


Article

Synthesis of Catechol Derived Rosamine Dyes and Their Reactivity toward Biogenic Amines

Filipe Monteiro-Silva ^{1,†}, Carla Queirós ^{2,†}, Andreia Leite ², María T. Rodríguez ³, María J. Rojo ³, Tomás Torroba ³, Rui C. Martins ¹, Ana M. G. Silva ^{2,*}  and Maria Rangel ⁴ 

¹ Centre for Applied Photonics, INESC TEC, Faculdade de Ciências da Universidade do Porto, Rua do Campo Alegre, 4169-007 Porto, Portugal; filipe.m.silva@inesctec.pt (F.M.-S.); rui.c.martins@inesctec.pt (R.C.M.)

² LAQV/REQUIMTE, Departamento de Química e Bioquímica, Faculdade de Ciências da Universidade do Porto, Rua do Campo Alegre, 4169-007 Porto, Portugal; carla.queiros@fc.up.pt (C.Q.); acleite@fc.up.pt (A.L.)

³ Departamento de Química, Facultad de Ciencias, Universidad de Burgos, 09001 Burgos, Spain; mtrod@ubu.es (M.T.R.); mjrocam@ubu.es (M.J.R.); ttorroba@ubu.es (T.T.)

⁴ LAQV/REQUIMTE, Instituto de Ciências Biomédicas de Abel Salazar, Rua Jorge de Viterbo Ferreira 228, 4050-313 Porto, Portugal; mrangel@icbas.up.pt

* Correspondence: ana.silva@fc.up.pt

† These authors contributed equally to this work.

Abstract: Functional organic dyes play a key role in many fields, namely in biotechnology and medical diagnosis. Herein, we report two novel 2,3- and 3,4-dihydroxyphenyl substituted rosamines (**3** and **4**, respectively) that were successfully synthesized through a microwave-assisted protocol. The best reaction yields were obtained for rosamine **4**, which also showed the most interesting photophysical properties, specially toward biogenic amines (BAs). Several amines including *n*- and *t*-butylamine, cadaverine, and putrescine cause spectral changes of **4**, in UV-Vis and fluorescence spectra, which are indicative of their potential application as an effective tool to detect amines in acetonitrile solutions. In the gas phase, the probe response is more expressive for spermine and putrescine. Additionally, we found that methanolic solutions of rosamine **4** and *n*-butylamine undergo a pink to yellow color change over time, which has been attributed to the formation of a new compound. The latter was isolated and identified as **5** (9-aminopyronin), whose solutions exhibit a remarkable increase in fluorescence intensity together with a shift toward more energetic wavelengths. Other 9-aminopyronins **6a**, **6b**, **7a**, and **7b** were obtained from methanolic solutions of **4** with putrescine and cadaverine, demonstrating the potential of this new xanthene entity to react with primary amines.

Keywords: microwave-assisted synthesis; catecholated rosamines; 9-aminopyronins; photophysical properties; biogenic amines detection



Citation: Monteiro-Silva, F.; Queirós, C.; Leite, A.; Rodríguez, M.T.; Rojo, M.J.; Torroba, T.; Martins, R.C.; Silva, A.M.G.; Rangel, M. Synthesis of Catechol Derived Rosamine Dyes and Their Reactivity toward Biogenic Amines. *Molecules* **2021**, *26*, 5082. <https://doi.org/10.3390/molecules26165082>

Academic Editor: Antonio Zuorro

Received: 26 July 2021

Accepted: 9 August 2021

Published: 22 August 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Catechols are biologically reactive molecules that are widespread in nature and largely used in biomedicine and nanotechnology [1–3]. Catechol-functionalized molecules and materials are involved in many areas of chemistry and materials science [4], especially for producing: (i) metallopolymer networks for water purification [5]; (ii) adhesive functional hydrogels for biomedical applications [6]; and (iii) polymers with antimicrobial properties [7]. Catechols can be easily oxidized via autoxidation in the presence of molecular oxygen, photooxidation, or through the addition of chemical agents or enzymatic oxidants [8–10]. *o*-Quinones are the most common oxidation products of catechols, being reactive intermediates in several reactions. They can also be attacked by nucleophiles such as amines, whose attack is dependent of the electron withdrawing or donating nature of the substituents present in the quinone skeleton [11], solvent, and pH.

Amines, on the other hand, are produced on the ton scale worldwide and are used in pharmaceutical, agricultural, and several industry-based applications [12]. Biogenic amines (BAs) are nitrogen-based compounds originating mainly from the decarboxylation of amino acids present in animals, microorganisms, and plants [13]. Amines can also display significant reactivities and their radical cations play a significant role in biological systems such as in the enzymatic metabolism of endogenous amines to the corresponding imines [14]. The detection of amines is a relevant research area with impact on the diagnosis of several diseases as well as industrial and environmental monitoring, besides food quality control. In this context, fluorescent probes have emerged as a rapid, simple, and accurate molecular tool to detect trace amounts of BAs. Optical detection of BAs is mainly based on: (i) the use of functional organic dyes such as phenanthridines [15], naphthalenes [16], benzothiazoles [17], porphyrinoids [18,19]; (ii) supramolecular hydrogels [20]; and (iii) boron complexes [21,22]. Recently revised by A. Gupta, the aggregation-induced emission (AIE) phenomenon by different materials presents itself as a method with high potential for the detection of amines by an increase or a quench of the fluorescence process [23].

In recent years, our group has synthesized several fluorescein and rhodamine-catechol conjugates and studied their interaction with transition metal ions from the coordination point of view, also exploring their ability to act as metal ion sensors [24,25]. Inspired by the protonation and redox behavior of catechols and by the excellent photophysical properties of Rhodamine B (RhB) and other xanthene derivatives [26], we present herein a simple design strategy to synthesize rosamines 1–4, with the purpose of detecting BAs by the direct conjugation of 2,3- and 3,4-dihydroxyphenyl rings with the xanthene platform (Figure 1).

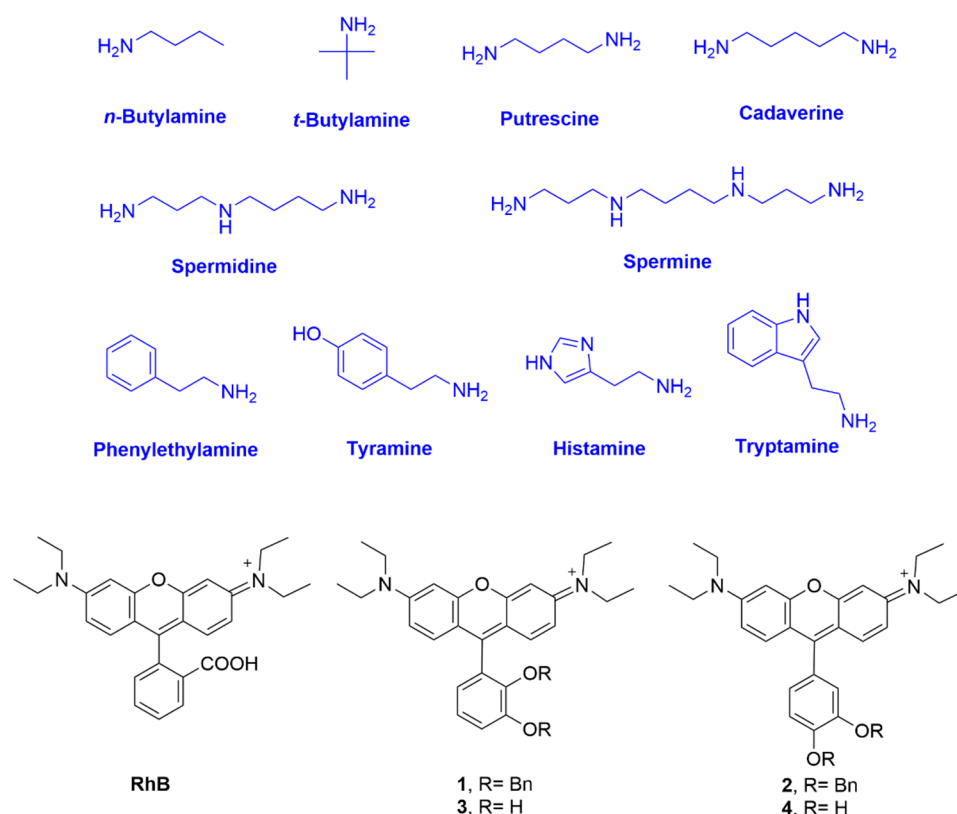
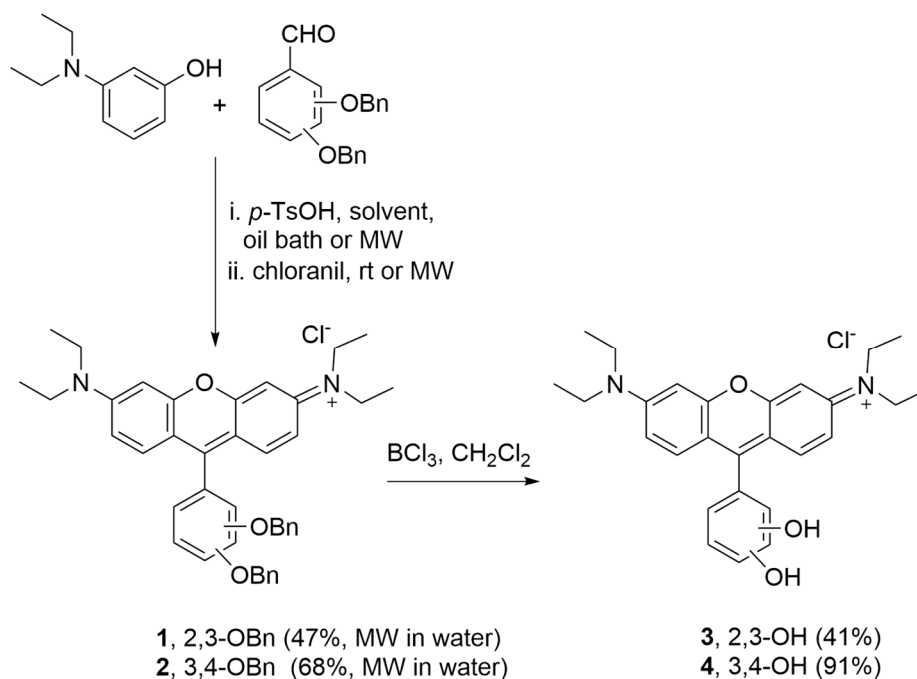


Figure 1. Chemical structures of the biogenic amines used in the studies, RhB and rosamines 1–4.

