# **Dynamic Contrast Enhanced MRI Mapping of Vascular Permeability for Evaluation of Breast Cancer Neoadjuvant Chemotherapy Response Using Image-to-Image Conditional Generative Adversarial Networks**

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**Abstract:** Dynamic contrast enhanced (DCE) MRI is a non-invasive imaging technique that has become a quantitative standard for assessing tumor microvascular permeability. Through the application of a pharmacokinetic (PK) model to a series of T1-weighed MR images acquired after an injection of a contrast agent, several vascular permeability parameters can be quantitatively estimated. These parameters, including  $K_{trans}$ , a measure of capillary permeability, have been widely implemented for assessing tumor vascular function as well as tumor therapeutic response. However, conventional PK modeling for translation of DCE MRI to PK vascular permeability parameter maps is complex and time-consuming for dynamic scans with thousands of pixels per image. In recent years, imageto-image conditional generative adversarial network (cGAN) is emerging as a robust approach in computer vision for complex cross-domain translation tasks. Through a sophisticated adversarial training process between two neural networks, image-to-image cGANs learn to effectively translate images from one domain to another, producing images that are indistinguishable from those in the target domain. In the present study, we have developed a novel image-to-image cGAN approach for mapping DCE MRI data to PK vascular permeability parameter maps. The DCE-to-PK cGAN not only generates high-quality parameter maps that closely resemble the ground truth, but also significantly reduces computation time over 1000-fold. The utility of the cGAN approach to map vascular permeability is validated using open-source breast cancer patient DCE MRI data provided by The Cancer Imaging Archive (TCIA). This data collection includes images and pathological analyses of breast cancer patients acquired before and after the first cycle of neoadjuvant chemotherapy (NACT). Importantly, in good agreement with previous studies leveraging this dataset, the percentage change of vascular permeability  $K_{trans}$  derived from the DCE-to-PK cGAN enables early prediction of responders to NACT.

**Keywords:** Dynamic contrast enhanced (DCE) MRI, vascular permeability, image-to-image conditional generative adversarial network (cGAN), breast cancer, neoadjuvant chemotherapy (NACT)

# **1. Introduction**

Neoadjuvant chemotherapy (NACT) has been established as a standard of care for patients with locally advanced breast cancer and has been expanded to include patients with early-stage breast cancer<sup>1</sup>. The primary goal of NACT is to downstage the tumor prior to surgery, enabling improved operability and conservation of healthy breast tissue<sup>2-5</sup>. Pathologic complete response (pCR) to NACT has been shown to be prognostic for survival<sup>1</sup>.

However, only 6-25% of breast cancer patients experience pCR to NACT<sup>3</sup>. Moreover, pathologic analysis must be performed after multiple cycles of NACT and surgery, limiting the ability to assess patient responses early in the treatment process. Currently, there is significant interest in non-invasive means to predict the pathologic response status to NACT prior to surgery, enabling clinicians to adjust personalized treatment regimens based on early responses, which in turn could lead to improved patient outcomes<sup>2</sup>.

To this end, various quantitative imaging modalities have been investigated for assessment of pathologic response status in breast cancer patients receiving NACT attributable to their non-invasive ability to assess tumor biology<sup>3,5,6</sup>. Early changes in underlying tumor biology characteristics including permeability, perfusion, metabolism, oxygenation, and cell density could enable early prediction of pathological response. In recent years, researchers have identified dynamic contrast enhanced (DCE) MRI and its resulting pharmacokinetic (PK) vascular permeability parameters as a potential biomarker for early assessment of NACT pathological response in breast cancer patients<sup>2,3,7</sup>. Tudorica et al. have shown that percentage changes in DCE MRI vascular permeability parameters are good to excellent predictors of pCR to NACT, with univariate logistic regression (ULR) c-statistic values ranging from 0.804-0.967 in a cohort of 28 invasive ductal carcinoma patients<sup>2</sup>. Namely, early changes in Ktrans, a measure of capillary permeability, immediately following the first cycle of NACT has shown the greatest promise for early prediction of pathologic response.

Despite its potential, the use of DCE MRI for monitoring treatment response has several limitations hindering its widespread clinical implementation. First, there is significant variability in image acquisition protocols and scanners across different institutions, which can lead to non-reproducible and inconsistent results in clinical oncology. Standardization of DCE MRI acquisition will enhance the reproducibility and consistency of DCE MRI studies. Moreover, DCE MRI requires sophisticated software for solving complex PK mathematical models that are used to generate vascular permeability parameters. There is also significant variation in not only PK models, but also the software tools/packages used to solve these models. It is well recognized that commercialization of a standardized software tool for generating vascular permeability parameters in DCE MRI will help establish the widespread clinical implementation of DCE MRI<sup>7</sup>. Furthermore, establishment of a simple software tool for generating vascular permeability parameters will enhance the ease of implementation and decrease the complexity and time required to solve these mathematical models.

To this end, several research groups including ours have investigated deep learning as a potential tool to standardize PK analysis and reduce computational complexity. These studies have applied deep learning approaches for quantification of vascular permeability in applications including brain cancer<sup>8-10</sup>, stroke<sup>11</sup>, pancreatic cancer<sup>12</sup>, and head and neck cancer<sup>13</sup>. The deep learning approaches explored in these studies include 1D convolutional neural network (CNN), 2D CNN, and recurrent neural networks (RNNs) such as gated recurrent unit (GRU) and long short-term memory (LSTM) networks. Although these studies have demonstrated the utility of deep learning for efficient quantification of vascular permeability, it is still uncertain if deep learning can reliably monitor tumor vascular permeability treatment responses with the same level of precision as conventional methods. Moreover, with the recent surge in the development of novel deep learning approaches, several innovative techniques have

emerged, offering potential improvements in both accuracy and robustness for quantifying vascular permeability in DCE MRI.

