

# SCIENTIFIC REPORTS



OPEN

## Novel Fluorescence Arginine Analogue as a Sensor for Direct Identification and Imaging of Nitric Oxide Synthase-like Enzymes in Plants

Received: 18 May 2016

Accepted: 21 July 2016

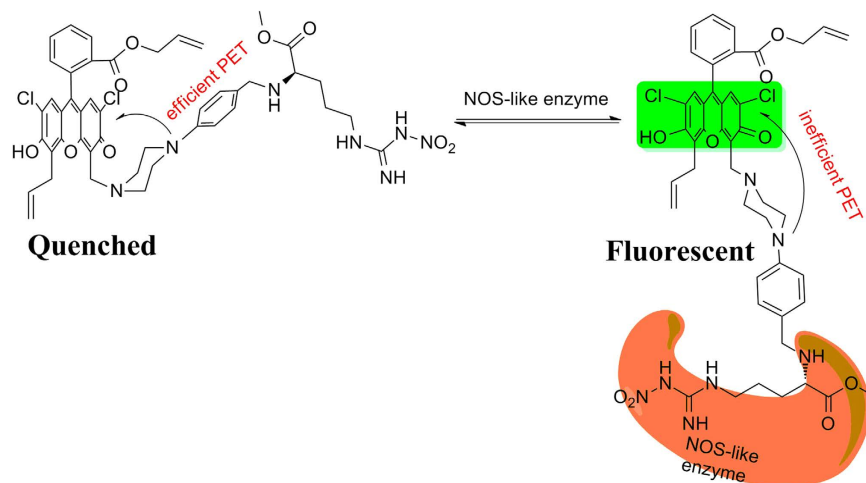
Published: 02 September 2016

Kang Chang<sup>1,\*</sup>, Tongtong Guo<sup>2,\*</sup>, Pengfei Li<sup>1</sup>, Yin Liu<sup>2</sup>, Yufang Xu<sup>1</sup>, Yuda Fang<sup>2</sup> & Xuhong Qian<sup>1</sup>

Nitric oxide synthase like enzyme (NOS-like enzyme), which produces nitric oxide, participates in many biological processes. However it remains unidentified and highly controversial that plants do possess a NOS-like enzyme. In this paper, a novel arginine analogue NP1 was designed and developed for the direct identification and real time tracking of NOS-like enzymes in plant by fluorescence sensing. It could bind NOS-like enzyme efficiently and enter the cell successfully. *In vivo* fluorescence response results directly proved that NOS-like enzymes did exist in tobacco leaf and would be stimulated by pathogen infection, which also provided a useful chemical tool for the study of the function of NOS-like enzyme in plants.

Nitric oxide (NO) is an important endogenous gaseous signalling molecule which universally exists in animals, plants and microbes. In animals, it regulates a variety of processes ranging from the control of blood pressure and smooth muscle relaxation to immune activation and neuronal signaling<sup>1</sup>. In plants, it modulates the growth and development, such as seed germination, stomatal closure, photosynthesis, respiration, cell death and stress resistance etc.<sup>2,3</sup>. When plants are attacked by biotic or abiotic stress, the signal molecule NO is produced, induces the production of other signal molecules including salicylic acid (SA), jasmonic acid (JA), ethylene (ET) and abscisic acid (ABA) etc.<sup>3-5</sup>, which participate in plant immunity including systemic acquired resistance (SAR) and induced systemic resistance (ISR)<sup>6-8</sup>. In animals, nitric oxide (NO) is usually produced by nitric oxide synthases (NOS)<sup>9</sup>, which has three different isoforms including inducible nitric oxide synthase (iNOS), endothelial nitric oxide synthase (eNOS) and neuronal nitric oxide synthase (nNOS)<sup>1</sup>. They catalyze the oxidation of L-arginine leading to L-citrulline and NO<sup>9</sup>. However, the generation of NO in plants is relatively complicated, the origin of NO can be grouped into two pathways: the nitrate reductase (NR) pathway and L-arginine dependent pathway<sup>4</sup>. In L-arginine pathway, plants also require nitric oxide synthase like enzyme (NOS-like enzyme). As one of the most important enzymes producing NO, NOS-like enzyme plays a vital role in plant resistance. It acts at the upstream of signalling pathway, participating in the establishment of SAR and ISR. However, the NOS-like enzyme remains unidentified in plants and the hypothesis that plants do possess a NOS-like enzyme is highly controversial<sup>10,11</sup>. But when the mammalian NOS inhibitors were applied in plants, the arginine-dependent NO synthesis activity was inhibited, and the production of NO significantly decreased, the expression of pathogen related-1 (PR-1) protein was also inhibited<sup>12,13</sup>. These implied the existence of NOS-like enzyme in plants. With increasing interest in exploring the existence of NOS-like enzyme and its function in plants, sensitive and selective detection techniques for monitoring endogenous NOS-like enzyme are urgently desirable, since the enzyme events in plant growth and development are still not fully understood. The current approaches for NOS-like enzyme detection in plants mainly are immunoassays and immunocytochemical analyses<sup>10,14</sup>. However, the

