





A Randomized, Double-Blind, 2-Treatment, 2-Period, Crossover Phase 1 Study to Compare the Pharmacokinetics, Safety and Tolerability of 60 IU/Kg of Abcertin and Cerezyme in Healthy Volunteers Following a Single Intravenous Administration

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ABSTRACT

Background: Imiglucerase (Cerezyme; Sanofi, Paris, France), an analogue of β -glucocerebrosidase produced by recombinant DNA technology, has been a safe and effective treatment for Gaucher disease (GD) for over 25 years. A new imiglucerase, Abcertin (Seongnam-si, Gyeonggi-do, Republic of Korea) has shown a similar safety and efficacy profile in previous clinical studies. This study compared the pharmacokinetics, immunogenicity, safety, and tolerability to EU-sourced Cerezyme following a single $60\,\mathrm{IU/kg}$ dose.

Methods: This phase 1, single-center, randomized, double-blind, two-way crossover study enrolled 36 healthy volunteers aged 18–45 years. Participants were randomly assigned to receive either Abcertin or Cerezyme in a predetermined sequence.

Results: Abcertin reached peak plasma concentrations at a median $t_{\rm max}$ of 61 min (range: 40–121 min). The mean $C_{\rm max}$, AUC_{0-last}, and AUC_{0-inf} were 115.4 mU/mL, 12,190 min·mU/mL, and 12,210 min mU/mL, respectively, indicating bioequivalence to Cerezyme. The mean $t_{\rm I/2}$, CL, and $V_{\rm z}$ were 6.88 min, 376.7 mL/min, and 3.62 L, respectively, and were comparable between the two treatments. One participant in the Cerezyme group developed anti-drug antibodies, which were non-neutralizing A total of 24 subjects experienced treatment-emergent adverse event (TEAE). The most common TEAE was headache (3 in the Abcertin group and 5 in the Cerezyme group), followed by general disorders and administration site condition (3 in Abcertin group and 5 in Cerezyme group). Two participants in the Cerezyme sequence experienced severe TEAEs: one had a urinary tract infection, and the other developed urticaria, which leading to study withdrawal.

Conclusion: Abcertin demonstrated pharmacokinetic equivalence to Cerezyme, with a comparable safety, immunogenicity, and tolerability profile. These findings support its potential as an affordable biosimilar for GD treatment.

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1 | Introduction

Gaucher disease (GD), an autosomal recessive disorder caused by a deficiency in the lysosomal enzyme glucocerebrosidase (GBA) encoded by GBA1 gene, is the most common glycolipid storage disorder, with an overall incidence of approximately 1 in 40,000 to 60,000 (Grabowski et al. 2004; Stirnemann et al. 2017). The incidence is significantly higher in individuals of Ashkenazi Jewish ancestry, affecting approximately 1 in 850 births due to a higher carrier frequency of the disease-causing gene (Horowitz et al. 1998). The accumulation of lipid glucocerebroside within the lysosomal compartment of macrophages causes functional impairment of internal organs, including the bone marrow, spleen, liver, and nervous system. The clinical phenotypes of GD are categorized into three forms, although the full disease spectrum is a continuum. Type 1 is characterized by the absence of neurological symptoms, whereas types 2 and 3 are characterized by neurological involvement (Charrow et al. 2000; Mistry et al. 2011; Tajima et al. 2009). The clinical presentation of type 1 GD varies widely, ranging from asymptomatic to severely debilitating (Biegstraaten et al. 2008; Tajima et al. 2009). Most symptomatic patients exhibit visceral, hematologic, and skeletal manifestations, including anemia, thrombocytopenia, hepatosplenomegaly, bone pain, and delayed growth and puberty. Type 3 GD, also known as subacute or chronic neuronopathic GD, presents with moderate systemic involvement, along with supranuclear saccadic horizontal gaze palsy, progressive myoclonus epilepsy, cerebellar ataxia, or spasticity (Daykin et al. 2021; Hwang et al. 2024; Kim et al. 2020; Roshan Lal and Sidransky 2017). Type 2 GD, the acute neuronopathic form, is characterized by early-onset and severe neurological deterioration, accompanied by systemic hepatosplenomegaly.

