

Identification of ceRNA network based on a RNA-seq shows prognostic lncRNA biomarkers in human lung adenocarcinoma

XING LI^{1,2}, BING LI¹, PIXIN RAN³ and LANYING WANG²

¹GMU-GIBH Joint School of Life Sciences, Guangzhou Medical University, Guangzhou, Guangdong 511400;

²Oncology Department, Gansu Provincial Hospital of Traditional Chinese Medicine, Lanzhou, Gansu 730050;

³State Key Laboratory of Respiratory Disease, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong 510030, P.R. China

Received October 21, 2017; Accepted July 5, 2018

DOI: 10.3892/ol.2018.9336

Abstract. Previous studies have emphasized the significant functions of long non-coding RNAs (lncRNAs) as competing endogenous RNAs (ceRNAs) in tumor biology. However, the functions of certain cancer lncRNAs in the lncRNA-related ceRNA network in lung adenocarcinoma (LUAD) are unknown. A systematic and integrative survey of RNA-seq data from The Cancer Genome Atlas (TCGA) was performed to identify candidate lncRNAs for the prognosis of LUAD. In total, 20,502 genes that contain 181 lncRNAs were evaluated in a cohort of 570 LUAD cases. Initially, 6,280 differentially expressed genes (fold-change >2, P<0.05) were obtained using R package, which includes 75 lncRNAs. Next, by univariate regression and multivariate Cox proportional hazards analysis, 32 genes were associated with survival in LUAD. Using these 29 mRNAs and 3 lncRNAs, a prognosis index (PI) was calculated to accurately estimate the survival in LUAD: $PI = \sum \exp_{\text{risk gene}} \times HR_{\text{risk gene}}$. Furthermore, the 32-gene signature was an independent prognostic indicator for LUAD (HR >1; P<0.05, by multivariate analysis). Weighted gene co-expression network analysis (WGCNA) of three risk lncRNAs-FAM138B, NHEG1 and TLX1NB-was performed, based on the P-values of the associated genes, and the top 27 miRNAs that bound to these lncRNAs were predicted by Miranda as target miRNAs.

Next, these target miRNAs were transferred to the TarBase, miRTarBase, miRecords and starBase v2.0 databases to obtain their target genes. According to the previous miRNA-mRNA and miRNA-lncRNA data, three lncRNA-miRNA-mRNA ceRNA networks were established, based on the 29 prognostic mRNAs, forming a regulatory network in LUAD. The present study provided insight into the lncRNA-related ceRNA network in LUAD and has identified potential diagnostic and prognostic biomarkers.

Introduction

Lung adenocarcinoma (LUAD) is the most well-known histological sub-type of non-small cell lung cancer (NSCLC), which is the most significant cause of cancer-associated mortality worldwide (1). Since LUAD tends to form metastases at an early stage, the prognosis for patients with LUAD is generally poor, with average 5-year survival rates of <20% (2). Although recent advances in molecular pathology and targeted therapies have improved clinical therapies, the 5-year overall survival of patients with LUAD remains low (3), requiring continued research to identify novel biomarkers and effective targeted molecular therapies.

Recent high-throughput transcriptome analyses have demonstrated that >90% of the transcriptome is transcribed into non-coding RNAs, among which microRNAs (miRNAs/miRs) and long non-coding RNAs (lncRNAs) have been revealed to be involved in the malignant behaviors of LUAD. By definition, lncRNAs are non-coding RNAs that are >200 base pairs (4). According to recent studies, lncRNAs are important in various biological processes, including gene expression, cell proliferation and differentiation, the immune response, apoptosis and human diseases, including various types of cancer (5-8).

It has been reported that lncRNAs serve as competing endogenous RNAs (ceRNAs) or miRNA sponges, inhibiting miRNA response elements (MREs) from exchanging with mRNAs by competing for common miRNAs during the tumorigenic process (9). The ceRNA hypothesis has been proposed as a novel regulatory mechanism between non-coding RNAs and coding RNAs (10), thereby contributing toward the occurrence and progression of cancer (11,12). Based on ceRNA theory,

Correspondence to: Dr Lanying Wang, Oncology Department, Gansu Provincial Hospital of Traditional Chinese Medicine, 418 Guazhou Road, Lanzhou, Gansu 730050, P.R. China
Email: wanglanying_1@163.com

Abbreviations: LUAD, lung adenocarcinoma; KEGG, Kyoto Encyclopedia of Genes and Genomes; lncRNA, long non-coding RNA; NSCLC, non-small cell lung cancer; GO, gene ontology; ROC, receiver operating characteristic; TCGA, The Cancer Genome Atlas; AUC, area under the curve; ceRNA, competing endogenous RNAs; WGCNA, weighted gene co-expression network analysis; MREs, miRNA response elements; CI, confidence intervals

Key words: long non-coding RNAs, lung adenocarcinoma, biomarker, prognosis index, overall survival

Akcakaya *et al* (13) demonstrated that miR-133b was significantly decreased in CRC and associated with overall survival and metastasis. Zhang *et al* (14) analyzed 372 lncRNAs from The Cancer Genome Atlas (TCGA) and NCBI GEO omnibus (GSE65485), and identified a complex cancer-specific ceRNA network in HCC. Zhang *et al* (15) revealed MALAT1 to be the direct target of miR-202 and reported that interfering with MALAT1 downregulates Gli2 through positive regulation of miR-202 to suppress the development of gastric cancer.

As a novel post-transcriptional regulatory system, ceRNAs provide a novel means of investigating the pathogenesis of cancer and detecting novel signatures with high accuracy in diagnosis. The development and survival of anomalous lncRNAs in different types of cancer (including LUAD) have been examined, revealing the potential of lncRNAs as prognostic cancer biomarkers (16–20). Nevertheless, the study of ceRNAs in LUAD is scarce, and there are few targets for tumor therapies. There are little data on large-scale samples, and microarray detection remains rare, and the association between cancer-specific lncRNAs and clinical features is unknown.

The TCGA database is a public platform that can be used to download the sequencing data of LUAD lncRNAs, miRNAs and mRNAs. Following obtaining RNA sequence data, lncRNAs, miRNAs and mRNAs with abnormal expression were identified, and an lncRNA/mRNA expression network was constructed. These results increase understanding of the molecular mechanisms and therapeutic targets of lncRNAs in the treatment of LUAD.

Materials and methods

TCGA dataset and analysis of the differentially expressed genes. LUAD individual RNA sequencing (RNA-Seq) data (level 3), computed on the Illumina HiSeq RNA-Seq platform, were obtained from the TCGA data portal (<https://tcga-data.nci.nih.gov/tcga/>), which contains 512 LUAD and 58 adjacent non-tumor lung tissue samples, until December 31, 2016. The raw RNA-Seq reads (lncRNA and mRNA) were post-processed and normalized on the TCGA RNASeqV2 system. The miRNA expression data (level 3) were collected by Illumina GA microRNA sequencing platforms (Illumina, Inc., Hayward, CA, USA), quantile-normalized prior to the analysis and expressed as reads per million (RPM).

The exclusion criteria were as follows: i) The histological diagnosis was not LUAD; ii) presence of a malignancy other than LUAD; and iii) patient samples lacking complete data for analysis.

A total of 493 patients with LUAD were included in the present study. lncRNA expression profiles for normal lung tissue samples were obtained from adjacent non-tumor lung tissues (n=58). In addition, there were 170 LUAD patients with lymph node metastases and 323 LUAD patients without lymph node metastases. According to the Union for International Cancer Control (UICC), there were 381 patients with stage I-II LUAD and 112 patients with stage III-IV LUAD (21).

The expression data of lncRNAs are displayed as RPM and the expression levels of each lncRNA were normalized by Deseq R package for further analysis. The data were obtained from TCGA, a community resource project that provides

data for community research. Following low-expression genes being filtered, the LUAD RNA-Seq coverage data contained 20,321 mRNA and 181 lncRNAs compared with the GENCODE database (<http://www.genencodegenes.org/>). Next, the differentially expressed lncRNAs were counted using EdgeR and the DEseq package of R (Padj <0.05 and absolute log₂FC>1). The final lncRNAs were selected using a combination of the two methods. Student's t-test (SPSS 20.0, Inc., IBM Corp., Armonk, NY, USA) was employed to analyze the difference in expression levels of these lncRNAs between LUAD and non-cancerous lung tissues. The present study meets the publication guidelines provided by TCGA (<http://cancergenome.nih.gov/publications/publicationguidelines>).

Functional enrichment analysis. The differentially expressed genes were analyzed by Gene Ontology (GO) (22) and Kyoto Encyclopedia of Genes and Genomes (KEGG) (23) pathway enrichment using the cluster Profiler package of R. P-values <0.05 and q values <0.05 were considered to indicate statistically significant differences, and were based on similar functional visualization and clustering.

Preparation of the prognostic signature of differentially expressed lncRNAs for LUAD. The screening threshold criteria for differentially expressed genes were as follows: log₂FC>1 and FDR<0.05. Additionally, the TCGA also collected clinical pathological features, including survival information. Cases with insufficient clinical data were omitted. Ultimately, the prognostic analysis included 493 samples, with 6,280 genes representing 6,205 mRNAs and 75 lncRNAs. The endpoints of the patients with LUAD in terms of overall survival (OS) were also studied. The mean follow-up period for these patients with LUAD was 28 months.

