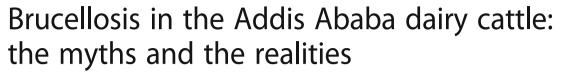
RESEARCH ARTICLE

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Abstract

Background: Bovine brucellosis is considered as an important disease among livestock and people in sub-Saharan African countries including Ethiopia. A cross-sectional study was conducted from November 2016 to May 2017 to estimate the prevalence and associated risk factors, and to assess knowledge-attitude and practices (KAP) of farm workers about bovine brucellosis in Addis Ababa dairy farms.

Results: A total of 1550 cattle from 127 dairy farms were serially tested using the Rose Bengal Plate Test (RBPT), Competitive Enzyme-Linked Immunosorbant Assay (c-ELISA) and Complement Fixation Test (CFT). Forty-three (2. 77%) of the collected sera were positive by the RBPT and only one of these was positive by c-ELISA (0.06%) and none was positive by CFT. The knowledge of farm workers towards the disease was very low and risk factors associated with *Brucella* infection were apparent in the study area.

Conclusion: Seropositivity for *Brucella* spp. was found in only a very small percentage by c-ELISA test, although risk factors for transmitting *Brucella* infection were present. The results suggest that bovine brucellosis is currently not a generalized problem in dairy cattle of Addis Ababa. Since this favorable disease situation is not the result of informed policy, there is no guarantee that it will continue unchanged. Setting clear policy in control of the disease and implementing "One Health" are the most constructive approaches recommended.

Keywords: Addis Ababa, Brucella, Dairy cattle, Ethiopia, Risk factors, Seroprevalence

Background

Ethiopia has one of the largest cattle populations in Africa [1] despite gaining minimum return from this resource as a result of various technical and non-technical factors, including infectious diseases. Bovine brucellosis is one of the infectious diseases hampering productivity of cattle and has been reported from several parts of the country [2] Bovine brucellosis is a zoonotic disease with economic and public health impact, particularly for human and animal populations in developing countries that rely mainly on livestock production [3]. The disease can generally cause significant loss of productivity through abortion, stillbirth, low herd fertility and low milk production [4].

Bovine brucellosis is considered to be predominantly caused by Brucella abortus; and, to a much lesser extent by B. melitensis, where cattle are kept together with infected goat or sheep. It is characterized clinically by abortion at first gestation, retained fetal membrane (RFM), metritis, orchitis and epididymitis [5, 6]. Sources of infection for the transmission of the bovine brucellosis are aborted fetuses, retained fetal membranes, and vaginal discharges and milk from infected animals. Direct contact with an aborting cow and the aborted fetus and indirect contact with contaminated fomites are the most common means of transmission of the disease in cattle. Ingestion of contaminated feed, fodder, water and grazing on contaminated pasture, may also play a secondary role in disease transmission [5, 7]. The disease has been eradicated from many developed countries; it however, remains a major public and animal health problem in many developing countries, where rural income relies mainly on livestock and dairy products [8]. Brucellosis in human often originates from domestic

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animal reservoir and associated with various risk factors and behavioral practices such as consumption of raw milk and milk products and close contact with infected animals [9, 10].

The risk factors that influence the spread and maintenance of brucellosis are related to management systems, the genetic content of susceptible animal population, biology of agents causing the disease and environmental factors. These include the size and composition of the herd, age of the animals, frequent contact between infected and susceptible herds, poor farm biosecurity and climate change [5, 11].

In Ethiopia there is no documented information on how and when bovine brucellosis was introduced and established. However, in the last two decades several serological surveys have showed that it is endemic and widespread [2, 12–26]. These studies in animals and humans were largely confined to serological surveys and commonly targeted cattle, occasionally sheep and goats, and rarely camels. So far, there was only one attempt to identify *Brucella* species in the country [27]; the distribution and proportion of their natural hosts was also not studied exhaustively [19]. According to [28], *Brucella* seroprevalence in dairy cattle of Ethiopia revealed highest prevalence in central Ethiopia followed by southern part whereas lowest prevalence was revealed in western part of the country.

