ.* **2020**, *728*, 138759. [[CrossRef](#)]
181. Skledar, D.G.; Carino, A.; Trontelj, J.; Troberg, J.; Distrutti, E.; Marchianò, S.; Tomašič, T.; Zega, A.; Finel, M.; Fiorucci, S.; et al. Endocrine Activities and Adipogenic Effects of Bisphenol AF and Its Main Metabolite. *Chemosphere* **2019**, *215*, 870–880. [[CrossRef](#)]
182. Stojanoska, M.M.; Milosevic, N.; Milic, N.; Abenavoli, L. The Influence of Phthalates and Bisphenol A on the Obesity Development and Glucose Metabolism Disorders. *Endocrine* **2017**, *55*, 666–681. [[CrossRef](#)] [[PubMed](#)]
183. Amin, M.M.; Ebrahimpour, K.; Parastar, S.; Shoshtari-Yeganeh, B.; Hashemi, M.; Mansourian, M.; Poursafa, P.; Fallah, Z.; Rafiei, N.; Kelishadi, R. Association of Urinary Concentrations of Phthalate Metabolites with Cardiometabolic Risk Factors and Obesity in Children and Adolescents. *Chemosphere* **2018**, *211*, 547–556. [[CrossRef](#)]
184. Hatch, E.E.; Nelson, J.W.; Qureshi, M.M.; Weinberg, J.; Moore, L.L.; Singer, M.; Webster, T.F. Association of Urinary Phthalate Metabolite Concentrations with Body Mass Index and Waist Circumference: A Cross-Sectional Study of NHANES Data, 1999–2002. *Environ. Health Glob. Access Sci. Source* **2008**, *7*, 27. [[CrossRef](#)]
185. Kim, J.H.; Park, H.; Lee, J.; Cho, G.; Choi, S.; Choi, G.; Kim, S.Y.; Eun, S.H.; Suh, E.; Kim, S.K.; et al. Association of Diethylhexyl Phthalate with Obesityrelated Markers and Body Mass Change from Birth to 3 Months of Age. *J. Epidemiol. Community Health* **2016**, *70*, 466–472. [[CrossRef](#)]
186. Mansouri, V.; Ebrahimpour, K.; Poursafa, P.; Riahi, R.; Shoshtari-Yeganeh, B.; Hystad, P.; Kelishadi, R. Exposure to Phthalates and Bisphenol A Is Associated with Higher Risk of Cardiometabolic Impairment in Normal Weight Children. *Environ. Sci. Pollut. Res.* **2019**, *26*, 18604–18614. [[CrossRef](#)]
187. Teitelbaum, S.L.; Mervish, N.; Moshier, L.E.; Vangeepuram, N.; Galvez, M.P.; Calafat, A.M.; Silva, M.J.; Brenner, L.B.; Wolff, M.S. Associations between Phthalate Metabolite Urinary Concentrations and Body Size Measures in New York City Children. *Environ. Res.* **2012**, *112*, 186–193. [[CrossRef](#)]
188. Desvergne, B.; Feige, J.N.; Casals-Casas, C. PPAR-Mediated Activity of Phthalates: A Link to the Obesity Epidemic? *Mol. Cell. Endocrinol.* **2009**, *304*, 43–48. [[CrossRef](#)]
189. Feige, J.N.; Gelman, L.; Rossi, D.; Zoete, V.; Métivier, R.; Tudor, C.; Anghel, S.I.; Grosdidier, A.; Lathion, C.; Engelborghs, Y.; et al. The Endocrine Disruptor Monoethyl-Hexyl-Phthalate Is a Selective Peroxisome Proliferator-Activated Receptor γ Modulator That Promotes Adipogenesis. *J. Biol. Chem.* **2007**, *282*, 19152–19166. [[CrossRef](#)] [[PubMed](#)]