The long-term safety and effectiveness of enzyme replacement therapy (ERT) in GD have been well established, particularly in improving bone, hematologic, and visceral disease parameters (Nabizadeh et al. 2018; Shemesh et al. 2015). However, rare manifestations of GD, such as mesenteric lymphadenopathy, Gaucheroma, interstitial pneumonitis, and neurological manifestations are not responsive to ERT (Kim et al. 2022; Ramaswami et al. 2021). Following the approval of an enzyme extracted from human placenta (alglucerase; Ceredase, Genzyme Corp., Cambridge, MA, USA) by the Food and Drug Administration (FDA) in 1991 for the treatment of type 1 GD, Imiglucerase (Cerezyme, Genzyme Corp), a recombinant DNA-produced analog of human GBA using Chinese Hamster Ovary (CHO) cells that can be produced in large quantities, was approved in 1994 (Barton et al. 1991; Zimran et al. 1995). Currently, two other recombinant enzymes have been developed: velaglucerase (Vpriv, Shire) produced using human fibroblasts and taliglucerase (Elelyso, Pfizer) produced using carrot cells (Zimran et al. 2010, 2011). A newly developed form of imiglucerase, Abcertin (ISU Abxis, Seongnam-si, Gyeonggi-do, Republic of Korea), has been characterized in vivo and in vitro for its structural, physicochemical, immunological, and biological properties. Previous clinical studies have demonstrated its safety and efficacy profiles to be comparable to those of Cerezyme (Choi et al. 2015; Lee et al. 2017). However, for Abcertin to be approved as a biosimilar under World Health Organization (WHO) or European Medicines Agency (EMA) guidelines, comparative pharmacokinetic (PK) studies with the reference product in healthy volunteers are required. To meet this regulatory requirement, the present study evaluated the PK, immunogenicity, short-term safety, and tolerability of Abcertin compared to Cerezyme following a single-dose (60 IU/kg) administration in healthy volunteers.

2 | Methods

2.1 | Ethical Compliance

This study was conducted in accordance with the ethical principles of the Declaration of Helsinki and the Guidelines of the International Council for Harmonization Guideline for Good Clinical Practice, and all applicable laws or regulations. Th study protocol was approved by the Bellberry Human Research Ethics Committee, and written informed consent was obtained from all participants before any study procedures were performed.

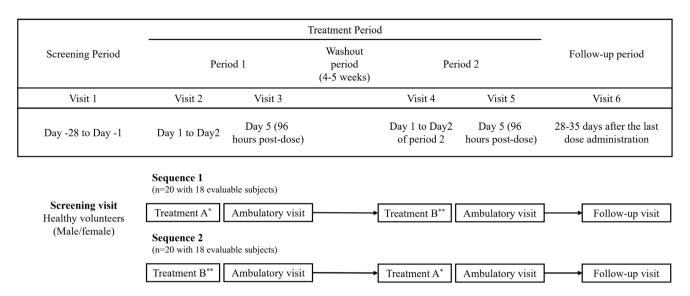
2.2 | Study Design

This was a phase 1, single-center, randomized, double-blind, two-way crossover study evaluating Abcertin and Cerezyme in healthy volunteers aged 18–45 years. In line with bioequivalence guidelines recommending studies using the highest strength, a single intravenous infusion of imiglucerase at 60 IU/kg—the highest approved initial dose for GD—was selected (Agency 2010). Participants were randomly assigned to receive either Treatment A (Abcertin, 60 IU/kg intravenously) followed by Treatment B (Cerezyme, 60 IU/kg intravenously) or vice versa, with a crossover design.