The prognostic value of the genes and the clinicopathological features were first evaluated by single-variable Cox proportional regression analysis (P<0.05). A multivariate Cox regression model with the survival and KMSurv packs (24) further confirmed the statistical indicators, including lncRNAs and clinicopathological features.

Based on the linear combination of multiple genes in the Cox regression model (β) and the linear combination of regression coefficients, the prognostic risk score was established based on a study undertaken by Tang *et al* (25). According to the definition of the median prognostic index (PI), patients with LUAD were classified by high and low scores, and the hazard ratio (HR) and 95% confidence interval (CI) were calculated. The time-dependent receiver operating characteristic (ROC) curve within 5 years was also analyzed as the defining point with the R package 'survival ROC' to evaluate the predictive accuracy of the prognostic model for the results of time-dependent diseases. Kaplan-Meier survival curves were drawn to assess the association between all parameters (clinical examination and prognostic risk score) and the OS of patients with LUAD (26).

Functional evaluation of lncRNAs with WGCNA. In analyzing the incorporation of differentially expressed lncRNAs into the network, WGCNA can describe the correlation patterns in gene expression profiles (27). WGCNA R package was used to assess the performance of an mRNA/lncRNA and

its module membership, and the correlation between each module and disease state was obtained through single-factor variance analysis. All mRNAs associated with an lncRNA in the module that were significantly associated with the disease were considered potential targets for the lncRNA. The function of an lncRNA was obtained by functional enrichment analysis of the target. Following an evaluation of the weighted correlations, the network feature was presented by Cytoscape 3.4.0. (<http://www.cytoscape.org/>).

lncRNA function prediction. The coding gene in the same WGCNA network module is a potential regulatory target gene for an lncRNA. The core enrichment lncRNA in the WGCNA network module, which is significantly associated with disease, is the risk lncRNA in the present study. GO and KEGG enrichment analyses of the risk lncRNAs and coding genes co-expressed in the same WGCNA module were performed using the R clusterProfiler package to obtain the functional information of disease risk lncRNAs.

Network analysis of ceRNA. lncRNAs can bind to miRNAs and affect ceRNA function. MiRanda (<http://miranda.org.uk/>) is used to predict the interaction between lncRNAs and miRNAs. miRanda is based mainly on the combination of circRNA and miRNA free energy size: The smaller the free energy, the stronger the binding capacity. Following application of filters of a threshold maximum energy ≤ -20 and score >160 , numerous microRNAs that bound to lncRNA were predicted, but only microRNAs with the top 10 scores were studied.

Based on the TarBase (<http://www.microrna.gr/tarbase>), miRTarBase (<http://mirtarbase.mbc.nctu.edu.tw/php/index.php>), miRecords (<http://c1.accurascience.com/miRecords/>) and starBase v2.0 (<http://starbase.sysu.edu.cn/>) databases, information on the downstream target genes of microRNAs in the ceRNA network was obtained. Cytoscape software was used to construct microRNA-target and microRNA-lncRNA integration networks that only showed the genes within the module being investigated. The mRNAs in the network were analyzed by GO BP enrichment using the clueGO plugins in Cytoscape software.

The correlations between LUAD-specific lncRNAs and clinical features. According to the bioinformatics analysis and the ceRNA network, prognostic lncRNAs were selected. Cox regression analysis was also performed on the results for the clinical factors, including sex, tumor pathological stage, Tumor-Node-Metastasis (TNM) stage (21) and the number of pack-years.

Statistical analysis. Data are presented as the mean \pm standard deviation and Student's t-tests, two-sided log-rank test, single-factor cox proportional risk regression, multi-factor cox proportional risk regression, R package 'survivalROC' and 'KMSurv' were used to perform statistical analysis. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Data source and pre-processing. The expression data on 20,502 genes were extracted, and R language (EdgeR and

Table I. Differentially expressed genes.

Category	Upregulated	Downregulated	Total
lncRNA	57	18	75
mRNA	4,311	1,894	6,205
Total	4,368	1,912	6,280

The differentially expressed genes were counted using EdgeR and the DESeq package of R ($P_{adj} < 0.05$ and absolute $\log_2FC > 1$). The final genes were selected using a combination of the two methods. Student's t-test was employed to analyze the difference in expression levels of genes between lung adenocarcinoma and non-cancerous lung tissues.

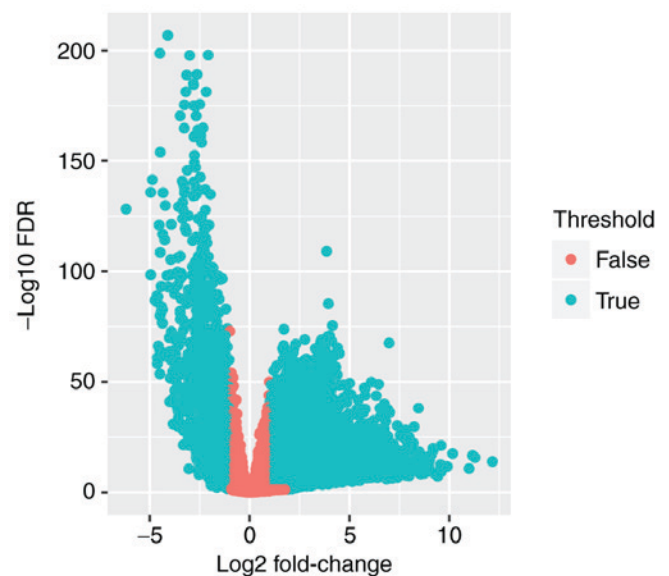


Figure 1. Volcano plot of the differentially expressed genes. The blue portion of the right side indicates high expression and left side indicates low expression. Red represents the long non-coding RNA expression with a $\log_2FC < 1$ and adjusted value < 0.05 . The x-axis represents an adjusted \log_2FC and the y-axis represents the P-value. This volcano plot was conducted by the ggplot2 package of R language.

DESeq) was used to calculate the expression data. The differentially expressed genes ($n=6280$; Table I) were selected to further investigate their prognostic value (Fig. 1).

Prognostic effect of differentially expressed genes. Following exclusion of those with insufficient survival data, the prognosis of 493 patients was obtained. Single-factor cox proportional hazard regression analysis revealed that 328 mRNAs and four lncRNAs had prognostic value in LUAD. Subsequently, multi-factor cox proportional hazard regression analysis was used to evaluate these results, and 32 genes (29 mRNAs and 3 lncRNAs; Table II) were independent prognostic indicators of LUAD ($HR > 1$; $P < 0.05$). The prognostic risk index PI was calculated for each of the 32 prognostic risk genes using a previously described formula, in which the expression of all risk genes is multiplied by the HR value and then summed (28,29). The Kaplan-Meier curve suggested that the median survival time of patients in the high-risk group was

Table II. 32 genes serving as prognostic indicators.

lncRNA	mRNA				
FAM138B	CRISP1	MYL10	NRG1	FAM9C	MYOC
	TP63	CRISP3	BC092501	KRTDAP	SLC16A7
TLX1NB	KRT9	STON1.GTF2A1L	SI	VSIG10L	IQCJ
	ADAM30	SPANXE	ACACB	FAM55A	F9
NHEG1	PLXNB3	ALDOB	BC040891	NRGN	MCC
	BAIAP2L2	FAM184B	C10orf27	RASGRF1	

Single-factor Cox proportional risk regression and multi-factor Cox proportional risk regression analysis was used to evaluate the prognostic indicators in lung adenocarcinoma.

37.8 months, significantly less than that of the low-risk group (47.4 months; $P=0.00409$; Fig. 2A). In addition, the present study also assessed the difference in expression levels of 32 genes between the high-risk and low-risk groups. The prognostic risk index aided in predicting the 5-year survival rate of patients with LUAD, with an AUC value of 0.69 (Fig. 2B). The median prognostic risk index of 'Risk score' and 'survival days' is the separation for low-risk and high-risk patients with LUAD. (Fig. 2C and D).

WGCNA module division and lncRNA target gene prediction.

The WGCNA network module was constructed for all differentially expressed mRNAs and lncRNAs, setting the number of genes in each module to be no less than 20. A total of 19 network modules were divided, and the correlation between the module and LUAD was analyzed by one-way analysis of variance. It was revealed that 11 modules were significantly correlated significantly with the disease. The present study focused on the network modules of the lncRNAs FAM138B (downregulated in LUAD), NHEG1 (downregulated in LUAD) and TLX1NB (upregulated in LUAD). FAM138B is located in the turquoise module containing 3,257 genes, 311 of which were co-expressed with FAM138B. We hypothesized that these 311 genes are potential FAM138B target genes. NHEG1 is located in the blue module containing 900 genes, 582 of which were co-expressed with NHEG1. Therefore, the 582 genes are potential NHEG1 target genes. TLX1NB lies in the turquoise module containing 3,257 genes, 2,262 of which are co-expressed with TLX1NB. We believe that these 2262 genes are potential TLX1NB target genes.