Market oriented new dairy industries are emerging in Ethiopia so as to contribute hugely towards filling the gap between an increasing national demand and supply for milk and milk products. Cow dairy development roadmap of the country is aimed at increasing the productivity of indigenous cattle through improvements in genetics, health and feeding to satisfy consumption demand and start export of cow milk and milk products. It is important that public and private dairy industries and cooperatives require up to date and consistent scientific data on bovine brucellosis. This would in turn assist in developing baseline information to establish nationwide bovine brucellosis intervention policies aimed at controlling and eradicating the disease.

Thus, this study was designed to estimate the seroprevalence of brucellosis in dairy cattle, to identify potential risk factors and assess knowledge, attitude and practices (KAP) of the farm workers towards the disease.

Results

Knowledge-attitude and practices (KAP) of the farm workers about brucellosis

Demographic characteristics

Out of 127 surveyed farms, 130 farm workers in 59 farms were willing to participate in the KAP study. Of the 130 farm workers responsible for the management of the farm, the majority (88%) was male, and more than

50% were between ages 25 to 60 years. Half of the participants were not married (51.5%) and most of them had family size below 10 people. Eighty percent of the respondents had attended only primary school and more than 5 % were illiterate (Table 1).

Risk factors

Most of the study participants reported several risk factors for acquiring bovine brucellosis. The majority of participants, 96% of farm workers in small scale, 100% in medium size and 92.6% in large herd sized farms were not aware of bovine brucellosis. Most respondents, 83.6% in small scale, 60% in medium size and 81.5% in large farms disposed of dead fetus/after birth to open dump in the environment and more than 5 % of participants in small herd sized farms fed aborted materials to dogs. Almost all participants in small scale and medium sized farms and 77.8% in large-scale farms practiced assisted parturition without wearing protective gloves or masks. More than 60 and 80% percent of farm workers in all farm sizes consume raw milk and meat, respectively (Table 2).

Farm characteristics

Of the 127 farms, 103, 17 and 7 were small, medium and large herd sized farms, respectively. Of the farms assessed by a questionnaire survey, more than 70% of medium and large farms, as well as 42.7% of small sized farms, had reproductive problems (abortion, stillbirth, retained fetal membrane and repeat breeding) in their farms. The majority of farms were using artificial

Table 1 Demographic characteristics of farm workers in the study area (n = 130)

| stady area (** 156) | | | | | |
|-----------------------------|-------------------|--------|---------|--|--|
| Demographic characteristics | Category | number | Percent | | |
| Gender | Female | 16 | 12.3 | | |
| | Male | 114 | 87.7 | | |
| Age | 12-24 | 54 | 41.5 | | |
| | 25–60 | 72 | 55.4 | | |
| | > 60 | 4 | 3.1 | | |
| Marital status | Single | 67 | 51.5 | | |
| | Married | 63 | 48.5 | | |
| Level of education | Illiterate | 9 | 6.9 | | |
| | Primary | 104 | 80 | | |
| | Secondary | 13 | 10 | | |
| | Technical/Diploma | 3 | 2.3 | | |
| | Degree | 1 | 0.8 | | |
| No of people in | 1–5 | 61 | 46.9 | | |
| the household ^a | 6–10 | 67 | 51.5 | | |
| | > 10 | 2 | 1.6 | | |

^aHousehold defined as family members regularly living together and sharing meals. *n* number

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Table 2 Knowledge-attitudes and practices of farm workers about *Brucella* infection in the study area