2. Results and Discussion

The synthetic route for rosamines 1–4 is simple and effective (Scheme 1). It consists of the condensation of the appropriate dibenzyloxybenzaldehyde and 3-(diethylamino)phenol in the presence of a catalytic amount of *p*-toluenesulfonic acid (*p*-TsOH), with subsequent oxidation with chloranil to give 1 and 2. The removal of the benzyl protecting groups was

performed using boron trichloride in dichloromethane to afford **3** and **4**. After screening different experimental conditions for condensation including (i) the heating method (oil bath or microwave irradiation, MW) and (ii) the reaction solvent (propionic acid or water), we verified that the combination of MW and water provided higher yields for compounds **1** and **2** (47% and 68%, respectively) in a remarkable shorter period of time (10 min) (please refer to the Materials and Methods section for more details).



Scheme 1. Synthetic route to obtain rosamines **1–4**.

Rosamines (**1–4**) were characterized by ^1H NMR, ^{13}C NMR, HRMS, UV–Vis, and fluorescence spectroscopy. These were obtained as a dark pink powder soluble in organic solvents, but were only sparingly soluble in an aqueous solution. Their photophysical properties including absorption extinction coefficients (ϵ) and fluorescence quantum yield (Φ_F) were studied in aprotic and protic solvents including chloroform, acetonitrile, and methanol and compared with **RhB**. The results are listed in Table 1.

The data showed that the molecular rigidity and the electron density of the substituent groups have remarkable effects on the photophysical properties of the compounds, namely: (i) in chloroform, the absorption bands of the catecholated compounds were in the shorter wavelength region than those of the benzylated ones; (ii) a marginally higher Φ_F value (0.60) was obtained for **1** in chloroform, where the orthogonal OBn substituent constrains the rotation of the 9-phenyl ring and thus enhances its fluorescence properties [27]; and (iii) the presence of the catechol substituent (**3** and **4**) remarkably decreased the Φ_F value of the molecules due to the intramolecular photoinduced electron transfer (PET), which occurs from the hydroxyl electron donating groups (D) to the xanthene electron acceptor fragment (A), producing the donor–acceptor (D–A) pair [28]. Although the catecholate derivatives are always less emissive than the respective benzylated derivatives, the emission of **3** and **4** (especially in terms of Φ_F values) was significantly higher in acetonitrile than in chloroform and methanol. Lower Stokes shifts were observed for rosamines **1** and **2** in chloroform, the same being true for **RhB**. On the other hand, for rosamines **3** and **4**, the solvent did not influence the Stokes shift values as significantly.

Table 1. Photophysical properties ($\lambda_{\max \text{ abs}}$, $\lambda_{\max \text{ em}}$, ϵ , Stokes shift, and Φ_F) of rosamines 1–4 and RhB in chloroform, acetonitrile, and methanol at 25 °C.

Dye	R	Solvent	$\lambda_{\max \text{ (abs)}}$ (nm)	$\epsilon \times 10^4$ ($M^{-1} \cdot \text{cm}^{-1}$)	$\lambda_{\max \text{ (em)}}$ (nm)	Stokes Shift (nm) (cm^{-1})	Φ_F
1	2,3-OBn	CHCl ₃	560	7.01	576	16 (496)	0.60
		CH ₃ CN	560	8.66	582	22 (675)	0.23
		CH ₃ OH	558	5.68	580	22 (680)	0.30
2	3,4-OBn	CHCl ₃	556	7.44	569	13 (411)	0.58
		CH ₃ CN	555	6.18	576	21 (657)	0.20
		CH ₃ OH	554	6.36	573	19 (689)	0.25
3	2,3-OH	CHCl ₃	546	10.5	566	20 (647)	0.04
		CH ₃ CN	557	4.50	580	23 (712)	0.12
		CH ₃ OH	550	9.38	570	20 (638)	0.02
4	3,4-OH	CHCl ₃	548	5.06	570	22 (704)	0.04
		CH ₃ CN	552	5.10	573	21 (664)	0.12
		CH ₃ OH	553	4.94	573	20 (631)	0.03
RhB	2-COOH	CHCl ₃	554	11.2	571	17 (537)	0.54
		CH ₃ CN	555	8.20	578	23 (717)	0.22
		CH ₃ OH	547	11.5	569	22 (707)	0.33

Taking into account the yields obtained in the synthesis and the peculiar spectral properties of the catechol derived compounds, we considered rosamine 4 as the most interesting dye in this series. Accordingly, we started by studying the influence of pH variation ($2 < \text{pH} < 12$) in the fluorescence intensity of rosamine 4. The graphical representation of the fluorescence intensity as a function of the pH value showed a decrease in intensity from 2 to approximately 10 (please refer to Figure S27 in the Supplementary Materials). In this range, two deprotonations can be observed both related to the deprotonation of the two hydroxyl groups, the first occurs for pH values from pH 2 to 6.5 (zwitterionic form) and the second deprotonation occurs for pH values above 6.5 until pH 9.90 (anionic form).

Subsequently, we investigated the color and fluorescence qualitative changes of acetonitrile solutions of 4 in the presence of the most common BAs diluted in water at room temperature using 2, 4, and 8 equiv. of amine. As shown in Figure S28 of the Supplementary Materials, by adding increasing amounts of amines, there was a change (quench of fluorescence) observed under ultraviolet radiation, indicating that rosamine 4 is sensitive to 2 equiv. of cadaverine, putrescine, spermidine, and spermine. Furthermore, upon addition of 4 and 8 equiv., a quench in fluorescence was also observed for *n*-butylamine and histamine, respectively.

We also tested the influence of selected amines, namely *n*- and *t*-butylamine, cadaverine, and putrescine on the spectroscopic behavior of rosamine 4 in acetonitrile, using UV–Vis and fluorescence (Figure 2). With the addition of 2 equiv. of amine, there was an absorbance decrease at 552 nm, more expressive for cadaverine. On the emission spectra, there was an intensity quench at 573 nm with the addition of 2 equiv. of amine, with the exception of *t*-butylamine, where a small increase was observed. In terms of percentage values, for the maximum absorbance, the decrease was in the order of 8%, 12%, 21%, and 29% for *n*-butylamine, *t*-butylamine, putrescine, and cadaverine, respectively, while for the emission intensity, the percentage values were 8%, 4% (increase), 27%, and 18%, respectively.

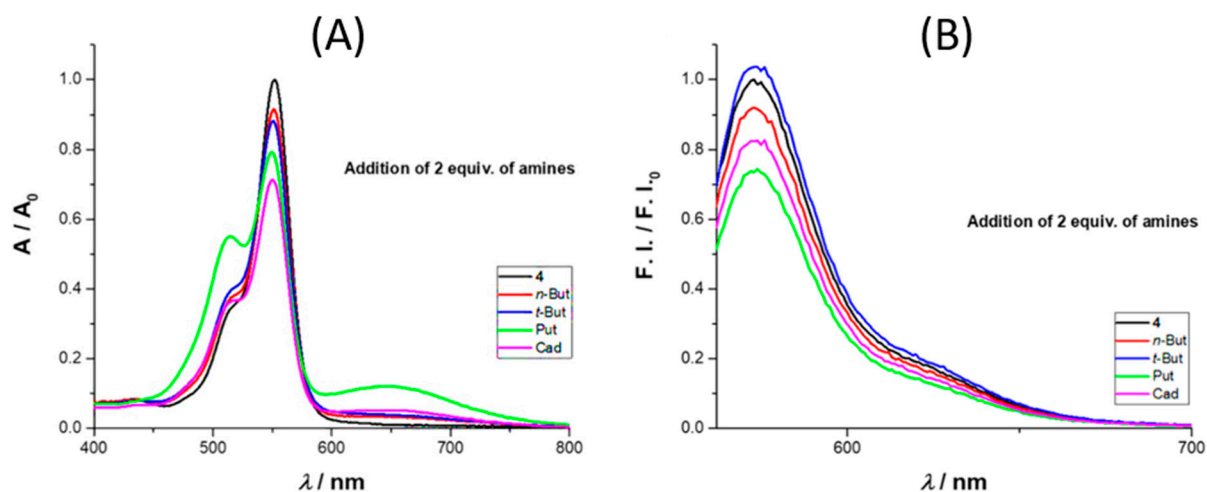


Figure 2. Comparison of the absorbance (A) and emission intensity (B) spectra of **4** toward 2 equiv. of *n*-butylamine (*n*-But, red line), *t*-butylamine (*t*-But, blue line), putrescine (Put, green line), and cadaverine (Cad, magenta line) in solution (acetonitrile, 25 °C).