One of these novel deep learning approaches, generative adversarial network (GAN), specifically imageto-image conditional GAN (cGAN), has proven to be a robust approach in computer vision for complex crossdomain translation tasks<sup>14,15</sup>. During training of an image-to-image cGAN, a Generator neural network learns to create synthetic images that are similar to a target domain using images from another domain. Simultaneously, a Discriminator neural network evaluates the synthetic images against real images and determines if they are real or fake, guiding the Generator to produce increasingly realistic images over time. By leveraging this sophisticated adversarial training scheme between two neural networks, image-to-image cGANs can generate high-quality images that closely resemble the ground truth. Several studies have revealed the robust capabilities of cGANs for complex medical imaging tasks, including synthetic image generation<sup>16,17</sup>, image de-noising<sup>18</sup>, segmentation<sup>19,20</sup>, superresolution<sup>21</sup>, and modality transfer<sup>22</sup>. Specifically, image-to-image cGANs have recently been successfully applied to challenging cross-domain translation tasks in medical imaging, such as synthesizing apparent diffusion coefficient (ADC) maps from diffusion-weighted imaging<sup>23</sup> and mapping cerebral blood volume (CBV) from standard MRI scans<sup>24</sup>.

Clearly, image-to-image cGANs are an attractive approach for translation of DCE MRI to PK vascular permeability parameter maps. However, to the best of our knowledge, an image-to-image cGAN has not been previously investigated for DCE-to-PK cross-domain translation tasks. In the present study, we have developed a novel DCE-to-PK cGAN approach for quantification of vascular permeability parameter maps in DCE MRI. The utility of the deep learning approach is validated in open-source breast cancer patient DCE MRI data provided by The Cancer Imaging Archive (TCIA). This data collection contains patient images and pathological analyses acquired before and after the first cycle of NACT from a multi-center data analysis challenge<sup>7,25</sup>. Moreover, the NACT induced early changes in vascular permeability as quantified by the DCE-to-PK cGAN has been correlated with pathological treatment response.

#### **2. Materials and Methods**

#### *2.1. Patient Data*

An open-source collection of breast cancer DCE MRI data provided by the Quantitative Imaging Network (QIN) and published on TCIA was used in the current study for training, testing, and validating the proposed imageto-image cGAN approach<sup>7,25</sup>. The data collection was used in a previous study investigating the response of breast cancer patients to NACT in a multi-QIN center data analysis challenge. As described previously<sup>7</sup>, 20 DCE MRI scans were acquired across ten breast cancer patients. Each patient was subject to two imaging exams, one acquired prior to the start of treatment, V1, and another acquired after the first cycle of treatment, V2. MRI studies were performed on a Siemens 3T TIM Trio system. DCE MRI was acquired using a 3D gradient echo-based timeresolved angiography with stochastic trajectories (TWIST) sequence (TR/TE: 6.2/2.9 msec; flip-angle: 10 degrees; field of view: 30 to 34 cm; in-plane matrix size:  $320 \times 320$ ; slice thickness: 1.4 mm; parallel imaging acceleration factor: 2; temporal resolution: 18 to 20 sec). Following the acquisition of two baseline images, a contrast agent

Gd(HP-D03A) [ProHance] was injected intravenously (0.1 mmol/kg at 2 ml/s) with a programmable power injector, followed by a 20 ml saline flush.

In addition to DCE MRI data, average tumor baseline longitudinal relaxation rate  $R_1$  ( $R_{10}$ ), populationaveraged arterial input function (AIF), tumor region of interest (ROI), and pathologic response status were provided for each patient for PK modeling and analysis. As described previously, tumor average  $R_{10}$  values were determined by comparing the signal intensity of baseline DCE images without contrast to spatially registered proton density images acquired immediately before DCE MRI<sup>26</sup>. The provided population-averaged AIF was obtained in a previous sagittal breast DCE MRI study, which employed the same contrast agent injection protocol<sup>27</sup>. Patient tumor ROIs were drawn by experienced breast radiologists at Oregon Health and Science University on post-contrast DCE MRI. Pathologic status was determined through pathologic comparisons pre-NACT and following the final cycle of NACT. A binary classification of pCR or non-pCR (pathologic nonresponse and pathologic partial response) was provided for each patient<sup>7</sup>.

# *2.2. DCE MRI Preprocessing*

DCE MRI scans were cropped to isolate patient tumor-bearing breasts and remove background anatomical structures including the lungs and heart, resulting in an in-plane matrix size of  $160 \times 160$ . The scans were truncated to twenty-eight frames to allow for all imaging datasets having the same temporal size. DCE images were then motion corrected using an in-house MATLAB co-registration algorithm.

#### *2.3. Pharmacokinetic Modeling*

Dynamic signal intensity changes in DCE MRI and patient-specific  $R_{10}$  values were used to determine  $C_t$ , the dynamic concentration of contrast within the tissue, according to the following equations:

$$
S(\tau) = S_0 \frac{\left(1 - e^{-TR \cdot R_1(\tau)}\right) \cdot \sin(\theta)}{1 - e^{-TR \cdot R_1(\tau)} \cdot \cos(\theta)} \tag{1}
$$

$$
R_1(\tau) = r_1 C_t(\tau) + R_{10} \tag{2}
$$

where S and  $S_0$  is the DCE MRI signal intensity with and without contrast agent, respectively,  $\theta$  is the flip angle of the DCE MRI sequence, and  $r_1$  is the relaxivity of the contrast agent. Following  $C_t$  mapping, the Extended Tofts Model (ETM) was applied to estimate vascular permeability parameters according to Eqn. 3:

$$
C_t(\tau) = V_p C_p(\tau) + K_{trans} \int_0^{\tau} C_p(\tau) e^{-k_{ep}(\tau - \tau')} d\tau
$$
\n(3)

where  $C_p$  is the dynamic concentration of contrast within the plasma, also known as the AIF.  $V_p$ ,  $K_{trans}$ , and  $k_{ep}$  are vascular permeability parameters that describe the fractional volume of blood plasma, the transfer rate of contrast agent from the blood plasma to the extravascular extracellular space (EES), and the reverse transfer rate of the contrast agent from the EES to the blood plasma, respectively<sup>10,28,29</sup>. A non-linear least squares (NLLS) curve-fitting algorithm was used to map the PK parameters to  $C_t$  and  $C_p$  for each voxel. The bounds for curve-fitting of each vascular parameter were:  $K_{trans} = [1e^{-8}, 2]$ ,  $k_{ep} = [1e^{-8}, 2]$ , and  $V_p = [1e^{-8}, 0.2]$ . All mathematical modeling algorithms were implemented in MATLAB R2024a with an Intel Xeon CPU @ 3.70 GHz and 32 GB RAM.



**Figure 1. Pathway for DCE-to-PK cGAN training**. A Generator neural network was trained to translate input  $R_{10}$  + DCE MRI to synthetic K<sub>trans</sub> maps. Simultaneously, a Discriminator neural network was trained to distinguish between the Generator synthetic  $K_{trans}$  maps and real ETM  $K_{trans}$  maps, conditioned on input  $R_{10}$  + DCE MRI. A custom least squares patch-based adversarial loss incorporating an L1-loss term was used to update both network parameters until the resulting synthetic  $K_{trans}$  maps generated by the Generator appeared indistinguishable from real ETM  $K_{trans}$  maps.

#### *2.4. DCE-to-PK cGAN Architecture*

The proposed cGAN architecture for translating DCE MRI to PK vascular permeability parameters is shown in **Figure 1**. Patient-specific  $R_{10}$  values were broadcasted to a matrix size of  $160 \times 160$  and concatenated with corresponding DCE MRI patient scans. The Generator neural network translates  $R_{10}$  + DCE MRI into synthetic vascular permeability  $K_{trans}$  maps. The Discriminator neural network determines if the input  $K_{trans}$  maps from the ETM or the Generator are real or fake, conditioned on corresponding  $R_{10}$  + DCE MRI. In the current study, we used a dual-pathway CNN from our previous work<sup>9,10</sup> for the Generator and a conditional PatchGAN for the Discriminator<sup>14</sup>.

For the Generator, an image input layer and convolutional layer were followed by the two parallel pathways, local and global, to capture multi-scale features. The local pathway consists of three convolutional non-dilated layers. The global pathway consists of three convolutional layers that were dilated by factors of 2, 4, and 8, respectively. The dual-pathway convolutional layers were designed with 128 filters with a size of  $3 \times 3$ . Local and global pathways were then concatenated and followed by four fully connected convolutional layers of 1024, 512, 128, and one filter, each with a size of  $1 \times 1$ . A ReLU activation layer followed each convolutional layer, except for the final convolutional layer, which was instead followed by a Tanh layer. Zero padding was applied to every convolutional operation.

For the Discriminator, a conditional PatchGAN was implemented to down-sample  $K_{trans}$  maps by a factor of eight and discriminate based on patches. The Discriminator was conditioned with corresponding  $R_{10}$  + DCE MRI used to create the real and synthetic K<sub>trans</sub> maps. An image input layer was followed by a fully convolutional architecture with five convolutional layers of 128, 256, 512, 1024, and one filter, each with a size of  $3 \times 3$ . Batch normalization was applied to all convolutional layers, except for the first and final convolutional layer. A leaky ReLU activation layer followed each convolutional layer, apart from the final convolutional layer, which was instead followed by a Sigmoid layer.

#### *2.5. Model Training*

DCE MRI and corresponding  $K_{trans}$  maps were normalized to [0 1] for model training. K-fold cross validation was performed to allow all patients to serve as testing data of the DCE-to-PK cGAN. To this end, imaging data from a single patient ( $n = 240$  DCE slices) from both pre and post-NACT (V1 and V2) were isolated and saved for network testing. The remaining patient data ( $n = 2,160$  DCE slices) were then randomly split into an 80:20 training to validation ratio. Using the Adam optimizer, hyper-parameters were manually adjusted to achieve the best network performance with minimum error: a learning rate of 1e<sup>-5</sup> for both the Generator and Discriminator, mini-batch size of 16, and maximum epochs of 200. Training data was shuffled at every epoch. A custom least squares cGAN-based approach<sup>30</sup> incorporating an L1-loss (mean absolute error) component was implemented for the Generator and Discriminator loss functions as follows:

$$
\mathcal{L}_D = \frac{1}{N} \sum_{i=1}^N [(D(x_i, y_i) - 1)^2 + D(x_i, G(x_i))^2]
$$
\n
$$
\mathcal{L}_G = \frac{1}{N} \sum_{i=1}^N [(D(x_i, G(x_i)) - 1)^2 + \lambda_{L1} * ||y_i - G(x_i)||_1]
$$
\n(5)

where N is the mini-batch size,  $x_i$  is the conditioned  $R_{10}$  + DCE MRI,  $y_i$  is the ground-truth ETM  $K_{trans}$  maps,  $G(x_i)$ is the Generator synthetic  $K_{trans}$  maps, and  $\lambda_{L1}$  is the weight of the L1 loss term, which was set to 1. Following training, the patient testing dataset was fed directly into the Generator to produce synthetic  $K_{trans}$  maps that were then rescaled back to their original distribution. This process was repeated 10-fold for all patients. All neural network algorithms were implemented in MATLAB R2024a with an NVIDIA GeForce RTX 3090.