<sup>1</sup>State Key Laboratory of Bioreactor Engineering, Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China. <sup>2</sup>National Key Laboratory of Plant Molecular Genetics; Shanghai Institute of Plant Physiology and Ecology; Shanghai Institutes for Biological Sciences; Chinese Academy of Sciences; Shanghai, China. \*These authors contributed equally to this work. Correspondence and requests for materials should be addressed to Y.X. (email: yfxu@ecust.edu.cn) or Y.F. (email: yfang@sibs.ac.cn) or X.Q. (email: xhqian@ecust.edu.cn)



**Figure 1.** Design idea of NP1 for the recognition of NOS-like enzymes.

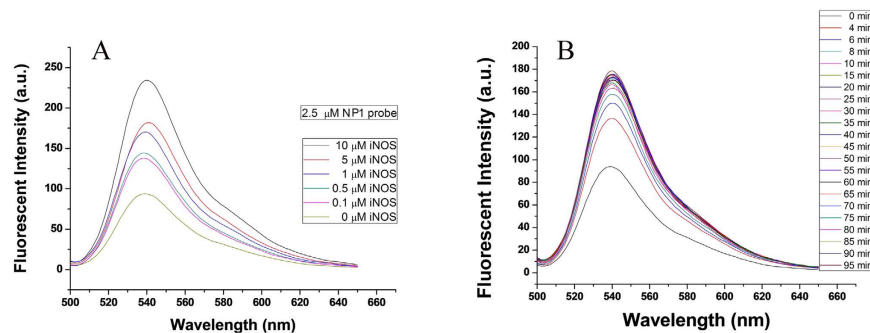
antibodies against mammalian NOS may recognize many other proteins not connected with NOS. These methods were also too time-consuming to be applied to prove the existence of NOS-like enzyme in plants. Therefore, it's of significant importance to find more convenient methods for the direct identification of NOS-like enzyme in plants. Based on these, we choose fluorescence-based assays as a new chemical method for the direct identification and real time tracking of NOS-like enzymes in plant. As we know, fluorescence-based assays could offer convenience, high sensitivity, non-destructiveness, as well as real-time imaging<sup>15</sup>. And fluorescence-based techniques have been widely applied in live cell imaging, medical imaging, drug screening, environmental monitoring and other fields<sup>16–18</sup>. Several small-molecule fluorescent sensors have been applied successfully in zooblast<sup>19–22</sup>. For example, Eric Deprez *et al.* have reported a two-photon fluorescent sensor which was used for the biological imaging of eNOS in living human umbilical vein endothelial cells<sup>23</sup>. However, there were much fewer reports about fluorescent sensors used in plants, one reason is that the existence of the plant cell wall, small molecule fluorescent sensor is difficult to enter the plant cell.

Herein, we expected to develop a fluorescent sensor which could be successfully used for the direct identification and real time tracking of NOS-like enzyme in plants. The designed fluorescent sensor **NP1** consisted of three moieties: a fluorophore, a quencher and a recognition receptor. In order to recognize NOS-like enzyme in plant efficiently, several NOS inhibitors were selected as recognition receptor<sup>24,25</sup>. According to the values of  $IC_{50}$  of different inhibitors (Supplementary Table S1), L-NNA is the most efficient NOS inhibitor, but according to the report<sup>26</sup>, the methyl ester of L-NNA, N-Nitro-L-arginine methyl ester (L-NAME) is more potent, possibly because of increased bioavailability<sup>11</sup>. So, L-NAME was chosen as the binding moiety for the recognition of NOS-like enzyme in plants. Fluorophores with absorption and emission in the near-infrared (NIR) region are tend to be selected in living animals due to interference of biomolecules<sup>27</sup>. By contrast, in plants, fluorophores with emission less than 650 nm are more desirable for *in vivo* imaging in order to avoid the interference of the fluorescence of chlorophyll<sup>28</sup>. So, in order to realize the real time tracking of NOS-like enzyme in plants, a 2',7'-dichlorofluorescein (DCF) fluorophore with emission less than 600 nm was chosen to give the fluorescence signal. Based on these, we illustrated our strategy in Fig. 1. A 2',7'-dichlorofluorescein (DCF) fluorophore coupled with a piperazine ring as an electron donor (quencher) was linked with the recognition receptor, N-Nitro-L-arginine methyl ester (L-NAME), which is a selective NOS inhibitor and widely used in the research of NOS-like enzyme both in mammals and plants<sup>29</sup>. In a free state, **NP1** kept the prior conformation in which the distal nitrogen atom of piperazine ring was proximal to xanthene ring of the fluorophore, causing the efficient quenching of the fluorescence. Upon the receptor of L-NAME combined with NOS-like enzyme, the conformation of piperazine would change from “boat form” to “chair form”, leading to the nitrogen atom away from the xanthene ring, thus photo-induced electron transfer (PET) effect was quenched and the fluorescence would be recovered. With this fluorescent arginine analogue **NP1** in hand, we can realize the direct identification and tracking of NOS-like enzyme in plants by fluorescence sensing using the mechanism of conformational photo-induced electron transfer (PET)<sup>10,14,30</sup>.

