



Complete Genome Sequence of *Sinorhizobium meliloti* S35m, a Salt-Tolerant Isolate from Alfalfa Rhizosphere in Soil Native to the Caucasus Region

Victoria S. Muntyan,^a Alexey M. Afonin,^b Maria E. Vladimirova,^a Alla S. Saksaganskaya,^a Emma S. Gribchenko,^b Olga Baturina,^c Marina L. Roumiantseva^a

^aAll-Russia Research Institute for Agricultural Microbiology (ARRIAM), Laboratory of Genetics and Breeding of Microorganisms, Saint Petersburg, Russia

^bAll-Russia Research Institute for Agricultural Microbiology (ARRIAM), Laboratory of Genetics of Plant-Microbe Interactions, Saint Petersburg, Russia

^cInstitute of Chemical Biology and Fundamental Medicine, Siberian Branch of the Russian Academy of Sciences (ICBFM SB RAS), Novosibirsk, Russia

ABSTRACT The genome of a symbiotically effective salt-tolerant strain, *Sinorhizobium meliloti* S35m, isolated from alfalfa rhizosphere in soil native to the Caucasus region, was sequenced. Genomic islands, prophages, and elements of a potential CRISPR/Cas I type (Cas3_0_I) system were identified in the genome.

The strain *Sinorhizobium meliloti* S35m was isolated from a root nodule of *Medicago sativa* subsp. *varia* (Martyn) Arcang. This nodule was formed on the roots of an alfalfa plant grown from a surface-sterilized seed (method from reference 1) inoculated with water extract from alfalfa rhizosphere in a soil sample from the Caucasus region (2, 3). The strain can grow on tryptone-yeast extract (TY) medium (4) with up to 0.75 M NaCl (5) and forms effective symbiosis with *Medicago sativa* subsp. *varia* var. “Vega 87” (6). The exact mechanisms behind the salt tolerance of this strain are still not fully understood.

This strain was stored in TY medium containing 20% (vol/vol) glycerol at -70°C . A single colony of S35m was grown overnight in TY broth (28°C , 180 rpm shaking). Genomic DNA (gDNA) was isolated using the phenol-chloroform extraction method and used for both the Nanopore and Illumina sequencing (7). To obtain fragments of about 600 bp, $1\ \mu\text{g}$ of gDNA was sheared in a microTUBE AFA fiber snap-cap tube Covaris S2 system. The paired-end library was produced using a NEBNext Ultra II DNA library prep kit for Illumina (New England Biolabs [NEB]) and dual-index NEBNext multiplex oligos (NEB). The library was sequenced on an Illumina MiSeq sequencer using the v3 reagent kit (2×300 bp) at the SB RAS Genomics Core Facility (ICBFM SB RAS), generating 371,608 reads. Adapter and low-quality sequences were removed using BBDuk with default parameters (8). Long-read sequencing using a MinION sequencer (Oxford Nanopore, United Kingdom) was done at the All-Russia Research Institute for Agricultural Microbiology (ARRIAM). The SQK-LSK109 kit and the 7th barcode from the EXP-NBD104 kit were used to make the library, omitting the DNA shearing. Guppy_basecaller v. 3.3.0 was used to base call and demultiplex the Nanopore reads.

The output long-read data set (N_{50} read length, 1,921 bp) was 1.5 Gb ($1.2 \cdot 10^6$ reads). The Flye pipeline v. 2.8-release (9) was used to assemble the Nanopore reads. Final assembly was polished 5 times consecutively using Racon v. 1.3.2 (10) with modifiers ($-m\ 8\ -x\ -6\ -g\ -8\ -w\ 500$), followed by a single polish using Medaka v. 1.0.3 (11) with default parameters. Short reads were used to polish the Nanopore assembly with three consecutive runs of Pilon v. 1.22 (12). The calculated coverage was $14\times$ for the Illumina reads and $221\times$ for the Nanopore reads. The 3 contigs correspond to the chromosome (3,610,844 bp; 62.8% GC content) and megaplasmids SMA (1,514,798 bp; 60.6% GC content) and SMB (1,659,814 bp; 62.2% GC content). PGAP v. 4.13 annotation

Citation Muntyan VS, Afonin AM, Vladimirova ME, Saksaganskaya AS, Gribchenko ES, Baturina O, Roumiantseva ML. 2021. Complete genome sequence of *Sinorhizobium meliloti* S35m, a salt-tolerant isolate from alfalfa rhizosphere in soil native to the Caucasus region. Microbiol Resour Announc 10:e01417-20. <https://doi.org/10.1128/MRA.01417-20>.

Editor David A. Baltrus, University of Arizona

Copyright © 2021 Muntyan et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Alexey M. Afonin, afoninalexeym@gmail.com.