## *2.6. Statistical Analysis*

Patient intratumoral individual pixel, slice mean, and tumor mean  $K_{trans}$  for both V1 and V2 were quantified using tumor ROI provided by the TCIA. Linear regression was applied to individual patients as well as an ensemble of patients K<sub>trans</sub> values for statistical correlation and significance. The root-mean-squared error (RMSE), mean absolute error (MAE), normalized RMSE ( $nRMSE$ ), normalized MAE ( $nMAE$ ), Pearson R-squared ( $R^2$ ), and concordance correlation coefficient (CCC) were determined for each patient pixel-by-pixel  $K_{trans}$  between the DCEto-PK cGAN and ETM. nRMSE and nMAE were normalized using patient standard deviation (SD) of target ETM Ktrans. ULR models were fit to compute the c-statistic, the area under the Receiver Operating Characteristic (ROC) curve, for V1  $K_{trans}$ , V2  $K_{trans}$ , and percentage  $K_{trans}$  change from V1 to V2 for both the ETM and cGAN approaches. Two-tailed and unpaired Student's t-tests were used to determine statistical significance between pre- and post-NACT.  $p < 0.05$  was considered to indicate statistical significance. Data were presented as mean  $\pm$  SD.



Figure 2. TCIA non-pCR patient tested using the DCE-to-PK cGAN. Two representative DCE MRI slices from a non-pCR patient revealed highly enhanced tumor lesions at both pre-treatment (V1) and post-NACT (V2). Notably, ETM and cGAN Ktrans maps showed excellent spatial correlation and high structural similarity (a). Quantification of tumor mean  $K_{trans}$  at V1 (n = 22 slices) and V2 (n = 21 slices) revealed a small decrease in vascular permeability at V2 by both the ETM and cGAN (b). Analysis of slice-based mean K<sub>trans</sub> (c) and pixelby-pixel K<sub>trans</sub> (**d**) showed strong correlations ( $R^2 = 0.98$ ) between the cGAN and ETM ( $p < 0.0001$ ). Mean  $\pm$ SD.

# **3. Results**

The DCE-to-PK cGAN was found to generate vascular permeability parameter maps in breast cancer TCIA patient datasets (n = 120 slices) in 2.5 seconds on average with an NVIDIA GeForce RTX 3090, significantly lowering the average computation time over 1000-fold relative to the ETM (150 min average on Intel Xeon CPU @ 3.70 GHz and 32 GB RAM). A representative non-pCR patient to NACT is depicted in **Figure 2**. DCE MRI revealed a large, highly enhanced tumor mass as well as multiple infiltrative lesions. These tumors were enhanced at both pre-treatment, V1, and post-NACT, V2. Vascular permeability K<sub>trans</sub> maps produced by the cGAN closely resembled ETM counterparts, exhibiting excellent spatial correlation and high structural similarity (**Figure 2a**). This patient was found to have elevated vascular permeability  $K_{trans}$  at V1 (ETM = 0.0233  $\pm$  0.0035 min<sup>-1</sup>; cGAN =



**Figure 3. TCIA pCR patient tested using the DCE-to-PK cGAN. Two representative DCE MRI slices from** a pCR patient revealed a highly enhanced tumor lesion at both pre-treatment, V1, and post-NACT, V2. Notably, Ktrans maps produced by the ETM and cGAN had excellent spatial correlation and high structural similarity (**a**). Quantification of tumor mean  $K_{trans}$  (**b**) at V1 (n = 19 slices) and V2 (n = 12 slices) revealed substantially decreased vascular permeability at V2 by both the ETM and cGAN ( $p < 0.0001$ ). Analysis of slice-based mean K<sub>trans</sub> (c) and pixel-by-pixel K<sub>trans</sub> (d) showed strong correlations ( $R^2 = 0.99$ ) between the cGAN and ETM ( $p <$ 0.0001). Mean  $\pm$  SD, \*\*\*\*p < 0.0001.

 $0.0230 \pm 0.0035$  min<sup>-1</sup>) that was largely retained at V2 (ETM =  $0.0213 \pm 0.0058$  min<sup>-1</sup>; cGAN =  $0.0200 \pm 0.0053$ min-1 ), with excellent agreement between the ETM and cGAN (**Figure 2b**). Linear regression (**Figure 2c and 2d**) revealed a significant linear correlation ( $p < 0.0001$ ) between the cGAN and ETM for both slice-based mean K<sub>trans</sub>  $(n = 43; R^2 = 0.98)$  and pixel-by-pixel  $K_{trans}$   $(n = 20,873; R^2 = 0.98)$  for this patient.

Similar to the non-pCR patient, as shown in **Figure 3**, DCE MRI of a representative pCR patient revealed a large, highly enhanced tumor at both V1 and V2. ETM and cGAN  $K_{trans}$  maps were almost indistinguishable, exhibiting excellent spatial correlation and high structural similarity (**Figure 3a**). While the patient was found to have elevated vascular permeability  $K_{trans}$  at V1 (ETM = 0.0383  $\pm$  0.0164 min<sup>-1</sup>; cGAN = 0.0380  $\pm$  0.0160 min<sup>-1</sup>), the patient had markedly decreased  $K_{trans}$  at V2 (ETM = 0.0155  $\pm$  0.0016 min<sup>-1</sup>; cGAN = 0.0156  $\pm$  0.0016 min<sup>-1</sup>) in