The absorbance and emission spectra of **4** after the addition of up to 8 equiv. of putrescine is presented in Figure 3 (further amine results are available in the Supplementary Materials). The absorbance spectra revealed three regions of interest, two with an intensity increase (512 and 649 nm) and the third (at 551 nm) where a quench was observed as the molar equivalents of putrescine increased. The intensity enhancement regions can be used to monitor the presence of putrescine in the concentration range (0 to 122 μM). The region between 649 and 657 nm is particularly interesting due to the enhancement of the absorbance; the same behavior was observed for cadaverine (please refer to the Supplementary Materials, Figure S29, and for different concentrations of cadaverine (0.1–40 equiv.) please refer to Figures S30 and S31). The excitation of **4** in the presence of 8 equiv. of putrescine at 649 nm did not lead to a new emission band, the same being true for the other BAs. Analyzing the emission spectra, a quench percentage of 55% was obtained for an addition of 8 equiv. of putrescine, much higher values than those observed for *n*-butylamine and cadaverine (12% and 32%, respectively), whereas for *t*-butylamine, a slight increase was obtained (6%).

To demonstrate that rosamine **4** can be used in the detection of volatile amine vapors, which are highly toxic, irritant, and corrosive, being important indicators of food quality, for example, in the assessment for fish freshness [29], we investigated its reactivity toward BAs in the gas phase. The study was carried out by injecting vapors of different commercial BAs in solutions of **4** dissolved in acetonitrile (see the Materials and Methods section for more details and the apparatus in Figure S32 in the Supplementary Materials). The spectrophotometrical measurements were carried at 533 nm. As shown in Figure 4, nitrogen, air, and water vapor did not affect rosamine **4**, since the value of fluorescence intensity did not change when they were injected. However, when the commercial BA vapor was introduced, a decrease in the fluorescence intensity was observed, which was more expressive upon spermidine addition, resulting in decreases greater than 90%, 87% for putrescine, 52% for spermidine, 47% for cadaverine, 32% for tryptamine, 21% for histamine, and 8% for tyramine. The results for putrescine and cadaverine confirmed rosamine **4** sensitivity toward these BAs with higher quenching percentages in the gas phase comparatively to those reported in solution for 8 equiv.

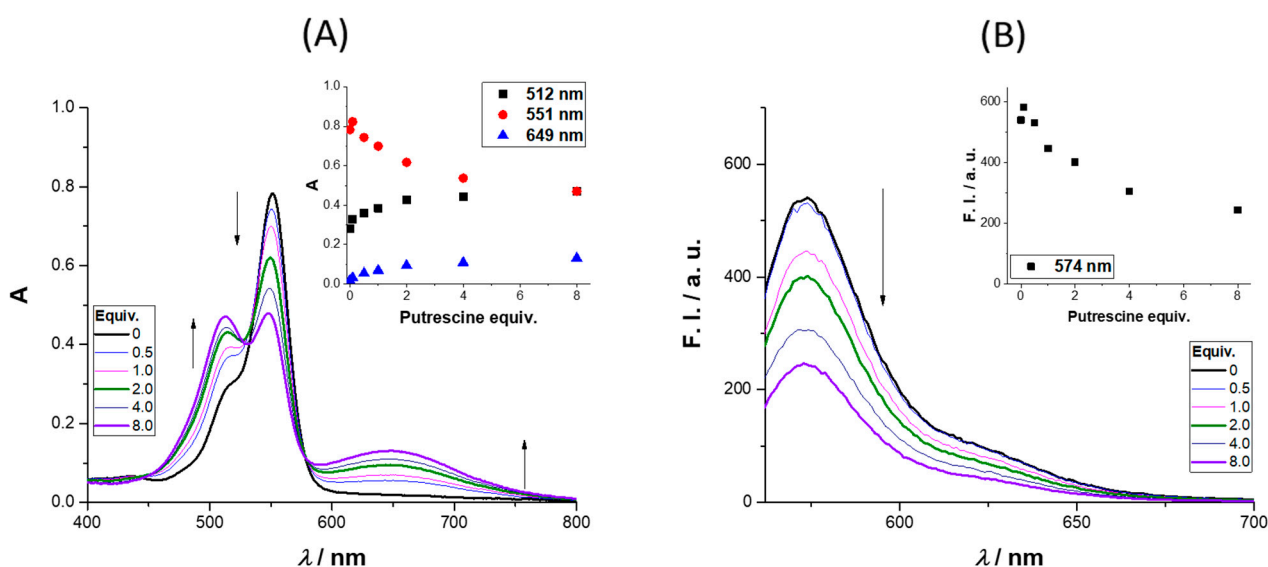


Figure 3. Absorption (A) and emission (B) spectra of **4** upon the addition (0–8 equiv.) of putrescine in CH₃CN (the inset graphs represent the variation in absorbance (left) and fluorescence intensity (right) for the wavelengths that display more intensity variation (512, 551, and 649 nm; 574 nm, respectively)). $[4] = 35 \mu\text{M}$ and $\lambda_{\text{exc}} = 551 \text{ nm}$.

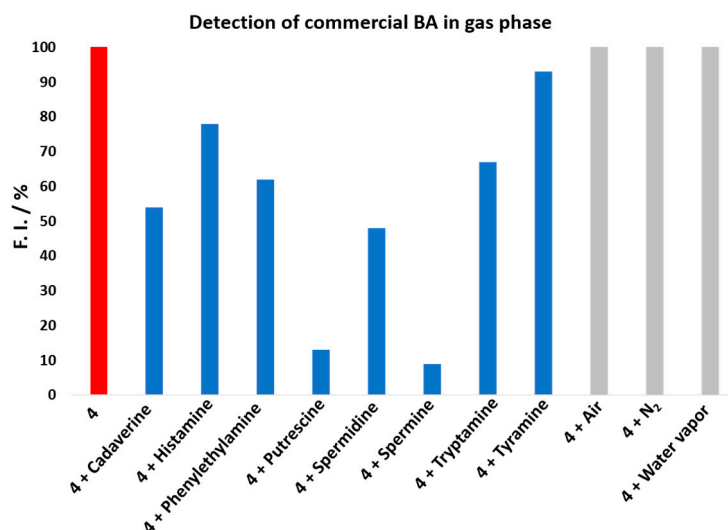


Figure 4. Comparison of the fluorescence intensity change of **4** toward BA in the gas phase at $\lambda_{\text{exc}} 533 \text{ nm}$. Probe **4** in red, Probe **4** + BA in blue, and Probe **4** + Controls (air, N₂ and water vapor) in grey.

During the experimental assays, when compound **4** was brought to the presence of amines, we observed that—particularly for methanolic solutions of **4** with *n*-butylamine—a color change from pink to yellow was observed over time, at room temperature and at 4 °C (refrigerator), which appeared to be due to the formation of a new compound. To verify the aforementioned hypothesis, a solution of rosamine **4** and *n*-butylamine (15 equiv.) in methanol was prepared, which was kept at 4 °C for 120 h. The TLC analysis of the resulting solution revealed the presence of a new compound with a bright yellow color, which was subsequently isolated by preparative thin layer chromatography and characterized by NMR and ESI-MS. The corresponding ¹H NMR spectrum (please refer to Figure 5B and Figure S33 of the Supplementary Materials) exhibited: (i) three set of signals from 6.73 to 8.12 ppm due to the resonance of the xanthene ring (no signals due to the catechol unit were detected indicating the absence of the catechol substituent in the molecule) and (ii) a new set of signals appeared at 1.05, 1.55, 1.93, and 4.02 ppm due to the entry of a butylamine moiety, suggesting the formation of 9-aminopyronin **5** (Figure 5A). A comparison of the ¹H NMR

spectra of compounds **4** and **5** is provided in the Supplementary Materials (please refer to Figure S35). Further evidences of **5** were obtained by ESI-MS, through the molecular ion peak $[M + H]^+$ at m/z 394.2851 (Figure 5C and Figure S39 in the Supplementary Materials).