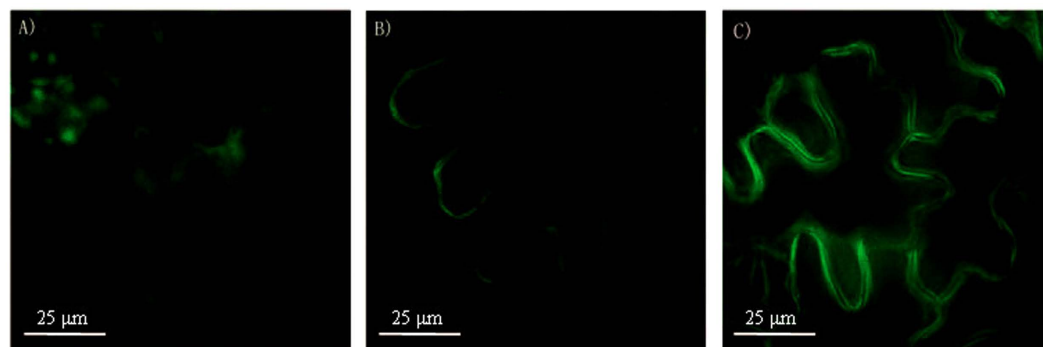
## Results and Discussion

**Synthesis and characterization of NP1.** The synthetic route toward **NP1** is depicted in Supplementary Figure S1. Starting from 2',7'-dichlorofluorescein, it reacted with allyl bromide to generate D-1, then rearranged to D-2 in diphenyl ether at the condition of reflux, then D-3 was formed by the condensation of D-2, formaldehyde and 1-Boc-piperazine, then deprotected by trifluoroacetic acid to prepare D-4, then by the condensation with p-fluorobenzaldehyde to prepare DCF1, finally, **NP1** was synthesized from DCF1 and L-NAME by condensation and reduction. **NP1** was characterized by <sup>1</sup>H NMR, <sup>13</sup>C NMR and HRMS (Supplementary Figures S3–S5). Then the designed fluorescent sensor **NP1** was employed for subsequent studies.

**Fluorescent properties of NP1.** Firstly, in order to determine the excitation and emission wavelength of **NP1**, the spectral characteristics of **NP1** were studied. The pH titration experiments of absorption and fluorescence



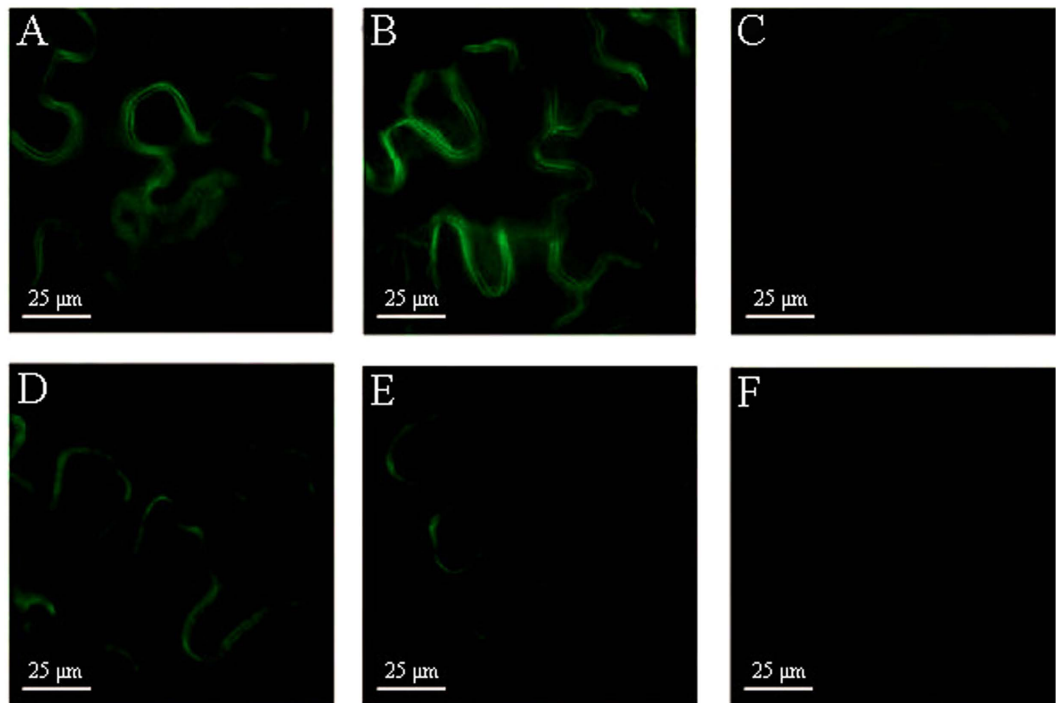
**Figure 2.** (A) The direct fluorescence response of NP1 to iNOS induced from mouse. Fluorescence responses of NP1 (2.5  $\mu\text{M}$ ) with 0–10  $\mu\text{M}$  iNOS in buffer (50 mM HEPES, pH 7.4, containing <1% DMSO) at 37  $^{\circ}\text{C}$ . Fluorescence excitation was provided at 480 nm. After adding iNOS to the NP1 buffer, the fluorescence data were acquired after certain time intervals (100 s) that are indicated in the figure and marked with different colors. (B) Time variation in the fluorescence intensity of NP1 (2.5  $\mu\text{M}$ ) at 538 nm with iNOS (5  $\mu\text{M}$ ) in buffer (50 mM HEPES, pH 7.4, containing <1% DMSO).