**Figure 4. DCE-to-PK cGAN deciphered intra-patient heterogeneous tumor response.** In a representative patient, two highly enhanced tumor lesions, lesion 1 (green arrow) and lesion 2 (red arrow), were identified on DCE images (a). Notably, ETM and cGAN K<sub>trans</sub> maps showed elevated vascular permeability at V1 for both lesions. Interestingly, both ETM and cGAN V2 K<sub>trans</sub> maps revealed markedly decreased vascular permeability for lesion 1 (**b**) and retained vascular permeability for lesion 2 (**c**). Slice-based mean Ktrans (**d**) and pixel-by-pixel K<sub>trans</sub> (**e**) analysis showed strong correlations ( $R^2 = 0.98$ ) between the cGAN and ETM ( $p < 0.0001$ ). Mean  $\pm$ SD, \*\*\*\*p < 0.0001.

good agreement by the ETM and cGAN (**Figure 3b**). Slice-based mean  $K_{trans}$  ( $n = 31$ ;  $R^2 = 0.99$ ) and pixel-by-pixel  $K_{trans}$  (n = 3,031;  $R^2$  = 0.99) revealed a significant linear correlation (p < 0.0001) between the cGAN and ETM for this patient (**Figure 3c and 3d**). Clearly, the DCE-to-PK cGAN accurately monitored vascular permeability responses in non-pCR (**Figure 2**) and pCR (**Figure 3**) patients with high structural similarity to the ETM.

The DCE-to-PK cGAN deciphered intra-patient heterogeneous tumor responses in good agreement to the ETM, further demonstrating its utility (**Figure 4**). Anatomical DCE images of a representative breast cancer patient revealed two tumor lesions, lesion 1 (green arrow) and lesion 2 (red arrow). Both lesions were largely enhanced at V1 and V2 (**Figure 4a**). ETM and cGAN maps showed elevated vascular permeability  $K_{trans}$  at V1 for lesion 1  $(ETM = 0.0247 \pm 0.0090 \text{ min}^{-1}; \text{ cGAN} = 0.02451 \pm 0.0091 \text{ min}^{-1})$  and lesion 2  $(ETM = 0.0283 \pm 0.0077 \text{ min}^{-1};$  $cGAN = 0.0287 \pm 0.0078$  min<sup>-1</sup>). In response to NACT, lesion 1 had markedly decreased  $K_{trans}$  (**Figure 4b**; ETM =  $0.0104 \pm 0.0030$  min<sup>-1</sup>;  $cGAN = 0.0102 \pm 0.0032$  min<sup>-1</sup>), whereas lesion 2 retained elevated permeability (**Figure** 4c; ETM =  $0.0232 \pm 0.0051$  min<sup>-1</sup>; cGAN =  $0.0239 \pm 0.0050$  min<sup>-1</sup>). Importantly, these heterogeneous responses between the two lesions were recapitulated with high accuracy by the cGAN. Moreover, there was significant linear

<b>Patient</b>	<b>RMSE</b>	<b>MAE</b>	nRMSE	nMAE	$\mathbf{R}^2$	CCC
Patient 01	1.98E-3	1.28E-3	0.167	0.108	0.981	0.985
Patient 05	$2.63E-3$	$1.62E-3$	0.113	0.069	0.988	0.993
Patient 06	1.15E-3	7.70E-4	0.181	0.120	0.973	0.985
Patient 08	$1.04E-3$	5.37E-4	0.100	0.052	0.990	0.995
Patient 10	2.18E-3	$1.33E-3$	0.264	0.161	0.947	0.968
Patient 12	3.19E-3	$1.11E-3$	0.316	0.110	0.954	0.959
Patient 13	1.95E-3	1.38E-3	0.223	0.157	0.981	0.978
Patient 14	1.97E-3	$1.26E - 3$	0.138	0.088	0.982	0.991
Patient 15	1.06E-3	$6.01E-4$	0.114	0.065	0.987	0.994
Patient 16	2.46E-3	$1.63E-3$	0.176	0.116	0.973	0.985

**Table 1. Predictive performance of DCE-to-PK cGAN**

correlation ( $p < 0.0001$ ) between the cGAN and ETM for both slice-based mean  $K_{trans}$  ( $n = 28$ ;  $R^2 = 0.99$ ) and pixelby-pixel  $K_{trans}$  ( $n = 6,471$ ;  $R^2 = 0.98$ ) for this patient (**Figure 4d and 4e**).

Error analysis (**Table 1**) across all patient's intratumoral pixel-by-pixel  $K_{trans}$  values showed low RMSE and MAE for the cGAN. Importantly, all patients had less predictive error than the SD seen on ETM maps (nRMSE and nMAE < 1).  $\mathbb{R}^2$  and CCC revealed strong correlations between the cGAN and ETM approaches for quantifying  $K_{trans}$  ( $R^2$  and CCC > 0.9). As shown in **Figure 5**, an intratumoral ensemble of tumor mean  $K_{trans}$  ( $n = 20$ ), slicebased mean  $K_{trans}$  (n = 359), and pixel-by-pixel  $K_{trans}$  (n = 143,640) across all patients demonstrated significant linear correlations ( $p < 0.0001$ ) between the ETM and cGAN ( $R^2 \ge 0.98$ ). Negligent over or under-prediction by the cGAN for tumor mean, slice-based mean, and pixel-by-pixel  $K_{trans}$  was found from Bland-Altman plot analysis (not shown here).

Pathological evaluations of individual patient responses to NACT were performed previously by Oregon Health & Science University in a study by Huang et al<sup>7</sup>. In this group of patients, seven were identified as non-pCR and three were pCR. Based on these classifications, we plotted tumor mean  $K_{trans}$  at V1 and V2 as well as percentage



**Figure 5. Patient ensemble linear regression analysis.** Tumor mean K<sub>trans</sub> (a) across all patients at both V1 and V2 (n = 20) revealed a significant linear correlation between the ETM and cGAN (p < 0.0001;  $R^2 = 0.99$ ). Similarly, intratumoral slice-based mean  $K_{trans}$  (**b**) and pixel-by-pixel  $K_{trans}$  (**c**) had strong linear correlations between the ETM and cGAN with  $R^2 = 0.99$  (n = 359) and 0.98 (n = 143,640), respectively (p < 0.0001).