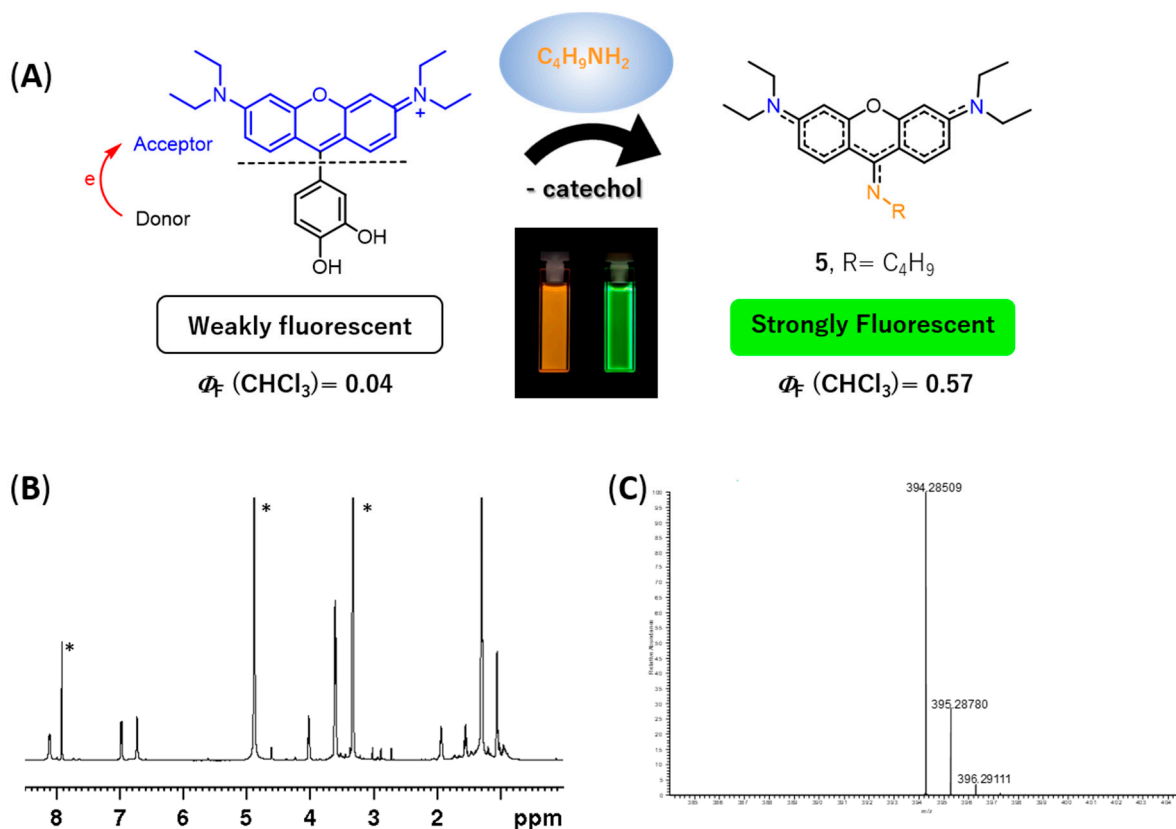
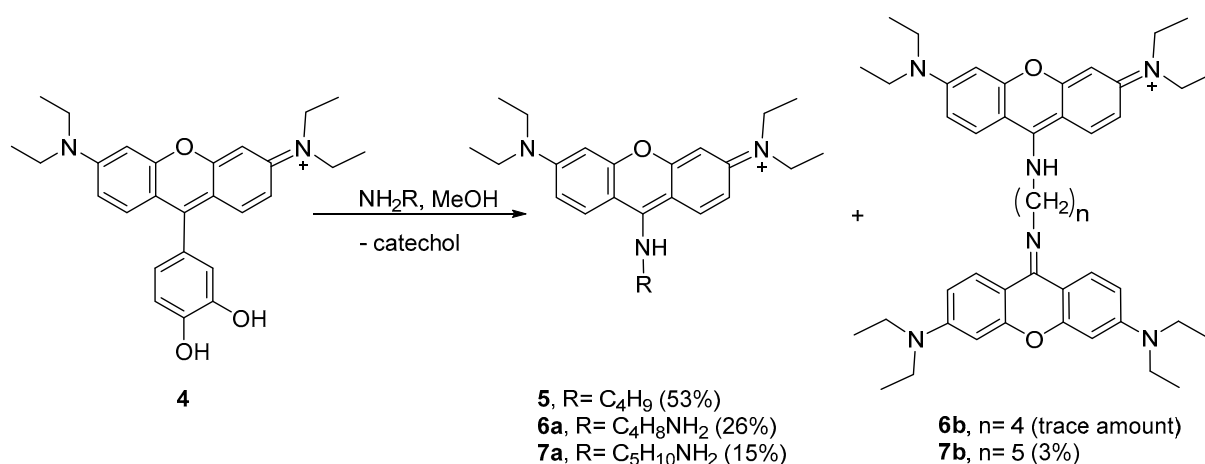


Figure 5. Reactivity of rosamine **4** with *n*-butylamine. (A) Scheme of the reaction of rosamine **4** with *n*-butylamine leading to the formation of 9-aminopyronin **5**. Rosamine **4** itself is weakly fluorescent due to the intramolecular PET quenching (left hand-side cuvette). However, after reaction with *n*-butylamine and chromatographic purification, 9-aminopyronin **5** can be isolated, which exhibits strong fluorescence (right hand-side cuvette); (B) 1H NMR spectrum of **5** in methanol- d_4 , * denote solvent peaks (chloroform and methanol); (C) ESI-MS spectrum of **5**.

In fact, 9-aminopyronin dyes are described in the literature as chemically stable dyes having high molar absorption coefficients and high fluorescence quantum yields. Being initially introduced by K. Burgess et al. [30], their synthesis typically involves the reaction of xanthenone derivatives with amines in the presence of trifluoromethanesulfonic anhydride or oxalyl chloride [31–34]. Alternatively, 9-aminopyronins can be prepared by reacting 9H-xanthen-9-thione with electron-withdrawing groups such as 4-aminopyridine and 2-aminoquinoline [35]. However, to the best of our knowledge, there are no preceding reports describing the synthesis of 9-aminopyronine dyes from catecholate xanthenes with amines.

Several attempts have been conducted in order to improve the yield of aminopyronin **5**, which included reactions at 4 °C using chloroform or methanol and the use of MW heating (*N*-methylpyrrolidine as solvent, 200 °C, 10 min; detailed procedures in the Materials and Methods section). The best result was obtained when methanol was used as a solvent (4 °C, for ca. 144 h) with a 53% yield.

Considering the results obtained with *n*-butylamine, we decided to study the peculiar reactivity of **4** using two other nucleophilic amines—putrescine and cadaverine. Using methanol as the solvent at 4 °C, the reactions took place in different extensions, leading to aminopyronins **6a,b** and **7a,b** (Scheme 2, see 1H NMR spectra in Figures S36–S38 in the Supplementary Materials).



Scheme 2. Reaction involving the nucleophilic attack of alkylamines to **4**, leading to aminopyronin **5**, **6a**, **6b**, **7a** and **7b**.

The reaction of **4** with putrescine mainly afforded the monomer **6a**, as supported by the NMR spectral analysis and MALDI-TOF mass spectrometry, showing $[M + H]^+$ at m/z 409.409 (theoretical $[M + H]^+ = 409.296$), while only a trace amount of dimer **6b** was obtained. Higher reaction extension was achieved when cadaverine was used, affording monomer **7a** together with the dimer **7b**. The ^1H NMR profile of **7a** follows the same profile as compound **5** while the MALDI-TOF spectrum showed the molecular ion $[M + H]^+$ at m/z 423.450 (theoretical $[M + H]^+ = 423.312$) supporting the structure **7a**. In the ^1H NMR spectrum of dimer **7b**, the presence of two equivalent xanthene rings per one cadaverine frame was confirmed by the 12 aromatic protons and the 10 aliphatic protons, which is consistent with the MALDI-TOF spectrum that showed $[M + H]^+$ at m/z 743.969 (theoretical $[M + H]^+ = 743.501$).

For comparison purposes, the photophysical properties of all isolated aminopyronins (**5**, **6a**, **7a**, **7b**) were accessed in chloroform, methanol, and acetonitrile (Table 2). All compounds presented an intense yellow color (naked eye), exhibiting absorption in the range of 430–440 nm with high extinction coefficients (10.6×10^4 – 4.35×10^4). Their emissions were in the range of 507–526 nm with large Stokes shifts (72–88 nm; 3265 – 3820 cm^{-1}), values around 5.4–6.6 times higher than those found for rosamines **1**–**4**. High fluorescence quantum yields were obtained, especially in chloroform ($\Phi_F = 0.57, 0.36, 0.58$, and 0.53 , respectively, for **5**, **6a**, **7a**, and **7b**), a fact in clear contrast with the starting rosamines **3** and **4** for which low Φ_F values were obtained in chloroform (Table 1).

Considering the possible similar reactivity of 2,3-dihydroxyphenyl substituted rosamine **3**, a similar study was performed from a solution of **3** and *n*-butylamine (15 equiv.) in methanol at 4 °C for 120 h. In this case, TLC analysis of the resulting solution showed a complex mixture of products very difficult to purify by chromatographic methods. It is very probable that, according to the generally accepted crosslinking chemistry of catechols and amines [11], the catechol unit would suffer different nucleophilic attacks of *n*-butylamine by either Michael-type addition or via the formation of a Schiff base, thus supporting the formation of many different products. However, in the case of the reaction of rosamine **4** with *n*-butylamine, 9-aminopyronin **5** was obtained as the major product of the reaction. The data suggest that the nucleophilic attack of the amine to the electrophilic 9-position of the xanthene takes place with the formation of the addition adduct, and that the loss of the catechol moiety occurs, potentially, during the isolation process.

Table 2. Photophysical properties ($\lambda_{\max \text{ abs}}$, $\lambda_{\max \text{ em}}$, ϵ , Stokes shift, and Φ_F) of aminopyronin 5, 6a, 7a, 7b in chloroform, acetonitrile, and methanol at 25 °C.

Dye.	Solvent	$\lambda_{\max \text{ abs}}$ (nm)	$\epsilon \times 10^4$ ($M^{-1} \cdot \text{cm}^{-1}$)	$\lambda_{\max \text{ em}}$ (nm)	Stokes Shift (nm) (cm^{-1})	Φ_F
5	CHCl ₃	435	5.31	507	72 (3265)	0.57
	CH ₃ CN	436	4.35	523	87 (3815)	0.10
	CH ₃ OH	438	5.43	525	87 (3783)	0.19
6a	CHCl ₃	430	7.44	507	77 (3532)	0.36
	CH ₃ CN	440	6.46	525	85 (3680)	0.17
	CH ₃ OH	439	6.75	522	83 (3622)	0.24
7a	CHCl ₃	434	9.57	512	78 (3510)	0.58
	CH ₃ CN	438	8.01	524	86 (3747)	0.11
	CH ₃ OH	438	8.91	522	84 (3674)	0.20
7b	CHCl ₃	435	10.8	508	73 (3303)	0.53
	CH ₃ CN	438	9.21	526	88 (3820)	0.16
	CH ₃ OH	440	10.6	524	84 (3643)	0.21