Functional analysis of the lncRNA target gene. From the WGCNA analysis of the FAM138B, NHEG1 and TLX1NB lncRNAs, the target genes of each lncRNA were obtained. The target genes of the lncRNAs were analyzed by GO and KEGG enrichment using the cluster Profiler package of R. The top three upregulated pathways of FAM138B and TLX1NB were ABC transporters, cAMP signaling and vascular smooth muscle contraction. The top five upregulated pathways of NHEG1 were cell cycle, DNA replication, homologous recombination, Fanconi anemia and p53 signaling (Fig. 3).

There were 14 prognostic risk genes among the target genes of FAM138B and 10 genes among the target genes of TLX1NB

(Table III). The two lncRNAs FAM138B and TLX1NB shared MYOC, PLXNB3 and ACACB among the eight prognostic risk genes as target genes (Fig. 4).

Construction of lncRNA-miRNA network. Miranda predicted microRNAs that were bound to the three lncRNAs, and 712 miRNAs were obtained at the thresholds. According to the predicted scores, the present study focused on the 27 miRNAs with the 10 highest scores (including duplicate scores) that bound to each lncRNA. The miRNA and lncRNA interaction network, which contained 715 nodes and 815 edges, was constructed (Fig. 5).

miRNA target gene prediction. Based on the TarBase, miRTarBase, miRecords and starBase v2.0 databases, the target genes of the aforementioned 27 key miRNAs were obtained, and the microRNA-mRNA network was constructed. The network contained 1,961 sides, 27 microRNAs and 1,229 mRNAs (816 upregulated and 413 downregulated; Fig. 6). Furthermore, GO BP enrichment analysis of the mRNAs in the network was performed (Fig. 7).

Construction of lncRNA-miRNA-mRNA ceRNA network.

The miRNA-mRNA and lncRNA-miRNA pairs were used to build the lncRNA-miRNA-mRNA ceRNA network. The 29 previous prognostic risk mRNAs and their microRNAs were separately extracted, and a regulatory network was established. The network also added lncRNAs, which regulate potential miRNAs, and constructed an important lncRNA-miRNA-mRNA network (Fig. 8). As demonstrated, hsa-miR-619-5p has a regulatory association with three prognostic risk factors. Three upregulated mRNAs and one downregulated mRNA were potentially affected by the lncRNA-ceRNA network.

Correlations between LUAD-specific lncRNAs and clinical features.

According to the single-factor and multi-factor Cox proportional risk regression analysis, three specific lncRNAs-FAM138B, NHEG1 and TLX1NB-were selected. Cox regression analysis for clinical factors, including sex, tumor pathological stage, TNM stage and number of pack-years, was then performed. The expression levels of these three lncRNAs were significantly altered by 'T' stage and primary tumor size ($P<0.05$; Table IV).

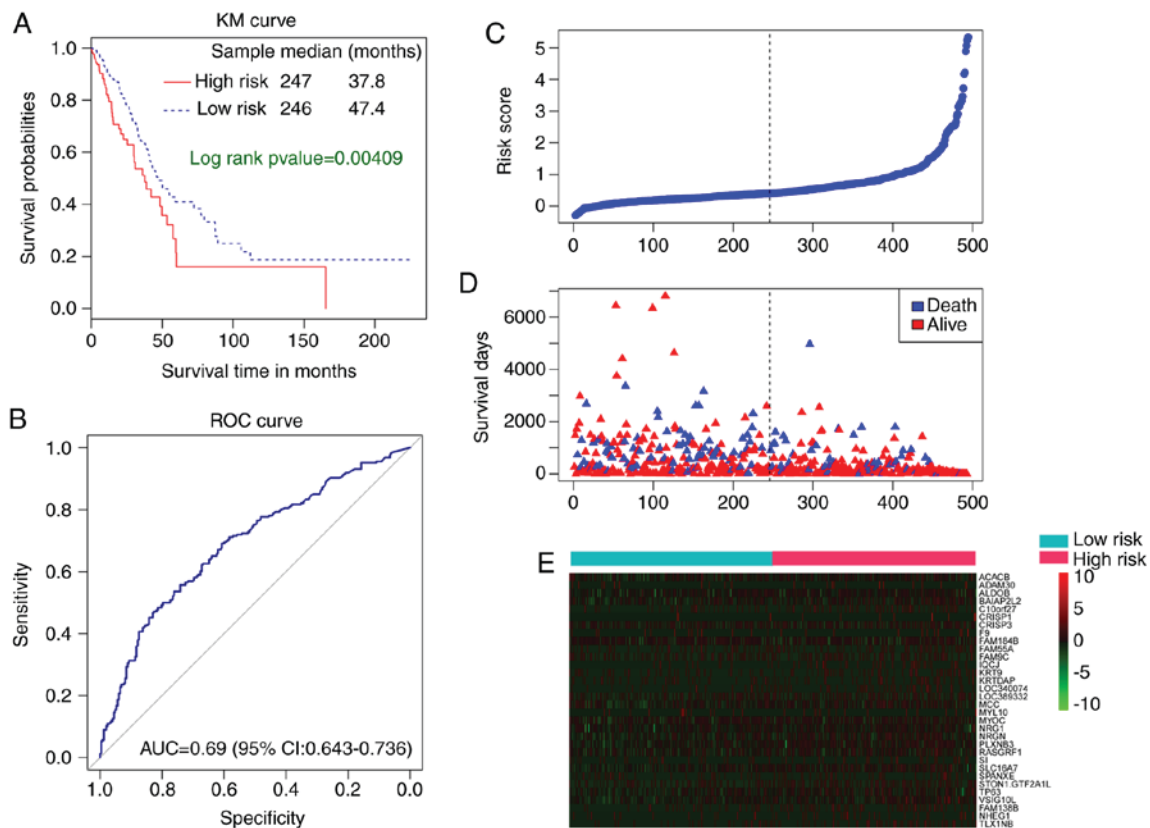


Figure 2. LncRNA risk score analysis and ROC Kaplan-Meier curves for the prognostic lncRNAs signature of patients with LUAD. (A) Kaplan-Meier survival curves showing overall survival outcomes according to relative high-risk and low-risk patients. P-values were calculated using the two-sided log-rank test. (B) Time-dependent ROC curve analysis for survival prediction by the 32 risk genes using the R package 'survivalROC'. (C) The low and high score group for the 32 risk genes in patients with LUAD. (D) The survival status and duration of LUAD cases. (E) Heat-map of the 32 risk genes expression in LUSC. The color gradient from green to red shows a trend from low expression to high expression. ROC, receiver operating characteristic; LUAC, lung adenocarcinoma.

GEO validation. The prognostic value of these three new lncRNAs was then assessed using GEO data. However, there were not enough survival data available in the GEO datasets. Therefore, the prognostic value of these lncRNA signatures requires further verification using other approaches in a larger number of samples.

Discussion

Lung cancer has the highest incidence and mortality rate of any disease worldwide (30), and lung glandular cell cancer is one of the most common types of LUAD (31). Surgery is considered the most effective method of curing LUAD. However, the majority of patients with LUAD in hospital have missed the opportunity for surgery (32). Chemotherapy and radiotherapy are the typical second-line treatments for LUAD, but their effects are not satisfactory (33). In previous years, the diagnosis of lung cancer, surgical techniques and novel drug treatments have improved, but the 5-year response rate for lung cancer continues to be maintained at 10% (34), mainly because lung cancer cannot be diagnosed in the early stages. Although numerous tests can support lung cancer examination and staging, including tissue biopsy and thoracoscopy, these methods are time-consuming and will delay early diagnosis and treatment (35). Therefore, to improve this situation, more attention is being paid to the occurrence and development of lung cancer-related genes and their regulatory mechanisms.

With the development of high-throughput sequencing technology and bioinformatics, lncRNAs have been revealed to be involved in the development of numerous diseases, particularly tumors (36-39). However, the study of lncRNA expression profiles in LUAD has gradually increased. Using TNM I stage lung adenocarcinoma samples in the TCGA database, Tian *et al* (40) demonstrated that FENDRR and LINC00312 had diagnostic value in patients with LUAD. A recent study has demonstrated that lncRNA can act as a ceRNA that adsorbs onto miRNAs to regulate mRNA expression (41). The ceRNA hypothesis is a novel regulatory mechanism that serves a role through miRNA competition (10,42). However, the study of ceRNAs in LUAD is not extensive. Li *et al* (43) revealed MIAT, MEG3 and LINC00115 to be the basic therapeutic targets of LUAD as ceRNAs that regulate miRNA-mRNA pairs. Sui *et al* (44) demonstrated that AFAP1-AS1 is associated with tumor pathological stage and that LINC00472 is associated with lymph node metastasis. Liu *et al* (45) presented lncRNA landscapes and revealed highly altered, differentially expressed lncRNAs in LUSC and LUAD. Two lncRNAs (IGF2BP2-AS1 and DGCR5) were correlated with an improved OS in LUSC, and three (MIR31HG, CDKN2A-AS1 and LINC01600) predicted a poor OS in LUAD.

