

Proteomic Biomarkers for Bisphenol A–Early Exposure and Women’s Thyroid Cancer

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Purpose

For the target treatment and prevention of women’s increased thyroid cancer, we focused on risks of environmental exposure to endocrine disrupting chemicals, particularly bisphenol A (BPA), and its high susceptible exposure-timing, particularly early exposure in lives.

Materials and Methods

Female ICR mice were exposed to BPA *in utero* and in early life (15, 75, and 300 mg/L of drinking water via pregnant mice and lactation). We identified BPA-responsive proteins in mice thyroid by two-dimensional gel electrophoresis, image analyses, and electrospray ionization quadrupole time-of-flight mass spectrometry. We further analyzed expression of the BPA-responsive proteins in women thyroid cancer patients (n=28).

Results

We found the altered 17 proteins in BPA dose-dependent manner among the thyroid tissues of offspring mice and identified nine proteins of them, including Anxa6, Atp5b, Hspa5, and Vcp, etc. In addition, we observed the positive association between blood BPA levels and mRNA expression of the ANXA6 and VCP not in normal but thyroid cancer tissues.

Conclusion

Our study provides ANXA6 and VCP as proteomic biomarkers for BPA–early life exposure and their potential for women’s thyroid cancer.

Key words

Bisphenol A, Endocrine disruptors, Thyroid neoplasms, Women, Maternal exposure, Proteomics, Annexin A6, Valosin-containing protein

Introduction

Bisphenol A [4,4'-(propane-2, 2-diyl)diphenol, BPA] has been speculated as an endocrine disrupting chemical (EDC); however, people are easily exposed to BPA in their daily lives, because of its wide use as glass substitutes [1]. BPA is detected in urine, blood, placenta, breast milk, and even umbilical cord [2]. Numerous studies also suggest that BPA perturbs thyroid functions through multiple mechanisms. For example, a recent Zebrafish study showed BPA interferences with thyroid specific gene expression [3]. In addition, BPA exposure seems to elevate triiodothyronine, while decrease thyroxin and thyroid-stimulating hormone (TSH) [4]. Therefore, thyroid can be a target organ of BPA toxicity [5]. We recently reported BPA is metabolized into toxic metabolites by cytochrome P450 2E1 [1]; however, its consistent mechanisms between *in vitro* or *in vivo* and human are not clear, yet.

Among thyroid diseases, thyroid cancer has been emphasized due to its incidence rate continuously increased across the world over the last three decades [6]. The dramatic increase in thyroid cancer incidences might be related to the advances of diagnostic technologies, overdiagnosis [7], environmental and lifestyle factors, etc. [8]. However, the increased medical surveillance or improved detection methods alone cannot fully explain the increase. Multiple factors might be associated together for risks of thyroid cancer [9]. Particularly, hormonal and reproductive factors have shown significant associations with thyroid cancer risks [10]. The estrogens have been conjectured to be one of the main causes of thyroid diseases [11] because of the higher thyroid cancer rate and an earlier peak rate of women than men. Therefore, environmental estrogens, i.e., EDCs, have been suspected as risk factors, but little is known about their risks for human thyroid cancer.

A few studies have explored the aspect of toxic end points of BPA. The results suggested the theory of 'two hits' or 'second hit' for instance hormone and BPA exposure as an alternative mechanism of BPA mutagenesis or carcinogenesis [12,13]. In addition, some studies suggested epigenetic alterations or genetic memory as BPA toxic mechanisms [14]. Particularly, the Developmental Origins of Health and Disease (DOHaD) hypothesis can be applied to the consequences of epigenetic memory or reprogramming. In other words, environmental and nutritional factors influence developmental plasticity during the critical periods of development, thereby altering susceptibility to diseases later in life [15]. Therefore, many researchers have highlighted the importance of exposure timing [16]. Therefore, we focused on risks of environmental exposure to BPA and its high susceptible exposure-timing, particularly early exposure in lives, for the

target treatment and prevention of women's increased thyroid cancer. We designed *in utero* and early life exposure to BPA in mice and identified BPA-responsive protein biomarkers in thyroid tissues of female offspring mice. We also confirmed these BPA exposure biomarkers in women thyroid cancer patients.