Ktrans change between V1 and V2 (**Figure 6**). Clearly, scatter plots of tumor mean  $K_{trans}$  for both the ETM (**Figure 6a**) and cGAN (**Figure 6b**) showed differential PK vascular permeability responses to NACT between non-pCR (red triangles) and pCR patients (black circles). The pCR patients had a more prominent decrease in tumor mean  $K_{trans}$ after the first cycle of NACT in comparison to non-pCR patients. There was great



**Figure 6. DCE-to-PK cGAN enables early prediction of responders to NACT.**  Scatter plots of tumor mean  $K_{trans}$  for both ETM (**a**) and cGAN (**b**) revealed differential vascular changes to NACT between non-pCR patients (red triangles) and pCR patients (black circles). Notably, in good agreement with previous work<sup>7</sup>,  $K_{trans}$  percentage changes (c) clearly distinguished the non-pCR from the pCR patients.

agreement in  $K_{trans}$  values and percentage  $K_{trans}$  change between the cGAN and ETM. Importantly, the DCE-to-PK cGAN quantified percentage  $K_{trans}$  change clearly distinguished non-pCR from pCR patients in good agreement to the ETM (Figure 6c). Furthermore, concurring with previous work leveraging this dataset<sup>7</sup>, ULR models found percentage  $K_{trans}$  change was an excellent **Table 2. ULR c-statistic values for early prediction of pathologic** 



DCE MRI is becoming increasingly common in standard-of-care cancer imaging regimens attributable to its noninvasive ability to assess tumor biology in the form of tumor microvascular permeability. However, DCE MRI has several limitations that still hinder its widespread clinical implementation. These limitations include variability in image acquisition protocols, the need for standardized PK analysis techniques, and the requirement for advanced computational tools. Addressing these issues could significantly enhance the clinical utility of DCE MRI in oncology. To this end, we have developed a novel DCE-to-PK cGAN approach for translation of DCE MRI to PK vascular permeability parameter maps.

An important finding from the present study is that the DCE-to-PK cGAN significantly lowered the computational time over 1000-fold relative the ETM, while generating synthetic  $K_{trans}$  maps that closely resembled real ETM K<sub>trans</sub> maps (**Figures 2-4**). In addition to the excellent spatial correlation and visual structural similarity between cGAN and ETM maps, we found low intratumoral error (nMAE and nRMSE < 1.0) and strong correlation  $(R^2$  and  $CCC > 0.9)$  across all patients (**Table 1**). Furthermore, an ensemble of tumor-bearing K<sub>trans</sub> values for all patients (**Figure 5**) revealed a significant linear correlation ( $p < 0.0001$ ) for tumor mean, slice-based mean, and

pixel-by-pixel  $K_{trans}$  ( $R^2 \ge 0.98$ ). Clearly, the DCE-to-PK cGAN can efficiently quantify vascular permeability in good agreement to conventional PK models.

Several previous studies have identified DCE MRI derived vascular permeability parameters as a potential biomarker for early assessment of NACT pathological response in breast cancer patients<sup>2,3,7</sup>. These studies have suggested that NACT-induced tumor function changes, particularly vascular function changes, precede changes in tumor size, which is the current standard of care for evaluating treatment response<sup>2,3,7</sup>. Furthermore, these early changes in vascular function are prognostic for pathological response and may serve as a useful surrogate for early monitoring of treatment response. Indeed, as shown in **Figure 2 and 3**, representative non-pCR and pCR patients from TCIA data collection as used in this study have minimal change in tumor size following the first cycle of NACT. In good agreement with previous work of this data collection<sup>7</sup>, the non-pCR patient was found to retain elevated vascular permeability based on the ETM, whereas the pCR patient exhibited a significant decrease in vascular permeability parameter  $K_{trans}$  (p < 0.0001). Importantly, the DCE-to-PK cGAN correctly quantified these early vascular permeability changes in both non-pCR and pCR patients in excellent agreement to the ETM.

It is noteworthy that tumors in both non-pCR (**Figure 2**) and pCR (**Figure 3**) breast cancer patients were highly enhanced on DCE MRI at both V1 and V2, indicating lesions were permeable both pre- and post-NACT. However, as discussed above, non-pCR and pCR patients showed differential vascular permeability  $K_{trans}$  changes in response to NACT.  $K_{trans}$  is related to the rate of enhancement on DCE MRI scans, indicating that the rate of enhancement may be more important for monitoring treatment response than the presence of high contrast enhancement. Another noteworthy finding is that the DCE-to-PK cGAN not only accurately quantified vascular permeability changes between individual patients, but also accurately deciphered intra-patient heterogeneous tumor responses, as evidenced in the patient shown in **Figure 4**. We anticipate the DCE-to-PK cGAN can serve as a useful tool for monitoring both inter-patient and intra-patient tumor vascular permeability responses, which can aid in clinical decision-making and personalized medicine.