The public lncRNA data were mined in the latest study (46,47) from TCGA, which identified risk lncRNAs and mRNAs to construct a ceRNA network. The present study first obtained tumor tissue and their corresponding normal tissue

Table III. lncRNA FAM138B and TLX1NB target gene.

lncRNA	Target gene	log2FC	FDR
FAM138B	MYOC	-4.357717939	0.0000000294
	PLXNB3	3.878550766	0.0000000191
	ACACB	-1.379164532	0.0000000121
	MCC	-1.27636241	0.0000000488
	RASGRF1	-2.055290402	0.0000000424
	VSIG10L	2.102105505	0.0000000327
	SLC16A7	1.372035277	0.0000000011
	NRGN	-1.203111742	0.00000000132
	NRG1	-1.17178188	0.000000029
	TLX1NB	2.102744205	0.0000335
	LOC340074	1.475322606	0.000465275
	STON1-GTF2A1L	-1.249288199	0.000666914
	MYL10	1.376749843	0.02548923
	C10orf27	1.038217387	0.03122004
TLX1NB	MYOC	-4.35771794	0.000000000272
	PLXNB3	3.87855077	0.000000000943
	ACACB	-1.37916453	0.000000000669
	MCC	-1.27636241	0.000000000432
	RASGRF1	-2.0552904	0.000000000571
	NRGN	-1.20311174	0.000000000444
	KRT9	2.93858028	0.00000755
	FAM138B	-1.11756478	0.00016011
	STON1-GTF2A1L	-1.2492882	0.000392098
	MYL10	1.37674984	0.018236455

lncRNA, long non-coding RNA.

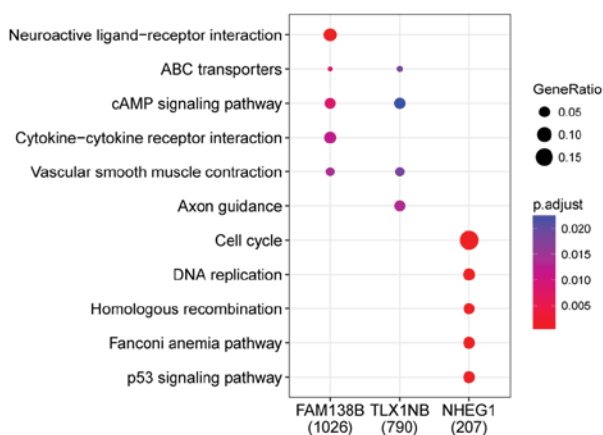


Figure 3. Significantly enriched Kyoto Encyclopedia of Genes and Genomes terms of the risk long non-coding RNA. The point color represents the P.adjust value, and the point size GeneRatio represents the ratio of the number of differential genes in the Gene Ontology item to the total number of differential genes.

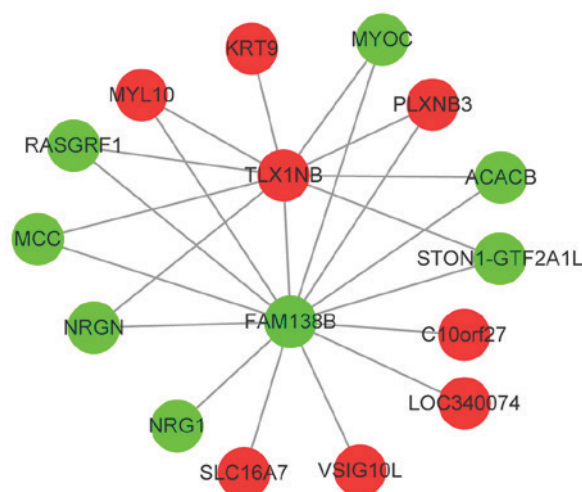


Figure 4. lncRNA FAM138B, lncRNA TLX1NB and their target risk genes. lncRNA, long non-coding RNA.

lncRNAs and mRNAs, and subsequently, those RNAs that tended to have prognostic value were selected by univariate and multivariate analysis. Next, 29 mRNAs and three lncRNAs were yielded to construct the PI value (25). Notably, the PI value proved to be an independent prognostic indicator of

LUAD. It was revealed that the PI, constructed from 32 differential genes, had good prognostic value in LUAD. The present study used TCGA data that were generated by RNA-Seq and used to assess the survival of a large number of patients with LUAD in 493 cases.

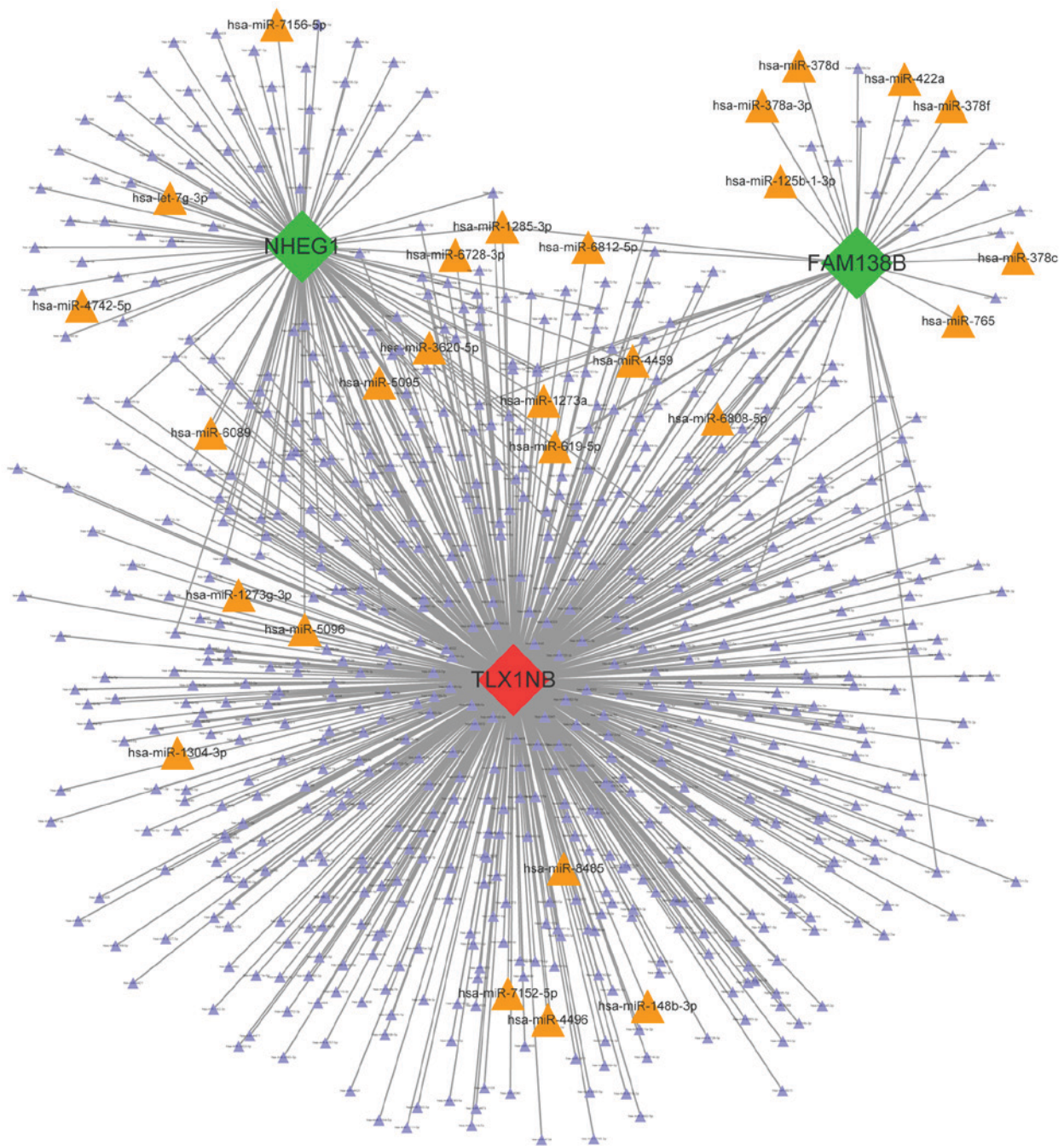


Figure 5. IncRNA-miRNA co-expression network (green for downregulated, red for upregulated, triangular for miRNA, orange for scoring TOP10 miRNA). IncRNA, long non-coding RNA; miRNA, microRNA.

A total of 29 mRNAs and three lncRNAs were involved in LUAD risk genes. The present study investigated the molecular mechanisms of these lncRNAs. However, none of these lncRNAs was identified in reference records, and little was known regarding the functional mechanisms of these three lncRNAs. Therefore, a WGCNA study on lncRNA-associated genes was conducted, introducing the 14 candidates that could be the most relevant genes in LUAD into the network. The present study focused on the three risk lncRNAs, and genes within these modules that had a weighted association. These genes were the lncRNA target genes. Subsequently, the bioinformatics of abnormally expressed mRNAs was studied, and the most highly

enriched KEGG pathway annotations could be the functions of lncRNAs. Based on the KEGG pathway analysis, several cancer-associated pathways were identified, including p53 signaling and the cell cycle.