Materials and Methods

1. Animal care and treatment

ICR mice in the first week of pregnancy were obtained from Orient Co. (Seoul, Korea) and housed under a 12-hr light/dark cycle (lights on 07:00-19:00) at 21-24°C and 40%-60% humidity with unlimited access to drinking water. The pregnant mice were fed with pellet food (Samtako, Seoul, Korea) *ad libitum*, and were administered three different concentrations of BPA (Sigma-Aldrich Co., St. Louis, MO), 15, 75, and 300 mg/L dissolved in 0.1% ethanol/water using glass bottles for 34-36 days, i.e., from day 7 of pregnancy to lactation (day 21 post-delivery). After the lactation, the offspring mice were allowed an access to untreated water. Fifteen or sixteen pregnant mice were administered with each concentration of BPA. In contrast, untreated control pregnant mice (n=18) were provided with 0.1% ethanol/water *ad libitum*. The 7-week-old female offspring were sacrificed in CO₂ chamber and obtained their thyroids. All animal experimental procedures were complied with the University Animal Care and Use Committee at Sookmyung Women's University.

2. Proteomic analyses

Total protein was extracted from 10 mg of the organs using our previous method [17] with minor modification. After centrifuging the homogenized samples (14,000 rpm for 10 minutes), we measured the protein concentrations using the Bradford assay. Two-dimensional gel electrophoresis (2D-PAGE) and image analyses were performed with the extracted 300 µg of each protein sample. The digitalized image was analyzed with PDQUEST software (ver. 6.1, Bio-Rad, Richmond, CA). After analysis of the nano-electrospray ionization on a quadrupole time-of-flight (Q-TOF) mass spectrometer (Micromass, Manchester, UK) with 30 µL of digested peptide mixture, all tandem mass spectrometry (MS/MS) spectra derived from spot were identified by NCBIInr and EST databases using the MASCOT search program [17]. The optical density of specific proteins was statistically analyzed with the Mann-Whitney test (significance at

$p < 0.05$). We duplicated the 2D gel analyses with two different mice for each treatment (control, 15, 75, and 300 mg BPA/L of water, total=8).

3. Thyroid tissues from cancer patients

Matched pairs of thyroid tumor and adjacent non-tumor tissues were collected from women thyroid cancer patients ($n=28$; age, 47.4 ± 11.6 years; papillary carcinoma, 100%; metastasis, 10.4% of the subjects), who received a surgical treatment at Hanyang University Hospital, Seoul, Korea. Their tissues were stored in sterile e-tubes and frozen at -80°C until RNA isolation. All of the procedures for the human study were approved by the Clinical Research Ethics Committee of Hanyang University Hospital.

4. Real-time polymerase chain reaction analysis

Total RNA was isolated from thyroid tumor and normal tissues in each patient (~ 20 mg) with Trizol reagent (Thermo Fisher Scientific, Carlsbad, CA) according to the manufacturer's instructions. The RNA quality was assessed by conventional agarose gel electrophoresis, and RNA concentration was determined using a Qubit fluorometer (Thermo Fisher Scientific). Total RNA ($1 \mu\text{g}$) from each sample was reverse transcribed to cDNA using a TaqMan RT Reagent kit (Thermo Fisher Scientific). We performed quantitative real-time polymerase chain reaction (PCR) analyses with TaqMan Gene Expression Assays [17]. Taqman qPCR primers and probes were as follows: *ATP5B* (Hs00969569_m1), *ANXA6* (Hs00241762_m1), *HSPA5* (Hs00607129_gH), and valosin-containing protein (*VCP*; Hs00997642_m1). The reaction was run on a 7500 Real-Time PCR System (Thermo Fisher Scientific) under the following conditions: 2 minutes at 50°C for AmpErase UNG activation and 10 minutes at 95°C for UNG inactivation, followed by 40 cycles of 15 seconds at 95°C for denaturing, and 1 minute at 60°C for annealing and extension. After constructing standard curves, we quantified mRNA expression levels of *ATP5B*, *ANXA6*, *HSPA5*, and *VCP*, and normalized with respect to the expression of 18S rRNA, of which primers and probes were obtained from Thermo Fisher Scientific.