3. Materials and Methods

Reagents and solvents were purchased as reagent-grade and used without further purification unless otherwise stated. NMR spectra were recorded on a Bruker Avance III 400 spectrometer (Bruker BioSpin GmbH, Rheinstetten, Germany; 400.15 MHz for ¹H and 100.63 MHz for ¹³C) or on a Bruker Avance III HD 600 spectrometer (Bruker BioSpin GmbH, Rheinstetten, Germany; 600.13 MHz for ¹H and 150.92 MHz for ¹³C). Chemical shifts (δ) were reported in ppm and coupling constants (J) in Hz; internal standard was TMS. Two-dimensional ¹H/¹H correlation spectra (COSY), gradient selected ¹H/¹³C heteronuclear single quantum coherence (HSQC), and ¹H/¹³C heteronuclear multiple bond coherence (HMBC) spectra were acquired using the standard Bruker software. Flash chromatography was carried out using silica gel (230–400 mesh, Merck, Kenilworth, NJ, USA). Microwave-assisted reactions were carried out in a CEM Discovery Labmate circular single-mode cavity instrument (300 W max magnetron power output) from CEM Corporation (CEM Microwave Technology Ltd., Buckingham, UK). Reactions were performed under closed-vessel conditions. MS analysis of compounds 1, 2, and 5 were carried out by electrospray ionization (ESI) in a LTQ-Orbitrap-XL instrument (Thermo Fischer Scientific, Winsford, UK) with the following ESI source parameters: electrospray needle voltage 3.1 kV, sheath gas nitrogen 6, capillary temperature 275 °C, capillary voltage 21 V, and tube lens voltage 55 V. Ionization polarity was adjusted according to sample. Mass spectra of rosamines 3 and 4 were acquired by Unidade De Espectrometria De Masas of Santiago de Compostela and microanalyses were acquired by Unidad De Análisis Elemental of Santiago de Compostela. Mass spectra of compounds 6a and 7a,b were acquired on a Bruker UltrafleXtreme MALDI-TOF/TOF equipped with a 200 Hz smartbeam laser (Bruker Daltonik GmbH, Bremen, Germany). Electronic absorption spectra were recorded with a Varian Cary bio50 spectrophotometer (Agilent Technologies, Santa Clara, CA, USA), equipped with a Varian Cary single-cell Peltier accessory, using 1 cm path-length quartz cells. Steady-state fluorescence measurements were carried out in a Varian spectrofluorometer, model Cary Eclipse (Agilent Technologies, Santa Clara, CA, USA), equipped with a constant-temperature cell holder (Peltier single-cell holder) with 5 nm slit width for excitation and emission. All photophysical assays were performed under controlled temperature conditions (25 °C), using the maximum λ_{exc} and the appropriate λ_{em} range, for each rosamine and considering the different solvents used. Rosamine stock solutions were prepared in anhydrous dimethylsulfoxide (DMSO) and DMSO percentage was kept under 1% regarding the reported studies with different solvents. Rhodamine B (RhB) was used as standard in the photophysical assays and the respective solutions were prepared in anhydrous ethanol. Solutions of the compounds, in appropriate concentration ranges,

were prepared previously to molar absorptivity coefficients (ϵ) and fluorescence quantum yield (Φ_F) determination assays. Generally, fluorescence quantum yield determination was done combining the literature-described [36,37], pH variation study was carried out from a stock solution of rosamine 4 in water, and dilutions were performed with ionic force adjustment (0.1 mol/L NaCl). Each pH adjustment from 2–12 was obtained by the addition of small amounts of concentrated HCl and NaOH, followed by UV–Vis and fluorescence measurements between 200–800 nm or 555–700 nm for UV–Vis and fluorescence, respectively.

Synthesis of benzylated precursors: To a solution of 2,3-dihydroxybenzaldehyde or 3,4-dihydroxybenzaldehyde (1.00 g, 7.24 mmol) and K_2CO_3 (2.20 g, 15.9 mmol) in DMF (20 mL) at 0 °C, benzyl bromide (1.89 mL, 15.9 mmol) was added drop-by-drop, under N_2 atmosphere. After stirring at 0 °C for 15 min, the reaction mixture was allowed to warm up to room temperature and the stirring was maintained for 4 h. After that time, the reaction mixture was precipitated into an ice/water mixture and neutralized using citric acid. The white solid obtained was filtered, washed with water, dissolved in chloroform, and crystallized in *n*-hexane to afford the respective products.

2,3-Dibenzyloxybenzaldehyde: Yield 75% (1.73 g). 1H NMR (400 MHz, $CDCl_3$) δ : 5.19 and 5.20 (4H, 2s, $2 \times CH_2C_6H_5$), 7.12 (1H, dt, *J* 8.0 Hz and *J* 0.6 Hz, H-Ar), 7.23–7.26, 7.32–7.43 and 7.47–7.49 (12H, 3m, H-Ar), 10.26 (1H, s, CHO). ^{13}C NMR ($CDCl_3$, 100.62 MHz) δ : 71.3 (CH_2), 76.5 (CH_2), 119.6, 119.9, 124.2, 127.6, 128.3, 128.5, 128.6, 128.7, 128.8, 130.5, 136.3, 151.5, 152.1, 190.2 (CHO). MS (EI) *m/z*: 318 M^+ . $C_{21}H_{18}O_3 \frac{1}{4} H_2O$ calcd. C 78.12, H 5.78; found C 78.46, H 5.34.

3,4-Dibenzyloxybenzaldehyde: Yield 84% (1.94 g). 1H NMR (400 MHz, $CDCl_3$) δ : 5.22 and 5.26 (4H, 2s, $2 \times CH_2C_6H_5$), 7.02 (1H, d, *J* 8.0 Hz, H-5), 7.32–7.49 (12H, m, H-Ar), 9.81 (1H, s, CHO) ppm. HRMS (ESI) *m/z*: calcd. for $C_{21}H_{19}O_3^+$, 319.129; found, 319.133.

3.1. Synthesis of Rosamines 1 and 2

(a) Via conventional heating using propionic acid as solvent: a solution of 3-(diethylamino)phenol (0.28 g, 1.72 mmol) with the appropriate benzaldehyde (0.27 g, 0.86 mmol) and *p*-TsOH (17.2 mg, 0.10 mmol) in propionic acid (10 mL) was placed in a reaction flask and heated to 65 °C for 16 h. After cooling to room temperature, the mixture was neutralized with NaOAc (3 mol/L, 100 mL). The resulting suspension was extracted with chloroform. The combined organic extracts were dried (Na_2SO_4 anhydrous) and the solvent evaporated. The resulting residue was dissolved in 40 mL of a mixture of $CHCl_3$ /MeOH (1:1), to which chloranil (0.10 g, 0.43 mmol) was added. The mixture was vigorously stirred for 2 h and concentrated in vacuum. The residue was purified by flash chromatography using a mixture of $CHCl_3$ /MeOH (9:1) and vacuum dried.

(b) Via MW using propionic acid or water as solvent: a solution of 3-(diethylamino)phenol (0.14 g, 0.86 mmol) with the appropriate benzaldehyde (0.13, 0.43 mmol) and *p*-TsOH (10.0 mg, 0.06 mmol) in water or propionic acid (5 mL) was placed in a 10 mL reaction vial, which was then sealed and placed in the cavity of a CEM microwave reactor. The reaction was irradiated at 80 °C (1 min ramp to 80 °C and 10 min hold at 80 °C, using 100 W maximum power). The solvent was decanted and the resulting solid was dissolved in 10 mL of a mixture of $CHCl_3$ /MeOH (1:1), to which chloranil (0.10 g, 0.43 mmol) was added. The mixture was placed into the cavity of the CEM microwave using 1 min ramp to 60 °C and 10 min hold at 60 °C, using 50 W maximum power. The residue was purified by flash chromatography using a mixture of $CHCl_3$ /MeOH (9:1) and vacuum dried.

Rosamine 1: (a) Yield 36% (95 mg); (b) Yields 43% (113 mg) and 47% (124 mg) in propionic acid and water, respectively. 1H NMR (400 MHz, $CDCl_3$) δ : 1.33 (12H, t, *J* 7.2 Hz, $4 \times CH_3$), 3.64 (8H, q, *J* 7.2 Hz, $4 \times CH_2$), 4.88 (2H, s, $2'$ - $CH_2C_6H_5$), 5.27 (2H, s, $3'$ - $CH_2C_6H_5$), 6.74–6.76 (3H, m, H-Ar), 6.77 (2H, d, *J* 2.4 Hz, H-4 and H-5), 6.82 (2H, dd, *J* 9.4 and *J* 2.4 Hz, H-2 and H-7), 6.93–7.03 (3H, m, H-Ar), 7.19 (2H, d, *J* 9.4 Hz, H-1 and H-8), 7.23–7.27 and 7.36–7.52 (7H, 2m, H-Ar) ppm. ^{13}C NMR (100 MHz, $CDCl_3$) δ : 12.7 (CH_3), 46.2 (CH_2), 71.2 ($3'$ - $CH_2C_6H_5$), 75.2 ($2'$ - $CH_2C_6H_5$), 96.2 (C-4 and C-5), 113.7, 114.0, 116.0 (C-2 and C-7),

122.4, 124.8, 126.7, 127.6, 127.8, 128.0, 128.4, 128.7, 132.2 (C-1 and C-8), 136.2, 136.6, 145.5 (C-2'), 152.3 (C-3'), 155.1 (C-9), 155.4 (C-3 and C-6), 157.7 (C-4a and C-5a) ppm. HRMS (ESI) m/z : calcd. for $C_{41}H_{43}N_2O_3^+$, 611.327; found 611.326.