The study included a total of 6 visits; two of which were inpatient visits (Visit 2 and Visit 4; 24h post-dose). The 2 treatment periods were separated by a washout of at least 28 days but not more than 35 days, a duration chosen to fully minimize potential pharmacodynamic and immunological carryover effects resulting from macrophage uptake, despite the short plasma halflife of imiglucerase. Follow-up visits occurred 28-35 days after administration during Treatment Period 2 (Figure 1). The two products were administered as a single intravenous infusion of 60 IU/kg over 2h at an infusion rate of 0.5 IU/kg/min. The start and end times of the intravenous infusions were documented in the case report forms, along with details of any interruptions (stop time/restart time), the volume infused, and any adjustments in the infusion rate. In case of infusion interruption, a PK blood sample was collected prior to restarting the infusion, followed by continuation of the designated time collection for the rest of the infusion. Infusions were administered on alternate arms of the participant across the 2 treatment periods.

2.3 | Study Participants

The study population consisted of healthy male and female volunteers who met all the inclusion criteria and none of the exclusion criteria. Participants were aged 18-45 years, with a body mass index of 18.5-30 kg/m 2 and a weight of 55-105 kg, as determined by the Principal Investigator based on medical history, physical examination, 12-lead electrocardiography (ECG), and



*Treatment A: Abcertin® (60 IU/kg single intravenous administration). **Treatment B: Cerezyme® (60 IU/kg single intravenous administration).

FIGURE 1 | Overview of the study structure.

laboratory evaluations (including hematology, blood chemistry, coagulation, and urinalysis tests). Participants were excluded if they were taking any investigational drugs in another study, or who were on extended follow-up, had ongoing symptoms of acute diseases within 28 days prior to Period 1, or had any medical history that could have affected the drug distribution, metabolism, and excretion. Additional inclusion and exclusion criteria are provided in Table S1.

2.4 | Pharmacokinetic and Safety Measurements

The PK analysis set included all randomized participants who received both Abcertin and Cerezyme according to protocol, had at least one quantifiable post-dose concentration, and had no major protocol deviations or events that could significantly affect PK analysis. Participants who tested positive for anti-drug antibodies (ADAs) during the pre-dose assessment of Treatment Period 1 were excluded. The schedule of assessments is shown in Table S2. The time points of assessment for PK studies of Abcertin and Cerezyme were within 30 min prior to infusion, and at 10, 20, 40, 60, 90, 120 min during infusion, and at 5, 10, 20, 30, 40, 50, 60, 70, 80, 90, and 120 min after the end of infusion. The primary PK endpoint was the area under the concentration–time curve (AUC) from time zero to infinity (AUC_{0-inf}). Secondary PK endpoints included maximum plasma concentration (C_{\max}), time to C_{\max} (t_{\max}), AUC from time zero to the last measurable plasma concentration (AUC $_{0-last}$), terminal half-life $(t_{1/2})$, total body clearance (CL), and volume of distribution in the terminal phase (V_z) .

Participants who received at least one dose of Abcertin or Cerezyme were included in the safety analysis. Safety evaluations were performed at pre- and post-dosing, and at every visit. These evaluations included a physical examination, measurements of vital signs, 12-lead ECG, clinical laboratory tests, injection site reactions and adverse events (AEs). AEs were coded according to the Medical Dictionary for Regulatory Activities

(MedDRA; version 22.1). For immunogenicity assessments, ADA levels were measured before dosing and at the follow-up visit. The ADA analysis set included all randomized participants who received both Abcertin and Cerezyme per protocol and had available ADA data. Participants who received treatment in only one period and subsequently withdrew were excluded. Samples that tested positive in the screening assay were subjected to a confirmatory assay. Those confirmed positive were further evaluated for antibody titer and neutralizing capacity. PK and neutralizing antibodies(NAbs) analyses were conducted using fluorescent enzyme activity assay platforms, ADA analyses were performed using an electrochemiluminescence (ECL)-based assay format.

