



CJC Open 3 (2021) 585-594

Original Article

Aortic Dimensions, Biophysical Properties, and Plasma Biomarkers in Children and Adults with Marfan or Loeys-Dietz Syndrome

Jason Z. Cui, PhD,^{a,b,‡} Kevin C. Harris, MD,^{c,‡} Koen Raedschelders, PhD,^d

Zsuzsanna Hollander, PhD,^e James E. Potts, PhD,^c Astrid De Souza, MSc,^c Marla Kiess, MD,^f

Bruce M. McManus, MD,^{e,g} Pascal Bernatchez, PhD,^h Leslie A. Raffin, MSN,^c Heidi Paine, MD,^c

Cornelis van Breemen, PhD,^a George G.S. Sandor, MD,^{c,§} and Mitra Esfandiarei, PhD^{a,i,§}

^a Department of Anesthesiology, Pharmacology and Therapeutics, British Columbia Children's Hospital Child and Family Research Institute, University of British Columbia, Vancouver, British Columbia, Canada

^b Department of Cardiothoracic Surgery, School of Medicine, Stanford University, Palo Alto, California, USA

^c Children's Heart Centre, British Columbia Children's Hospital, University of British Columbia, Vancouver, British Columbia, Canada

^d Advanced Clinical Biosystems Research Institute at Smidt Heart Institute, Los Angeles, California, USA

^e UBC James Hogg Research Centre, University of British Columbia, Vancouver, British Columbia, Canada

^fDivision of Cardiology, St Paul's Hospital, Vancouver, British Columbia, Canada

^g Department of Pathology and Laboratory Medicine, University of British Columbia, Vancouver, British Columbia, Canada

^bDepartment of Anesthesiology, Pharmacology and Therapeutics, Centre for Heart and Lung Innovation, St Paul's Hospital, University of British Columbia, Vancouver, British Columbia, Canada

ⁱ Department of Biomedical Sciences, College of Graduate Studies, Midwestern University, Glendale, Arizona, USA

ABSTRACT

Background: Aortic dilation, stiffening, and dissection are common and potentially lethal complications of Marfan syndrome (MFS) and Loeys-Dietz syndrome (LDS), which involve abnormal transforming growth factor beta (TGF- β) signalling. The relation of aortic dimensions, stiffness, and biomarker levels is unknown. The objective of this study was to measure aortic dimensions, stiffness, TGF- β and matrix metalloproteinase (MMP) levels, and endothelial function in patients with MFS, and to compare TGF- β levels in patients with MFS receiving different therapeutic regimens.

Marfan syndrome (MFS) and Loeys-Dietz syndrome (LDS) are hereditary aortopathies resulting from abnormal transforming

[‡]These authors contributed equally to this work.

E-mail: kharris2@cw.bc.ca

See page 593 for disclosure information.

RÉSUMÉ

Contexte : La dilatation, la rigidification et la dissection de l'aorte sont des complications fréquentes et parfois mortelles du syndrome de Marfan (SM) et du syndrome de Loeys-Dietz (SLD), qui sont tous deux dûs à une anomalie de la voie de signalisation du facteur de croissance transformant bêta (TGF- β). On ne connaît pas la relation entre les dimensions et la rigidité de l'aorte et la présence de biomarqueurs. Notre étude visait à mesurer les dimensions et la rigidité de l'aorte, les taux de TGF- β et de métalloprotéases matricielles (MMP) et la fonction endothéliale chez des patients atteints du SM, et à les comparer aux

growth factor beta (TGF- β) signalling.^{1,2} The clinical features of both are pleiotropic and they share phenotypic features and, most significantly, aortic dilation and dissection leading to early death.³ However, with recent improvements in the management of MFS, the average life expectancy has increased to almost normal.⁴ The abnormal cellular signalling mechanisms in these syndromes are complex.⁵⁻⁷ The prevailing theory stipulates that fibrillin-1 mutations cause the release of fibrillin-1-bound latent TGF- β , resulting in increased levels of circulating TGF- β , which initiates intracellular cascades with downstream effects that account for microfibrillar proteolysis, elastin breakage leading to aortic dilatation, dissection, and other clinical changes observed in these conditions.^{8,9} Increased

https://doi.org/10.1016/j.cjco.2020.12.018

2589-790X/© 2021 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/ by-nc-nd/4.0/).

Received for publication October 16, 2020. Accepted December 20, 2020.

Ethics Statement: The research reported has adhered to the relevant ethical guidelines.

[§]Co-senior authors.

Corresponding author: Dr Kevin C. Harris, Children's Heart Center, 1F27 - 4480 Oak St, Vancouver, British Columbia V6H 3V4, Canada. Tel.: 604-875-3878.

Methods: This was a cohort study of 40 MFS and 4 LDS patients and 87 control participants. Aortic dimension and stiffness indexes, including pulse wave velocity (PWV), were measured using echocardiography and Doppler. Total and free TGF- β and MMP blood levels were measured using Quantikine (R&D Systems, Inc, Minneapolis, MN) and Quanterix (Billerica, MA) kits. Endothelial function was measured using brachial artery flow-mediated dilation.

Results: PWV was increased in patients with MFS. There were increased MMP-2 levels in those with MFS but no increase in free or total TGF- β or MMP-9 levels compared with control participants. There was no difference in TGF- β levels between MFS patients receiving no medications, angiotensin receptor blockers, and β -blockers. PWV correlated most strongly with age. Endothelial function showed premature gradual decline in patients with MFS.