Lu *et al* (48) assessed the expression and function of the lncRNA SOX21 antisense RNA 1 (sox21-as1) in 68 pairs of LUAD tissues and the corresponding non-tumor tissues, and demonstrated that sox21-as1 is associated with the development of LUAD. Furthermore, the knockdown of sox21-as1 inhibited the proliferation of LUAD cells *in vitro* and *in vivo*, inducing cell cycle stagnation and apoptosis. Wu *et al* (49) integrated two datasets to search for novel target genes and pathways to explain the pathogenicity of LUAD, and discovered that

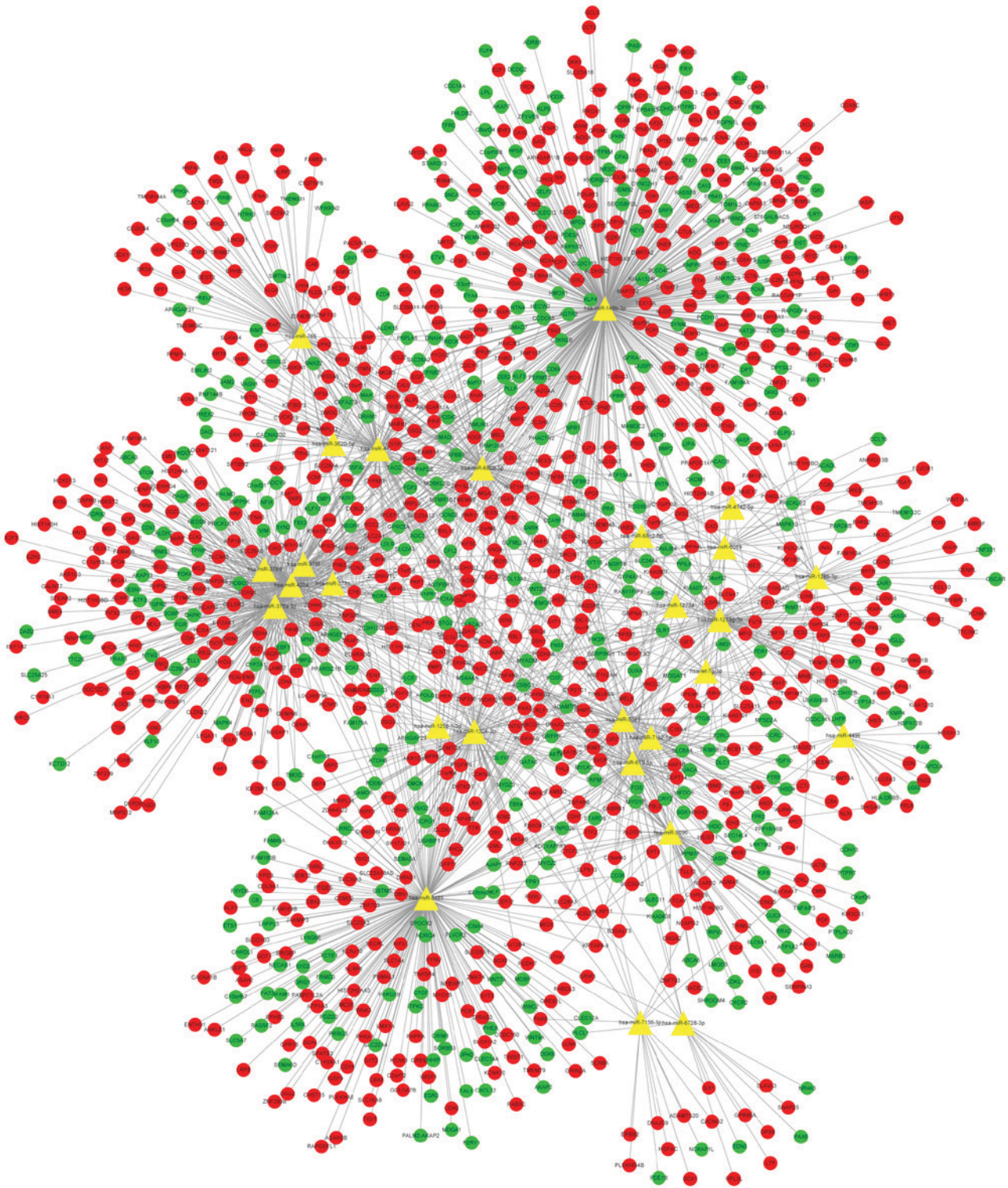


Figure 6. miRNA-mRNA co-expression network. Red for upregulated genes, green for downregulated genes, triangular for miRNAs and circular for mRNAs. Due to the large number of miRNA target genes, the figure only includes the differentially expressed mRNAs. miRNA, microRNA.

PPM1D and GADD45B could modulate the progression of LUAD through p53 signaling.

Furthermore, the present study investigated the target genes of three lncRNAs. Fourteen of the FAM138B target genes were prognostic risk genes, as were 10 TLX1NB target genes-8 of which were shared. In the absence of calcium, NRG1, which combines calmodulin with synaptic protein kinase substrates, is reduced in monkeys and rats following exposure to welding

smoke (50). MCC is a candidate colorectal tumor suppressor gene that is thought to negatively modulate cell cycle progression (51) and encodes guanine nucleotide exchange factor (GEF). RASGRF1 stimulates the dissociation of GDP from RAS. This gene is expressed in various tissues other than the brain, including the lung and pancreas, and several tumor cell lines (52). KRT9 serves an important role in mature palm and plantar skin tissue and in the morphological processes that

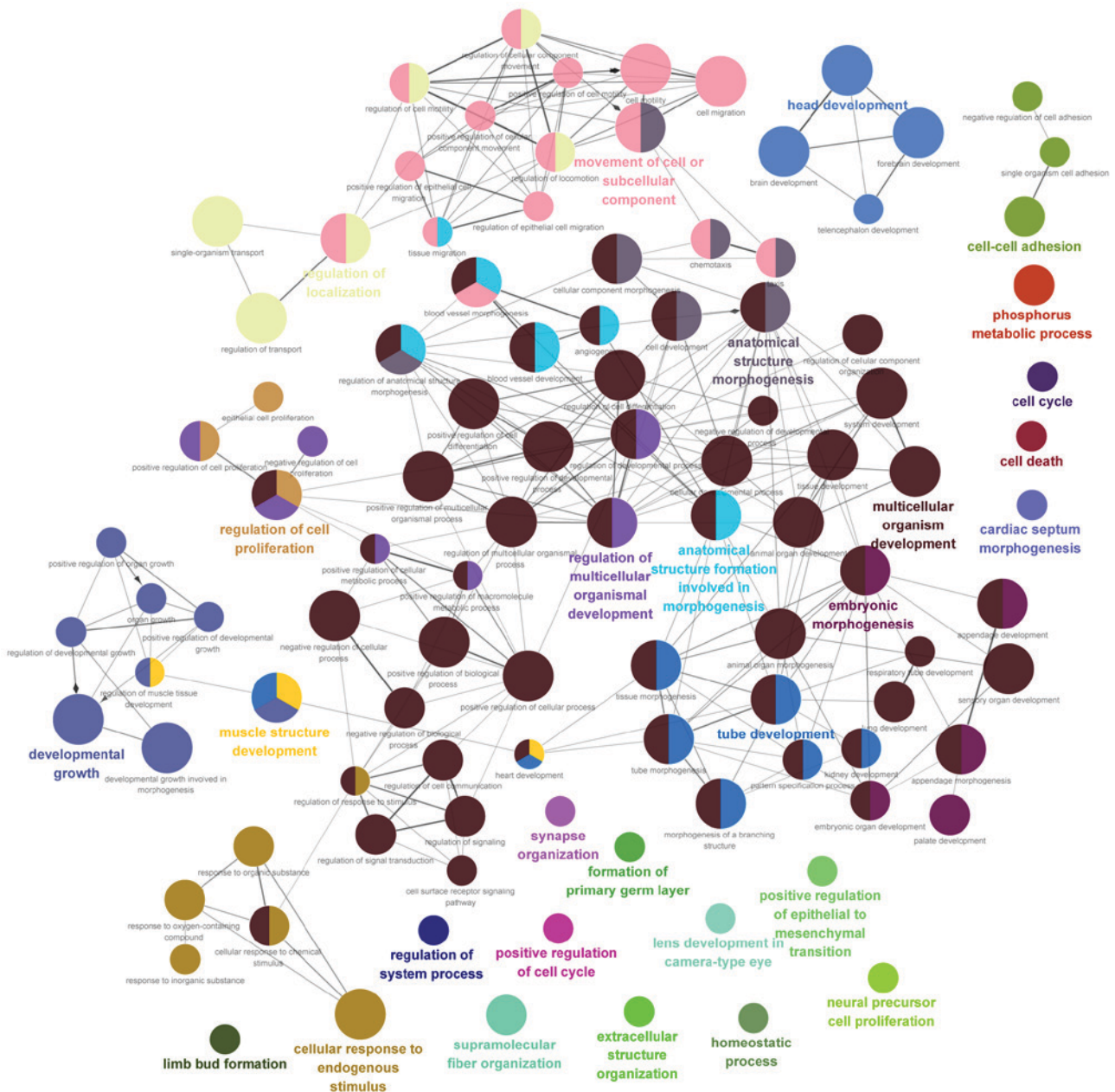


Figure 7. miRNA-target genes of Gene Ontology Biological Process enrichment results. The results were calculated as $-\log_{10}$ (P-value) ($P < 0.05$). miRNA, microRNA.