2.5 | Statistical Analysis

All analyses were performed using SAS version 9.3 or later in a secure and validated environment. PK analyses were conducted using Phoenix WinNonLin (WNL) version 8.0 or later (Certara USA Inc., Princeton, NJ). Descriptive statistics for the primary endpoint (AUC_{0-inf}) were presented by treatment and period, including the number of participants (N), mean, standard deviation (SD), percent coefficient of variation (%CV), median, minimum and maximum values, geometric mean, and geometric %CV. PK biosimilarity between Abcertin and Cerezyme was assessed using an analysis of variance (ANOVA) on log-transformed $\mathrm{AUC}_{0-\mathrm{inf}}$ values. The ANOVA model included fixed effects for treatment, period, sequence, and participants nested within the sequence. The point estimates and 90% confidence intervals (CIs) for the difference among treatments were constructed. The point estimates and 90% CIs were then exponentially back-transformed to provide point and 90% CI estimates for the ratio (for logarithmicallytransformed data) of geometric least squares (LS) means for imiglucerase $\mathrm{AUC}_{0-\mathrm{inf}}$ for Abcertin versus Cerezyme. PK biosimilarity of Abcertin was concluded relative to each source of Cerezyme if the 90% CI fell entirely within 80.00% to 125.00% for AUC_{0-inf}. As a sensitivity analysis, an additional analysis was

performed using subject within sequence as a random effect instead of as a fixed effect. Descriptive statistics for each secondary PK endpoint were summarized by treatment and period, and the same ANOVA model was applied to $C_{\rm max}$ and ${\rm AUC}_{\rm 0-last}$. A sensitivity analysis using a random effect for participants nested within the sequence was also conducted.

3 | Results

3.1 | Baseline Characteristics

Of the 84 participants screened, 42 were randomized, with 38 completing the study; 2 discontinued due to consent withdrawal, 1 due to ineligibility, and 1 due to severe urticaria. The numbers of participants in the safety, PK, and ADA populations were 42,

TABLE 1 | The demographic and baseline characteristics of participants.

	Total (N=42)
Age (years)	26.6 ± 7.49
Sex (Male, N, %)	29 (69.05)
Race (N, %)	
White	32 (76.19)
Australian Native	0 (0)
Asian	8 (19.05)
Other Pacific Islander	0 (0)
Black	0 (0)
Other	2 (4.76)
Height (cm)	173.96 ± 9.64
Weight (kg)	72.42 ± 12.04
$BMI (kg/m^2)$	23.86 ± 2.87

Abbreviation: BMI, body mass index.

37, and 38, respectively. Demographic and baseline characteristics are summarized in Table 1.

3.2 | Pharmacokinetic Evaluations

One participant experienced an interrupted infusion and an unscheduled sampling time point during Treatment Period 2, resulting in the exclusion of their PK data from the summary statistics. As a result, 37 participants were included in the PK analysis set. The mean concentrations of Abcertin and Cerezyme over 4 h post-dose are shown in Figure 2.

Overall, PK parameters were comparable between Abcertin and Cerezyme (Table 2). For Abcertin, peak plasma concentrations were reached at a median $t_{\rm max}$ of 61.0 min (range, 40–120 min). The mean values for C_{max} , $AUC_{0-\text{inf}}$, and $AUC_{0-\text{last}}$ of Abcertin $115.4 \pm 23.8 \,\text{mU/mL}$, $12,210 \pm 2483 \,\text{min} \cdot \text{mU/mL}$, 12,190 ± 2480 min·mU/mL, respectively. The mean values for $t_{1/2}$, CL, and V_z were $6.88 \pm 1.14 \,\mathrm{min}$, $363.7 \pm 124.3 \,\mathrm{mL/min}$, and 3.62 ± 0.87 L, respectively. Inter-subjective variability for C_{max} , $\mathrm{AUC}_{0-\mathrm{inf}}$, and $\mathrm{AUC}_{0-\mathrm{last}}$ were 20.6%, 20.3%, and 20.3%, respectively. For Cerezyme, the median $t_{\rm max}$ was 60 min (range, 20-120 min). The mean values for $C_{\rm max}$, ${\rm AUC}_{\rm 0-inf}$, and ${\rm AUC}_{\rm 0-last}$ of Cerezyme were 117.5 ± 24.7 mU/mL, 12,440 ± 2514 min·mU/mL, and 12,320 ± 2513 min·mU/mL, respectively. The mean values for $t_1/_2$, CL, and V_2 were $7.20 \pm 0.91 \,\mathrm{min}$, $368.2 \pm 116.0 \,\mathrm{mL/min}$, and 3.77 ± 1.04 L, respectively. Inter-subject variabilities for C_{max} , AUC_{0-inf} , and AUC_{0-last} were 21.1%, 20.2%, and 20.2%, respectively.