Rosamine 2: (a) Yield 12% (32 mg); (b) Yields 61% (160 mg) and 68% (179 mg) in propionic acid and water, respectively. 1H NMR (400 MHz, $CDCl_3$) δ : 1.32 (12H, t, J 7.2 Hz, $4 \times CH_3$), 3.63 (8H, q, J 7.2 Hz, $4 \times CH_2$), 5.24 (2H, s, $3'-CH_2C_6H_5$), 5.31 (2H, s, $4'-CH_2C_6H_5$), 6.77 (2H, d, J 2.4 Hz, H-4 and H-5), 6.80 (2H, dd, J 9.6 and J 2.4 Hz, H-2 and H-7), 6.89 (1H, d, J 2.0 Hz, H-2'), 6.92 (1H, dd, J 8.0 and J 2.0 Hz, H-6'), 7.17 (1H, d, J 8.0 Hz, H-5'), 7.27 (2H, d, J 9.6 Hz, H-1 and H-8), 7.31–7.45 (8H, m, H-Ar), 7.53 (2H, d, J 7.2 Hz, H-Ar) ppm. ^{13}C NMR (100 MHz, $CDCl_3$) δ : 12.8 (CH_3), 46.2 (CH_2), 71.2 and 71.3 ($CH_2C_6H_5$), 96.5 (C-4 and C-5), 113.2 (C-1a and C-8a), 114.1 (C-2 and C-7), 114.4 (C-5'), 116.9 (C-2'), 123.8 (C-6'), 124.2 (C-1'), 127.37, 127.41, 128.0, 128.2, 128.76, 128.79, 132.2 (C-1 and C-8), 136.6, 136.7, 148.4 (C-3'), 151.1 (C-4'), 155.4 (C-3 and C-6), 157.1 (C-9), 158.0 (C-4a and C-5a) ppm. HRMS (ESI) m/z : calcd. for $C_{41}H_{43}N_2O_3^+$, 611.327; found, 611.326.

3.2. Synthesis of Rosamines 3 and 4

A 1 mol/L solution of boron trichloride in dichloromethane (2 mL) was dropped slowly into an ice-bath cooled suspension of rosamine 1 or 2 (85.6 mg, 0.14 mmol) in dry dichloromethane (8 mL) under a N_2 atmosphere. The mixture was stirred at room temperature for 18 h. Methanol (20 mL) was added to stop the reaction, followed by several washings with acetone and methanol. After removal of the solvent in vacuum, the residue was precipitated with methanol/acetone.

Rosamine 3: Yield 41% (25 mg). 1H NMR (400 MHz, $DMSO-d_6$) δ : 1.21 (12H, t, J 7.2 Hz, $4 \times CH_3$), 3.65 (8H, q, J 6.8 Hz, $4 \times CH_2$), 6.66 (1H, d, J 7.6 Hz, H-6'), 6.89 (1H, dd, J 8.0 and J 7.6 Hz, H-5'), 6.96 (2H, d, J 2.2 Hz, H-4 and H-5), 7.07 (1H, d, J 8.0 Hz, H-4'), 7.14 (2H, dd, J 9.4 and J 2.2 Hz, H-2 and H-7), 7.26 (2H, d, J 9.4 Hz, H-1 and H-8), 9.00 (1H, s, OH), 9.96 (1H, s, OH) ppm. ^{13}C NMR (100 MHz, $DMSO-d_6$) δ : 12.5 (CH_3), 45.3 (CH_2), 95.8 (C-4 and C-5), 113.0 (C-1a and C-8a), 114.3 (C-2 and C-7), 117.2 (C-4'), 119.3 (C-1'), 119.5 (C-5'), 120.4 (C-6'), 131.9 (C-1 and C-8), 143.0 (C-2'), 145.8 (C-3'), 155.1 (C-3 and C-6), 155.6 (C-9), 157.4 (C-4a and C-5a) ppm. MS (ESI) m/z : 431.2 M^+ . $C_{27}H_{31}ClN_2O_3 \cdot 3/2CH_2Cl_2$ calcd. C, 57.59, H, 5.77, N, 4.71; found C, 58.07, H, 5.46, N, 4.75.

Rosamine 4: Yield 91% (55 mg). 1H NMR (400 MHz, $DMSO-d_6$) δ : 1.22 (12H, t, J 7.2 Hz, $4 \times CH_3$), 3.65 (8H, q, J 6.8 Hz, $4 \times CH_2$), 6.79 (1H, dd, J 8.0 and J 2.1 Hz, H-6'), 6.94 (3H, d, J 2.1 Hz, H-4, H-5 and H-2'), 7.05 (1H, d, J 8.0 Hz, H-5'), 7.16 (2H, dd, J 9.8 and J 2.1 Hz, H-2 and H-7), 7.47 (2H, d, J 9.8 Hz, H-1 and H-8), 9.77 (2H, s broad, $2 \times OH$) ppm. ^{13}C NMR (100 MHz, $DMSO-d_6$) δ : 12.5 (CH_3), 45.3 (CH_2), 95.9 (C-4 and C-5), 112.5 (C-1a and C-8a), 114.2 (C-2 and C-7), 116.0 (C-5'), 117.3 (C-2'), 121.7 (C-6'), 122.3 (C-1'), 132.1 (C-1 and C-8), 145.6 (C-3'), 147.8 (C-4'), 154.9 (C-3 and C-6), 157.3 (C-9), 157.4 (C-4a and C-5a) ppm. MS (ESI) m/z : 431.2 M^+ . $C_{27}H_{31}ClN_2O_3 \cdot \frac{1}{2}CH_2Cl_2$ calcd. C, 64.83, H, 6.33, N, 5.50; found C, 64.41, H, 6.28, N, 5.42.

3.3. Reaction of Rosamine 4 with Amines

(a) Protocol at 4 °C with *n*-butylamine: To an ice-bath cooled solution of rosamine 4 (9.9 mg, 23 μ mol) in chloroform (500 μ L), *n*-butylamine (34.3 μ L, 0.35 mol, 15 equiv.) was added dropwise and the mixture was stirred at 0 °C for 2 h. Afterward, the reaction mixture was placed in the refrigerator for 22 h. After that time, the reaction mixture was washed with diethyl ether to remove the *n*-butylamine excess and purified by preparative TLC using a chloroform/methanol mixture (9:1). Amino-pyronin 5 was obtained as a yellow solid (24% yield, 2.2 mg), whereas initial rosamine 4 (22%, 2.4 mg) was recovered. A similar procedure was performed using methanol as the solvent, with an extended reaction time to six days. Amino-pyronin 5 was obtained as a yellow solid (53% yield, 4.8 mg), whereas no rosamine 4 was recovered. A similar experiment using rosamine 3 in chloroform at 4 °C was performed but, in this case, a complex mixture of products was obtained.

(b) Protocol under MW with *n*-butylamine: A solution of rosamine **4** (9.9 mg, 23 mmol) in NMP (*N*-methylpyrrolidine, 0.1 mL) was placed in a 10 mL reaction vial, to which *n*-butylamine (34.0 μ L, 0.35 mol, 15 equiv.) was added. The vial was then sealed and placed in the cavity of a CEM microwave reactor. The reaction was irradiated at 200 °C (1 min ramp to 200 °C and 35 min hold at 200 °C, using 150 W maximum power). The reaction was controlled by TLC at 5, 20, and 35 min. The resulting mixture was purified by preparative TLC using a mixture of chloroform/methanol (8:2) and vacuum dried, affording amino-pyronin **5** (10% yield, 0.9 mg), whereas rosamine **4** (7% yield, 0.7 mg) was recovered.

Amino-pyronin **5**: (a) Yield 53% (4.8 mg) and (b) Yield 10% (0.9 mg) ^1H NMR (600 MHz, MeOD- d_4) δ : 1.05 (3H, t, J 7.4 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_3$), 1.29 (12H, t, J 6.8 Hz, $4\times\text{CH}_2\text{CH}_3$), 1.55 (2H, q, J 7.4 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_3$), 1.93 (2H, t, J 7.4 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_3$), 3.60 (8H, q, J 6.8 Hz, $4\times\text{CH}_2\text{CH}_3$), 4.02 (2H, t, J 7.4 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_3$), 6.73 (2H, s broad, H-4 and H-5), 6.98 (2H, dd, J 9.0 and J 1.8 Hz, H-2 and H-7), 8.12 (2H, d, J 8.4 Hz, H-1 and H-8) ppm. ^{13}C NMR (150 MHz, MeOD- d_4) δ : 11.3 ($\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_3$), 12.6 (CH_2CH_3), 19.7 and 31.2 and 47.2–48.2 (CH_2 , where one of the signals is under the signal of MeOD- d_4), 44.5 (CH_2CH_3), 67.7, 81.7, 96.2 (C-4 and C-5), 110.7 (C-2 and C-7), 128.5, 153.4, 154.6. HRMS (ESI) m/z : calcd. for $\text{C}_{25}\text{H}_{36}\text{N}_3\text{O}^+$, 394.3285; found, 394.285.

(c) Protocol at 4 °C with putrescine: To an ice-bath solution of rosamine **4** (10.2 mg, 2.4×10^{-2} mol) in chloroform (500 μ L), putrescine (17.8 μ L, 0.18 mol, 7.5 equiv.) was added dropwise and the mixture was stirred at 0 °C for 2 h. Then, the reaction mixture was placed in the refrigerator for 22 h. After that time, the reaction mixture was washed with diethyl ether to remove the amine excess and purified by preparative TLC using a chloroform/methanol mixture (9:1), to give amino-pyronin **6a** (26% yield, 2.5 mg) as a yellow solid, and only trace amount of **6b** (dimer).

Amino-pyronin **6a**: Yield 26% (2.5 mg) ^1H NMR (600 MHz, MeOD- d_4) δ : 1.28 (12H, t, J 6.8 Hz, $4\times\text{CH}_2\text{CH}_3$), 1.83 and 2.01 (4H, 2t, J 7.2 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 3.02 (2H, t, J 7.2 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 3.59 (8H, q, J 7.2 Hz, $4\times\text{CH}_2\text{CH}_3$), 4.08 (2H, t, J 7.2 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 6.73 (2H, s broad, H-4 and H-5), 6.97 (2H, dd, J 9.3 and J 2.4 Hz, H-2 and H-7), 8.13 (2H, d, J 9.3 Hz, H-1 and H-8) ppm. MS (MALDI) m/z : calcd for $\text{C}_{25}\text{H}_{37}\text{N}_4\text{O}^+$, 409.296; found, 409.409.