190. Hurst, C.H.; Waxman, D.J. Activation of PPARalpha and PPARgamma by Environmental Phthalate Monoesters. *Toxicol. Sci.* **2003**, *74*, 297–308. [[CrossRef](#)] [[PubMed](#)]
191. Yin, L.; Yu, K.S.; Lu, K.; Yu, X. Benzyl Butyl Phthalate Promotes Adipogenesis in 3T3-L1 Preadipocytes: A High Content Cellomics and Metabolomic Analysis. *Toxicol. In Vitro* **2016**, *32*, 297–309. [[CrossRef](#)] [[PubMed](#)]
192. Sakuma, S.; Sumida, M.; Endoh, Y.; Kurita, A.; Yamaguchi, A.; Watanabe, T.; Kohda, T.; Tsukiyama, Y.; Fujimoto, Y. Curcumin Inhibits Adipogenesis Induced by Benzyl Butyl Phthalate in 3T3-L1 Cells. *Toxicol. Appl. Pharmacol.* **2017**, *329*, 158–164. [[CrossRef](#)]
193. Zhang, L.; Sun, W.; Duan, X.; Duan, Y.; Sun, H. Promoting Differentiation and Lipid Metabolism Are the Primary Effects for DINP Exposure on 3T3-L1 Preadipocytes. *Environ. Pollut.* **2019**, *255*, 113154. [[CrossRef](#)]

194. Chiu, C.Y.; Sun, S.C.; Chiang, C.K.; Wang, C.C.; Chan, D.C.; Chen, H.J.; Liu, S.H.; Yang, R. Sen Plasticizer Di(2-Ethylhexyl)Phthalate Interferes with Osteoblastogenesis and Adipogenesis in a Mouse Model. *J. Orthop. Res.* **2018**, *36*, 1124–1134. [[CrossRef](#)]
195. Hao, X.; Guan, X.; Zhao, X.; Ji, M.; Wen, X.; Chen, P.; Chen, F.; Yang, J.; Lian, Q.; Ye, L.; et al. Phthalate Inhibits Leydig Cell Differentiation and Promotes Adipocyte Differentiation. *Chemosphere* **2021**, *262*, 127855. [[CrossRef](#)] [[PubMed](#)]
196. Sargis, R.M.; Johnson, D.N.; Choudhury, R.A.; Brady, M.J. Environmental Endocrine Disruptors Promote Adipogenesis in the 3T3-L1 Cell Line through Glucocorticoid Receptor Activation. *Obesity* **2010**, *18*, 1283–1288. [[PubMed](#)]
197. Singh, N.; Dalal, V.; Kumar, P. Molecular Docking and Simulation Analysis for Elucidation of Toxic Effects of Dicyclohexyl Phthalate (DCHP) in Glucocorticoid Receptor-Mediated Adipogenesis. *Mol. Simul.* **2020**, *46*, 9–21. [[CrossRef](#)]
198. Schmidt, J.-S.; Schaedlich, K.; Fiandanese, N.; Pocar, P.; Fischer, B. Effects of Di(2-Ethylhexyl) Phthalate (DEHP) on Female Fertility and Adipogenesis in C3H/N Mice. *Environ. Health Perspect.* **2012**, *120*, 1123–1129. [[CrossRef](#)] [[PubMed](#)]
199. Chiang, H.C.; Wang, C.H.; Yeh, S.C.; Lin, Y.H.; Kuo, Y.T.; Liao, C.W.; Tsai, F.Y.; Lin, W.Y.; Chuang, W.H.; Tsou, T.C. Comparative Microarray Analyses of Mono(2-Ethylhexyl)Phthalate Impacts on Fat Cell Bioenergetics and Adipokine Network. *Cell Biol. Toxicol.* **2017**, *33*, 511–526. [[CrossRef](#)] [[PubMed](#)]
200. Ellero-Simatos, S.; Claus, S.P.; Benelli, C.; Forest, C.; Letourneur, F.; Cagnard, N.; Beaune, P.H.; de Waziers, I. Combined Transcriptomic-1H NMR Metabonomic Study Reveals That Monoethylhexyl Phthalate Stimulates Adipogenesis and Glyceroneogenesis in Human Adipocytes. *J. Proteome Res.* **2011**, *10*, 5493–5502.