5. Analyses of blood BPA in thyroid cancer patients

BPA levels in blood specimens were analyzed using reverse phase-HPLC/FLD via liquid-liquid extraction with minor modifications [2]. In brief, $500 \mu\text{L}$ of blood sample and $50 \mu\text{L}$ of $4.13 \mu\text{M}$ bisphenol B (internal standard) were mixed with $30 \mu\text{L}$ of 2.0 M sodium acetate (pH 5.0), and hydrolyzed with $20 \mu\text{L}$ of β -glucuronidase ($2,000 \text{ U}$) for 5 hours at 37°C in a shaking water bath. After hydrolysis, $100 \mu\text{L}$ of 2.0 M

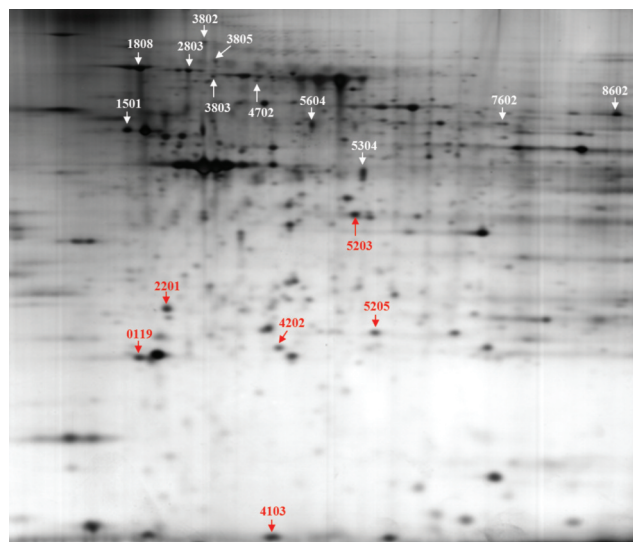


Fig. 1. The 2D gel image shows 17 protein spots were altered in bisphenol A (BPA) dose-dependent manner on thyroid tissue of female offspring by prenatal exposure to BPA.

HCl was added and the mixtures were extracted twice with 5 mL of ethylacetate. Six milliliters of the supernatant was evaporated in a Savant SpeedVac centrifuge (Savant Instruments Inc., Holbrook, NY) and dissolved in $300 \mu\text{L}$ of 60% acetonitrile. One hundred microliters of the resultant was injected into the HPLC/FLD apparatus. The HPLC/FLD system was the same with the previous one [2]. Samples were analyzed with gradient elution with water (solvent A) and acetonitrile (solvent B) (ratio of A to B, 0-35 minutes, 75:25 to 52:48; 35-40 minutes, 52:48 to 0:100; 40-45 minutes, 0:100-0:100; 45-55 minutes, 0:100 to 75:25; 55-70 minutes, 75:25). The samples were analyzed at 225 nm (excitation) and 305 nm (emission) with 1 mL/min of flow.

6. Statistical analysis

Shapiro-Wilk W test was used to examine the degree of normal distribution of BPA levels. The normality of the expression data was verified using the Kolmogorov-Smirnov test. The Wilcoxon signed rank test was used to compare levels of gene expression of *ANXA6*, *ATP5B*, *HSPA5*, and *VCP* between tumor and adjacent non-tumor tissues. Pearson's correlation analysis was used to determine whether there were differences in correlation of the gene expressions between tumor and non-tumor tissues. All statistical analyses were conducted using the JMP ver. 4.0.2 package (SAS Institute Inc., Cary, NC).

