



Complete Genome Sequences of Four *Parageobacillus* Strains Isolated from Soil in Japan

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ABSTRACT We isolated four *Parageobacillus* strains from soil in Japan and completely sequenced their genomes. Three of four strains showed $\geq 98.9\%$ average nucleotide identity (ANI) to *Parageobacillus caldoxylosilyticus* S1812^T, while one strain, designated KH3-4, showed the highest ANI (91%) to *Parageobacillus thermantarcticus* M1^T, suggesting the species novelty of KH3-4.

Parageobacillus is a genus of betaproteobacteria in the family *Burkholderiaceae* that is Gram-positive and a facultatively anaerobic thermophile. *Parageobacillus* species have great biotechnological potential (1), for example, as a source for thermophilic enzymes (2), fuel production (3, 4), and the bioremediation of environmental pollutants (5). At the time of writing, there are six validly named species in the genus *Parageobacillus* (<https://lpsn.dsmz.de/genus/parageobacillus>). So far, seven complete genome sequences have been reported for *Parageobacillus*, including for *Parageobacillus caldoxylosilyticus* (1 strain), *Parageobacillus thermoglucosidasius* (4 strains), and *Parageobacillus toebii* (2 strains).

We collected soil samples from the city of Tsukuba, Japan. The samples were suspended in distilled water and spread over Lennox LB agar (1.6% [wt/vol]) plates. After incubation at 65°C overnight, dozens of well-separated single colonies were isolated; colony PCR was conducted to analyze the 16S rRNA genes using a set of primers, Bac8f (C) and UN1542r (6). Among the colonies, four strains, designated KH1-5, KH1-6, KH3-4, and KH3-5, which were expected to belong to the genus *Parageobacillus*, were subjected to complete genome analysis.

To prepare the genomic DNA, cells were grown in 5 mL LB broth at 65°C for 24 h with vigorous shaking (200 rpm). The genomic DNA was purified using a blood and cell culture DNA mini kit (Qiagen). For long-read sequencing, unsheared genomic DNA (1 μ g) was treated using a short-read eliminator kit (Circulomics) to remove fragments of <10 Kbp, and a library was constructed using a ligation sequencing kit (Oxford Nanopore Technologies [ONT]). Sequencing was performed using a GridION X5 system on a FLO-MIN106 R9.41 revD flow cell (ONT). Base calling was conducted using Guppy v.4.0.11. The raw sequencing data (Table 1) were filtered (Q < 10; length, <1,000 bases) using NanoFilt v.2.7.1 (7). For short-read sequencing, a library was constructed using an MGIEasy FS PCR free DNA library prep set (MGI) with a \sim 400 to 500-bp insert. Paired-end sequencing (2 \times 150 bases) was then performed on a DNBSEQ-400 instrument (MGI). The raw sequencing data (Table 1) were filtered (Q < 30; length, <20 bases) using fastp v.0.20.1 (8). The trimmed long- and short-read data were assembled using Unicycler v.0.4.8 (9), and the assembly was polished using Pilon v.1.24 (10). Each strain contained a single circular chromosome, and KH3-5 contained one circular plasmid; the circularity was confirmed using Unicycler.

Automatic annotation was conducted using DFAST v.1.2.15 (11); the genomic features are summarized in Table 1. A JSpecies analysis (12) revealed that KH1-5, KH1-6,

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TABLE 1 Sequencing metrics for the four *Parageobacillus* strains in this study

Strain	BioSample accession no.	Chromosome or plasmid	DNBSEQ (short-read) data			GridION (long-read) data					GenBank accession no.	
			No. of paired-end reads	Total length (Mb)	SRA accession no.	No. of reads	N_{50} (bp)	Total length (Mb)	SRA accession no.	Length (bp)		GC content (%)
<i>P. caldxylosilyticus</i> KH1-5	SAMD00442691	Chromosome	7,592,538	1,139	DRR346603	136,964	5,757	552	DRR346607	3,850,765	44.3	AP025623
<i>P. caldxylosilyticus</i> KH1-6	SAMD00442692	Chromosome	7,161,769	1,074	DRR346604	179,515	9,837	1,244	DRR346608	3,850,773	44.3	AP025624
<i>Parageobacillus</i> sp. KH3-4	SAMD00442693	Chromosome	9,226,524	1,384	DRR346605	981,529	4,531	3,090	DRR346609	3,816,932	43.0	AP025627
<i>P. caldxylosilyticus</i> KH3-5	SAMD00442694	Chromosome	7,158,442	1,074	DRR346606	1,107,180	4,644	3,572	DRR346610	3,832,285	44.2	AP025625
		Plasmid (pPcaKH3-5b)								6,889	51.7	AP025626

and KH3-5 showed $\geq 98.9\%$ average nucleotide identity (ANI) to each other and to the type strain of *P. caldxylosilyticus* (strain S1812; GenBank accession number [GCF_019272935.1](https://doi.org/10.1093/bioinformatics/bty149)), while KH3-4 showed the highest ANI (91.9%) to the type strain of *P. thermantarcticus* (strain M1; [GCF_900111865.1](https://doi.org/10.1093/bioinformatics/bty560)), suggesting the species novelty of KH3-4 (95% ANI being the cutoff for the delineation of a species). For all software, default parameters were used.

Data availability. All four *Parageobacillus* strains reported in this paper are associated with BioProject accession number [PRJDB12551](https://doi.org/10.1093/bioinformatics/bty560). The BioSample accession numbers, genome sequences, and raw sequencing data are available under the accession numbers listed in Table 1.

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REFERENCES

- Hussein AH, Lisowska BK, Leak DJ. 2015. The genus *Geobacillus* and their biotechnological potential. *Adv Appl Microbiol* 92:1–48. <https://doi.org/10.1016/bs.aambs.2015.03.001>.
- De Maayer P, Brumm PJ, Mead DA, Cowan DA. 2014. Comparative analysis of the *Geobacillus* hemicellulose utilization locus reveals a highly variable target for improved hemicellulolysis. *BMC Genomics* 15:836. <https://doi.org/10.1186/1471-2164-15-836>.
- Cripps RE, Eley K, Leak DJ, Rudd B, Taylor M, Todd M, Boakes S, Martin S, Atkinson T. 2009. Metabolic engineering of *Geobacillus thermoglucosidasius* for high yield ethanol production. *Metab Eng* 11:398–408. <https://doi.org/10.1016/j.jymben.2009.08.005>.
- Zhou J, Wu K, Rao CV. 2016. Evolutionary engineering of *Geobacillus thermoglucosidasius* for improved ethanol production. *Biotechnol Bioeng* 113:2156–2167. <https://doi.org/10.1002/bit.25983>.
- Moxley E, Puerta-Fernández E, Gómez EJ, Gonzalez JM. 2019. Influence of abiotic factors temperature and water content on bacterial 2-chlorophenol biodegradation in soils. *Front Environ Sci* 7:41. <https://doi.org/10.3389/fenvs.2019.00041>.
- Miyazaki K, Sato M, Tsukuda M. 2017. PCR primer design for 16S rRNAs for experimental horizontal gene transfer test in *Escherichia coli*. *Front Bioeng Biotechnol* 5:14. <https://doi.org/10.3389/fbioe.2017.00014>.
- De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Tanizawa Y, Fujisawa T, Kaminuma E, Nakamura Y, Arita M. 2016. DFAST and DAGA: Web-based integrated genome annotation tools and resources. *Biosci Microbiota Food Health* 35:173–184. <https://doi.org/10.12938/bmfh.16-003>.
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.