Abcertin (60 IU/kg) was bioequivalent to Cerezyme (60 IU/kg) with respect to $\mathrm{AUC}_{0-\mathrm{inf}}$, $\mathrm{AUC}_{0-\mathrm{last}}$, and C_{max} , as the corresponding 90% CIs for geometric mean ratios (GMRs) were entirely within the predefined bioequivalence range of 80%–125% (Table 3).

In a sensitivity analysis, the test/reference GMRs for AUC_{0-inf} AUC_{0-last}, and C_{max} were 98.18%, 98.15%, and 98.05%, respectively, with their corresponding 90% CIs fully contained within the bioequivalence range of 80%–125% (94.57%–101.92% for AUC_{0-inf} , 94.55%–101.90% for AUC_{0-last} , and 93.86%–102.42% for C_{max}).

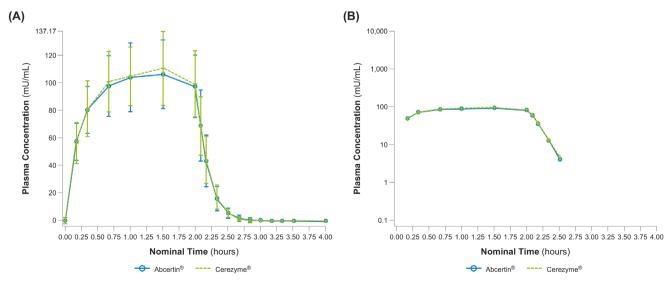


FIGURE 2 | Pharmacokinetic Results. (A) Arithmetic mean ± SD (linear scale) and (B) geometric mean (semi-logarithmic scale).

TABLE 2 | Plasma pharmacokinetic parameters.

			Abc	Abcertin						Cer	Cerezyme			
	Mean±SD	CV(%)a	CV(%) ^a Median	Min	Max	Geo mean	Geo CV (%)	Mean±SD	CV(%)a	CV(%) ^a Median	Min	Max	Geo mean	Geo CV (%)
$C_{ m max} \left({ m mU/mL} ight)$	115.4 ± 23.8	20.6	115.0	68.1	172.0	112.8	22.1	117.5 ± 24.7	21.1	118.0	72.5	184.0	115.0	21.1
$t_{ m max} ({ m min})$			61.0	40.0	121.0					0.09				
$\mathrm{AUC}_{\mathrm{0-inf}}$ (min·mU/mL)	$12,210\pm 2483$	20.3	12480.0	7300.0	19000.0	11960.0	21.5	$12,440 \pm 2514$	20.2	12260.0	7480.0	19200.0	12190.0	20.9
${ m AUC}_{0-{ m last}}$ (min·mU/mL)	$12,190 \pm 2480$	20.3	12470.0	7290.0	189000.0	11940.0	21.5	$12,320\pm2513$	20.2	12230.0	7460.0	19200.0	12170.0	20.9
$t_{1/2}$ (min)	6.882 ± 1.139	16.6	8.9	5.1	10.0	6.797	15.9	7.204 ± 0.9135	12.7	7.3	5.5	9.4	7.147	12.9
CL (mL/min)	363.7 ± 124.3	33	323.7	235.0	705.0	360.0	30.2	368.2 ± 116.0	31.5	317.7	203.0	715.0	353.2	28.9
$V_{ m z}({ m L})$	3.624 ± 0.8695	24	3.2	2.4	5.7	3.5	23.2	3.770 ± 1.039	27.6	3.4	2.2	7.1	3.6	26.8
Abbreviations: AUC, as area under the concentration-time curve from pre-dose to infinite time; AUC, area under the plasma drug concentration-time curve from time zero to last measurable concentration; CL, total body	area under the conc	entration-time	curve from pre	-dose to infir	nite time; AUC,	area und	er the plasm	a drug concentration-	time curve fr	om time zero t	o last measu	rable concen	tration; CL, to	tal body