**Figure 3.** Fluorescence images of tobacco leaf epidermal cells incubated with NP1 (10  $\mu\text{M}$ ) containing pathogen of *Pst* DC3000 (OD<sub>600</sub> = 0.2) or not by syringe injection for 6 h at room temperature taken by a Delta Vision PersonalDV system (Applied Precision) using an Olympus UPLANAPO water-immersion objective lens (60  $\times$ /1.20 numerical aperture)<sup>36</sup>. Filters used for NP1 were exciter (470/40 nm/nm) and emitter (520/40 nm/nm). (A) Tobacco leaves incubated with *Pst* DC3000. (B) Tobacco leaves incubated with NP1. (C) Tobacco leaves incubated with NP1 and *Pst* DC3000.

were performed in buffer system (50 mM HEPES buffer, pH = 7.4, containing <1% DMSO) at 37  $^{\circ}\text{C}$ . The results showed that the absorption and emission maxima were at 518 nm and 538 nm (Supplementary Figure S2), respectively. In the pH range from 6 to 8, which is consistent with the physiological environment, the emission intensities were quite low and almost unchanged (Supplementary Figure S2). So, NP1 is quite suitable for imaging in plants. Considering the influence of the shorter Stokes Shift of NP1 itself, the subsequent fluorescent experiments were performed at an excitation wavelength of 480 nm. Then, the fluorescence properties of NP1 (2.5  $\mu\text{M}$ ) in the absence and presence of iNOS were determined. Figure 2A depicted elevated fluorescence intensities with increasing the amounts of iNOS (0–10  $\mu\text{M}$ ). This was due to the specific binding of NP1 to iNOS, leading to its conformational change, thereby blocking the PET (Photo electric transfer) quenching. Besides, the dynamic change of fluorescence intensity of NP1 with iNOS was also tested, as shown in Fig. 2B, the fluorescence intensity in reaction of NP1 with iNOS reached the maximum value within approximately 10 min. *In vitro* fluorescence data showed NP1 bound NOS efficiently, and the combination of NP1 to iNOS was fast and firm, proving that NP1 was sensitive and stable enough to be applied. With the encouraging data in hands, the imaging experiments in living tobacco leaves were carried out afterwards.

**Detection of NOS-like enzyme in tobacco leaves.** We thereafter assessed the potential utility of NP1 to monitor NOS-like enzyme in living tobacco leaves. The four-week old tobacco leaves were chosen for the imaging experiments because of their high endogenous expression of NOS-like enzyme during plant defense<sup>31</sup>. Tobacco leaves injected with the free NP1 (10  $\mu\text{M}$ ) showed relatively weak fluorescence emission (Fig. 3B) after incubated in phytotron for 6 hours. In contrast, upon injected of NP1 and *Pst* DC3000 to the tobacco leaves at the same time, due to the overexpression of NOS-like enzyme during pathogen infection, a strong green fluorescence was observed (Fig. 3C) after incubated in phytotron for 6 hours. In order to exclude the interference of *Pst* DC3000, we observed the fluorescence of leaves injected with *Pst* DC3000, and found it also showed a weak fluorescence



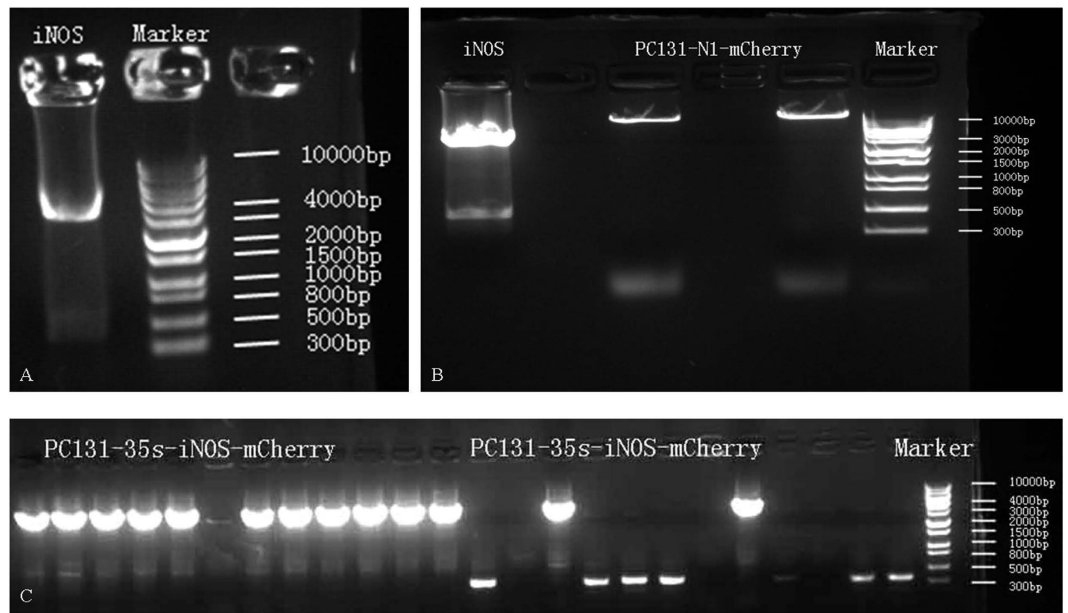
**Figure 4.** Fluorescence images of tobacco leaf cells incubated with NP1 (10  $\mu$ M) containing pathogen of *Pst* DC3000 (OD<sub>600</sub> = 0.2) or not by syringe injection at room temperature taken by a DeltaVision PersonalDV system (Applied Precision) using an Olympus UPLANAPO water-immersion objective lens (60  $\times$ /1.20 numerical aperture). Filters used for GFP were exciter (470/40 nm/nm) and emitter (520/40 nm/nm). (A–C) showed tobacco leaves incubated with NP1 and *Pst* DC3000 for 4 h, 6 h and 8 h, respectively. (D–F) showed tobacco leaves incubated with NP1 for 4 h, 6 h and 8 h, respectively.