Table 1. Identification of differentially expressed proteins by BPA-dose dependent manner in mice

Spot No. ^{a)}	Accession No. ^{b)}	Protein name (description) ^{c)}	Molecular function ^{c)}
Up-regulated			
4702	gi 148701559	Anxa6 (annexin A6)	Calcium-dependent phospholipid binding, calcium ion bonding
5205	gi 27753960	Abhd14b (abhydrolase domain containing 14b)	Cytoplasm hydrolase activity, metabolic process
5604	gi 3169729	Hls2 (hematopoietic lineage switch 2)	Plasma glutamate carboxypeptidase
8602	gi 453155	Krt9 (keratin 9)	Intermediate filament, perinuclear region of cytoplasm
1808	gi 121570	Hspa5 (78 kDa glucose-regulated protein)	Protein binding, ER overload response, cellular response to IL-4
2803	gi 119531	Pdia4 (protein disulfide-isomerase A4)	Protein disulfide isomerase
3802	gi 6005942	Vcp (valosin-containing protein)	Protein binding ubiquitin-dependent protein catabolic process
Down-regulated			
1501	gi 104	Atp5b (ATP synthase beta subunit)	Lipid metabolic process, MHC class 1 protein binding
4202	gi 10946936	Ak1 (adenylate kinase 1)	ATP metabolic process, nucleoside diphosphate biosynthetic process

ER, estrogen receptor; IL-4, interleukin 4. ^{a)}Location of the protein resolved by two-dimensional gel electrophoresis (Fig. 1) and identified by matrix assisted laser desorption ionization–time of flight mass spectrometry, ^{b)}Protein accession number from the NCBI nonredundant database, ^{c)}Annotation from the NCBI non-redundant database.

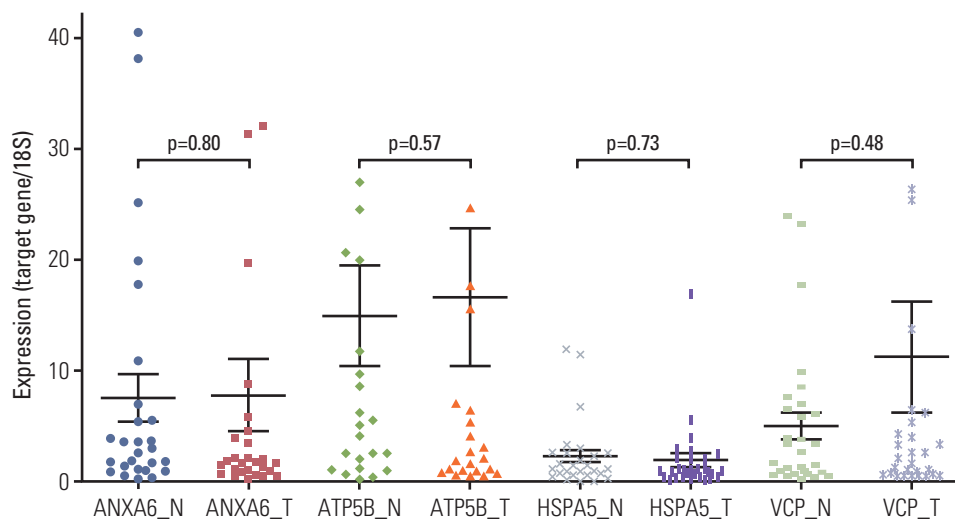


Fig. 2. No differences in expression of *ANXA6*, *ATP5B*, *HSPA5*, or *VCP* between normal and tumor thyroid tissues in the women thyroid patients. N, normal; T, tumor.

Results

When the pregnant ICR mice were administered 15, 75, and 300 mg of BPA/L in water from day 7 of pregnancy to lacta-

tion, the BPA intake for each group was estimated at approximately 8.9±1.8 mg/kg/day, 47.1±5.8 mg/kg/day, and 171.1±16.8 mg/kg/day, respectively. These levels were established by the lowest-observed adverse-effect level (50 mg/kg/day) of BPA [18]. There were no statistically significant differences