201. Qi, W.; Xu, Q.; Xu, Y.; Wang, Z.; Yang, L.; Guo, S.; Shi, Y.; Zhao, T.; Zhou, L.; Ye, L. Effect of Notch Pathway on Lipid Accumulation Induced by Mono-2-Ethylhexyl Phthalate on 3T3-L1 Cells. *Ecotoxicol. Environ. Saf.* **2021**, *208*, 111472. [[CrossRef](#)]
202. Qi, W.; Zhou, L.; Zhao, T.; Ding, S.; Xu, Q.; Han, X.; Zhao, Y.; Song, X.; Zhao, T.; Zhang, X.; et al. Effect of the TYK-2/STAT-3 Pathway on Lipid Accumulation Induced by Mono-2-Ethylhexyl Phthalate. *Mol. Cell. Endocrinol.* **2019**, *484*, 52–58. [[CrossRef](#)]
203. Hsu, J.W.; Nien, C.Y.; Yeh, S.C.; Tsai, F.Y.; Chen, H.W.; Lee, T.S.; Chen, S.L.; Kao, Y.H.; Tsou, T.C. Phthalate Exposure Causes Browning-like Effects on Adipocytes in Vitro and in Vivo. *Food Chem. Toxicol.* **2020**, *142*, 111487. [[CrossRef](#)]
204. Chiang, H.c.; Kuo, Y.T.; Shen, C.C.; Lin, Y.H.; Wang, S.L.; Tsou, T.C. Mono(2-Ethylhexyl)Phthalate Accumulation Disturbs Energy Metabolism of Fat Cells. *Arch. Toxicol.* **2016**, *90*, 589–601. [[CrossRef](#)]
205. Hao, C. Perinatal Exposure to Diethyl-Hexyl-Phthalate Induces Obesity in Mice. *Front. Biosci.* **2013**, *E5*, E653. [[CrossRef](#)]
206. Hao, C.; Cheng, X.; Xia, H.; Ma, X. The Endocrine Disruptor Mono-(2-Ethylhexyl) Phthalate Promotes Adipocyte Differentiation and Induces Obesity in Mice. *Biosci. Rep.* **2012**, *32*, 619–629. [[CrossRef](#)]
207. Gu, H.; Liu, Y.; Wang, W.; Ding, L.; Teng, W.; Liu, L. In Utero Exposure to Di-(2-Ethylhexyl) Phthalate Induces Metabolic Disorder and Increases Fat Accumulation in Visceral Depots of C57BL/6J Mice Offspring. *Exp. Ther. Med.* **2016**, *12*, 3806–3812. [[CrossRef](#)]
208. Lee, K.I.; Chiang, C.W.; Lin, H.C.; Zhao, J.F.; Li, C.T.; Shyue, S.K.; Lee, T.S. Maternal Exposure to Di-(2-Ethylhexyl) Phthalate Exposure Deregulates Blood Pressure, Adiposity, Cholesterol Metabolism and Social Interaction in Mouse Offspring. *Arch. Toxicol.* **2016**, *90*, 1211–1224. [[CrossRef](#)]
209. Rajesh, P.; Sathish, S.; Srinivasan, C.; Selvaraj, J.; Balasubramanian, K. Phthalate Is Associated with Insulin Resistance in Adipose Tissue of Male Rat: Role of Antioxidant Vitamins. *J. Cell Biochem.* **2013**, *114*, 558–569. [[CrossRef](#)]
210. Zhang, J.; Powell, C.A.; Kay, M.K.; Park, M.H.; Meruvu, S.; Sonkar, R.; Choudhury, M. A Moderate Physiological Dose of Benzyl Butyl Phthalate Exacerbates the High Fat Diet-Induced Diabesity in Male Mice. *Toxicol. Res.* **2020**, *9*, 353–370. [[CrossRef](#)]
211. Zhou, L.; Chen, H.; Xu, Q.; Han, X.; Zhao, Y.; Song, X.; Zhao, T.; Ye, L. The Effect of Di-2-Ethylhexyl Phthalate on Inflammation and Lipid Metabolic Disorder in Rats. *Ecotoxicol. Environ. Saf.* **2019**, *170*, 391–398. [[CrossRef](#)]
212. Campioli, E.; Martinez-Arguelles, D.B.; Papadopoulos, V. In Utero Exposure to the Endocrine Disruptor Di-(2-Ethylhexyl) Phthalate Promotes Local Adipose and Systemic Inflammation in Adult Male Offspring. *Nutr. Diabetes* **2014**, *4*, e115. [[CrossRef](#)]