emission (Fig. 3A). This suggested that the fluorescence enhancement was indeed due to the expression of endogenous NOS-like enzyme during pathogen infection.

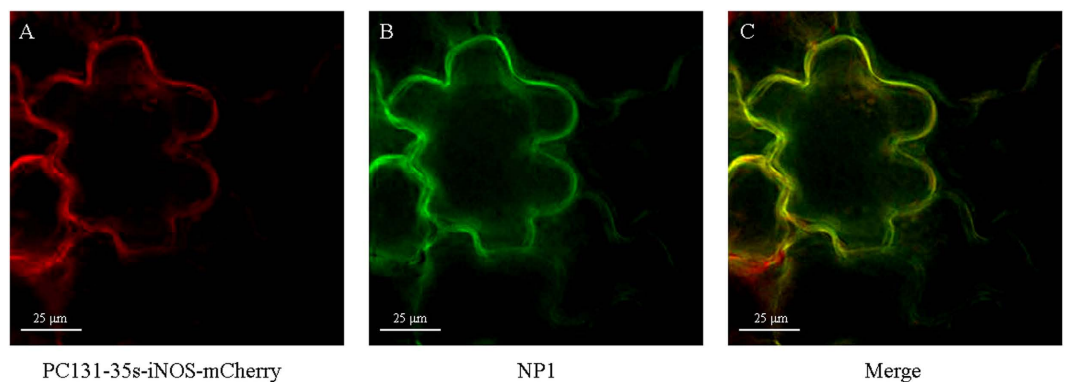
Subsequently, the dynamic change of fluorescence intensity in tobacco leaves was investigated (Fig. 4) for the study of the burst of NOS-like enzyme during pathogen infection. Fluorescence images were taken in 4 to 8 hours respectively after injection of NP1 and *Pst* DC3000. A notable fluorescence change was observed, the fluorescence intensity has become stronger 4 hours (Fig. 4A) after the infection of *Pst* DC3000, an increase from 4 to 6 hours (Fig. 4A,B) and a decrease from 6 to 8 hours (Fig. 4B,C) in fluorescence intensity of infected leaves were observed respectively. The decrease of fluorescence intensity may be due to the diffusion of the molecule itself. According to the data, the fluorescence response began 4 hours after infection, much earlier than the onset of SAR which usually expressed PR proteins 1–3 days after invasion<sup>7</sup>. So, the expression of NOS-like enzyme was at the upstream of plant defense and regulated the signal network by producing NO. Therefore, NP1 could be used as a useful chemical tool for the detection of NOS-like enzyme *in vitro* and in living tobacco leaves, and furthermore, it could also be an indicator to judge the onset of defense response of plants.

Next, in order to further study the subcellular localization of NP1 and NOS-like enzyme, and the specificity of NP1 to NOS-like enzyme, a plasmid PC131-35s-iNOS-mCherry including genes of iNOS and a red fluorescent protein named mCherry was constructed<sup>32,33</sup>. The length of gene of iNOS was 3438 bp, and the experimental results were consistent with it (Fig. 5). Then it was transformed into agrobacterium tumefaciens and injected into leaves of tobacco. Two days later, the plasmid was expressed completely in tobacco leaves. After that, NP1 was injected into tobacco leaves, and half an hour later, the images were taken by a fluorescence microscope (Fig. 6). The plasmid was visualized as red fluorescence from the mCherry channel, showing that the inherent localization of NOS-like enzyme in plants was in the cytoplasm of cells (Fig. 6A), and NP1 was visualized as strong green fluorescence from GFP channel (Fig. 6B). The localization of NP1 was confirmed by the merged images of the plasmid and NP1. It showed a noticeable yellow area (Fig. 6C), indicating that NP1 was localized in the cytoplasm, where it bonded to iNOS and showed a stronger fluorescence emission.