(d) Protocol at 4 °C with cadaverine: To an ice-bath cooled solution of rosamine **4** (10.2 mg, 2.4×10^{-2} mol) in chloroform (500 μ L), cadaverine (20.8 μ L, 0.18 mol, 7.5 equiv.) was added dropwise and the mixture was stirred at 0 °C for 2 h. Afterward, the reaction mixture was placed in the refrigerator for 22 h. After that time, the reaction mixture was washed with diethyl ether to remove the amine excess and purified by preparative TLC using a chloroform/methanol mixture (9:1) to give amino-pyronin **7a** (15% yield, 1.5 mg) and **7b** (0.6 mg, 3%).

Amino-pyronin **7a**: Yield 15% (1.5 mg) ^1H NMR (600 MHz, MeOD- d_4) δ : 1.29 (12H, t, J 6.8 Hz, $4\times\text{CH}_2\text{CH}_3$), 1.60, 1.78 and 2.01 (6H, 3t, J 7.3 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 2.99 (2H, t, J 7.3 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}_2$), 3.61 (8H, q, J 6.8 Hz, $4\times\text{CH}_2\text{CH}_3$), 4.06 (2H, t, J 7.3 Hz, $\text{NHCH}_2(\text{CH}_2)_4\text{NH}_2$), 6.74 (2H, s broad, H-4 and H-5), 6.99 (2H, dd, J 9.0 and J 2.1 Hz, H-2 and H-7), 8.15 (2H, d, J 9.0 Hz, H-1 and H-8) ppm. MS (MALDI) m/z : calcd. for $\text{C}_{26}\text{H}_{39}\text{N}_4\text{O}^+$, 423.312; found, 423.450.

Amino-pyronin **7b**: Yield 3% (0.6 mg) ^1H NMR (400 MHz, MeOD- d_4) δ : 1.26 (24H, t, J 7.2 Hz, $8\times\text{CH}_2\text{CH}_3$), 1.71 (2H, t, J 7.3 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}$), 2.04 (4H, t, J 7.3 Hz, $\text{NHCH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{NH}$), 3.57 (16H, q, J 7.2 Hz, $8\times\text{CH}_2\text{CH}_3$), 4.05 (4H, t, J 7.3 Hz, $\text{NHCH}_2(\text{CH}_2)_3\text{CH}_2\text{NH}_2$), 6.66 (4H, d, J 2.5 Hz, H-4 and H-5), 6.89 (4H, dd, J 9.6 and J 2.5 Hz, H-2 and H-7), 8.07 (4H, d, J 9.6 Hz, H-1 and H-8) ppm. MS (MALDI) m/z : calcd. for $\text{C}_{47}\text{H}_{63}\text{N}_6\text{O}_2^+$, 743.501; found, 743.969.

3.4. Sensing Study of Rosamine **4** with Different Biogenic Amines

(a) Preliminary studies in solution. Nine samples were prepared from 500 μ L of rosamine **4** dissolved in acetonitrile with a concentration of 10^{-5} M. Then, 2, 4, and up

to 8 equiv. of 5 μ L solutions of biogenic amines (1-histamine, 2-tyramine, 3-cadaverine, 4-putrescine, 5-phenylethylamine, 6-spermidine, 7-spermine, 8-*n*-butylamine, and 9-rosamine **4**) dissolved in water were added. The changes were observed under visible and UV light and registered.

(b) UV-Vis and fluorescence studies in solution. A stock solution of rosamine **4** in DMSO was diluted to 35 μ M in acetonitrile. Then 0.1, 0.5, 1.0, 2.0, 4.0, and 8.0 equiv. of amine solutions (*n*-butylamine, *t*-butylamine, cadaverine, and putrescine) in acetonitrile were added. UV-Vis and fluorescence spectra were recorded in the appropriate concentration and wavelength ranges between 225 and 800 nm (absorbance) and 561–800 nm (emission; $\lambda_{\text{exc}} = 551$ nm, 5 nm slit width for excitation and emission and 650 V) at 25 °C.

(c) Studies in gas phase. The study was carried out using 2.5 mL of rosamine **4** dissolved in acetonitrile and with a concentration of 5 μ M. To dispose the amines in the gas phase, a small amount of each commercial biogenic amine (cadaverine, spermidine, spermine, phenylethylamine, histamine, tyramine, tryptamine, and putrescine) was placed in 10 mL vials, closed with a septum, and heated on a heating plate, as shown in Figure S32, at different temperatures depending on the boiling point of each amine. The vapor produced was transferred with a syringe and bubbled into a cuvette containing 2.5 mL of **4**. Afterward, the measurement was carried out in the spectrophotometer at 533 nm. The same procedure was carried out using nitrogen, air, and water vapor to check any putative effects on probe **4**.

4. Conclusions

Two catechol-derived rosamine dyes were synthesized through a simple and straightforward strategy involving the microwave-assisted synthesis of rosamines **1** and **2**, with subsequent debenzylation using boron trichloride to give derivatives **3** and **4**. Significant changes in the UV-Vis spectra were observed for the catechol derived rosamine **4** toward biogenic amines (BAs), especially for putrescine, where two new bands appeared at 512 and 649 nm and a quench was observed at 551 nm. The highest fluorescence intensity quenching was observed for the addition of 8 equiv. of putrescine (55%).

Furthermore, we found that methanolic solutions of rosamine **4** and *n*-butylamine showed an expressive color change over time, which was attributed to the formation of the 9-aminopyronin **5**. Different parameters including solvent, temperature, and reaction time influence the outcome of this transformation; the best yield was obtained in methanol at 4 °C for ca. 144 h, affording **5** in 53% yield. Other 9-aminopyronins **6a**, **6b**, **7a**, and **7b** were obtained from methanolic solutions of **4** with putrescine and cadaverine. All 9-aminopyronins exhibited exceptional spectroscopic properties with blue shifts in absorption and emission spectra as well as a high increase in the fluorescence quantum yield, comparatively to the pristine rosamine **4**.

The present results corroborate the high reactivity of the catechol derivative **4** toward amines, and hint that this rosamine can be considered in the future as a biogenic amine sensor in solution (acetonitrile) and in the gas phase.

Supplementary Materials: The following are available online: MS, ^1H , ^{13}C , and 2D NMR spectra of the new compounds; additional UV-Vis and fluorescence measurements including the fluorescence intensity of **4** with pH variation and detection of biogenic amines in solution and in the gas phase.

Author Contributions: F.M.-S. and C.Q. carried out the synthesis and structural characterization of all compounds. M.T.R., M.J.R. and T.T. performed the sensing studies with biogenic amines. A.L. performed the analyses of the physicochemical studies, R.C.M. reviewed the manuscript and content, M.R. analyzed the physicochemical and sensing studies, and A.M.G.S. designed and supervised the synthetic work and manuscript writing. All authors have read and agreed to the published version of the manuscript.

Funding: This work received financial support from PT national funds (FCT/MCTES, Fundação para a Ciência e a Tecnologia and Ministério da Ciência, Tecnologia e Ensino Superior) through the projects UIDB/50006/2020, PTDC/QUI-QIN/28142/2017, and PTDC/QUI-QOR/29426/2017. This

work also received financial support from the Junta de Castilla y León, Consejería de Educación y Cultura y Fondo Social Europeo (Grant BU263P18). F. M. -S. gratefully acknowledges the financial support provided by FCT (Portugal's Foundation for Science and Technology) within grant DFA/BD/9136/2020. A.M.G.S. and A.L. thank FCT for funding through program DL 57/2016 – Norma transitória.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are contained within the article and are also available in the Supplementary Materials.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of compounds 1–7 are available from the authors.