213. Manteiga, S.; Lee, K. Monoethylhexyl Phthalate Elicits an Inflammatory Response in Adipocytes Characterized by Alterations in Lipid and Cytokine Pathways. *Environ. Health Perspect.* **2017**, *125*, 615–622. [[CrossRef](#)]
214. Sonkar, R.; Powell, C.A.; Choudhury, M. Benzyl Butyl Phthalate Induces Epigenetic Stress to Enhance Adipogenesis in Mesenchymal Stem Cells. *Mol. Cell. Endocrinol.* **2016**, *431*, 109–122. [[CrossRef](#)]
215. Xu, Q.; Qi, W.; Zhang, Y.; Wang, Q.; Ding, S.; Han, X.; Zhao, Y.; Song, X.; Zhao, T.; Zhou, L.; et al. DNA Methylation of JAK3/STAT5/PPAR γ Regulated the Changes of Lipid Levels Induced by Di (2-Ethylhexyl) Phthalate and High-Fat Diet in Adolescent Rats. *Environ. Sci. Pollut. Res.* **2020**, *27*, 30232–30242. [[CrossRef](#)] [[PubMed](#)]
216. Moody, L.; Kougiyas, D.; Jung, P.M.; Digan, I.; Hong, A.; Gorski, A.; Chen, H.; Juraska, J.; Pan, Y.X. Perinatal Phthalate and High-Fat Diet Exposure Induce Sex-Specific Changes in Adipocyte Size and DNA Methylation. *J. Nutr. Biochem.* **2019**, *65*, 15–25. [[CrossRef](#)]
217. Su, H.; Yuan, P.; Lei, H.; Zhang, L.; Deng, D.; Zhang, L.; Chen, X. Long-Term Chronic Exposure to Di-(2-Ethylhexyl)-Phthalate Induces Obesity via Disruption of Host Lipid Metabolism and Gut Microbiota in Mice. *Chemosphere* **2022**, *287*, 132414. [[CrossRef](#)] [[PubMed](#)]
218. Hao, C.J.; Cheng, X.J.; Xia, H.F.; Ma, X. The Endocrine Disruptor Diethylstilbestrol Induces Adipocyte Differentiation and Promotes Obesity in Mice. *Toxicol. Appl. Pharmacol.* **2012**, *263*, 102–110. [[CrossRef](#)]
219. Newbold, R.R.; Padilla-Banks, E.; Jefferson, W.N. Environmental Estrogens and Obesity. *Mol. Cell. Endocrinol.* **2009**, *304*, 84–89. [[CrossRef](#)] [[PubMed](#)]
220. Hatch, E.E.; Troisi, R.; Palmer, J.R.; Wise, L.A.; Titus, L.; Strohsnitter, W.C.; Ricker, W.; Hyer, M.; Hoover, R.N. Prenatal Diethylstilbestrol Exposure and Risk of Obesity in Adult Women. *J. Dev. Orig. Health Dis.* **2014**, *6*, 201–207. [[CrossRef](#)]

221. Jensen, E.T.; Longnecker, M.P. Pharmacologic Sex Hormones in Pregnancy in Relation to Offspring Obesity. *Obesity* **2014**, *22*, 2406–2412. [[CrossRef](#)]
222. Biasiotto, G.; Zanella, I.; Masserdotti, A.; Pedrazzani, R.; Papa, M.; Caimi, L.; Di Lorenzo, D. Municipal Wastewater Affects Adipose Deposition in Male Mice and Increases 3T3-L1 Cell Differentiation. *Toxicol. Appl. Pharmacol.* **2016**, *297*, 32–40. [[CrossRef](#)]
223. Tsou, T.C.; Yeh, S.C.; Hsu, J.W.; Tsai, F.Y. Estrogenic Chemicals at Body Burden Levels Attenuate Energy Metabolism in 3T3-L1 Adipocytes. *J. Appl. Toxicol.* **2017**, *37*, 1537–1546. [[CrossRef](#)]
224. Bhardwaj, P.; Ikeda, T.; Zhou, X.K.; Wang, H.; Zheng, X.E.; Giri, D.D.; Elemento, O.; Verma, A.; Miyazawa, M.; Mukherjee, S.; et al. Supplemental Estrogen and Caloric Restriction Reduce Obesity-Induced Periprostatic White Adipose Inflammation in Mice. *Carcinogenesis* **2019**, *40*, 914–923. [[CrossRef](#)] [[PubMed](#)]
225. Escher, B.I.; Stapleton, H.M.; Schymanski, E.L. Tracking Complex Mixtures of Chemicals in Our Changing Environment. *Science* **2020**, *367*, 388–392. [[CrossRef](#)] [[PubMed](#)]