Conclusions: Despite the increased PWV, monitoring aortic stiffness or TGF- β levels would not be helpful in patients with MFS. TGF- β levels were not increased and the increased MMP-2 levels suggest consideration of a different therapeutic target.

levels of circulating TGF- β have been found in patients with MFS and to a lesser extent in those receiving medical treatment.^{10,11} It has been proposed that circulating TGF- β might serve as a prognostic biomarker for aortic disease and aid in the decision for elective surgery. However, no association between aortic root size and circulating TGF- β was reported in a Japanese study of MFS patients, ¹² and aortic dissection has been reported in some MFS and LDS patients with a low-risk aortic root dimension.¹³ The primary role of TGF- β signalling in the causation of aortic aneurysms in these genetic syndromes has been challenged recently.^{14,15} Other common biomarkers have not been extensively studied in these conditions although abnormal matrix metalloproteinase (MMP) levels have been reported in a mouse model of MFS; changes in apolipoprotein levels have been detected in animal studies and in patients with abdominal aneurysms. However, the mechanistic significance of potential biomarkers has not been established.^{6,16,}

In addition to aortic dilatation, human and animal evidence shows increased aortic wall stiffness in those with MFS, and dysfunction of smooth muscle and endothelial cells.^{7,18-20} Echocardiography is used for monitoring of aortic root size, and can be used to assess pulse wave velocity (PWV), stiffness, and vascular impedance.^{21,22} Increased aortic stiffness has been shown in MFS patients even in the absence of aortic dilatation.¹⁹ Unfortunately, to our knowledge, there have been no studies to investigate whether serum biomarker levels correlate with the biophysical properties of the aorta in patients with these forms of genetic aortopathy or whether monitoring these might be clinically useful. Hence, the purpose of this study was to measure the aortic root dimensions, the biophysical properties of the aorta, and serum biomarkers and determine if there is any correlation in patients with MFS and LDS. taux de TGF- β observés chez des patients également atteints de SM, mais recevant un autre traitement.

Méthodologie : Il s'agissait d'une étude de cohorte menée auprès de 40 patients atteints du SM et de quatre patients atteints du SLD, ainsi que de 87 témoins. Les indices des dimensions et de la rigidité aortiques, y compris la vitesse d'onde de pouls (VOP), ont été mesurés par échocardiographie et par échographie Doppler. Les taux sanguins de TGF- β et de MMP totaux et libres ont été mesurés à l'aide de trousses Quantikine (R&D Systems, Inc, Minneapolis, MN) et Quanterix (Billerica, MA). La fonction endothéliale a été mesurée par dilatation liée au flux dans l'artère brachiale.

Résultats : La VOP était plus élevée chez les patients atteints du SM. On a aussi observé une hausse des taux de MMP-2 chez les patients atteints de SM, mais aucune augmentation des taux de TGF- β ou de MMP-9 libres ou totaux comparativement aux témoins. Il n'y avait pas de différence entre les taux de TGF- β chez les patients atteints de SM ne recevant aucun traitement, ceux qui prenaient un antagoniste des récepteurs de l'angiotensine et ceux qui prenaient un bêtabloquant. La VOP été plus fortement corrélée avec l'âge. La fonction endothéliale a affiché un déclin progressif prématuré chez les patients atteints du SM.

Conclusions : Malgré l'augmentation de la VOP, il ne semble pas utile de surveiller la rigidité aortique ni les taux de TGF- β en cas de SM. Les taux de TGF- β n'étaient pas plus élevés chez les patients atteints du SM, et la hausse des taux de MMP-2 indique qu'il conviendrait de choisir une autre cible thérapeutique.

Methods

Patients

The study protocol was approved by the University of British Columbia Clinical Research Ethics Board and the Children's and Women's Health Centre of British Columbia's Research Review Committee. Consecutive patients who met the criteria for MFS on the basis of the Ghent nosology²³ were recruited at British Columbia Children's Hospital and St Paul's Hospital, in Vancouver, Canada. All patients underwent physical examination and genetic screening before enrollment. The inclusion criteria were: subjects must (1) conform to the diagnostic criteria for MFS or LDS; (2) be between 8 and 60 years old; (3) had not undergone aortic root surgery; (4) have technically suitable echocardiographic windows to obtain the required images; and (5) provide informed consent and/or assent. The exclusion criteria were: subjects (1) with significant aortic or mitral valve regurgitation; (2) who were pregnant; and (3) who use illicit drugs. We did subgroup analyses on patients younger than 20 years and older than 20 years. Patients were matched 1:2 to healthy control participants from our institutional database and older subjects (older than 20 years) recruited prospectively who met the following criteria: subjects must (1) be between 8 and 60 years old; (2) be in good health with no acute or chronic illness; (3) have technically suitable echocardiographic windows to obtain the required images (imaging was attempted in all potential participants); and (4) provide informed consent. The control subject exclusion criteria were: subjects who (1) had undergone cardiothoracic surgery or whose aortic dimensions were abnormal; (2) were pregnant; and (3) use nicotine products or illicit drugs.

After consent was obtained, height, weight, arm span, blood pressure (BP), medication history, and genetic test data were recorded. All patients had full echocardiography. Flowmediated dilation (FMD), was performed in all but 7 patients with MFS. Nitroglycerin challenge was performed in adults who met the BP criteria of systolic BP of 100 mm Hg or more. Blood for biomarkers was obtained in all but 5 MFS patients, and adult control subjects who consented to blood analyses. The interpreting physicians and laboratories were blinded to the patient's identity and disease or control category.

Echocardiography

Standard 2-dimensional, M-mode, and Doppler echocardiography was performed on all patients. Left ventricular volumes, ejection fraction, aortic dimensions, and cardiac output were calculated. Our laboratory protocol for measuring the biophysical properties of the aorta was undertaken as described previously.¹⁹ The methods and calculations used to derive all of the indexes of vascular function are described in Supplemental Appendix S1.

Endothelial function

Brachial artery FMD was performed according to the standard protocol. The images were analyzed by the Cardiovascular Imaging Research Core Laboratory at the Vancouver Hospital and Health Sciences Centre.

Blood sample collection and processing

The blood sample collection, processing, and storage protocol has been previously described.²⁴ Freeze-thaw cycles were avoided to ensure the accuracy and reliability of the measurements.

Plasma biomarker measurement using enzyme-linked immunosorbent assay

For the quantitative determination of circulating levels of TGF- β 1, standard enzyme-linked immunosorbent assay (ELISA) testing was performed using a commercially available kit, Human TGF- β 1 Quantikine ELISA Kit DB100B (R&D Systems, Inc, Minneapolis, MN). MMP-2 and MMP-9 concentrations were measured following similar procedures for TGF- β 1, except for the sample activation, using Total MMP-2 Quantikine ELISA Kit MMP200 (R&D Systems, Inc) and Human MMP-9 Quantikine ELISA Kit DMP900 (R&D Systems, Inc), respectively. The plasma levels of TGF- β 1 and MMP-2/-9 of 44 patients were compared with that of 37 healthy control participants.