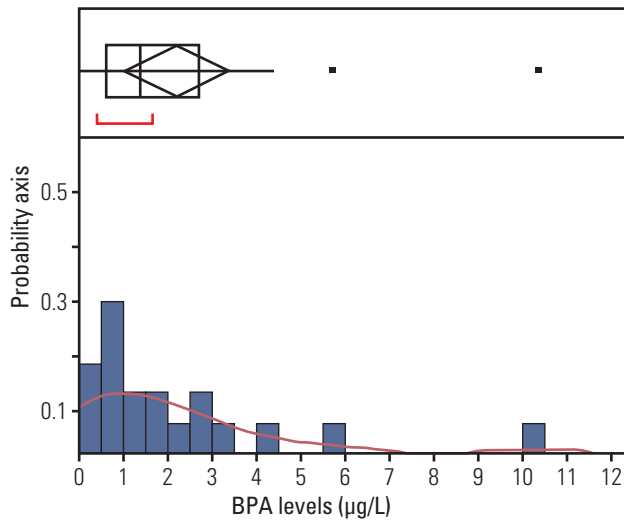


Fig. 3. Distribution of blood bisphenol A (BPA) levels. Upper part of the figure shows an outlier box plot with the square in the box showing the interquartile range: median, 1.38 µg/L.

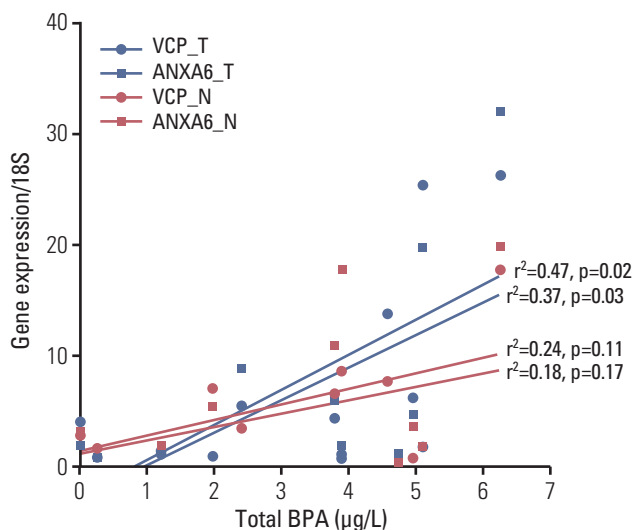


Fig. 4. The positive associations between blood bisphenol A (BPA) levels and gene expressions of *ANXA6* or *VCP* in tumor rather than normal tissues among the women thyroid cancer patients. Upper blue line, *VCP* in tumor tissues; lower blue line, *ANXA6* in tumor tissues; upper red line, *VCP* in normal tissue; lower red line, *ANXA6* in normal tissues; N, normal; T, tumor; For the view of figure, we excluded an outlier, who had 10.32 µg/L of blood BPA by outlier test of Grubbs [19].

in offspring's body weight due to BPA exposure.

Fig. 1 shows a representative 2D-PAGE gel of the proteomic analysis. Comparison of abundant proteins on 2D gels using the PDQUEST program revealed that the expression of 17 proteins was significantly changed in a dose-dependent manner (Fig. 1): 11 proteins were upregulated, and six downregulated by BPA exposure. However, we identified nine proteins out of the 17 proteins using the electrospray ionization Q-TOF mass spectrometry analysis (Table 1).

Women's resected thyroid tumor and adjacent nontumor tissues were used to compare the expression of BPA-responsive proteomic biomarkers. The protein biomarkers identified in mice were examined for human. Four genes, *ANXA6*, *ATP5B*, *HSPA5*, and *VCP*, were selected based on their molecular functions as potential BPA-responsive markers in human. Three genes, *ANXA6*, *HSPA5*, and *VCP*, were up-regulated by BPA in mice thyroid tissues (Table 1). Expression of the four genes was assessed using matched tumor and adjacent nontumor tissues (n=28). There were no significant differences in the gene expression between the tumor and adjacent nontumor tissues ($p > 0.05$) (Fig. 2), although the mean expression level of *VCP* was somewhat higher in the tumor tissues than the adjacent non-tumor tissues.

BPA was detected in 90% of patients' blood with the HPLC/FLD system. The limit of quantification was 0.39 ng/mL. The range of total BPA in the blood was 0-10.32 ng/mL (Fig. 3). The validity of BPA protein biomarkers from mice was examined in thyroid cancer patients by correlating BPA levels and gene expression. The results revealed strong associations between blood BPA levels and expression of the *VCP* or *ANXA6* in thyroid tumor tissues (Fig. 4). In contrast, this trend was not shown in thyroid normal tissues.