226. Zhang, Y.; Dong, T.; Hu, W.; Wang, X.; Xu, B.; Lin, Z.; Hofer, T.; Stefanoff, P.; Chen, Y.; Wang, X.; et al. Association between Exposure to a Mixture of Phenols, Pesticides, and Phthalates and Obesity: Comparison of Three Statistical Models. *Environ. Int.* **2019**, *123*, 325–336. [[CrossRef](#)] [[PubMed](#)]
227. Lyche, J.L.; Nourizadeh-Lillabadi, R.; Almaas, C.; Stavik, B.; Berg, V.; Skåre, J.U.; Alestrøm, P.; Ropstad, E. Natural Mixtures of Persistent Organic Pollutants (POP) Increase Weight Gain, Advance Puberty, and Induce Changes in Gene Expression Associated with Steroid Hormones and Obesity in Female Zebrafish. *J. Toxicol. Environ. Health Part A* **2010**, *73*, 1032–1057. [[CrossRef](#)] [[PubMed](#)]
228. Berntsen, H.F.; Berg, V.; Thomsen, C.; Ropstad, E.; Zimmer, K.E. The Design of an Environmentally Relevant Mixture of Persistent Organic Pollutants for Use in *In Vivo* and *In Vitro* Studies. *J. Toxicol. Environ. Health Part A* **2017**, *80*, 1002–1016. [[CrossRef](#)]
229. Ma, X.; Wang, D.; Zhao, W.; Xu, L. Deciphering the Roles of PPAR γ in Adipocytes via Dynamic Change of Transcription Complex. *Front. Endocrinol.* **2018**, *9*, 473. [[CrossRef](#)]
230. Rosen, E.D.; Sarraf, P.; Troy, A.E.; Bradwin, G.; Moore, K.; Milstone, D.S.; Spiegelman, B.M.; Mortensen, R.M. PPAR γ Is Required for the Differentiation of Adipose Tissue *In Vivo* and *In Vitro*. *Mol. Cell* **1999**, *4*, 611–617. [[CrossRef](#)]
231. Wang, Q.A.; Zhang, F.; Jiang, L.; Ye, R.; An, Y.; Shao, M.; Tao, C.; Gupta, R.K.; Scherer, P.E. Peroxisome Proliferator-Activated Receptor γ and Its Role in Adipocyte Homeostasis and Thiazolidinedione-Mediated Insulin Sensitization. *Mol. Cell Biol.* **2018**, *38*. [[CrossRef](#)]
232. Ros Pérez, M.; Medina-Gómez, G. Obesity, Adipogenesis and Insulin Resistance. *Endocrinol. Nutr.* **2011**, *58*, 360–369. [[CrossRef](#)]
233. Farmer, S.R. Regulation of PPAR γ Activity during Adipogenesis. *Int. J. Obes.* **2005**, *29*, S13–S16. [[CrossRef](#)]
234. Lefterova, M.I.; Haakonsson, A.K.; Lazar, M.A.; Mandrup, S. PPAR γ and the Global Map of Adipogenesis and Beyond. *Trends Endocrinol. Metab.* **2014**, *25*, 293–302. [[CrossRef](#)] [[PubMed](#)]
235. Rosen, E.D.; Hsu, C.-H.; Wang, X.; Sakai, S.; Freeman, M.W.; Gonzalez, F.J.; Spiegelman, B.M. C/EBP α Induces Adipogenesis through PPAR γ : A Unified Pathway. *Genes Dev.* **2002**, *16*, 22–26. [[CrossRef](#)] [[PubMed](#)]
236. Lee, J.-E.; Ge, K. Transcriptional and Epigenetic Regulation of PPAR γ Expression during Adipogenesis. *Cell Biosci.* **2014**, *4*, 29. [[CrossRef](#)] [[PubMed](#)]
237. Fujiki, K.; Kano, F.; Shiota, K.; Murata, M. Expression of the Peroxisome Proliferator Activated Receptor γ Gene Is Repressed by DNA Methylation in Visceral Adipose Tissue of Mouse Models of Diabetes. *BMC Biol.* **2009**, *7*, 38. [[CrossRef](#)] [[PubMed](#)]