clearance; C_{\max} , maximum plasma concentration; CV, coefficient of variation; Geo CV, geometric coefficient of variation; Geo Mean, geometric mean; t_{i2} , terminal half-life; t_{\max} , time to achieve C_{\max} ; V_z , volume of distribution, a CV (%) = $(SD/Mean) \times 100$. b Geo CV (%) = sqrt(exp(v)—1) × 100, where v is the variance of ln(concentration).

One participant (number 10118) experienced an interrupted infusion but restarted infusion approximately 10 min during Treatment Period 2. A sensitivity analysis excluding this participant's data to assess its impact on the overall PK analysis yielded the same bioequivalence conclusion, as the bioequivalence criteria for AUC $_{\rm 0-inf}$, AUC $_{\rm 0-last}$, and $C_{\rm max}$ remained satisfied. The test/reference GMRs were 98.37%, 98.34%, and 98.12%, respectively, with 90% CIs fully within the bioequivalence range of 80%–125% (94.67%–102.21% for AUC $_{\rm 0-inf}$, 94.64%–102.19% for AUC $_{\rm 0-last}$, and 93.81%–102.63% for $C_{\rm max}$).

3.3 | Immunogenicity Evaluation

In immunogenicity evaluation, all 19 participants who received Abcertin in Sequence 1 were negative for ADA presence at Day 1 pre-dose in Treatment Period 2. On the other hand, 1 out of 19 participants who received Cerezyme in Sequence 1 were positive for treatment-induced ADA at Day 1 pre-dose in Treatment Period 2, which was not positive in NAbs. All 35 follow-up immunogenicity samples were negative for ADA.

3.4 | Safety and Tolerability

The safety analysis set included all participants (N=42) who received at least one dose of Abcertin or Cerezyme. All reported AEs were mild or moderate in severity, and no serious adverse event occurred during the study. Overall, 24 (57.14%) treatment-emergent adverse events (TEAEs) were reported: 18 (42.86%) in the Abcertin sequence and 18 (42.86%) in the Cerezyme sequence. Table 4 summarizes the TEAEs related to the treatment. Fifteen participants (14 in the Abcertin group and 5 in the Cerezyme group) experienced mild TEAEs, while seven participants (4 in the Abcertin group and 3 in the Cerezyme group) experienced moderate TEAEs. Two severe TEAEs were in the Cerezyme sequence: one participant developed a severe urinary tract infection, and another experienced severe urticaria, leading to study withdrawal. No AEs resulted in death. No clinically significant changes were observed in vital signs, laboratory values, or ECG findings, and no signs of hepatotoxicity were detected.

4 | Discussion

Abcertin was approved in Iran, Peru, Bolivia, Colombia, Ecuador, Venezuela, Algeria and South Korea for the treatment of GD through a national regulatory pathway that permitted the submission of Phase 3 data post-approval. The clinical efficacy, PK, and safety data for Abcertin were reported in published Phase 2 and Phase 3 studies. However, according to WHO and EMA guidelines, comparative PK studies in healthy volunteers are mandatory for biosimilar approval.

This study evaluated the PK equivalence of Abcertin and Cerezyme in healthy adults. Following intravenous administration of a single 60 IU/kg dose, the PK parameters demonstrated bioequivalence between Abcertin and Cerezyme, based on the primary PK endpoint, $\mathrm{AUC}_{0-\mathrm{inf}}$, with an Abcertin/Cerezyme GMR of 98.18% and 90% CIs within the predefined bioequivalence range of 80%–125% (94.57%–101.92% for $\mathrm{AUC}_{0-\mathrm{inf}}$).