form these tissues, and is upregulated (53,54) in platycodin D (PD)-treated HepG2 cells. PLXNB3, a part of the coding family of plexiform proteins, acts as a receptor for signal 5A and serves a role in invasive growth, axon guidance and cell migration. Plexin and B3 serve vital roles in the invasion and metastasis of gastric cancer (55). ACACB may be involved in the regulation of fatty acid oxidation. It inhibits the synthesis of fatty acids and tumor development in non-small cell lung cancer in preclinical models (56).

lncRNAs can function as ceRNAs by binding miRNAs, regulating the expression of genes. LUAD-associated lncRNA-related ceRNA networks provide vital hints in detecting crucial RNAs of ceRNA-mediated gene regulation networks during the initiation and growth of LUAD. There is increasing evidence that lncRNAs are a key part of ceRNA networks by regulating the transcription of other

RNAs (5,57-61). For example, HOTAIR can use miR-331-3p as endogenous lodging to eliminate the inhibitory activity of the miRNA induced by HER2 3'-UTR and increase its post-transcriptional levels (62). Therefore, as potential connections to lncRNA, miRNA and mRNA may be involved in the initiation, as well as the development, of LUAD. The present study predicted a network involving three types of risk lncRNAs and 712 possible target miRNAs. To facilitate the next step, 27 miRNAs with a score of TOPI0 were selected. Based on the TarBase, miRTarBase, miRecords and starBase v2.0 databases, 1,229 objective genes of these 27 miRNAs were obtained, and GO enrichment analysis was performed on these target genes. The GO enrichment results included positive regulation of the epithelial-to-mesenchymal transition, regulation of cell proliferation, positive regulation of the cell cycle and cell death, and multicellular organism development.

Table IV. The correlations between lung adenocarcinoma specific lncRNAs and clinical features.

Id	Coef	Secoef	HR	Lower.95.CI	Upper.95.CI	P-value
Sex	0.00980122	0.08968855	1.0098494	0.84705885	1.2039256	0.99727594
Smoke	0.00000723	0.00211715	1.0000072	0.99586625	1	0.99727594
Stage	0.48304592	0.47140751	1.6210043	0.64345974	4.08363557	0.30551012
Metastasis	-0.1477427	0.24226305	0.8626531	0.53656395	1.38691821	0.93402361
Node	-0.0575892	0.11703873	0.9440377	0.75052609	1.18744332	0.93402361
Tumor	1.29785383	0.19225169	3.6614302	2.51192012	5.34	0.000000000147

Single-factor Cox regression analysis was used to evaluate the correlations with the R survival package (P<0.05).

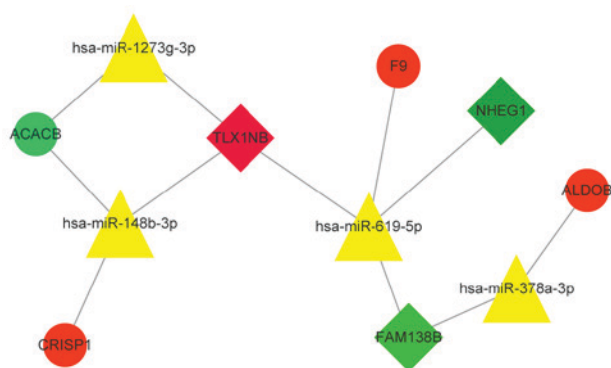


Figure 8. lncRNA-miRNA-mRNA ceRNA network. Red represents upregulated, green represents downregulated, and yellow triangle represents miRNAs. lncRNA, long non-coding RNA; miRNA, microRNA; ceRNA, competing endogenous RNA.

Among the 1,229 genes, there were four prognostic risk genes. ALDOB is associated with the GO term phosphorus metabolic process. The main function is to catalyze the reversible transformation of fructose-1, 6-diphosphite to dihydroxyacetone and glyceraldehyde 3-phosphate (63). Abnormal changes in ALDOB are associated with a number of diseases, including hereditary fructose intolerance, hepatitis, cirrhosis and cancer (64). Using data mined from the Gene Expression Omnibus for the colorectal cancer transcriptome (GSE35452) database, Tian *et al* (65) reported that ALDOB is associated with glycolysis and is the most significantly increased transcript (GO: 0006096). It may serve a significant role in colorectal cancer progression and the response to neoadjuvant concurrent chemoradiotherapy, and as a novel prognostic biomarker. ACACB positively regulates organ growth. It catalyzes the ATP-dependent carboxylation of acetyl-CoA to malonyl-CoA (66). Recent studies have reported that ACACB is increased in a variety of human cancer types, most likely to promote fat generation and meet the requirement for rapid growth and proliferation (67-69). Jones *et al* (70) demonstrated that knockout of ACACB changes the lipid content in glioblastoma multiform U87 cells, increases caspase activity and inhibits the growth of fat to generate tumor treatment effects. The F9 gene encodes vitamin K-dependent coagulation factor IX, which circulates in the blood as an inactive zymogen (71). Alterations in this gene cause blood coagulation factor IX deficiency, which is a recessive X-linked

disorder, also called hemophilia B or Christmas disease. The presentation of hemophilia B is consistent with easy bruising, urinary tract bleeding and nosebleeds. We hypothesized that tumor-associated mutations in F9 may exacerbate bleeding symptoms due to necrosis of the tumor. CRISP1, also known as astrocyte elevated gene-1 (AEG-1), is an oncogene that is overexpressed in a wide range of cancer types and serves a key role in regulating carcinogenesis (72). AEG-1 serves an important role in regulating hepatocellular carcinoma (HCC). AEG-1-overexpression at the mRNA and protein expression levels has been observed in a high percentage (>90%) of patients with HCC. A progressive increase in the levels of AEG-1 has been observed in patients with HCC, correlating directly with disease stage, a higher rate of recurrence and a poorer overall survival (73,74).

The present study predicted a network consisting of lncRNAs, miRNAs and target genes, and these target genes are also cancer-related genes regulated by lncRNAs through the ceRNA network. The lncRNA FAM138B regulates ALDOB through hsa-miR-378a-3p and regulates F9 through hsa-miR-619-5p; the lncRNA NHEG1 also regulates F9 through hsa-miR-619-5p. The lncRNA TLX1NB regulates CRISP1 through hsa-miR-148b-3p and regulates ACACB through hsa-miR-1273g-3p. Therefore, hsa-miR-619-5p is at the center of three prognostic risk lncRNA ceRNA regulation networks and may be a key miRNA.

Therefore, three lncRNAs may be candidate prognostic biomarkers for LUAD patients with significant clinical significance. In summary, the present study examined lncRNAs in 493 patients with LUAD from TCGA and constructed three ceRNA networks as novel prognostic indicators of LUAD. However, further experiments are required to verify the clinical and biological functions of these three lncRNAs.

Acknowledgements

The authors would like to thank Dr Sean Kim (Yale University) for editing the English text of the first draft of this manuscript.

Funding

The present study was supported by funds from the Gansu Provincial Health Bureau Scientific Research Project (grant no. GSWSKY2018-09).

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

PR and LW contributed toward the conception and design of the study; BL analyzed and interpreted the data; and LW and XL designed the experiments, drafted, and revised the manuscript..