238. Ahmadian, M.; Suh, J.M.; Hah, N.; Liddle, C.; Atkins, A.R.; Downes, M.; Evans, R.M. PPAR γ Signaling and Metabolism: The Good, the Bad and the Future. *Nat. Med.* **2013**, *19*, 557–566. [[CrossRef](#)]
239. Janesick, A.; Blumberg, B. Minireview: PPAR γ as the Target of Obesogens. *J. Steroid Biochem. Mol. Biol.* **2011**, *127*, 4–8. [[CrossRef](#)] [[PubMed](#)]
240. Pallottini, V.; Bulzomi, P.; Galluzzo, P.; Martini, C.; Marino, M. Estrogen Regulation of Adipose Tissue Functions: Involvement of Estrogen Receptor Isoforms. *IDDT* **2008**, *8*, 52–60. [[CrossRef](#)]
241. Foryst-Ludwig, A.; Clemenz, M.; Hohmann, S.; Hartge, M.; Sprang, C.; Frost, N.; Krikov, M.; Bhanot, S.; Barros, R.; Morani, A.; et al. Metabolic Actions of Estrogen Receptor Beta (ER β) Are Mediated by a Negative Cross-Talk with PPAR γ . *PLoS Genet.* **2008**, *4*, e1000108. [[CrossRef](#)]
242. Jeong, S.; Yoon, M. 17 β -Estradiol Inhibition of PPAR γ -Induced Adipogenesis and Adipocyte-Specific Gene Expression. *Acta Pharm. Sin.* **2011**, *32*, 230–238. [[CrossRef](#)]
243. Yepuru, M.; Eswaraka, J.; Kearbey, J.D.; Barrett, C.M.; Raghov, S.; Veverka, K.A.; Miller, D.D.; Dalton, J.T.; Narayanan, R. Estrogen Receptor- β -Selective Ligands Alleviate High-Fat Diet- and Ovariectomy-Induced Obesity in Mice. *J. Biol. Chem.* **2010**, *285*, 31292–31303. [[CrossRef](#)]
244. González-Granillo, M.; Savva, C.; Li, X.; Fitch, M.; Pedrelli, M.; Hellerstein, M.; Parini, P.; Korach-André, M.; Gustafsson, J.-Å. ER β Activation in Obesity Improves Whole Body Metabolism via Adipose Tissue Function and Enhanced Mitochondria Biogenesis. *Mol. Cell. Endocrinol.* **2019**, *479*, 147–158. [[CrossRef](#)]
245. Bian, X.; Liu, T.; Zhou, M.; He, G.; Ma, Y.; Shi, Y.; Wang, Y.; Tang, H.; Kang, X.; Yang, M.; et al. Absence of Estrogen Receptor Beta Leads to Abnormal Adipogenesis during Early Tendon Healing by an Up-regulation of PPAR γ Signalling. *J. Cell Mol. Med.* **2019**, *23*, 7406–7416. [[CrossRef](#)] [[PubMed](#)]
246. Dieudonné, M.N.; Leneuve, M.C.; Giudicelli, Y.; Pecquery, R. Evidence for Functional Estrogen Receptors α and β in Human Adipose Cells: Regional Specificities and Regulation by Estrogens. *Am. J. Physiol.-Cell Physiol.* **2004**, *286*, C655–C661. [[CrossRef](#)]

247. Davis, K.E.; Neinast, D.M.; Sun, K.; Skiles, M.W.; Bills, D.J.; Zehr, A.J.; Zeve, D.; Hahner, D.L.; Cox, W.D.; Gent, M.L.; et al. The Sexually Dimorphic Role of Adipose and Adipocyte Estrogen Receptors in Modulating Adipose Tissue Expansion, Inflammation, and Fibrosis. *Mol. Metab.* **2013**, *2*, 227–242. [[CrossRef](#)] [[PubMed](#)]
248. Zhou, Z.; Moore, T.M.; Drew, B.G.; Ribas, V.; Wanagat, J.; Civelek, M.; Segawa, M.; Wolf, D.M.; Norheim, F.; Seldin, M.M.; et al. Estrogen Receptor α Controls Metabolism in White and Brown Adipocytes by Regulating *Polg1* and Mitochondrial Remodeling. *Sci. Transl. Med.* **2020**, *12*, eaax8096. [[CrossRef](#)] [[PubMed](#)]
249. Santos, R.S.; Frank, A.P.; Fátima, L.A.; Palmer, B.F.; Öz, O.K.; Clegg, D.J. Activation of Estrogen Receptor Alpha Induces Beiging of Adipocytes. *Mol. Metab.* **2018**, *18*, 51–59. [[CrossRef](#)]