In summary, we have developed a novel fluorescent arginine analogue NP1 as a chemical tool for the direct identification and real time tracking of NOS-like enzyme in living tobacco leaves by fluorescence sensing. To our knowledge, this is the first time to use fluorescence sensing for the detection of NOS-like enzyme in plants. Unlike traditional methods, NP1 was more convenient to be applied and time saving. It enabled fluorescence imaging of endogenous NOS-like enzyme induced by pathogen infection in tobacco leaves, and further proved the existence of NOS-like enzyme in plants. We also found that NP1 was localized in the cytoplasm and the burst of NOS-like enzyme was at the upstream of defense response in tobacco. Thus NP1 shows the potential to be used as a valuable research tool in studying biological roles of NOS-like enzyme in plants. Furthermore, NP1 could also be an



**Figure 5.** (A) The product of iNOS polymerase chain reaction (PCR). (B) The product of digestion of iNOS and PC131-35s-N1-mCherry by *Sal*I and *Spe*I. (C) The product of colony PCR of PC131-35s-iNOS-mCherry.



**Figure 6.** Fluorescence images of tobacco leaf cells incubated with NP1 (10 μM) and PC131-35s-iNOS-mCherry by syringe injection at room temperature taken by a DeltaVision PersonalDV system (Applied Precision) using an Olympus UPLANAPO water-immersion objective lens (60 ×/1.20 numerical aperture). Filters used for images were mCherry and GFP respectively.

indicator to judge the onset of defense response of plants, and in the future, NP1 has the potential to be exploited as a platform for the screening of plant activators<sup>34,35</sup>.

## Methods

**Fluorescent imaging in tobacco leaves.** Four-week-old tobacco was chosen for the following experiments. Test condition: NP1: 100 μM/L, *Pst* DC 3000: OD<sub>600</sub> = 0.2. Then NP1, *Pst* DC3000 and the mix of NP1 and *Pst* DC3000 were injected into three tobacco leaves by injectors to completely infiltrate leaves. Then incubated in phytotron for 4 hours, and taking images every 2 hours by a fluorescence microscope. Fluorescent image of tobacco leaf cells were acquired at room temperature with a DeltaVision PersonalDV system (Applied Precision) consisting of an IX70 inverted microscope (Olympus) equipped with an UPLANAPO water immersion objective lens (60 × 1.20 numerical aperture; Olympus) and a Photometrics (Roper Scientific) CoolSnap ES<sub>2</sub> camera with Applied Precision customizations and drivers. Filters used for GFP were exciter (470/40 nm/nm) and emitter (520/40 nm/nm).

**Expression of PC131-35s-iNOS-mCherry in tobacco leaves.** All enzymes used in this experiment were purchased from TaKaRa Biotechnology Co., Ltd.

**PCR of iNOS.** Because that iNOS is induced from mouse, the cDNA was purchased from Guangzhou FunenGen Co., Ltd. PCR Condition: primer1: 1 μL, primer 2: 1 μL, cDNA: 1 μL, dNTP: 4 μL, H<sub>2</sub>O<sub>2</sub>: 32 μL, primerSTAR HS



DNA polymerase: 1  $\mu$ L, 5  $\times$  primerSTAR buffer: 10  $\mu$ L. After reaction, the result was confirmed by agarose chromatography (Fig. 5A) and then the PCR product was recycled by a kit purchased from Shanghai Generay Biotech Co., Ltd. Then, the product was dissolved in 30  $\mu$ L water for the following experiments.

**Digestion of iNOS and PC131-35s-N1-mCherry.** Condition for iNOS: iNOS PCR product: 30  $\mu$ L, Sal 1 enzyme: 0.75  $\mu$ L, Spe 1 enzyme: 0.75  $\mu$ L, 10  $\times$  H Buffer: 4  $\mu$ L, H<sub>2</sub>O<sub>2</sub>: 4.5  $\mu$ L. Condition for PC131-35s-N1-mCherry (vector): vector: 4  $\mu$ L, Sal 1 enzyme: 0.75  $\mu$ L, Spe 1 enzyme: 0.75  $\mu$ L, 10  $\times$  H Buffer: 2  $\mu$ L, H<sub>2</sub>O<sub>2</sub>: 12.5  $\mu$ L. Putting them in incubator of 37  $^{\circ}$ C for 2 hours. After that, the mix were separated by agarose chromatography (Fig. 5B) and the right bands were cut off and recycled by corresponding kit purchased from Shanghai Generay Biotech Co., Ltd.

**Connection of iNOS with vector.** Condition: recycled iNOS: 6  $\mu$ L, recycled vector (PC131-35s-N1-mCherry): 2  $\mu$ L, T4 ligase: 1  $\mu$ L, T4 Buffer: 1  $\mu$ L. Mixing thoroughly, then placing it in incubator of 22  $^{\circ}$ C for 4 hours. After that, the connection product was put into coli, ice-bath 30 minutes, then stimulated at water bath of 42  $^{\circ}$ C for 90 seconds, after that, adding 800  $\mu$ L LB culture medium, and putting it in 37  $^{\circ}$ C swing bed for 1 hour. Then placing the mix on the culture dish, and cultivated at 37  $^{\circ}$ C overnight. The connection efficacy was shown in Fig. 5C.

**Plasmid extraction.** Put the bacterial colony into 4 mL LB culture medium with kanamycin, cultivated in 37  $^{\circ}$ C swing bed overnight. Then extract the plasmid according the corresponding kit purchased from Shanghai Generay Biotech Co., Ltd. And at last, the plasmid was confirmed by sequencing.