References

1. Yang, J.; Cohen Stuart, M.A.; Kamperman, M. Jack of all trades: Versatile catechol crosslinking mechanisms. *Chem. Soc. Rev.* **2014**, *43*, 8271–8298. [[CrossRef](#)] [[PubMed](#)]
2. Kim, J.; Lee, C.; Ryu, J.H. Adhesive Catechol-Conjugated Hyaluronic Acid for Biomedical Applications: A Mini Review. *Appl. Sci.* **2020**, *11*, 21. [[CrossRef](#)]
3. Katir, N.; Marcotte, N.; Michlewska, S.; Ionov, M.; El Brahm, N.; Bousmina, M.; Majoral, J.P.; Bryszewska, M.; El Kadib, A. Dendrimer for Templating the Growth of Porous Catechol-Coordinated Titanium Dioxide Frameworks: Toward Hemocompatible Nanomaterials. *ACS Appl. Nano Mater.* **2019**, *2*, 2979–2990. [[CrossRef](#)]
4. Sedó, J.; Saiz-Poseu, J.; Busqué, F.; Ruiz-Molina, D. Catechol-Based Biomimetic Functional Materials. *Adv. Mater.* **2013**, *25*, 653–701. [[CrossRef](#)]
5. Joshi, S.; Kathuria, H.; Verma, S.; Valiyaveetil, S. Functional Catechol-Metal Polymers via Interfacial Polymerization for Applications in Water Purification. *ACS Appl. Mater. Interfaces* **2020**, *12*, 19044–19053. [[CrossRef](#)]
6. Zhang, W.; Wang, R.; Sun, Z.; Zhu, X.; Zhao, Q.; Zhang, T.; Cholewinski, A.; Yang, F.K.; Zhao, B.; Pinnaratip, R.; et al. Catechol-functionalized hydrogels: Biomimetic design, adhesion mechanism, and biomedical applications. *Chem. Soc. Rev.* **2020**, *49*, 433–464. [[CrossRef](#)]
7. Razaviamri, S.; Wang, K.; Liu, B.; Lee, B.P. Catechol-Based Antimicrobial Polymers. *Molecules* **2021**, *26*, 559. [[CrossRef](#)]
8. Maier, G.P.; Bernt, C.M.; Butler, A. Catechol oxidation: Considerations in the design of wet adhesive materials. *Biomater. Sci.* **2018**, *6*, 332–339. [[CrossRef](#)]
9. Pinnataip, R.; Lee, B.P. Oxidation Chemistry of Catechol Utilized in Designing Stimuli-Responsive Adhesives and Antipathogenic Biomaterials. *ACS Omega* **2021**, *6*, 5113–5118. [[CrossRef](#)] [[PubMed](#)]
10. Ito, S.; Sugumaran, M.; Wakamatsu, K. Chemical Reactivities of ortho-Quinones Produced in Living Organisms: Fate of Quinonoid Products Formed by Tyrosinase and Phenoloxidase Action on Phenols and Catechols. *Int. J. Mol. Sci.* **2020**, *21*, 6080. [[CrossRef](#)]
11. Yang, J.; Saggiomo, V.; Velders, A.H.; Cohen Stuart, M.A.; Kamperman, M. Reaction Pathways in Catechol/Primary Amine Mixtures: A Window on Crosslinking Chemistry. *PLoS ONE* **2016**, *11*, e0166490. [[CrossRef](#)] [[PubMed](#)]
12. Timmer, B.; Olthuis, W.; van den Berg, A. Ammonia sensors and their applications—A review. *Sens. Actuators B Chem.* **2005**, *107*, 666–677. [[CrossRef](#)]
13. Danchuk, A.I.; Komova, N.S.; Mobarez, S.N.; Doronin, S.Y.; Burmistrova, N.A.; Markin, A.V.; Duerkop, A. Optical sensors for determination of biogenic amines in food. *Anal. Bioanal. Chem.* **2020**, *412*, 4023–4036. [[CrossRef](#)] [[PubMed](#)]
14. Wójcik, W.; Łukasiewicz, M.; Puppel, K. Biogenic amines: Formation, action and toxicity—A review. *J. Sci. Food Agric.* **2021**, *101*, 2634–2640. [[CrossRef](#)] [[PubMed](#)]
15. Saravanakumar, M.; Umamahesh, B.; Selvakumar, R.; Dhanapal, J.; Ashok kumar, S.K.; Sathiyarayanan, K.I. A colorimetric and ratiometric fluorescent sensor for biogenic primary amines based on dicyanovinyl substituted phenanthridine conjugated probe. *Dyes Pigm.* **2020**, *178*, 108346. [[CrossRef](#)]
16. Mallick, S.; Chandra, F.; Koner, A.L. A ratiometric fluorescent probe for detection of biogenic primary amines with nanomolar sensitivity. *Analyst* **2016**, *141*, 827–831. [[CrossRef](#)]
17. Bao, C.; Shao, S.; Zhou, H.; Han, Y. A New ESIPT-Based Fluorescent Probe for the Highly Sensitive Detection of Amine Vapors. *New J. Chem.* **2021**, *45*, 10735–10740. [[CrossRef](#)]
18. Castillero, P.; Roales, J.; Lopes-Costa, T.; Sánchez-Valencia, J.; Barranco, A.; González-Elipe, A.; Pedrosa, J. Optical Gas Sensing of Ammonia and Amines Based on Protonated Porphyrin/TiO₂ Composite Thin Films. *Sensors* **2016**, *17*, 24. [[CrossRef](#)] [[PubMed](#)]
19. Sousaraei, A.; Queirós, C.; Moscoso, F.G.; Silva, A.M.G.; Lopes-Costa, T.; Pedrosa, J.M.; Cunha-Silva, L.; Cabanillas-Gonzalez, J. Reversible Protonation of Porphyrinic Metal-Organic Frameworks Embedded in Nanoporous Polydimethylsiloxane for Colorimetric Sensing. *Adv. Mater. Interfaces* **2021**, *8*, 2001759. [[CrossRef](#)]
20. Ikeda, M.; Yoshii, T.; Matsui, T.; Tanida, T.; Komatsu, H.; Hamachi, I. Montmorillonite–Supramolecular Hydrogel Hybrid for Fluorocolorimetric Sensing of Polyamines. *J. Am. Chem. Soc.* **2011**, *133*, 1670–1673. [[CrossRef](#)]

21. Li, Z.; Song, Y.; Lu, Z.; Li, Z.; Li, R.; Li, Y.; Hou, S.; Zhu, Y.P.; Guo, H. Novel difluoroboron complexes of curcumin analogues as “dual-dual” sensing materials for volatile acid and amine vapors. *Dyes Pigm.* **2020**, *179*, 108406. [[CrossRef](#)]
22. VanDenburgh, K.L.; Liu, Y.; Sadhukhan, T.; Benson, C.R.; Cox, N.M.; Erbas-Cakmak, S.; Qiao, B.; Gao, X.; Pink, M.; Raghavachari, K.; et al. Multi-state amine sensing by electron transfers in a BODIPY probe. *Org. Biomol. Chem.* **2020**, *18*, 431–440. [[CrossRef](#)]
23. Gupta, A. Aggregation-Induced Emission: A Tool for Sensitive Detection of Amines. *ChemistrySelect* **2019**, *4*, 12848–12860. [[CrossRef](#)]
24. Queirós, C.; Silva, A.M.G.; Lopes, S.C.; Ivanova, G.; Gameiro, P.; Rangel, M. A novel fluorescein-based dye containing a catechol chelating unit to sense iron(III). *Dyes Pigm.* **2012**, *93*, 1447–1455. [[CrossRef](#)]
25. Queirós, C.; Leite, A.; Couto, M.G.M.; Cunha-Silva, L.; Barone, G.; de Castro, B.; Rangel, M.; Silva, A.M.N.; Silva, A.M.G. The Influence of the Amide Linkage in the Fe III -Binding Properties of Catechol-Modified Rosamine Derivatives. *Chem. Eur. J.* **2015**, *21*, 15692–15704. [[CrossRef](#)] [[PubMed](#)]
26. Leite, A.; Cunha-Silva, L.; Silva, D.; Lobo Ferreira, A.I.M.C.; Santos, L.M.N.B.F.; Cardoso, I.C.S.; Silva, V.L.M.; Rangel, M.; Silva, A.M.G. Synthesis of Pyridyl and N-Methylpyridinium Analogues of Rosamines: Relevance of Solvent and Charge on Their Photophysical Properties. *Chem. Eur. J.* **2019**, *25*, 15073–15082. [[CrossRef](#)]
27. Wu, L.; Burgess, K. Synthesis and spectroscopic properties of rosamines with cyclic amine substituents. *J. Org. Chem.* **2008**, *73*, 8711–8718. [[CrossRef](#)]
28. Savarese, M.; Aliberti, A.; De Santo, I.; Battista, E.; Causa, F.; Netti, P.A.; Rega, N. Fluorescence lifetimes and quantum yields of rhodamine derivatives: New insights from theory and experiment. *J. Phys. Chem. A* **2012**, *116*, 7491–7497. [[CrossRef](#)]
29. García-Tojal, J.; Cuevas, J.V.; Rojo, M.J.; Díaz de Greñu, B.; Hernando-Muñoz, C.; García-Calvo, J.; Salgado, M.M.; Torroba, T. Synthesis of fluorogenic arylureas and amides and their interaction with amines: A competition between turn-on fluorescence and organic radicals on the way to a smart label for fish freshness. *Molecules* **2021**, *26*, 1404. [[CrossRef](#)]
30. Wu, L.; Burgess, K. Fluorescent amino- and thiopyronin dyes. *Org. Lett.* **2008**, *10*, 1779–1782. [[CrossRef](#)] [[PubMed](#)]
31. Song, Y.; Zhang, H.; Wang, X.; Geng, X.; Sun, Y.; Liu, J.; Li, Z. One stone, three birds: PH triggered transformation of aminopyronine and iminopyronine based lysosome targeting viscosity probe for cancer visualization. *Anal. Chem.* **2021**, *93*, 1786–1791. [[CrossRef](#)] [[PubMed](#)]
32. Zhang, H.; Liu, J.; Sun, Y.Q.; Huo, Y.; Li, Y.; Liu, W.; Wu, X.; Zhu, N.; Shi, Y.; Guo, W. A mitochondria-targetable fluorescent probe for peroxynitrite: Fast response and high selectivity. *Chem. Commun.* **2015**, *51*, 2721–2724. [[CrossRef](#)] [[PubMed](#)]
33. Sun, Y.Q.; Liu, J.; Zhang, H.; Huo, Y.; Lv, X.; Shi, Y.; Guo, W. A mitochondria-targetable fluorescent probe for dual-channel no imaging assisted by intracellular cysteine and glutathione. *J. Am. Chem. Soc.* **2014**, *136*, 12520–12523. [[CrossRef](#)] [[PubMed](#)]
34. Horváth, P.; Šebej, P.; Kovář, D.; Damborský, J.; Prokop, Z.; Klán, P. Fluorescent pH Indicators for Neutral to Near-Alkaline Conditions Based on 9-Iminopyronin Derivatives. *ACS Omega* **2019**, *4*, 5479–5485. [[CrossRef](#)]
35. Guo, X.; Wei, X.R.; Sun, R.; Xu, Y.J.; Chen, Y.; Ge, J.F. The optical properties of 9-amino-9H-xanthene derivatives in different pH and their application for biomarkers in lysosome and mitochondria. *Sens. Actuators B Chem.* **2019**, *296*, 126621. [[CrossRef](#)]
36. Williams, A.T.R.; Winfield, S.A.; Miller, J.N. Relative fluorescence quantum yields using a computer-controlled luminescence spectrometer. *Analyst* **2004**, *108*, 1067. [[CrossRef](#)]
37. Fery-Forgues, S.; Lavabre, D. Are Fluorescence Quantum Yields So Tricky to Measure? A Demonstration Using Familiar Stationery Products. *J. Chem. Educ.* **2009**, *76*, 1260. [[CrossRef](#)]