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- Siegel R, Naishadham D and Jemal A: Cancer statistics, 2012. *CA Cancer J Clin* 62: 10-29, 2012.
- Lin JJ, Cardarella S, Lydon CA, Dahlberg SE, Jackman DM, Jänne PA and Johnson BE: Five-Year survival in EGFR-Mutant metastatic lung adenocarcinoma treated with EGFR-TKIs. *J Thorac Oncol* 11: 556-565, 2016.
- Qi L, Li Y, Qin Y, Shi G, Li T, Wang J, Chen L, Gu Y, Zhao W and Guo Z: An individualised signature for predicting response with concordant survival benefit for lung adenocarcinoma patients receiving platinum-based chemotherapy. *Br J Cancer* 115: 1513-1519, 2016.
- Lee JT: Epigenetic regulation by long noncoding RNAs. *Science* 338: 1435-1439, 2012.
- Zhou M, Wang X, Shi H, Cheng L, Wang Z, Zhao H, Yang L and Sun J: Characterization of long non-coding RNA-associated ceRNA network to reveal potential prognostic lncRNA biomarkers in human ovarian cancer. *Oncotarget* 7: 12598-12611, 2016.
- Wang PL, Liu B, Xia Y, Pan CF, Ma T and Chen YJ: Long non-coding RNA-Low expression in tumor inhibits the invasion and metastasis of esophageal squamous cell carcinoma by regulating p53 expression. *Mol Med Rep* 13: 3074-3082, 2016.
- Shang C, Guo Y, Zhang J and Huang B: Silence of long noncoding RNA UCA1 inhibits malignant proliferation and chemotherapy resistance to adriamycin in gastric cancer. *Cancer Chemother Pharmacol* 77: 1061-1067, 2016.
- Wang Y, Zhang Y, Yang T, Zhao W, Wang N, Li P, Zeng X and Zhang W: Long non-coding RNA MALAT1 for promoting metastasis and proliferation by acting as a ceRNA of miR-144-3p in osteosarcoma cells. *Oncotarget* 8: 59417-59434, 2017.
- Wang P, Zhi H, Zhang Y, Liu Y, Zhang J, Gao Y, Guo M, Ning S and Li X: miRSponge: A manually curated database for experimentally supported miRNA sponges and ceRNAs. *Database (Oxford)* 2015: pii: bav098, 2015.
- Salmela L, Poliseno L, Tay Y, Kats L and Pandolfi PP: A ceRNA hypothesis: the Rosetta stone of a hidden RNA language? *Cell* 146: 353-358, 2011.
- Tan JY, Sirey T, Honti F, Graham B, Piovesan A, Merckenschlager M, Webber C, Ponting CP and Marques AC: Extensive microRNA-mediated crosstalk between lncRNAs and mRNAs in mouse embryonic stem cells. *Genome Res* 25: 655-666, 2015.
- Karreth FA and Pandolfi PP: ceRNA cross-talk in cancer: When ce-bling rivalries go awry. *Cancer Discov* 3: 1113-1121, 2013.
- Akcakaya P, Ekelund S, Kolosenko I, Caramuta S, Ozata DM, Xie H, Lindfors U, Olivecrona H and Lui WO: miR-185 and miR-133b deregulation is associated with overall survival and metastasis in colorectal cancer. *Int J Oncol* 39: 311-318, 2011.
- Zhang J, Fan D, Jian Z, Chen GG and Lai PB: Cancer specific long noncoding RNAs show differential expression patterns and competing endogenous RNA potential in hepatocellular carcinoma. *PLoS One* 10: e0141042, 2015.
- Zhang Y, Chen Z, Li MJ, Guo HY and Jing NC: Long non-coding RNA metastasis-associated lung adenocarcinoma transcript 1 regulates the expression of Gli2 by miR-202 to strengthen gastric cancer progression. *Biomed Pharmacother* 85: 264-271, 2017.
- Liu T, Zhang X, Yang YM, Du LT and Wang CX: Increased expression of the long noncoding RNA CRNDE-h indicates a poor prognosis in colorectal cancer, and is positively correlated with IRX5 mRNA expression. *Onco Targets Ther* 9: 1437-1448, 2016.
- Han D, Gao X, Wang M, Qiao Y, Xu Y, Yang J, Dong N, He J, Sun Q, Lv G, *et al*: Long noncoding RNA H19 indicates a poor prognosis of colorectal cancer and promotes tumor growth by recruiting and binding to eIF4A3. *Oncotarget* 7: 22159-22173, 2016.
- Qiu JJ and Yan JB: Long non-coding RNA LINC01296 is a potential prognostic biomarker in patients with colorectal cancer. *Tumour Biol* 36: 7175-7183, 2015.
- Yin DD, Liu ZJ, Zhang E, Kong R, Zhang ZH and Guo RH: Decreased expression of long noncoding RNA MEG3 affects cell proliferation and predicts a poor prognosis in patients with colorectal cancer. *Tumour Biol* 36: 4851-4859, 2015.
- Liu Y, Zhang M, Liang L, Li J and Chen YX: Over-expression of lncRNA DANCR is associated with advanced tumor progression and poor prognosis in patients with colorectal cancer. *Int J Clin Exp Pathol* 8: 11480-11484, 2015.
- Edge SB and Compton CC: The American Joint Committee on Cancer: The 7th edition of the AJCC cancer staging manual and the future of TNM. *Ann Surg Oncol* 17: 1471-1474, 2010.
- Gene Ontology Consortium: The gene ontology: Enhancements for 2011. *Nucleic Acids Res* 40 (Database Issue): D559-D564, 2012.
- Kanehisa M: The KEGG database. *Novartis Found Symp* 247: 91-103, 101-103, 119-128, 244-152, 2002.
- Foulger RE, Osumi-Sutherland D, McIntosh BK, Hulo C, Masson P, Poux S, Le Mercier P and Lomax J: Representing virus-host interactions and other multi-organism processes in the Gene Ontology. *BMC Microbiol* 15: 146, 2015.
- Tang RX, Chen WJ, He RQ, Zeng JH, Liang L, Li SK, Ma J, Luo DZ and Chen G: Identification of a RNA-Seq based prognostic signature with five lncRNAs for lung squamous cell carcinoma. *Oncotarget* 8: 50761-50773, 2017.
- Zhou M, Sun Y, Sun Y, Xu W, Zhang Z, Zhao H, Zhong Z and Sun J: Comprehensive analysis of lncRNA expression profiles reveals a novel lncRNA signature to discriminate nonequivalent outcomes in patients with ovarian cancer. *Oncotarget* 7: 32433-32448, 2016.
- Presson AP, Sobel EM, Papp JC, Suarez CJ, Whistler T, Rajeevan MS, Vernon SD and Horvath S: Integrated weighted gene co-expression network analysis with an application to chronic fatigue syndrome. *BMC Syst Biol* 2: 95, 2008.
- Lossos IS, Czerwinski DK, Alizadeh AA, Wechsler MA, Tibshirani R, Botstein D and Levy R: Prediction of survival in diffuse large-B-cell lymphoma based on the expression of six genes. *N Engl J Med* 350: 1828-1837, 2004.
- Alizadeh AA, Gentles AJ, Alencar AJ, Liu CL, Kohrt HE, Houot R, Goldstein MJ, Zhao S, Natkunam Y, Advani RH, *et al*: Prediction of survival in diffuse large B-cell lymphoma based on the expression of 2 genes reflecting tumor and microenvironment. *Blood* 118: 1350-1358, 2011.
- Meza R, Meernik C, Jeon J and Cote ML: Lung cancer incidence trends by gender, race and histology in the United States, 1973-2010. *PLoS One* 10: e0121323, 2015.
- Kerr KM: Pulmonary adenocarcinomas: Classification and reporting. *Histopathology* 54: 12-27, 2009.
- Aokage K, Yoshida J, Hishida T, Tsuboi M, Saji H, Okada M, Suzuki K, Watanabe S and Asamura H: Limited resection for early-stage non-small cell lung cancer as function-preserving radical surgery: A review. *Jpn J Clin Oncol* 47: 7-11, 2017.
- Provencio M, Isla D, Sanchez A and Cantos B: Inoperable stage III non-small cell lung cancer: Current treatment and role of vinorelbine. *J Thorac Dis* 3: 197-204, 2011.