Discussion

BPA showed proteomic alteration in various organs. For example, BPA negatively affected sperm motility, viability, mitochondrial functions, and intracellular ATP levels by activating the mitogen-activated protein kinase, phosphatidylinositol 3-kinase, and protein kinase-A pathways and induced differential expressions of 24 proteins *in vitro* [20]. We also reported that some of BPA-responsive proteins, Apo-AI, DppIII, Set and Vat, were involved in the anti-inflammatory response and tumorigenesis in immune organs, e.g., the thymus and spleen of mice [17,21]. In addition, some *in vivo* studies showed the impact of BPA on the thyroid. For example, BPA exposure in pregnant mice induced CpG hypomethylation of the promoter region of *Cdk5* activator-binding protein (*Cabp^{LAP}*) [22] and *Cdk5*

activity via Rb is known to be critical to tumorigenesis and progression of medullary thyroid carcinoma [23]. However, these findings are not confirmed in human, yet. Therefore, the present study was performed to fill the gap between humans and animals and was focused on how BPA interferes thyroid cancer via BPA-responsive proteins.

We identified the BPA-responsive proteins in the thyroids of pregnant mice (Table 1). Among these, nine proteins are involved in the regulation of lipid metabolism, protein catabolism, and ubiquitin/proteasome-degradation pathways. We further corroborated these findings by using thyroid tumor tissues from women thyroid cancer patients to determine whether BPA exposure affects thyroid cancer. Our study revealed a significant correlation between the levels of BPA in blood and the gene expression of the BPA responsive proteins, e.g., *ANXA6* and *VCP*, in thyroid tissues. Annexin A6 belongs to the highly conserved annexin protein family. The function of *ANXA6* is linked to its ability to bind phospholipids in a Ca^{2+} -dependent manner, thereby interacting with cellular membranes in a dynamic, reversible, and regulated fashion. It is also considered to be a tumor suppressor that is downregulated in squamous cervical cancer [24], breast cancer [25] via *RAS/MAPK* or *EGFR* signaling pathways. *ANXA6* also increased anchorage-independent growth in breast cancer cells [26]. In the thyroid gland, TSH modulated ATP-induced Ca^{2+} signaling via *ANXA6* in thyrocytes [27]. Similarly, Xiong et al. [28] suggested that *ANXA6* is a potential biomarker for papillary thyroid carcinoma with lymph node metastasis via proteomic analyses in patients. Therefore, our results suggest an involvement of BPA in thyroid carcinogenesis via *ANXA6*.

The protein levels of *VCP*, an inflammation regulator, also called various cellular activities (AAA+) chaperone p97, were up-regulated in mice and its mRNA expression in women's

thyroid tumor tissues (Fig. 4). *VCP* is known to govern critical steps in ubiquitin-dependent protein quality control and intracellular signaling pathways [29]. In addition, *VCP* expression was correlated with lymph node metastasis and low survival rate in thyroid cancer [30]. Therefore, the up-regulation of *Vcp* by BPA (Table 1) suggests that thyroid function may be disrupted via *VCP*-mediated inflammatory pathways. However, there were no statistically significant differences between non-tumor and tumor tissues in the expression levels of the *ANXA6* or *VCP* in the present study. Therefore, BPA may accelerate tumor progress rather than tumor incidence via *ANXA6* or *VCP*. Our previous study showed that BPA is not mutagenic but stimulates MNNG mutagenicity [13]. In addition, the second hit theory or two-hit hypothesis can explain BPA-involvement in thyroid cancer via epigenetic memory with clear carcinogens [12,13].

In the near future, we will confirm the present results in enlarged number of human subjects to overcome the study limitation, particularly small size of the human tissues, and to obtain reproducibility and reliability for the above BPA-responsive biomarkers.

In conclusion, our study provides *ANXA6* and *VCP* as proteomic biomarkers for BPA-early life exposure and their potential for women's thyroid cancer.

Conflicts of Interest

Conflict of interest relevant to this article was not reported.

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