TABLE 3 | Bioequivalence between Abcertin and Cerezyme.

	Geometric lea	st square mean	Geometric mean ratio (%)	
Parameters	Abcertin	Cerezyme	(Abcertin/Cerezyme)×100	90% CI
AUC _{0-inf} (min*mU/mL)	11,950	12,170	98.18	94.57–101.92
AUC _{0-last} (min*mU/mL)	11,930	12,160	98.15	94.55-101.90
$C_{\rm max}$ (mU/mL)	112.7	115	98.05	93.86-102.42

TABLE 4 | Summary of treatment-emergent adverse events (TEAEs) related to the investigational product.

	Abcertin	Cerezyme	Total $(N=42)$
Participants with ≥1 TEAE	5 (11.9)	9 (21.4)	13 (31.0)
Nervous system disorders	2 (4.8)	5 (11.9)	7 (16.7)
Headache	2 (100)	5 (100)	7 (100)
Gastrointestinal disorders	2 (4.8)	2 (4.8)	3 (7.1)
Dyspepsia	1 (50.0)	1 (50.0)	2 (66.7)
Vomiting	1 (50.0)	1 (50.0)	1 (33.3)
Diarrhea	0	1 (50.0)	1 (33.3)
Skin and subcutaneous tissue disorders	1 (2.4)	1 (2.4)	2 (4.8)
Rash	1 (100)	0	1 (50.0)
Urticaria	0	1 (100)	1 (50.0)
Blood and lymphatic system disorders	0	1 (2.4)	1 (2.4)
Lymphopenia	0	1 (100)	1 (100)
General disorders and administration site conditions	0	1 (2.4)	1 (2.4)
Injection site erythema	0	1 (100)	1 (100)

Additional PK parameters, $\mathrm{AUC}_{0-\mathrm{last}}$ and C_{max} , also met bioequivalence criteria using the same ANOVA model, with 90% CIs similarly contained within the predefined range. A sensitivity analysis, using an ANOVA model with subject within sequence as a random effect, confirmed these results. Other PK parameters, including t_{max} , $t_{\mathrm{l/2}}$, CL, and V_{z} , were also similar between Abcertin and Cerezyme.

Improvements in hematological, visceral, and bone parameters, as well as quality of life, have been well established following imiglucerase therapy in patients with GD (Serratrice et al. 2016; Weinreb et al. 2013; Zimran et al. 2018). The efficacy of Abcertin in patients with type 1 GD was comparable to that of Cerezyme in a Phase 2, multicenter, open-label, switch-over trial (Choi et al. 2015). Additionally, a multicenter, open-label Phase 3 study demonstrated the efficacy of Abcertin by significantly increasing hemoglobin concentration and platelet count while decreasing organ volume, acid phosphatase, and chemokine ligand 18 levels (Lee et al. 2017). Also, long-term combination therapy with Abcertin 60 IU/kg and high-dose ambroxol (35 mg/kg/day) has shown promising results in type 3 GD (Hwang et al. 2024; Kim et al. 2020).

In a phase 2 study, the PK profile of Abcertin after intravenous administration of 30–60 U/kg over 90 min demonstrated a $T_{\rm max}$

of $1.34 \pm 0.32 \, h$ (range, $1.00 - 1.67 \, h$) and a clearance half-life of $0.11 \pm 0.01 \, \text{h}$ (range, $0.09 - 0.12 \, \text{h}$) (Choi et al. 2015). The elimination half-life, plasma clearance, and volume of distribution of imiglucerase are independent of infusion time and dose within a range of 7.5-60 U/kg (Edmunds 2005). The plasma activity half-life is short, ranging from 3.6 to 10.4 min, which is advantageous due to the rapid inactivation of glucocerebrosidase at the neutral pH of blood and the stabilization of enzymatic activity for lysosomal trafficking via rapid endocytosis (Weinreb 2008). The uptake and tissue distribution of iodine-123-labeled imiglucerase or aglucerase in eight patients with type 1 GD and one healthy control showed rapid blood clearance with a half-life of 4–7 min and significant uptake in the liver (30%), spleen (15%), and bone marrow. Bone marrow clearance had a half-life of 14.1 h, whereas visceral clearance followed a biphasic pattern, with half-lives of 1-2h and 34-42h (Mistry et al. 1996).