To comply with the suggestion that free TGF- β 1 should be measured,²⁵ free and total TGF- β 1 were measured using a Simoa HD-1 analyzer (Quanterix, Billerica, MA) and the Quanterix TGF- β assay.

The detailed description of the blood sampling and storing procedure, Elisa measurement using the Quantikine (R&D Systems, Inc) and Quanterix methods is given in Supplemental Appendix S1.

Statistical analysis

On the basis of previous work from our laboratory, a difference in PWV of 50 cm/s was considered clinically 587

	Young	ter than 20 years		Olc	ler than 20 years			All patients	
Characteristic	Control $(n = 51)$	Marfan (n = 27)	Ρ	Control $(n = 36)$	Marfan (n = 17)	Р	Control $(n = 87)$	Marfan (n $= 44$)	Р
Age, years	13.6 (9.6-17.7)	14.0 (9.7-17.4)	NS	41.3 (28.8-45.6)	39.8 (29.5-48.4)	NS	18.2 (12.5-35.3)	17.9 (12.9-35.5)	NS
Sex, male:female	28:23	14:13	NS	14:13	7:10	NS	42:45	21:23	NS
Height, cm	161.3 (135.9-167.5)	173 (151-185)	0.003	169 (164.1-177.2)	180.7 (171.2-188.5)	0.001	165.8 (153.5-171.1)	176 (164-186)	< 0.001
Weight, kg	51.3 (31.3-61.6)	57.3 (40-68.7)	NS	71.0 (64.9-77.6)	76.4 (62.1-85.7)	NS	63.1 (42.6-76.4)	63.1 (42.6-76.4)	NS
BSA, m ²	1.55 (1.10-1.69)	1.79 (1.28-1.89)	NS	1.83 (1.69-1.96)	1.95 (1.79-2.13)	NS	1.67(1.42-1.84)	1.83 (1.43-1.95)	0.04
BMI	20.0 (17.1-22.6)	17.4 (15.8-19.5)	0.005	24.7 (22.3-27.2)	22.1 (19.4-26.9)	NS	22.2 (19.5-25.5)	18.6 (16.6-22.4)	0.002
SBP, mm Hg	107 (100-115)	101 (97-110)	NS	113 (106-120)	115 (111-124)	NS	110 (100-116)	110 (100-115)	NS
DBP, mm Hg	60 (58-69)	63 (57-66)	NS	68 (63-74)	68 (61-74)	NS	65 (60-71)	64 (59-70)	NS
PP, mm Hg	42 (40-48)	42 (35-49)	NS	45 (38-49)	50 (48-55)	0.008	43 (39-48)	47 (36-50)	NS

BSA, body surface area; DBP, diastolic blood pressure; LDS, Loeys-Dietz syndrome; MFS, Marfan syndrome; NS, not significant; PP, pulse pressure; SBP, systolic blood pressure. mass index; (all gc) BMI, body

	Young	ger than 20 years		Older	than 20 years		All	l patients	
	Control $(n = 51)$	Marfan (n = 27)	P	Control $(n = 36)$	Marfan (n = 17)	Р	Control $(n = 87)$	Marfan (n = 44)	P
Ao annulus, cm	1.9 (1.7-2.0)	2.1 (1.8-2.3)	0.006	2.0 (1.9-2.2)	2.3 (2.1-2.3)	0.004	1.9 (1.8-2.1)	2.2 (1.9-2.3)	< 0.001
SoV, cm	2.5 (2.4-2.8)	3.5 (2.9-3.8)	< 0.001	2.9 (2.6-3.3)	3.6(3.4-4.2)	< 0.001	2.7 (2.4-3.0)	3.6(3.2-3.9)	< 0.001
SoV z-score	0.16 (-0.28 to 0.93)	3.0(2.3-4.3)	< 0.001	-0.52 (-1.24 to 0.38)	1.4(0.2-3.1)	< 0.001	-0.19 (-0.76 to 0.72)	2.7(1.4-4.0)	< 0.001
ST junction, cm	2.1 (2.0-2.3)	2.4 (2.3-2.8)	< 0.001	2.5 (2.3-2.8)	2.8 (2.5-3.1)	0.014	2.3 (2.1-2.5)	2.6(2.3-3.0)	< 0.001
SoV:annulus	1.33 (1.26-1.42)	1.70 (1.46-1.84)	< 0.001	1.45 (1.33-1.57)	1.70 (1.51-1.84)	< 0.001	1.36 (1.28-1.47)	1.70 (1.50-1.84)	< 0.001
Comparison of	the diameters of the aortic	(Ao) annulus, sinus of	Valsalva (SoV)	, SoV z-score, sinotubular (S	(T) junction, and ratio	of SoV:annulus	in Marfan syndrome patient	s vs age- and sex-matcl	ned control

participant on the basis of their ages of younger or older than 20 years.

Table 2. Measurements of the aorta

CJC Open Volume 3 2021

significant.¹⁹ Separate subgroup analyses were performed on younger (younger than 20 years) vs older groups (older than 20 years) as well as medication status (no-medication vs atenolol vs losartan). Free and total TGF- β measured using Quantikine (R&D Systems, Inc) were compared with TGF- β obtained using Quanterix assays. Frequency tables were generated for all categorical variables. Median (interquartile range) values are reported for all continuous variables. A Mann-Whitney U test was used to determine differences between 2 groups. A Kruskal-Wallis test with a post hoc pairwise Mann-Whitney U test was used to compare difference between more than 2 groups. A Spearman ρ was calculated to determine significant correlations between variables. A K statistic was used to calculate the level of agreement between the 2 methods used to calculate TGF- β 1. All tests were 2-sided and P values < 0.05 were considered statistically significant. All analyses were done using SPSS Version 25 (IBM Corp, Armonk, NY).