| Variables | Proportion of respondents (n) Herd size | | | |
|-----------------------|--|--------------------------|-------------------------|--|
| | | | | |
| | Small (<i>n</i> = 73) n (%) | Medium (n = 30) n (%) | Large (n = 27) n (%) | |
| Awareness about br | ucellosis | | | |
| Yes | 3 (4) | 0 (0) | 2 (7.4) | |
| No | 70 (96) | 30(100) | 25 (92.6) | |
| Dead fetus/After birt | :h disposal | | | |
| Burning/Burying | 7 (9.6) | 12 (40) | 5 (18.5) | |
| Open dump | 61 (83.6) | 18 (60) | 22 (81.5) | |
| Give to dog | 5 (6.8) | 0 (0) | 0 (0) | |
| Assist parturition | | | | |
| Yes | 72 (98.6) | 30(100) | 21 (77.8) | |
| No | 1 (1.4) | 0 (0) | 6 (22.2) | |
| Consume raw milk | | | | |
| Yes | 50 (68.5) | 18 (60) | 20 (74) | |
| No | 23 (31.5) | 12 (40) | 7 (26) | |
| Consume raw meat | | | | |
| Yes | 60 (82.2) | 25(83.3) | 24 (88.9) | |
| No | 13 (17.8) | 5 (16.7) | 3 (11.1.) | |

n number

insemination (AI) for breeding purposes and 71.4% of large sized farms raised their own replacement animals whereas most of small and medium sized farms used both (raised their own stock and purchase). The practices of provision of separate pens for parturition and aborted animals were 28.6 and 14.3% in large sized farms, respectively, whereas there were no such practice in small and medium sized farms and almost all farms used flushing with tap water to clean pens after parturition. The majority (85.7%) of the large herd sized farms, and more than 40% of medium and small-scale farms, used separate feed and water supply for each animal. Reproductive problems and age were prominent culling criteria in all farms and the majority of farms (> 70%) in the study area did not report frequent contact of dairy animals with other species (sheep and goat) (Table 3).

Seroprevalence of bovine brucellosis in dairy cattle

A total of 1550 dairy cattle were tested with RBT and 43 (2.77%) of them were positive in this test. The RBT positive sera samples were further tested using c-ELISA and CFT. Only one animal was confirmed seropositive for bovine brucellosis in the study area based on c-ELISA and no sero reactor animal was found by CFT. In addition to RBT positive sera samples, equal number of randomly selected RBT negative sera were shipped to APHA, Weybridge, UK and further tested using RBT and C-ELISA.

However, the result was the same. The seroprevalence of bovine brucellosis in Addis Ababa dairy farms was thus 0.06% (1/1550) based on c-ELISA test. The herd level seroprevalence of brucellosis based on c-ELISA was 0.8% (1/127). The prevalence of antibodies to *Brucella* spp. in small, medium and large sized cattle farms was 0, 5.8% (1/17) and 0%, respectively (Table 4).

Discussion

Improvement of knowledge-attitudes and practices among urban dairy farm workers could have a significant impact on the reduction of many zoonotic infections, including brucellosis. The results of the KAP study show that the majority of farm workers in the studied dairy farms were not aware of bovine brucellosis (96.1%). Farm workers with a primary and lower level of education were less likely to have heard of brucellosis when compared to those with secondary and higher level of education. Animal attendants with a primary and lower level of education are hence likely at a higher risk of exposure to the disease. Similar findings were reported by a study conducted in Tajikistan [29]. Low awareness of the disease in the study area might be explained by the low prevalence of brucellosis in dairy cows. The majority (88%) of farm workers were male. This could be due to the fact that farm works in urban and peri-urban intensive diary is labor demanding, as a result of which most farm owners prefer to employ male farm workers.

Practices posing a high risk of *Brucella* transmission are very common: most participants reported assisting in animal parturition, disposing aborted fetuses/after birth in open environment without protective gloves or masks and consumption of raw meat and milk. The reason could be poor knowledge of the disease and risks of transmission but also lack of resources used for personal protection such as gloves, aprons and antiseptics. Creation of awareness of the farm owners and attendants is important to control brucellosis in the area even though the prevalence in animals was low in this serological survey.