The long-term safety and immunogenicity of imiglucerase have been reported with no serious adverse events reported in clinical trials or in the International Cooperative Gaucher Group (ICCG) Gaucher registry (Starzyk et al. 2007; Weinreb 2008). Good tolerance to imiglucerase has also been confirmed in switch and non-inferiority studies comparing other ERTs, including velaglucerase and taliglucerase (Elstein et al. 2015; Pastores et al. 2016; Smith et al. 2016). Overall, ERT is considered safe for

all ages and GD subtypes, with no reported death or irreversible damage related to treatment (Gupta and Pastores 2018; Starzyk et al. 2007; Zimran et al. 1995). A few cases of anaphylactic reactions have been reported, typically during the first or second infusion, and were efficiently managed by premedication, slowing the infusion rate, or switching to another ERT. Most AEs, including chills, fever, pruritus, rashes, urticaria, and dyspnea, were mild to moderate in severity, transient, and did not lead to discontinuation of therapy. In this study, 24 participants (57.14%) experienced at least one TEAE. Headache was the most frequently reported TEAE considered treatment-related by the investigator. All adverse events observed with Abcertin treatment were mild or moderate in severity, with no clinically relevant abnormalities observed in laboratory tests, vital signs, or ECG. Two severe TEAEs, considered by the investigator to be treatment-related, were reported in the Cerezyme treatment sequence. Notably, one participant experienced urticaria, a special-interest adverse event, leading to study withdrawal. The incidence of treatment-induced ADAs was low following a single intravenous administration of Abcertin or Cerezyme; only 1 participant after receiving Cerezyme in sequence 2 was positive in ADA without neutralizing capacity. In a comparative trial, ADAs developed in 6 of 15 patients receiving alglucerase and in 3 of 15 patients receiving imiglucerase. While antibodies appeared within 3-6 months in the imiglucerase group, no major immunological effects or diminished therapeutic responses were observed in either group (Grabowski et al. 1995).

5 | Conclusion

Cerezyme, an ERT for GD, has been shown to be effective and safe for disease management. However, its high cost significantly limits access, particularly in resource-limited settings. Introducing biosimilars could improve treatment affordability and availability, thereby expanding access to effective therapies. By meeting key biosimilarity criteria, Abcertin may offer a more cost-effective alternative, increasing accessibility for patients with GD.

Following the administration of a single dose to healthy adults, Abcertin demonstrated PK equivalence to Cerezyme based on the study's primary endpoints. Secondary PK parameters, immunogenicity, and safety profiles were also comparable between the two groups. These findings provide evidence that Abcertin could serve as a potential biosimilar to Cerezyme.

Author Contributions

C.L., J.W., Y.L., H.W.Y., B.H.L. contributed to the study design. C.L., J.W., Y.L. contributed to data collection. E.K., C.L., J.W., Y.L., and B.H.L. contributed to data analysis or interpretation. E.K. and B.H.L. drafted the manuscript. All authors critically reviewed and revised the manuscript, approved the final manuscript for publication.

Ethics Statement

The study was conducted in accordance with the ethical principles of the Declaration of Helsinki and the Guidelines of the International Council for Harmonization Guideline for Good Clinical Practice, and all applicable laws or regulations.

Consent

This study was approved by the Bellberry Human Research Ethics Committee and written informed consent was obtained from all participants before any study procedures.

Conflicts of Interest

Y.L. is an employee of ISU ABXIS. E.K., D.K., S.H., C.L., J.W., H.W.Y., and B.H.L. declare no conflicts of interest.

Data Availability Statement

Anonymized data not included in this article will be shared by request from any qualified investigator.

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Supporting Information

Additional supporting information can be found online in the Supporting Information section.