Results

Forty-four patients, 40 with MFS and 4 patients with LDS (21 male and 23 female) and 87 control participants (42 male and 45 female) were enrolled. Of these, 27 MFS and 51 control participants were younger than 20 years. Table 1 shows the complete demographic data. For ease of description, we have included the LDS patients in the category of MFS in the subsequent text and tables. As expected, the heights of the whole MFS group and MFS subgroups were higher than for control participants (P < 0.01). The weights and body surface areas were not different across the whole or MFS subgroups but the body mass indexes (BMIs) were higher for the control participants in the whole and younger age groups than the corresponding MFS groups (P = 0.005), and was similar between the older MFS and control subgroups. Thus, the older MFS subgroup showed relatively higher levels of obesity with age. The BP results show that only pulse pressure was higher in the older MFS subgroup compared with control participants (P = 0.008).

Aortic root measurements are shown in Table 2. As expected, the diameters of all of the aortic measures were greater in the whole MFS and MFS subgroups than in control participants (P < 0.01).

The results of the indexes of aortic stiffness are shown in Table 3. PWV was greater in the whole MFS cohort and younger MFS subgroup than in control participants but not in the older MFS subgroup compared with control participants (P < 0.01). There was no difference in the input or characteristic impedance between the MFS groups and control participants. The elastic strain modulus was greater in the MFS subgroups than in control participants (P < 0.05); the β -index was greater in the whole MFS cohort and younger subgroup than in control participants (P < 0.01), but the older MFS subgroup was not different from control participants. Thus, stiffness indexes converged in the older MFS patients compared with control participants.

Table 4 shows the results of plasma biomarkers measured using both assays. Plasma MMP-2 levels were significantly higher in patients with MFS compared with control participants (P = 0.004). However, there were no significant

4arfan (n = 27)	Р	Control $(n = 36)$	Marfan $(n = 17)$	Р	Control $(n = 87)$	Marfan (n = 44)	Р
439 (391-542)	< 0.001	454 (423-491)	500 (397-608)	NS	409 (344-455)	450 (393-576)	0.003
194 (158-252)	NS	187 (142-208)	178 (155-220)	NS	192 (162-221)	184 (158-231)	NS
136 (113-195)	NS	152 (130-176)	130(114-164)	NS	144 (122-175)	136 (113-169)	NS
338 (262-488)	0.003	455 (325-529)	560 (418-899)	0.04	314 (235-444)	429 (302-650)	0.002
60 (2.36-2.98)	< 0.001	2.82 (2.57-2.97)	2.89 (2.68-3.36)	NS	2.45 (2.22-2.81)	2.77 (2.47-3.33)	0.001
エーキ じ じ ぶ ぶ	arfan $(n = 27)$ 39 (391-542) 34 (158-252) 36 (113-195) 38 (262-488) 50 (2.36-2.98)	arfan (n = 27) P 39 (391-542) < 0.001 94 (158-252)NS 36 (113-195)NS 38 (262-488) 0.003 50 (2.36-2.98) < 0.001	arfan (n = 27)PControl (n = 36) 39 (391-542)< 0.001	arfan (n = 27)PControl (n = 36)Marfan (n = 17) 39 (391-542)< 0.001	arfan (n = 27)PControl (n = 36)Marfan (n = 17)P 39 (391-542)< 0.001	arfan (n = 27)PControl (n = 36)Marfan (n = 17)PControl (n = 87) 39 (391-542)< 0.001	arfan (n = 27)PControl (n = 36)Marfan (n = 17)PControl (n = 87)Marfan (n = 44)39 (391-542)< 0.001

Table 3. Measures of aortic stiffness

differences in plasma MMP-9 levels, or free or total TGF- β 1 levels in MFS vs control participants. There was a high level of agreement between the 2 methods used to measure total TGF- β 1 (P < 0.001; Table 5). The subgroup analysis was predicated on the results of the comparison of the total MFS group with the control participants and, because they were so similar, we believe it was not justified.

The comparison of the demographic characteristics, aortic diameters, and vascular function of the no medication subgroup or subgroups treated with either atenolol or losartan is shown in Supplemental Table S1. The no medication subgroup was older, taller, heavier, had greater body surface area and BMI than those treated with either of the medications (P < 0.05). The annulus, sinus of Valsalva (SoV), and sinotubular junction were not different between the groups but the SoV z-score was lower in the no medication subgroup (P = 0.02). There were no differences in PWV, FMD, or free or total TGF- β levels between the no medication, β -blocker and angiotensin receptor blocker (ARB) subgroups. Median FMD for younger MFS patients was 10.1 vs 6.4 for older MFS patients (P < 0.001). There were no significant differences in input impedance, characteristic impedance, strain modulus, β -index, systolic BP, diastolic BP, pulse pressure, or heart rate for the no medication vs β -blocker vs ARB groups, which are not shown.

The results of correlation analyses are shown in Tables 6 and 7. PWV strongly correlated with age in MFS and control participants and age-related measures of aortic diameter in control participants but less so in MFS patients. In MFS patients, FMD also correlated with age. Figure 1 shows the correlation between PWV and age in control participants and MFS patients across all age groups. Figure 2 shows subanalysis of the effect of age on FMD in MFS patients across all age groups.

Discussion

Our study showed a difference in PWV between the younger MFS subgroup and control participants; age-related arterial stiffening occurs in both groups, abolishing the difference between them. The finding of decreased endothelial function is consistent with previous human and mouse model studies of MFS.^{7,20} In this study we did not find that total or free TGF- β 1 was increased in our MFS patients. The biomarker results showing increased MMP-2 levels but not TGF- β 1 or MMP-9 are novel findings.

As expected, the heights, sinus of Valsalva and sinotubular junction dimensions were significantly larger in the MFS patient group and subgroups than in control participants. Of interest was the finding that although the BMI in MFS patients and the younger MFS subgroup was less than in control participants, the BMI in older MFS subgroup was not, suggesting a relative increase in weight.