In the present study, bovine brucellosis at individual animal level was 0.06% (1/1550) and herd level prevalence was 0.8% (1/127) using c-ELISA whereas no seropositive animal was found on CFT in dairy farms of Addis Ababa. This observation is consistent with previous reports made by [30] in Eastern and Western Showa zones of central Ethiopia using Rose Bengal Plate test (RBPT), serum agglutination test (SAT) and complement fixation test (CFT) (n = 564). This report is also consistent with [31], who reported no positive reactors in intensive dairy farms of the Addis Ababa area (n = 747). Similarly, a study by [23] could not find positive reactors in Adama, central Ethiopia (n = 52) and northern Ethiopia (Mekele and Gondar) (n = 252). A study

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Table 3 Summary of characteristics of dairy farms in the study area

| Variables | Herd size | | | | | |
|--------------------------------------|--|--|--------------------------------|--|--|--|
| | Small (<i>n</i> = 103) Frequency (%) | Medium (<i>n</i> = 17) Frequency (%) | Large (n = 7) Frequency (%) | | | |
| Reproductive problems | | | | | | |
| Yes | 44 (42.7) | 12 (70.6) | 5 (71.4) | | | |
| No | 59 (57.3) | 5 (29.4) | 2 (28.6) | | | |
| Service type | | | | | | |
| Al | 71 (69) | 9 (53) | 5 (71.4) | | | |
| Bull Both | 9 (8.7) 23 (22.3) | 4 (23.5) 4 (23.5) | 0 (0) 2 (28.6) | | | |
| Replacement stock | | | | | | |
| Raised own | 31 (30.1) | 8 (47) | 5 (71.4) | | | |
| Purchased Both | 5 (4.9) 67 (65) | 1 (6) 8 (47) | 0 (0) 2 (28.6) | | | |
| Parturition pen | | | | | | |
| Yes | 0 (0) | 0 (0) | 2 (28.6) | | | |
| No | 103 (100) | 17 (100) | 5 (71.4) | | | |
| Cleaning pen after parturition | | | | | | |
| Flushed with tap water | 101 (97) | 17 (100) | 7 (100) | | | |
| Flushed with water and disinfect | 2 (3) | 0(0) | 0 (0) | | | |
| Separate pen for aborted cow | | | | | | |
| Yes | 1 (1) | 0 (0) | 1 (14.3) | | | |
| No | 102 (99) | 17 (100) | 6 (85.7) | | | |
| Feed and water supply | | | | | | |
| Own | 50 (48.5) | 7 (41.2) | 6 (85.7) | | | |
| Communal | 5 (4.9) | 4 (23.5) | 0 (0) | | | |
| Both | 48 (46.6) | 6 (35.3) | 1 (14.3) | | | |
| Culling criteria | | | | | | |
| Reproductive problem | 57 (55.3) | 6 (35.3) | 4 (57.1) | | | |
| Logistics | 7 (6.8) | 3 (17.7) | 0 (0) | | | |
| Age | 39 (37.9) | 8 (47) | 3 (42.9) | | | |
| Contact with other spp. ^a | | | | | | |
| Yes | 12 (11.6) | 4 (23.5) | 2 (28.6) | | | |
| No | 91 (88.4) | 13 (76.5) | 5 (71.4) | | | |

Contact with other spp. ^a Sheep and goat

Table 4 Individual animal level and herd level prevalence of bovine brucellosis in dairy farms of Addis Ababa

| Farm type | Individual farm | | Herd level | | | | | |
|--------------|-----------------|------------------------|--|------------------------------------|-----------|------------------------------------|--|------------------------------------|
| | No tested | RBT Positive nª (%) | C-ELISA Positive n ^a (%) | CFT Positive n ^a (%) | No Tested | RBT Positive n ^a (%) | C-ELISA Positive n ^a (%) | CFT Positive n ^a (%) |
| Small | 821 | 5(0.6) | 0(0) | 0(0) | 103 | 5(4.8) | 0(0) | 0(0) |
| Medium | 363 | 9(2.5) | 1(0.27) | 0(0) | 17 | 6(35.3) | 1(5.8) | 0(0) |
| Large | 366 | 29(7.9) | 0(0) | 0(0) | 7 | 3(42.9) | 0(0) | 0(0) |
| Total | 1550 | 43(2.77) | 1(0.06) | 0(0) | 127 | 14(11) | 1(0.8) | 0(0) |

n^a number positive

conducted in Debrebirhan and Ambo towns by [32] reported that there was only one sero-reactor animal to Brucella infection using CFT(n = 415).