34. Ortea I, Rodriguez-Ariza A, Chicano-Galvez E, Arenas Vacas MS and Jurado Gamez B: Discovery of potential protein biomarkers of lung adenocarcinoma in bronchoalveolar lavage fluid by SWATH MS data-independent acquisition and targeted data extraction. *J Proteomics* 138: 106-114, 2016.
35. Navani N, Nankivell M, Lawrence DR, Lock S, Makker H, Baldwin DR, Stephens RJ, Parmar MK, Spiro SG, Morris S, *et al*: Lung cancer diagnosis and staging with endobronchial ultrasound-guided transbronchial needle aspiration compared with conventional approaches: An open-label, pragmatic, randomised controlled trial. *Lancet Respir Med* 3: 282-289, 2015.
36. Liu Y, Yang Y, Li L, Liu Y, Geng P, Li G and Song H: LncRNA SNHG1 enhances cell proliferation, migration and invasion in cervical cancer. *Biochem Cell Biol* 96: 38-43, 2018.
37. Zhao M, Sun D, Li X, Xu Y, Zhang H, Qin Y and Xia M: Overexpression of long noncoding RNA PEG10 promotes proliferation, invasion and metastasis of hypopharyngeal squamous cell carcinoma. *Oncol Lett* 14: 2919-2925, 2017.
38. Bao H, Guo CG, Qiu PC, Zhang XL, Dong Q and Wang YK: Long non-coding RNA Igf2as controls hepatocellular carcinoma progression through the ERK/MAPK signaling pathway. *Oncol Lett* 14: 2831-2837, 2017.
39. Lai Y, Xu P, Liu J, Li Q, Ren D, Zhang J and Wang J: Decreased expression of the long non-coding RNA MLLT4 antisense RNA 1 is a potential biomarker and an indicator of a poor prognosis for gastric cancer. *Oncol Lett* 14: 2629-2634, 2017.
40. Tian Z, Wen S, Zhang Y, Shi X, Zhu Y, Xu Y, Lv H and Wang G: Identification of dysregulated long non-coding RNAs/microRNAs/mRNAs in TNM I stage lung adenocarcinoma. *Oncotarget* 8: 51703-51718, 2017.
41. Chen G, Peng L, Zhu Z, Du C, Shen Z, Zang R, Su Y, Xia Y and Tang W: LncRNA AFAP1-AS functions as a competing endogenous RNA to regulate RAP1B expression by sponging miR-181a in the HSCR. *Int J Med Sci* 14: 1022-1030, 2017.
42. Tay Y, Rinn J and Pandolfi PP: The multilayered complexity of ceRNA crosstalk and competition. *Nature* 505: 344-352, 2014.
43. Li DS, Ainiwaer JL, Sheyhiding I, Zhang Z and Zhang LW: Identification of key long non-coding RNAs as competing endogenous RNAs for miRNA-mRNA in lung adenocarcinoma. *Eur Rev Med Pharmacol Sci* 20: 2285-2295, 2016.
44. Sui J, Li YH, Zhang YQ, Li CY, Shen X, Yao WZ, Peng H, Hong WW, Yin LH, Pu YP and Liang GY: Integrated analysis of long non-coding RNA-associated ceRNA network reveals potential lncRNA biomarkers in human lung adenocarcinoma. *Int J Oncol* 49: 2023-2036, 2016.
45. Liu B, Chen Y and Yang J: LncRNAs are altered in lung squamous cell carcinoma and lung adenocarcinoma. *Oncotarget* 8: 24275-24291, 2017.
46. Zhang Y, Wen DY, Zhang R, Huang JC, Lin P, Ren FH, Wang X, He Y, Yang H, Chen G and Luo DZ: A preliminary investigation of PVT1 on the effect and mechanisms of hepatocellular carcinoma: Evidence from clinical data, a meta-analysis of 840 cases, and in vivo validation. *Cell Physiol Biochem* 47: 2216-2232, 2018.
47. Jiang R, Zhao C, Gao B, Xu J, Song W and Shi P: Mixomics analysis of breast cancer: Long non-coding RNA linc01561 acts as ceRNA involved in the progression of breast cancer. *Int J Biochem Cell Biol* 102: 1-9, 2018.
48. Lu X, Huang C, He X, Liu X, Ji J, Zhang E, Wang W and Guo R: A novel long Non-coding RNA, SOX21-AS1, indicates a poor prognosis and promotes lung adenocarcinoma proliferation. *Cell Physiol Biochem* 42: 1857-1869, 2017.
49. Wu X, Zang W, Cui S and Wang M: Bioinformatics analysis of two microarray gene-expression data sets to select lung adenocarcinoma marker genes. *Eur Rev Med Pharmacol Sci* 16: 1582-1587, 2012.
50. Heo JD, Oh JH, Lee K, Kim CY, Song CW, Yoon S, Han JS and Yu IJ: Gene expression profiling in the lung tissue of cynomolgus monkeys in response to repeated exposure to welding fumes. *Arch Toxicol* 84: 191-203, 2010.
51. Fukuyama R, Niculaita R, Ng KP, Obusez E, Sanchez J, Kalady M, Aung PP, Casey G and Sizemore N: Mutated in colorectal cancer, a putative tumor suppressor for serrated colorectal cancer, selectively represses beta-catenin-dependent transcription. *Oncogene* 27: 6044-6055, 2008.
52. Guerrero C, Rojas JM, Chedid M, Esteban LM, Zimonjic DB, Popescu NC, Font de Mora J and Santos E: Expression of alternative forms of Ras exchange factors GRF and SOS1 in different human tissues and cell lines. *Oncogene* 12: 1097-1107, 1996.
53. Langbein L, Heid HW, Moll I and Franke WW: Molecular characterization of the body site-specific human epidermal cytokeratin 9: cDNA cloning, amino acid sequence, and tissue specificity of gene expression. *Differentiation* 55: 57-71, 1993.
54. Lu JJ, Lu DZ, Chen YF, Dong YT, Zhang JR, Li T, Tang ZH and Yang Z: Proteomic analysis of hepatocellular carcinoma HepG2 cells treated with platycodin D. *Chin J Nat Med* 13: 673-679, 2015.
55. Pan GQ, Ren HZ, Zhang SF, Wang XM and Wen JF: Expression of semaphorin 5A and its receptor plexin B3 contributes to invasion and metastasis of gastric carcinoma. *World J Gastroenterol* 15: 2800-2804, 2009.
56. Svensson RU, Parker SJ, Eichner LJ, Kolar MJ, Wallace M, Brun SN, Lombardo PS, Van Nostrand JL, Hutchins A, Vera L, *et al*: Inhibition of acetyl-CoA carboxylase suppresses fatty acid synthesis and tumor growth of non-small-cell lung cancer in preclinical models. *Nat Med* 22: 1108-1119, 2016.
57. Li CY, Liang GY, Yao WZ, Sui J, Shen X, Zhang YQ, Peng H, Hong WW, Ye YC, Zhang ZY, *et al*: Integrated analysis of long non-coding RNA competing interactions reveals the potential role in progression of human gastric cancer. *Int J Oncol* 48: 1965-1976, 2016.
58. Guo LL, Song CH, Wang P, Dai LP, Zhang JY and Wang KJ: Competing endogenous RNA networks and gastric cancer. *World J Gastroenterol* 21: 11680-11687, 2015.
59. Hu Y, Tian H, Xu J and Fang JY: Roles of competing endogenous RNAs in gastric cancer. *Brief Funct Genomics* 15: 266-273, 2016.
60. Wu Q, Guo L, Jiang F, Li L, Li Z and Chen F: Analysis of the miRNA-mRNA-lncRNA networks in ER+ and ER-breast cancer cell lines. *J Cell Mol Med* 19: 2874-2887, 2015.
61. Song X, Cao G, Jing L, Lin S, Wang X, Zhang J, Wang M, Liu W and Lv C: Analysing the relationship between lncRNA and protein-coding gene and the role of lncRNA as ceRNA in pulmonary fibrosis. *J Cell Mol Med* 18: 991-1003, 2014.
62. Liu XH, Sun M, Nie FQ, Ge YB, Zhang EB, Yin DD, Kong R, Xia R, Lu KH, Li JH, De W, *et al*: Lnc RNA HOTAIR functions as a competing endogenous RNA to regulate HER2 expression by sponging miR-331-3p in gastric cancer. *Mol Cancer* 13: 92, 2014.
63. Penhoet E, Rajkumar T and Rutter WJ: Multiple forms of fructose diphosphate aldolase in mammalian tissues. *Proc Natl Acad Sci USA* 56: 1275-1282, 1966.
64. Asaka M, Kimura T, Meguro T, Kato M, Kudo M, Miyazaki T and Alpert E: Alteration of aldolase isozymes in serum and tissues of patients with cancer and other diseases. *J Clin Lab Anal* 8: 144-148, 1994.
65. Tian YF, Hsieh PL, Lin CY, Sun DP, Sheu MJ, Yang CC, Lin LC, He HL, Solórzano J, Li CF and Chang IW: High expression of Aldolase B confers a poor prognosis for rectal cancer patients receiving neoadjuvant chemoradiotherapy. *J Cancer* 8: 1197-1204, 2017.
66. Wakil SJ and Abu-Elheiga LA: Fatty acid metabolism: Target for metabolic syndrome. *J Lipid Res* 50 (Suppl): S138-S143, 2009.
67. Milgraum LZ, Witters LA, Pasternack GR and Kuhajda FP: Enzymes of the fatty acid synthesis pathway are highly expressed in *in situ* breast carcinoma. *Clin Cancer Res* 3: 2115-2120, 1997.
68. Yahagi N, Shimano H, Hasegawa K, Ohashi K, Matsuzaka T, Najima Y, Sekiya M, Tomita S, Okazaki H, Tamura Y, *et al*: Co-ordinate activation of lipogenic enzymes in hepatocellular carcinoma. *Eur J Cancer* 41: 1316-1322, 2005.
69. Swinnen JV, Brusselmans K and Verhoeven G: Increased lipogenesis in cancer cells: New players, novel targets. *Curr Opin Clin Nutr Metab Care* 9: 358-365, 2006.
70. Jones JE, Esler WP, Patel R, Lanba A, Vera NB, Pfefferkorn JA and Vernochet C: Inhibition of Acetyl-CoA carboxylase 1 (ACC1) and 2 (ACC2) reduces proliferation and De Novo lipogenesis of EGFRvIII Human Glioblastoma cells. *PLoS One* 12: e0169566, 2017.
71. Simioni P, Tormene D, Tognin G, Gavasso S, Bulato C, Iacobelli NP, Finn JD, Spiezia L, Radu C and Arruda VR: X-linked thrombophilia with a mutant factor IX (factor IX Padua). *N Engl J Med* 361: 1671-1675, 2009.
72. Sarkar D and Fisher PB: AEG-1/MTDH/LYRIC: Clinical significance. *Adv Cancer Res* 120: 39-74, 2013.
73. Yoo BK, Emdad L, Su ZZ, Villanueva A, Chiang DY, Mukhopadhyay ND, Mills AS, Waxman S, Fisher RA, Llovet JM, *et al*: Astrocyte elevated gene-1 regulates hepatocellular carcinoma development and progression. *J Clin Invest* 119: 465-477, 2009.
74. Zhu K, Dai Z, Pan Q, Wang Z, Yang GH, Yu L, Ding ZB, Shi GM, Ke AW, Yang XR, *et al*: Metadherin promotes hepatocellular carcinoma metastasis through induction of epithelial-mesenchymal transition. *Clin Cancer Res* 17: 7294-7302, 2011.