In a previous work leveraging this data collection by Huang et al., a multi-QIN center data analysis challenge revealed significant variability of vascular permeability parameters derived from twelve different DCE MRI computational modeling algorithms<sup>7</sup>. The authors found up to a two-fold to four-fold parameter value difference between software tools, albeit controlling for tumor ROI definition as well as  $R_{10}$  and AIF measurements. A separate study similarly revealed a 10-fold to 100-fold difference in vascular permeability parameters derived from different software tools in DCE MRI<sup>31</sup>. These differences in vascular permeability parameters based on different PK models and software tools could lead to non-reproducible results in DCE MRI studies across different institutions. Huang's study showed strong agreement in percentage change of vascular permeability parameter  $K_{trans}$ for nearly all software tools, of which 11 out of 12 were good (0.8  $\leq$  c-statistic  $\leq$  0.9) to excellent (c-statistic  $\geq$  0.9) predictors of pathologic response by ULR analysis<sup>7</sup>.

In the present study leveraging the same dataset from Huang's study<sup>7</sup>, we found our in-house ETM software returned smaller vascular permeability  $K_{trans}$  values than the majority of the other computational modeling algorithms used in the data analysis challenge. Our vascular permeability  $K_{trans}$  values produced by in-house ETM

software were similar to the BWH-3D Slicer TM software tool7,32 used in the data analysis challenge (**Figure 6a**). The DCE-to-PK cGAN approach also predicted smaller vascular permeability  $K_{trans}$  values relative to the other software tools as the deep learning approach was trained to recapitulate our ETM software (**Figure 6b**). Nevertheless, percentage K<sub>trans</sub> changes for both our ETM software and cGAN are in excellent agreement to all software tools as used in Huang's study (**Figure 6c**) 7 . Similarly, we show that both our ETM software and cGAN approach are excellent predictors of pathologic response (c-statistic  $= 1.0$ ) when considering percentage change of vascular permeability parameter  $K_{trans}$  (**Table 2**).

As revealed in the present study, an image-to-image cGAN may serve as a useful tool in DCE MRI for monitoring treatment response. However, it is worth noting that the current sample size of the study cohort is small. Including additional training and testing cases will further elucidate the long-term utility of the DCE-to-PK cGAN approach. To this end, with the recent surge in the development of novel artificial intelligence (AI) approaches, specifically generative AI, it would be interesting to evaluate the ability of these approaches to generate synthetic DCE MRI. Such synthetic data could be used to augment and expand training cases, potentially improving the performance of the DCE-to-PK cGAN. Moreover, a direct comparison of the DCE-to-PK cGAN to previously developed deep learning algorithms for efficient DCE MRI mapping was not performed in the current study but is warranted. It would similarly be interesting to evaluate an ensemble learning approach to combine the knowledge of alternative deep learning frameworks, which may lead to enhanced predictive accuracy over implementation of a single deep learning model.

# **5. Conclusion**

In summary, we have developed a novel image-to-image cGAN approach for efficient translation of DCE MRI to PK vascular permeability parameter maps. The DCE-to-PK cGAN can generate vascular permeability parameter maps that closely resemble real parameter maps and reduce the computational time over 1000-fold compared to conventional computational tools. Importantly, percentage change of vascular permeability parameter K<sub>trans</sub> maps derived from the cGAN approach were excellent early predictors of pathologic response in breast cancer patients to NACT, highlighting its utility for monitoring treatment response. We anticipate the DCE-to-PK cGAN will serve as a useful tool to standardize PK analysis and reduce computational complexity in both general oncologic imaging and clinical trials.

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#### **Data Availability**

The dataset used in the present study are publicly available through The Cancer Imaging Archive<sup>7,25</sup>.