In contrast, there are reports of a higher prevalence of antibodies to *Brucella* spp. in Addis Ababa dairy farms, 1.5% [11] and 2.2% [23]. A similar study by [26] reported 1.4% in Asella and Bishoftu towns using the card test (CT), RBPT, indirect Enzyme-Linked Immuno Sorbant Assay (i-ELISA) and Complement Fixation Test (CFT). In Ethiopia, brucellosis in animals has been reported from different localities of the country; with prevalence ranging from 0.4 to 8% particularly associated with cattle in both intensive and extensive management systems [2, 11–15, 18–25]. A high seroprevalence of brucellosis (38%) in cattle in western Ethiopia has been reported [33], while most of the studies suggested a low seroprevalence (below 5%) in cattle.

In the present study only one sero reactor animal to Brucella spp. was found in cattle populations of Addis Ababa dairy farms. According to the individual animal record in the farm, this seroreactor animal in the farm was purchased from outside the capital and had late abortion history at first calving. This is remarkable, as bovine brucellosis is considered the world's most common bacterial zoonosis [34] and listed among top five zoonotic diseases in Ethiopia [35]. The hypothesis that brucellosis is endemic in the investigated dairy farms of Addis Ababa could thereby not be confirmed in the present study. However, the presence of one or more positive reactors in the herd is a reliable predictor of the presence of infection [5]. Seropositivity for Brucella spp. was found in only a very small percentage by c-ELISA test, although risk factors for obtaining *Brucella* infection such as, age variety, origin of animals, different level of parity, history of abortion, and herd size composition were present as revealed by farm characteristics analysis.

In epidemiological studies, the use of two or more tests applied serially is recommended to maximize the accuracy of test results. RBPT is highly sensitive test and c-ELISA and CFT are highly specific and also sensitive and usually used as confirmatory tests [36]. The combination of these tests in this study could therefore maximize the accuracy of the findings. False positive serological reactors in RBT are due to cross-reactions with Smooth Lipopolysaccharide (S-LPS) antigens of other bacteria. As there has never been history of vaccination, seropositivity in this case is due to natural infection.

The difference in test results of C-ELISA and CFT is due to the variation in sensitivity of the tests. The C-ELISA test is more sensitive as compared to CFT for the diagnosis of brucellosis even though both tests have similar specificities of 99.9% [36]. Moreover, CFT is prone to prozone effect (low dilutions of some titrated

sera from infected animals do not fix complement) that could lead to a false negative result [37, 38].

The very low prevalence could be explained by the cross sectional study design if informal culling practices suggested by [17] had been instituted. These include culling of cows with abortion history of at least two times and above for any reason and removing seropositive reactors from the herd for economic reasons. The other possible explanation might be absence of infectious foci, such as *Brucella*-infected dairy farms or ranches in the surrounding areas, which could spread the disease among contact herds. Movement of infected animals to susceptible herds is a common route of transmission [5]. The random selection used in our study design should have detected clustered infection if it were common [39].

Conclusions

Urban and peri-urban dairy farming offers an important opportunity to improve the livelihood of people in low-income countries. There was only one seropositive animal found for bovine brucellosis in the present study by the c-ELISA test. This study showed poor knowledge of brucellosis and abundant high-risk behaviors among the farm workers. Poor knowledge and high-risk practices strengthens the logic for including health education as part of control programs. At present, there is no officially coordinated program for control of bovine brucellosis in Ethiopia. The disease is unlikely to be a significant limiter of dairy production in the Addis Ababa area due to the low prevalence, but it may be present in other animal types. Since this favorable disease situation is not the result of informed policy, there is no guarantee that it will continue unchanged.

The current study warrants the need for constant surveillance program in case the prevalence rates do change. Implementing a "test and slaughter" program in the zero grazing system to eliminate the existing low risk of brucellosis could avert the cost-related limitation of brucellosis control. Modernization of husbandry systems and testing of new animals before introducing to dairy farms should be encouraged. A multi-sectorial framework should be promoted involving all stakeholders working in public and animal health in the context of a "One Health" approach. Since the current study was limited to cross sectional study design, future studies in the dairy farms should follow longitudinal study types to ascertain actual burden of the disease in the study area.