250. Blüher, M. Importance of Estrogen Receptors in Adipose Tissue Function. *Mol. Metab.* **2013**, *2*, 130–132. [[CrossRef](#)] [[PubMed](#)]
251. Ahluwalia, A.; Hoa, N.; Ge, L.; Blumberg, B.; Levin, E.R. Mechanisms by Which Membrane and Nuclear ER Alpha Inhibit Adipogenesis in Cells Isolated From Female Mice. *Endocrinology* **2020**, *161*, bqaa175. [[CrossRef](#)]
252. Bitirim, C.V.; Ozer, Z.B.; Akcali, K.C. Estrogen Receptor Alpha Regulates the Expression of Adipogenic Genes Genetically and Epigenetically in Rat Bone Marrow-Derived Mesenchymal Stem Cells. *PeerJ* **2021**, *9*, e12071. [[CrossRef](#)] [[PubMed](#)]
253. Shin, J.-H.; Hur, J.-Y.; Seo, H.S.; Jeong, Y.-A.; Lee, J.K.; Oh, M.-J.; Kim, T.; Saw, H.S.; Kim, S.H. The Ratio of Estrogen Receptor α to Estrogen Receptor β in Adipose Tissue Is Associated with Leptin Production and Obesity. *Steroids* **2007**, *72*, 592–599. [[CrossRef](#)]
254. Le Magueresse-Battistoni, B. Adipose Tissue and Endocrine-Disrupting Chemicals: Does Sex Matter? *Int. J. Environ. Res. Public Health* **2020**, *17*, 9403. [[CrossRef](#)]
255. Lee, R.A.; Harris, C.A.; Wang, J.-C. Glucocorticoid Receptor and Adipocyte Biology. *Nucl. Recept. Res.* **2018**, *5*. [[CrossRef](#)]
256. Abdou, H.-S.; Atlas, E.; Haché, R.J.G. A Positive Regulatory Domain in CCAAT/Enhancer Binding Protein β (C/EBP β) Is Required for the Glucocorticoid-Mediated Displacement of Histone Deacetylase 1 (HDAC1) from the C/Ebp α Promoter and Maximum Adipogenesis. *Endocrinology* **2013**, *154*, 1454–1464. [[CrossRef](#)]
257. Steger, D.J.; Grant, G.R.; Schupp, M.; Tomaru, T.; Lefterova, M.I.; Schug, J.; Manduchi, E.; Stoeckert, C.J.; Lazar, M.A. Propagation of Adipogenic Signals through an Epigenomic Transition State. *Genes Dev.* **2010**, *24*, 1035–1044. [[CrossRef](#)] [[PubMed](#)]
258. Merrett, J.E.; Bo, T.; Psaltis, P.J.; Proud, C.G. Identification of DNA Response Elements Regulating Expression of CCAAT/Enhancer-Binding Protein (C/EBP) β and δ and MAP Kinase-Interacting Kinases during Early Adipogenesis. *Adipocyte* **2020**, *9*, 427–442. [[CrossRef](#)] [[PubMed](#)]
259. Asada, M.; Rauch, A.; Shimizu, H.; Maruyama, H.; Miyaki, S.; Shibamori, M.; Kawasome, H.; Ishiyama, H.; Tuckermann, J.; Asahara, H. DNA Binding-Dependent Glucocorticoid Receptor Activity Promotes Adipogenesis via Krüppel-like Factor 15 Gene Expression. *Lab. Invest.* **2011**, *91*, 203–215. [[CrossRef](#)] [[PubMed](#)]
260. Bauerle, K.T.; Hutson, I.; Scheller, E.L.; Harris, C.A. Glucocorticoid Receptor Signaling Is Not Required for In Vivo Adipogenesis. *Endocrinology* **2018**, *159*, 2050–2061. [[CrossRef](#)]
261. Park, Y.-K.; Ge, K. Glucocorticoid Receptor Accelerates, but Is Dispensable for, Adipogenesis. *Mol. Cell Biol.* **2017**, *37*. [[CrossRef](#)]
262. Desarzens, S.; Faresse, N. Adipocyte Glucocorticoid Receptor Has a Minor Contribution in Adipose Tissue Growth. *J. Endocrinol.* **2016**, *230*, 1–11. [[CrossRef](#)]