Different methods of measuring arterial stiffening have been used in a number of studies of MFS.^{19,26-28} Because the elastic aorta acts as a conduit and a cushion for pulsatile arterial flow, stiffness of the aorta is manifested by increased PWV. It is important to differentiate between studies that measured PWV directly and those that measured aortic dimensional change to derive stiffness or its reciprocal, distensibility. The latter are suboptimal because they are on

 Table 4. Plasma biomarkers measurement

Biomarker	Control $(n = 37)$	Marfan (n $=$ 39)	Р
MMP-2	171.5 (154.3-193.9)	195.7 (170.1-228.8)	0.006
MMP-9	44.3 (27.9-96.4)	58.9 (42.2-83.8)	NS
TGF-β 1,2,3, pg/mL	882.2 (711.6-1166.9)	930.4 (811.3-1285.1)	NS
TGF-β (Quanterix*)	1191.2 (979.8-1455.4)	1274.5 (1060.6-1429.21)	NS
TGF-β Free	1.29 (0.47-2.17)	0.88 (0.50-1.93)	NS

Comparison of the plasma levels of MMP-2, MMP-9, and free TGF- β 1 in Marfan syndrome patients vs age- and sex-matched control participants. MMP, matrix metalloproteinase; TGF, transforming growth factor.

* Measurements obtained using a Quanterix $\mbox{\ensuremath{\mathbb{R}}}$ (Billerica, MA) Simoa HD-1 analyzer and TGF- $\mbox{\ensuremath{\beta}}$ kit.

the basis of the stress/strain relationship. A larger aorta, as in MFS, will distend less to absorb a similar stroke volume than a normal-sized aorta and, on the basis of the formula, strain will be less and stiffness will always compute to be increased. The same criticism can be made of methods that derive PWV from aortic dimensions. Increased PWV has been shown to be the earliest marker of cardiovascular risk in adults.²⁹

A limited number of studies have measured PWV directly, using magnetic resonance imaging, carotid-femoral or echocardiographic methods, and all report increased PWV in patients with MFS, implying increased stiffness.^{19,26-28} This study, which has a wide age range of subjects, confirms these findings, but only in the younger patients. Despite the fact that the aortas in the older MFS subgroup were less dilated, it was interesting to find that as the MFS and control groups aged, their PWV increased and their 2 slopes converged. Agerelated increases in PWV in the normal population are well reported but it is not clear why there was convergence of the slopes between MFS patients and control participants in this study. Because most patients who require surgery for aortic root dilation are in the 20 plus age group, this finding suggests that monitoring of arterial stiffness does not offer any clinical benefit in the follow-up of these patients. However, to conclusively assess this, a larger study using this technique with a number of surgical and nonsurgical patients would be needed.

To our knowledge, our study is the first to show that there was early onset and progressive decline in FMD with age in MFS patients. In the normal population, FMD has been shown to remain constant until the fourth decade in men, and the fifth decade in women after which a steady decline develops.³⁰ These observations are consistent with the results of human studies²⁰ and in the MFS mouse model,⁷ which showed endothelial dysfunction.

In contrast with previous studies,^{10,11} we did not find increased levels of TGF- β 1, or decreased levels of MMP-9³¹ among our MFS cohort. Rather we found increased levels of MMP-2. Because of the role of maladaptive TGF- β signalling in MFS and LDS, circulating levels of this centrally important cytokine have been proposed as a prognostic and therapeutic marker.^{10,11} The ELISA method and the TGF- β 1 assay we used was the same Quantikine immunoassay as those studies.³²⁻³⁴ To confirm this result, and in keeping with the suggestion that free TGF- β 1 may be a more relevant marker,³⁵ we analyzed free and total TGF- β 1 levels using the Quanterix immunoassay. Free TGF- β 1 was not increased and total TGF- β 1 levels were very similar using both assays. Therefore, it is unlikely that the discrepancy between our results and those reporting increased TGF- β 1 levels is rooted in the analysis itself. Instead, our results are consistent with other studies that reported no increased levels of circulating TGF- β 1 in patients with MFS. A Japanese study also reported no significant difference in the mean plasma TGF-B1 level between the MFS group and control participants.¹² Those investigators suggested that circulating TGF-B1 is not a diagnostic and therapeutic marker for Japanese MFS patients, although their findings did not exclude the association of TGF- β with the pathogenesis of MFS. In contrast, another Japanese study showed that circulating TGF-B1 levels are approximately fivefold higher in patients with acute aortic dissection compared with control participants,³⁶ with type B dissections showing an approximately twofold elevation in circulating TGF- β 1 levels compared with type A dissections. This suggested that circulating TGF- β 1 levels might be a biomarker in acute aortic dissection, but not for disease status, dilation, or risk prediction in patients with MFS. A recent study reported that a number of MFS patients who had emergency surgery for dissection had smaller aortic root sizes than that quoted in the guidelines for surgery, underscoring the need for better predictors of dissection.

Our findings suggest that dysregulated TGF- β signalling is not reflected in circulating free or total TGF- β concentrations. TGF- β signalling is complex and modulated by a broad number of auxiliary cascades in a tissue-, cell-, and contextspecific manner. Because of the diversity of the biological processes it affects and regulates, we speculate that circulating TGF- β concentrations are generally maintained within a relatively narrow range. By extension, the perturbations in TGF- β signalling that characterize MFS and LDS might result

Table 5. Comparison of TGF- β measured using different methods

	TGF-β 1,2,3, pg/mL	TGF-β (Quanterix*)	TGF-β free
Control TGF-β 1,2,3, pg/mL TGF-β (Quanterix*) TGF-β free	0.967^{\dagger} 0.822^{\dagger}	0.887^{\dagger} - 0.887^{\dagger}	0.822^{\dagger} 0.887^{\dagger}
Marfan TGF-β 1,2,3, pg/mL TGF-β (Quanterix*) TGF-β free	$0.970^{\dagger} \\ 0.875^{\dagger}$	0.970^{\dagger} 0.958^{\dagger}	$0.875^{\dagger} \\ 0.958^{\dagger} \\ -$

TGF, transforming growth factor.

* Measurements obtained using a Quanterix® (Billerica, MA) Simoa HD-1 analyzer and TGF- β kit.