**Transformation.** 0.6  $\mu$ L plasmid was added into agrobacterium, mixed thoroughly, stimulated by electric shock. Then, the mix was added into 700  $\mu$ L LB culture medium, cultivated in 30  $^{\circ}$ C swing bed for 1 hour. Then the mix was placed on the culture dish, and cultivated at 30  $^{\circ}$ C for 2 days. Then, the bacterial colony was collected, and suspended in 1 mL water and injected into tobacco leaves with an injector to completely infiltrate leaves. Two days later, the protein was completely expressed in tobacco leaves.

## References

- Mukherjee, P., Cinelli, M. A., Kang, S. & Silverman, R. B. Development of nitric oxide synthase inhibitors for neurodegeneration and neuropathic pain. *Chem Soc Rev.* **43**, 6814–6838, doi: 10.1039/c3cs60467e (2014).
- Misra, A. N., Misra, M. & Singh, R. Nitric oxide biochemistry, mode of action and signaling in plants. *J Med Plants Res.* **4**, 2729–2739 (2010).
- Shi, H. T. *et al.* *In vivo* role of nitric oxide in plant response to abiotic and biotic stress. *Plant signaling & behavior* **7**, 438–440, doi: 10.4161/psb.19219 (2012).
- Besson-Bard, A., Pugin, A. & Wendehenne, D. New insights into nitric oxide signaling in plants. *Annual Review of Plant Biology* **59**, 21–39, doi: 10.1146/annurev.arplant.59.032607.092830 (2008).
- Hong, J. K. *et al.* Nitric oxide function and signalling in plant disease resistance. *J Exp Bot.* **59**, 147–154, doi: 10.1093/jxb/erm244 (2008).
- Pieterse, C. M., Leon-Reyes, A., Van der Ent, S. & Van Wees, S. C. Networking by small-molecule hormones in plant immunity. *Nature chemical biology* **5**, 308–316, doi: 10.1038/nchembio.164 (2009).
- Durrant, W. E. & Dong, X. Systemic acquired resistance. *Annu Rev Phytopathol.* **42**, 185–209, doi: 10.1146/annurev.phyto.42.040803.140421 (2004).
- Pieterse, C. M. J. *et al.* Induced Systemic Resistance by Beneficial Microbes. *Annu Rev Phytopathol.* **52**, 347–375, doi: 10.1146/annurev-phyto-082712-102340 (2014).
- Jachymova, M. *et al.* Recruitment of governing elements for electron transfer in the nitric oxide synthase family. *P Natl Acad Sci USA* **102**, 15833–15838, doi: 10.1073/pnas.0506522102 (2005).
- Guo, F. Q., Okamoto, M. & Crawford, N. M. Identification of a plant nitric oxide synthase gene involved in hormonal signaling. *Science* **302**, 100–103 (2003).
- Eckardt, N. A. A Functional Nitric Oxide Synthase in *Ostreococcus tauri*. *The Plant Cell* **22**, 3507, doi: 10.1105/tpc.110.221112 (2010).
- Klessig, D. F. *et al.* Nitric oxide and salicylic acid signaling in plant defense. *Proc Natl Acad Sci USA* **97**, 8849–8855 (2000).
- Durner, J., Wendehenne, D. & Klessig, D. F. Defense gene induction in tobacco by nitric oxide, cyclic GMP, and cyclic ADP-ribose. *Proceedings of the National Academy of Sciences* **95**, 10328–10333, doi: 10.1073/pnas.95.17.10328 (1998).
- Stohr, C. & Ullrich, W. R. Generation and possible roles of NO in plant roots and their apoplastic space. *J Exp Bot.* **53**, 2293–2303, doi: 10.1093/jxb/Er110 (2002).
- Zhang, L. *et al.* A highly selective and sensitive near-infrared fluorescent probe for imaging of hydrogen sulphide in living cells and mice. *Scientific Reports* **6**, 18868, doi: 10.1038/srep18868 <http://www.nature.com/articles/srep18868#supplementary-information> (2016).
- Vendrell, M., Zhai, D., Er, J. C. & Chang, Y.-T. Combinatorial Strategies in Fluorescent Probe Development. *Chemical Reviews*, doi: 10.1021/cr200355j (2012).
- Razgulina, A., Ma, N. & Rao, J. Strategies for *in vivo* imaging of enzyme activity: an overview and recent advances. *Chem Soc Rev.* **40**, 4186–4216, doi: 10.1039/c1cs15035a (2011).
- Qian, X. *et al.* “Alive” dyes as fluorescent sensors: fluorophore, mechanism, receptor and images in living cells. *Chem Commun.* **46**, 6418–6436, doi: 10.1039/c0cc00686f (2010).
- Yu, F. B. A. *et al.* A Near-IR Reversible Fluorescent Probe Modulated by Selenium for Monitoring Peroxynitrite and Imaging in Living Cells. *J Am Chem Soc.* **133**, 11030–11033, doi: 10.1021/ja202582x (2011).
- Yadav, A. K. *et al.* Fluorescence-Quenched Substrates for Live Cell Imaging of Human Glucocerebrosidase Activity. *J Am Chem Soc.* doi: 10.1021/ja5106738 (2015).
- Chen, W. *et al.* The Development of Fluorescent Probes for Visualizing Intracellular Hydrogen Polysulfides. *Angewandte Chemie International Edition* **54**, 13961–13965, doi: 10.1002/anie.201506887 (2015).
- Kojima, R. *et al.* Development of a Sensitive Bioluminescent Probe for Imaging Highly Reactive Oxygen Species in Living Rats. *Angewandte Chemie International Edition* n/a–n/a, doi: 10.1002/anie.201507530 (2015).
- Li, Y. *et al.* Rational design of a fluorescent NADPH derivative imaging constitutive nitric-oxide synthases upon two-photon excitation. *Proc Natl Acad Sci USA* **109**, 12526–12531, doi: 10.1073/pnas.1205645109 (2012).
- Li, H. & Poulos, T. L. Structure–function studies on nitric oxide synthases. *Journal of inorganic biochemistry* **99**, 293–305, doi: <http://dx.doi.org/10.1016/j.jinorgbio.2004.10.016> (2005).