Received 9 December 2020

Accepted 9 March 2021

Published 18 March 2021

(13) of all three replicons resulted in 3 *rrn-rrl* operons, 54 tRNAs, and 6,166 predicted protein-encoding open reading frames (ORFs). Prophages, genomic islands (GI), and CRISPR/Cas sequences were determined using PHASTER (14), IslandViewer (15), and CRISPR-Cas++ (16), respectively.

Two genomic islands (9 and 11 kb), an intact prophage of 54 kb similar to *Sinorhizobium* phage phiLM21 (GenBank accession number [NC_029046](#)), an incomplete prophage of 25 kb similar to *Enterobacteria* phage phi92 (GenBank accession number [NC_023693](#)), and 9 potential CRISPR cassettes with one or two spacers and four genes encoding potential Cas proteins (Cas3_0_1) similar to the CRISPR/Cas type I system were found in the genome; similarity was determined using the BLASTn algorithm (17).

Data availability. The accession numbers in NCBI are [CP065020.1](#) to [CP065022.1](#) (assemblies), [PRJNA678757](#) (BioProject), and [SAMN16812329](#) (BioSample); [SRR13084429](#) and [SRR13084428](#) are the accession numbers for raw short-read and long-read data in the NCBI SRA. This announcement is for the first version of the S35m genome assembly.

ACKNOWLEDGMENT

This work was supported by the RSF grant 20-16-00105.

REFERENCES

1. Arraouadi S, Badri M, Jaleel CA, Djébalı N, Ilahi H, Huguet T, Aouani ME. 2009. Analysis of genetic variation in natural populations of *Medicago truncatula* of southern Tunisian ecological areas, using morphological traits and SSR markers. *Trop Plant Biol* 2:122–132. <https://doi.org/10.1007/s12042-009-9034-5>.
2. Ivanov AI. 1980. Alfalfa. In Brezhnev D (ed), *Scientific works of the Academy of Agricultural Sciences*. State Publishing House, Moscow, Russia. (In Russian.)
3. Nakhutsrishvili G, Abdaladze O. 2017. Plant diversity of the Central Great Caucasus, p 17–111. In Nakhutsrishvili G, Abdaladze O, Batsatsashvili K, Spehn E, Körner C (ed), *Plant diversity in the Central Great Caucasus: a quantitative assessment*. Geobotany studies. Springer International Publishing AG, Cham, Switzerland. https://doi.org/10.1007/978-3-319-55777-9_3.
4. Beringer JE. 1974. R factor transfer in *Rhizobium leguminosarum*. *J Gen Microbiol* 84:188–198. <https://doi.org/10.1099/00221287-84-1-188>.
5. Roumiantseva ML, Saksaganskaia AS, Muntyan VS, Cherkasova ME, Simarov BV. 2018. Structural polymorphism of *Sinorhizobium meliloti* genes related to virulence and salt tolerance. *Russ J Genet* 54:525–535. (In Russian.) <https://doi.org/10.7868/S001667581805003X>.
6. Roumiantseva ML, Muntyan VS. 2015. Root nodule bacteria *Sinorhizobium meliloti*: tolerance to salinity and bacterial genetic determinants. *Microbiology* 84:303–318. (In Russian.) <https://doi.org/10.1134/S0026261715030170>.
7. Green MR, Sambrook J. 2017. Isolation of high-molecular-weight DNA using organic solvents. *Cold Spring Harb Protoc* 2017.pdb.prot093450. <https://doi.org/10.1101/pdb.prot093450>.
8. Bushnell B. 2016. BMAP short-read aligner, and other bioinformatics tools. <http://sourceforge.net/projects/bbmap/>.
9. Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>.
10. Vaser R, Sović I, Nagarajan N, Šikić M. 2017. Fast and accurate de novo genome assembly from long uncorrected reads. *Genome Res* 27:737–746. <https://doi.org/10.1101/gr.214270.116>.
11. Oxford Nanopore Technologies. 2018. Medaka. <https://github.com/nanoporetech/medaka>.
12. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
13. Tatusova T, Dicuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
14. Arndt D, Grant J, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. *Nucleic Acids Res* 44:W16–W21. <https://doi.org/10.1093/nar/gkw387>.
15. Bertelli C, Laird MR, Williams KP, Lau BY, Hoad G, Winsor GL, Brinkman FSL, Simon Fraser University Research Computing Group. 2017. IslandViewer 4: expanded prediction of genomic islands for larger-scale datasets. *Nucleic Acids Res* 45:W30–W35. <https://doi.org/10.1093/nar/gkx343>.
16. Couvin D, Bernheim A, Toffano-Nioche C, Touchon M, Michalik J, Néron B, Rocha EPC, Vergnaud G, Gautheret D, Pourcel C. 2018. CRISPRCasFinder, an update of CRISPRFinder, includes a portable version, enhanced performance and integrates search for Cas proteins. *Nucleic Acids Res* 46:W246–W251. <https://doi.org/10.1093/nar/gky425>.
17. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421–429. <https://doi.org/10.1186/1471-2105-10-421>.