 $^{\dagger}P < 0.001.$

Table 6.	Correlation	analysis	of	aortic	stiffness
----------	-------------	----------	----	--------	-----------

		Younger than	20 years	years Older than 20 years			All pa	tients				
		Control	Ma	arfan	Con	trol	Ν	Iarfan	C	ontrol	Ma	rfan
	ρ	Р	ρ	P	ρ	Р	ρ	Р	ρ	Р	ρ	Р
PWV vs age	0.26	NS (0.068)	0.47	0.013	0.49	0.002	0.44	NS (0.07)	0.59	< 0.001	0.41	0.006
PWV vs Ao Ann	0.23	NS	0.25	NS	0.27	NS	0.23	NS	0.36	0.001	0.28	NS
PWV vs SoV	0.49	0.001	0.37	0.04	0.27	NS	-0.17	NS	0.55	< 0.001	0.20	NS
PWV vs SoV z-score	0.33	0.033	0.14	NS	-0.03	NS	-0.40	NS	0.06	NS	-0.28	NS
PWV vs SoV:Ann	0.31	0.04	0.22	NS	0.03	NS	-0.41	NS	0.34	0.002	-0.02	NS
PWV vs sinotubular junction	0.39	0.008	0.35	NS	0.46	0.005	0.01	NS	0.64	< 0.001	0.30	0.049

Correlation among pulse wave velocity (PWV) and biophysical properties of the aorta in Marfan syndrome patients vs age- and sex-matched control participants. Groups separated according to age younger or older than 20 years (Spearman ρ calculated).

Ann, annulus; Ao, aortic; NS, not significant; SoV, sinus of Valsalva; sinotubular junction, sinotubular junction.

from compensatory and/or parallel signalling cascades within the context of the vascular microenvironment. Indeed, our findings might support the notion that there is an interplay of complex processes that yield the vascular phenotype of MFS.

Our finding of endothelial dysfunction suggests accelerated ageing in those with MFS. In an MFS mouse model, mesenteric arterial endothelium-dependent relaxation stimulated by acetylcholine was significantly decreased, suggesting an impairment of nitric oxide release.³⁸ Endothelial vasoactive mediators regulate smooth muscle contractility and vascular smooth muscle tone.³⁹⁻⁴² Decreased endothelial function in those with MFS is also associated with increased plasma levels of homocysteine,^{43,44} and can be compromised through increased arterial wall stiffness.⁴⁵ Nitric oxide bioavailability is decreased due to aging.

To our knowledge, the current study is the first to examine plasma levels of MMP-2 and MMP-9 in patients with MFS and those with LDS. Our findings of increased MMP-2 is consistent with increased expression of MMPs, which has been observed in human abdominal aneurysm tissue.^{46,4} MMPs constitute a tightly regulated family of zincdependent endopeptidases that proteolytically cleave most components of the base membrane and the extracellular matrix. They play an important role in the regulation of a variety of physiological processes, including angiogenesis and vascular remodelling, and are also involved in vascular diseases such as hypertension, atherosclerosis, and aortic aneurysm. Increased MMP activity is accompanied by a widespread deterioration of microfibrils, and elastic and collagenous fibres, which eventually leads to the loss of extracellular matrix integrity, endothelial dysfunction, and reduction of smooth muscle contractility.⁴⁸ Our previous studies showed that progression of aortic aneurysm in a mouse model of MFS is associated with upregulation of MMP-2 and MMP-9.6,7 Interestingly, in a recent study it was reported that serum MMP-2 levels correlated positively, whereas MMP-9 levels did not correlate and showed greater variation in patients with aortic root/ ascending aortic aneurysms.⁴⁹ In the mouse model of MFS, we have previously shown that doxycycline, an antibiotic and a general inhibitor of MMPs, could correct aortic elastin fibre structure and organization and was more effective than atenolol in preventing thoracic aortic aneurysm.^{50,51} Another study in our laboratory reported that drug therapy with losartan and doxycycline completely suppressed aneurysm formation in MFS mice, improved elastic fibre organization, downregulated MMP-2/-9 and TGF- β 1 expression, and normalized aortic contractile and relaxation function to control values.⁵² Although the implication of the findings in this study is to treat these patients with MMP inhibitors, the unsuccessful results of clinical trials of ARBs have shown that there needs to be caution in extrapolating animal model results to humans.^{53,54}

The comparison of the results of patients not receiving medications with those receiving either a β -blocker or ARB is complicated by the fact that the no medication subgroup was older and had relatively smaller SoV z-scores, an approach consistent with recent treatment guidelines.⁵⁵ Of note, there was no difference in the PWV or FMD between the 3 groups. Of interest also is that there was no difference in free or total TGF- β levels between the 3 groups.

Limitations

The results of this study might have been affected by the total number of patients studied, which is relatively small compared with major therapeutic trials, especially in the number of older MFS patients. We recognize that our study might not have been sufficiently powered to detect differences in TGF- β 1. Because PWV increases with age in the normal population, it would have been preferable to have more older patients and control participants to further parse out the relationship between PWV and age. A larger number of patients might have also shed more information about the loss of endothelial function and the role of MMP-2 in determining the aortic size and vascular function.

Table 7	7.	Correlation	analysis	of	FMD
---------	----	-------------	----------	----	-----

FMD vs	Marfan (n = 37)	Р
Age	-0.50	0.002
Height	-0.21	NS
Weight	-0.42	0.011
BSA	-0.36	0.03
BMI	-0.43	0.008
PWV	-0.29	NS (0.08)
SoV z-score	0.333	0.046

Correlation among flow-mediated dilation (FMD) and demographic parameters, pulse wave velocity (PWV), and sinus of Valsalva (SoV) z-score in Marfan syndrome patients.

BMI, body mass index; BSA, body surface area.



Figure 1. Correlation of pulse wave velocity (PWV) vs age in control participants and Marfan patients across all age groups.