25. Alderton, W. K., Cooper, C. E. & Knowles, R. G. Nitric oxide synthases: structure, function and inhibition. *Biochemical Journal* **357**, 593–615 (2001).
26. Marletta, M. A. Approaches toward Selective Inhibition of Nitric Oxide Synthase. *J Med Chem.* **37**, 1899–1907, doi: 10.1021/jm00039a001 (1994).
27. Yuan, L., Lin, W., Zheng, K., He, L. & Huang, W. Far-red to near infrared analyte-responsive fluorescent probes based on organic fluorophore platforms for fluorescence imaging. *Chem Soc Rev.* **42**, 622–661, doi: 10.1039/c2cs35313j (2013).
28. Karapetyan, N. V. *et al.* Long-wavelength chlorophylls in photosystem I of cyanobacteria: Origin, localization, and functions. *Biochemistry (Moscow)* **79**, 213–220, doi: 10.1134/s0006297914030067 (2014).
29. Yargıçoğlu, P. *et al.* Effects of N-nitro l-arginine methyl ester (l-NAME), a potent nitric oxide synthase inhibitor, on visual evoked potentials of rats exposed to different experimental stress models. *Acta Physiologica Scandinavica* **180**, 307–316, doi: 10.1111/j.0001-6772.2003.01254.x (2004).
30. Zhang, S. *et al.* A highly selective space-folded photo-induced electron transfer fluorescent probe for carbonic anhydrase isozymes IX and its applications for biological imaging. *Chem Commun (Camb)* **47**, 8301–8303, doi: 10.1039/c1cc12386f (2011).
31. Furfine, E. S., Harmon, M. F., Paith, J. E. & Garvey, E. P. Selective-Inhibition of Constitutive Nitric-Oxide Synthase by L-N(G)-Nitroarginine. *Biochemistry* **32**, 8512–8517, doi: 10.1021/Bi00084a017 (1993).
32. Liu, Y. & Fang, Y. Nucleolus-tethering system (NoTS). *Nucleus* **5**, 293–297, doi: 10.4161/nucl.29837 (2014).
33. Liu, Y., Liu, Q., Yan, Q., Shi, L. & Fang, Y. Nucleolus-tethering system (NoTS) reveals that assembly of photobodies follows a self-organization model. *Mol Biol Cell* **25**, 1366–1373, doi: 10.1091/mbc.E13-09-0527 (2014).
34. Du, Q., Zhu, W., Zhao, Z., Qian, X. & Xu, Y. Novel Benzo-1,2,3-thiadiazole-7-carboxylate Derivatives As Plant Activators and the Development of Their Agricultural Applications. *J Agr Food Chem.* **60**, 346–353, doi: 10.1021/jf203974p (2011).
35. Gozzo, F. Systemic Acquired Resistance in Crop Protection: From Nature to a Chemical Approach. *J Agr Food Chem.* **51**, 4487–4503, doi: 10.1021/jf030025s (2003).
36. Fang, Y. & Spector, D. L. Live cell imaging of plants. *Cold Spring Harbor protocols* **2010**, pdb top68, doi: 10.1101/pdb.top68 (2010).

## Acknowledgements

This work is financially supported by the National Natural Science Foundation of China (Grants 21236002), the National High Technology Research and Development Program of China (863 Program 2011AA10A207) and the National Key Laboratory of Plant Molecular Genetics.

## Author Contributions

Y.X., X.Q. and Y.F. conceived the idea and directed the work. K.C. and P.L. performed the synthesis. P.L. performed the *in vitro* fluorescence tests. K.C., T.G. and Y.L. performed the *in vivo* imaging. All authors reviewed the manuscript.

## Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

**Competing financial interests:** The authors declare no competing financial interests.

**How to cite this article:** Chang, K. *et al.* Novel Fluorescence Arginine Analogue as a Sensor for Direct Identification and Imaging of Nitric Oxide Synthase-like Enzymes in Plants. *Sci. Rep.* **6**, 32630; doi: 10.1038/srep32630 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016