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# **References**

- 1 Asaoka, M., Gandhi, S., Ishikawa, T. & Takabe, K. Neoadjuvant Chemotherapy for Breast Cancer: Past, Present, and Future. *Breast Cancer (Auckl)* **14**, 1178223420980377, doi:10.1177/1178223420980377 (2020).
- 2 Tudorica, A. *et al.* Early Prediction and Evaluation of Breast Cancer Response to Neoadjuvant Chemotherapy Using Quantitative DCE-MRI. *Transl Oncol* **9**, 8-17, doi:10.1016/j.tranon.2015.11.016 (2016).
- 3 Cheng, Q. *et al.* The Diagnostic Performance of DCE-MRI in Evaluating the Pathological Response to Neoadjuvant Chemotherapy in Breast Cancer: A Meta-Analysis. *Front Oncol* **10**, 93, doi:10.3389/fonc.2020.00093 (2020).
- 4 Kaufmann, M. *et al.* Recommendations from an international consensus conference on the current status and future of neoadjuvant systemic therapy in primary breast cancer. *Ann Surg Oncol* **19**, 1508-1516, doi:10.1245/s10434-011-2108-2 (2012).
- 5 Reig, B. *et al.* Breast MRI for Evaluation of Response to Neoadjuvant Therapy. *Radiographics* **41**, 665- 679, doi:10.1148/rg.2021200134 (2021).
- 6 Suo, S. *et al.* Diffusion-weighted MRI for predicting pathologic response to neoadjuvant chemotherapy in breast cancer: evaluation with mono-, bi-, and stretched-exponential models. *J Transl Med* **19**, 236, doi:10.1186/s12967-021-02886-3 (2021).
- 7 Huang, W. *et al.* Variations of dynamic contrast-enhanced magnetic resonance imaging in evaluation of breast cancer therapy response: a multicenter data analysis challenge. *Transl Oncol* **7**, 153-166, doi:10.1593/tlo.13838 (2014).
- 8 Fang, K. *et al.* Convolutional neural network for accelerating the computation of the extended Tofts model in dynamic contrast-enhanced magnetic resonance imaging. *J Magn Reson Imaging* **53**, 1898- 1910, doi:10.1002/jmri.27495 (2021).
- 9 Arledge, C. A. *et al.* Deep learning quantification of vascular pharmacokinetic parameters in mouse brain tumor models. *Front Biosci (Landmark Ed)* **27**, 99, doi:10.31083/j.fbl2703099 (2022).
- 10 Arledge, C. A. *et al.* Transfer Learning Approach to Vascular Permeability Changes in Brain Metastasis Post-Whole-Brain Radiotherapy. *Cancers (Basel)* **15**, doi:10.3390/cancers15102703 (2023).
- 11 Ulas, C. *et al.* Convolutional Neural Networks for Direct Inference of Pharmacokinetic Parameters: Application to Stroke Dynamic Contrast-Enhanced MRI. *Front Neurol* **9**, 1147, doi:10.3389/fneur.2018.01147 (2018).
- 12 Ottens, T. *et al.* Deep learning DCE-MRI parameter estimation: Application in pancreatic cancer. *Med Image Anal* **80**, 102512, doi:10.1016/j.media.2022.102512 (2022).
- 13 Zou, J., Balter, J. M. & Cao, Y. Estimation of pharmacokinetic parameters from DCE-MRI by extracting long and short time-dependent features using an LSTM network. *Med Phys* **47**, 3447-3457, doi:10.1002/mp.14222 (2020).
- 14 Isola, P., Zhu, J. Y., Zhou, T. & Efros, A. A. Image-to-Image Translation with Conditional Adversarial Networks. *arXiv:1611.07004v3* (2016).
- 15 Chrysos, G. G., Kossaifi, J. & Zafeiriou, S. RoCGAN: Robust Conditional GAN. *Int J Comput Vision* **128**, 2665-2683, doi:10.1007/s11263-020-01348-5 (2020).
- 16 Yi, X., Walia, E. & Babyn, P. Generative adversarial network in medical imaging: A review. *Med Image Anal* **58**, 101552, doi:10.1016/j.media.2019.101552 (2019).
- 17 Han, C. *et al.* GAN-based synthetic brain MR image generation. *IEEE 15th International Symposium on Biomedical Imaging (ISBI 2018)*, 734-738 (2018).
- 18 Li, Y., Zhang, K., Shi, W., Miao, Y. & Jiang, Z. A Novel Medical Image Denoising Method Based on Conditional Generative Adversarial Network. *Comput Math Methods Med* **2021**, 9974017, doi:10.1155/2021/9974017 (2021).
- 19 Xun, S. *et al.* Generative adversarial networks in medical image segmentation: A review. *Comput Biol Med* **140**, 105063, doi:10.1016/j.compbiomed.2021.105063 (2021).
- 20 Xue, Y., Xu, T., Zhang, H., Long, R. & Huang, X. SegAN: Adversarial Network with Multi-scale L1 Loss for Medical Image Segmentation. *arXiv:1706.01805* (2017).

- 21 Nasser, S. A., Shamsi, S., Bundele, V., Garg, B. & Sethi, A. Perceptual cGAN for MRI Super-resolution. *Annu Int Conf IEEE Eng Med Biol Soc* **2022**, 3035-3038, doi:10.1109/EMBC48229.2022.9871832 (2022).
- 22 Yang, Q. *et al.* MRI Cross-Modality Image-to-Image Translation. *Sci Rep* **10**, 3753, doi:10.1038/s41598- 020-60520-6 (2020).
- 23 Hu, L. *et al.* Calculation of Apparent Diffusion Coefficients in Prostate Cancer Using Deep Learning Algorithms: A Pilot Study. *Frontiers in Oncology* **11**, doi:10.3389/fonc.2021.697721 (2021).
- 24 Wang, B. *et al.* Quantitative Cerebral Blood Volume Image Synthesis from Standard MRI Using Imageto-Image Translation for Brain Tumors. *Radiology* **308**, e222471, doi:10.1148/radiol.222471 (2023).
- 25 Huang, W. *et al.* Variations of dynamic contrast-enhanced magnetic resonance imaging in evaluation of breast cancer therapy response: a multicenter data analysis challenge (QIN Breast DCE-MRI) (Version 2) [Data set]. *The Cancer Imaging Archive*, doi:https://doi.org/10.7937/k9/tcia.2014.a2n1ixox (2014).
- 26 Tudorica, L. A. *et al.* A feasible high spatiotemporal resolution breast DCE-MRI protocol for clinical settings. *Magn Reson Imaging* **30**, 1257-1267, doi:10.1016/j.mri.2012.04.009 (2012).
- 27 Li, X. *et al.* Dynamic NMR effects in breast cancer dynamic-contrast-enhanced MRI. *Proc Natl Acad Sci U S A* **105**, 17937-17942, doi:10.1073/pnas.0804224105 (2008).
- 28 Tofts, P. S. Modeling tracer kinetics in dynamic Gd-DTPA MR imaging. *J Magn Reson Imaging* **7**, 91- 101, doi:10.1002/jmri.1880070113 (1997).
- 29 Crowe, W. *et al.* MRI evaluation of the effects of whole brain radiotherapy on breast cancer brain metastasis. *Int J Radiat Biol* **95**, 338-346, doi:10.1080/09553002.2019.1554920 (2019).
- 30 Mao, X. *et al.* On the Effectiveness of Least Squares Generative Adversarial Networks. *IEEE Trans Pattern Anal Mach Intell* **41**, 2947-2960, doi:10.1109/TPAMI.2018.2872043 (2019).
- 31 Heye, T. *et al.* Reproducibility of dynamic contrast-enhanced MR imaging. Part I. Perfusion characteristics in the female pelvis by using multiple computer-aided diagnosis perfusion analysis solutions. *Radiology* **266**, 801-811, doi:10.1148/radiol.12120278 (2013).
- 32 Fedorov, A. *et al.* 3D Slicer as an image computing platform for the Quantitative Imaging Network. *Magnetic Resonance Imaging* **30**, 1323-1341, doi:10.1016/j.mri.2012.05.001 (2012).