Conclusions

In MFS patients, PWV was increased and was associated with age, independently of aortic root size. We showed premature age-related decline in FMD in MFS patients. Our study confirms the elevation of serum MMP-2 in MFS patients but not MMP-9, and total or free TGF- β 1. The clinical implications are that: (1) routine evaluation of aortic stiffness or serum TGF- β 1 do not help in monitoring MFS; and (2) therapies that inhibit MMP-2 with or without treatment to improve endothelial function warrant evaluation in patients with MFS.



Figure 2. Correlation of flow-mediated dilation (FMD) vs age in Marfan syndrome patients across all age groups.

Funding Sources

This work was funded by the Canadian Institute for Health Research, Ottawa, Ontario, Canada (grant MOP111266).

Disclosures

The authors have no conflicts of interest to disclose.

References

- 1. Dietz HC, Cutting GR, Pyeritz RE, et al. Marfan syndrome caused by a recurrent de novo missense mutation in the fibrillin gene. Nature 1991;352:337-9.
- Loeys BL, Chen J, Neptune ER, et al. A syndrome of altered cardiovascular, craniofacial, neurocognitive and skeletal development caused by mutations in TGFBR1 or TGFBR2. Nat Genet 2005;37:275-81.
- Loeys BL, Schwarze U, Holm T, et al. Aneurysm syndromes caused by mutations in the TGF-beta receptor. N Engl J Med 2006;355:788-98.
- Judge DP, Dietz HC. Therapy of Marfan syndrome. Annu Rev Med 2008;59:43-59.
- Gallo EM, Loch DC, Habashi JP, et al. Angiotensin II-dependent TGFbeta signaling contributes to Loeys-Dietz syndrome vascular pathogenesis. J Clin Invest 2014;124:448-60.
- 6. Chung AW, Au Yeung K, Sandor GG, et al. Loss of elastic fiber integrity and reduction of vascular smooth muscle contraction resulting from the upregulated activities of matrix metalloproteinase-2 and -9 in the thoracic aortic aneurysm in Marfan syndrome. Circ Res 2007;101:512-22.
- Chung AW, Au Yeung K, Cortes SF, et al. Endothelial dysfunction and compromised eNOS/Akt signaling in the thoracic aorta during the progression of Marfan syndrome. Br J Pharmacol 2007;150:1075-83.
- Habashi JP, Judge DP, Holm TM, et al. Losartan, an AT1 antagonist, prevents aortic aneurysm in a mouse model of Marfan syndrome. Science 2006;312:117-21.
- Lavoie P, Robitaille G, Agharazii M, et al. Neutralization of transforming growth factor-beta attenuates hypertension and prevents renal injury in uremic rats. J Hypertens 2005;23:1895-903.
- Franken R, den Hartog AW, de Waard V, et al. Circulating transforming growth factor-beta as a prognostic biomarker in Marfan syndrome. Int J Cardiol 2013;168:2441-6.
- 11. Matt P, Schoenhoff F, Habashi J, et al. Circulating transforming growth factor-beta in Marfan syndrome. Circulation 2009;120:526-32.
- Ogawa N, Imai Y, Nishimura H, et al. Circulating transforming growth factor beta-1 level in Japanese patients with Marfan syndrome. Int Heart J 2013;54:23-6.
- Legget ME, Unger TA, O'Sullivan CK, et al. Aortic root complications in Marfan's syndrome: identification of a lower risk group. Heart 1996;75: 389-95.
- Mallat Z, Ait-Oufella H, Tedgui A. The pathogenic transforming growth factor-beta overdrive hypothesis in aortic aneurysms and dissections: a mirage? Circ Res 2017;120:1718-20.
- Wei H, Hu JH, Angelov SN, et al. Aortopathy in a mouse model of Marfan syndrome is not mediated by altered transforming growth factor beta signaling. J Am Heart Assoc 2017;6:e004968.
- Ahnstrom J, Gottsater A, Lindblad B, Dahlback B. Plasma concentrations of apolipoproteins A-I, B and M in patients with abdominal aortic aneurysms. Clin Biochem 2010;43:407-10.

- Wang YX, Martin-McNulty B, Freay AD, et al. Angiotensin II increases urokinase-type plasminogen activator expression and induces aneurysm in the abdominal aorta of apolipoprotein E-deficient mice. Am J Pathol 2001;159:1455-64.
- Lee L, Cui JZ, Cua M, et al. Aortic and cardiac structure and function using high-resolution echocardiography and optical coherence tomography in a mouse model of Marfan syndrome. PLoS One 2016;11: e0164778.
- Bradley TJ, Potts JE, Potts MT, DeSouza AM, Sandor GG. Echocardiographic Doppler assessment of the biophysical properties of the aorta in pediatric patients with the Marfan syndrome. Am J Cardiol 2005;96: 1317-21.
- Wilson DG, Bellamy MF, Ramsey MW, et al. Endothelial function in Marfan syndrome: selective impairment of flow-mediated vasodilation. Circulation 1999;99:909-15.
- O'Rourke MF, Staessen JA, Vlachopoulos C, Duprez D, Plante GE. Clinical applications of arterial stiffness; definitions and reference values. Am J Hypertens 2002;15:426-44.
- Oliver JJ, Webb DJ. Noninvasive assessment of arterial stiffness and risk of atherosclerotic events. Arterioscler Thromb Vasc Biol 2003;23:554-66.
- Loeys BL, Dietz HC, Braverman AC, et al. The revised Ghent nosology for the Marfan syndrome. J Med Genet 2010;47:476-85.
- Quon BS, Dai DL, Hollander Z, et al. Discovery of novel plasma protein biomarkers to predict imminent cystic fibrosis pulmonary exacerbations using multiple reaction monitoring mass spectrometry. Thorax 2016;71: 216-22.
- Grainger DJ, Mosedale DE, Metcalfe JC. TGF-beta in blood: a complex problem. Cytokine Growth Factor Rev 2000;11:133-45.
- 26. Kiotsekoglou A, Moggridge JC, Saha SK, et al. Assessment of aortic stiffness in Marfan syndrome using two-dimensional and Doppler echocardiography. Echocardiography 2011;28:29-37.
- Harada K, Yasuoka K, Shimada Y. Usefulness of tissue doppler imaging for assessing aortic wall stiffness in children with the Marfan syndrome. Am J Cardiol 2004;93:1072-5.
- Nollen GJ, Groenink M, Tijssen JG, Van Der Wall EE, Mulder BJ. Aortic stiffness and diameter predict progressive aortic dilatation in patients with Marfan syndrome. Eur Heart J 2004;25:1146-52.
- Vlachopoulos C, Aznaouridis K, Stefanadis C. Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. J Am Coll Cardiol 2010;55:1318-27.
- Celermajer DS, Sorensen KE, Spiegelhalter DJ, et al. Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. J Am Coll Cardiol 1994;24:471-6.
- Ikonomidis JS, Jones JA, Barbour JR, et al. Expression of matrix metalloproteinases and endogenous inhibitors within ascending aortic aneurysms of patients with Marfan syndrome. Circulation 2006;114: 1365-70.
- 32. Coupes BM, Williams S, Roberts IS, Short CD, Brenchley PE. Plasma transforming growth factor beta(1) and platelet activation: implications for studies in transplant recipients. Nephrol Dial Transplant 2001;16: 361-7.
- Lee MJ. Heparin inhibits activation of latent transforming growth factorbeta1. Pharmacology 2013;92:238-44.
- 34. Kim TH, Lee EK, Lee MJ, Kim JH, Yang WS. Fucoidan inhibits activation and receptor binding of transforming growth factor-beta1. Biochem Biophys Res Commun 2013;432:163-8.

- Khan SA, Joyce J, Tsuda T. Quantification of active and total transforming growth factor-beta levels in serum and solid organ tissues by bioassay. BMC Res Notes 2012;5:636.
- Suzuki T, Trimarchi S, Sawaki D, et al. Circulating transforming growth factor-beta levels in acute aortic dissection. J Am Coll Cardiol 2011;58:775.
- Desai MY, Kalahasti V, Hutt Centeno E, et al. Adult patients with Marfan syndrome and ascending aortic surgery. J Am Coll Cardiol 2019;73:733-4.
- Syyong HT, Chung AW, Yang HH, van Breemen C. Dysfunction of endothelial and smooth muscle cells in small arteries of a mouse model of Marfan syndrome. Br J Pharmacol 2009;158:1597-608.
- **39.** Ramsey MW, Goodfellow J, Jones CJ, et al. Endothelial control of arterial distensibility is impaired in chronic heart failure. Circulation 1995;92:3212-9.
- Boutouyrie P, Bezie Y, Lacolley P, et al. In vivo/in vitro comparison of rat abdominal aorta wall viscosity. Influence of endothelial function. Arterioscler Thromb Vasc Biol 1997;17:1346-55.
- 41. Wilkinson IB, Qasem A, McEniery CM, et al. Nitric oxide regulates local arterial distensibility in vivo. Circulation 2002;105:213-7.
- Vanhoutte PM. Endothelium-dependent hyperpolarizations: the history. Pharmacol Res 2004;49:503-8.
- 43. Giusti B, Porciani MC, Brunelli T, et al. Phenotypic variability of cardiovascular manifestations in Marfan syndrome. Possible role of hyperhomocysteinemia and C677T MTHFR gene polymorphism. Eur Heart J 2003;24:2038-45.
- 44. Jiang X, Yang F, Tan H, et al. Hyperhomocystinemia impairs endothelial function and eNOS activity via PKC activation. Arterioscler Thromb Vasc Biol 2005;25:2515-21.
- Peng X, Haldar S, Deshpande S, Irani K, Kass DA. Wall stiffness suppresses Akt/eNOS and cytoprotection in pulse-perfused endothelium. Hypertension 2003;41:378-81.
- 46. Davis V, Persidskaia R, Baca-Regen L, et al. Matrix metalloproteinase-2 production and its binding to the matrix are increased in abdominal aortic aneurysms. Arterioscler Thromb Vasc Biol 1998;18:1625-33.

- Freestone T, Turner RJ, Coady A, et al. Inflammation and matrix metalloproteinases in the enlarging abdominal aortic aneurysm. Arterioscler Thromb Vasc Biol 1995;15:1145-51.
- 48. Chung AW, Yang HH, Yeung KA, van Breemen C. Mechanical and pharmacological approaches to investigate the pathogenesis of Marfan syndrome in the abdominal aorta. J Vasc Res 2008;45: 314-22.
- 49. Meffert P, Tscheuschler A, Beyersdorf F, et al. Characterization of serum matrix metalloproteinase 2/9 levels in patients with ascending aortic aneurysms. Interact Cardiovasc Thorac Surg 2017;24:20-6.
- 50. Chung AW, Yang HH, Radomski MW, van Breemen C. Long-term doxycycline is more effective than atenolol to prevent thoracic aortic aneurysm in Marfan syndrome through the inhibition of matrix metalloproteinase-2 and -9. Circ Res 2008;102:e73-85.
- Cui JZ, Lee L, Sheng X, et al. In vivo characterization of doxycyclinemediated protection of aortic function and structure in a mouse model of Marfan syndrome-associated aortic aneurysm. Sci Rep 2019;9: 2071.
- 52. Yang HH, Kim JM, Chum E, van Breemen C, Chung AW. Effectiveness of combination of losartan potassium and doxycycline versus single-drug treatments in the secondary prevention of thoracic aortic aneurysm in Marfan syndrome. J Thorac Cardiovasc Surg 2010;140. 305-12.e2.
- Forteza A, Evangelista A, Sanchez V, et al. Efficacy of losartan vs. atenolol for the prevention of aortic dilation in Marfan syndrome: a randomized clinical trial. Eur Heart J 2016;37:978-85.
- Lacro RV, Dietz HC, Sleeper LA, et al. Atenolol versus losartan in children and young adults with Marfan's syndrome. N Engl J Med 2014;371:2061-71.
- Singh MN, Lacro RV. Recent clinical drug trials evidence in Marfan syndrome and clinical implications. Can J Cardiol 2016;32: 66-77.

Supplementary Material

To access the supplementary material accompanying this article, visit *CJC Open* at https://www.cjcopen.ca/ and at https://doi.org/10.1016/j.cjco.2020.12.018.