Methods

Description of the study area

The study was conducted in Addis Ababa dairy farms. Addis Ababa, the capital city of Ethiopia, lies at an elevation of 2300 m (7,500 ft) above sea level and is featured by a grassland biome. It is geographically located at 9°1′

48"N latitude and 38°44'24"E longitude. It has a typical highland climate with temperature ranging from $11\,^{\circ}\text{C}$ - $24\,^{\circ}\text{C}$. Addis Ababa has a mean annual rainfall of 1300 mm with bimodal distribution. The city is divided into 10 boroughs (Fig. 1), called sub cities, and 99 wards (kebeles) [40].

Dairy cattle production systems in Ethiopia are classified into commercial dairy systems, urban and peri-urban smallholder dairy, rural smallholder (mixed crop and livestock production), and pastoral and agro pastoral [23]. The dairy systems hold mainly exotic breeds or crosses with the local zebu breeds, while the rural husbandry systems stock mainly zebus. This study focused on urban and peri-urban smallholder farms, which produce milk for home use and sale, and commercial dairy systems, which are producing milk as a full-time business. These dairies constitute the main dairy source for the capital and produce milk for sale [26]. Within these systems dairy farms were classified based on size of the herd and herd management into large scale farms, with more than 50 animals, medium

scale farms (20 to 50 animals) and smallholder farms (< 20 animals).

Variables collected

Explanatory variables that were hypothesized to be risk factors for the disease were assessed at both individual animal and farm level. Information related to herd structure was extracted from individual herd records and when this was not the case from farm owners or managers interview using a pre tested structured questionnaires. Herd level parameters studied include herd size as described above, the presence of reproductive problems such as abortion, retained fetal membranes and still birth in the farm, separate calving pen, brucellosis testing in the farm, frequent contact with animals in other herds and species, which were categorized as yes or no variables. The major reasons for culling were coded either as reproductive problems, old age or logistics. Breeding strategy was artificial insemination, bull or both. The afterbirth (aborted fetuses and fetal membranes) disposal method was also categorized into burying, open dump or feeding to dogs.

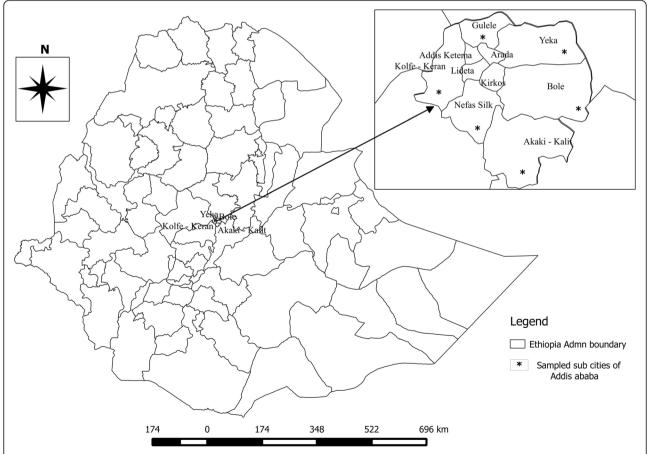


Fig. 1 Map of Addis Ababa showing studied/sampled sub cities. The map depicted in Fig. 1 is our own developed from Ethiopian shape files using QGIS software

The feeding and water supply strategies were classified into communal and own.

Individual animals were categorized as young 6–17 months or adult (≥17 months), breed as Holstein-Friesian (HF), cross, or local. The origin of each individual animal was defined as either own stock or purchased. The clinical reproductive disorder experience, such as abortion, still-birth and retained fetal membrane were categorized as either yes or no variables. Parity number and frequency of some of the aforementioned disorders were also recorded. Physiological status of an animal was categorized as pregnant, non-pregnant, lactating and lactating-pregnant. Repeat breeding was also assessed based on the animal owner's general observation. Accordingly, cows that demanded 3 or more services per pregnancy were categorized as a repeat breeder otherwise they were categorized as a non-repeat breeder.

Study population

The target study populations were dairy cattle above six months of age, which consist of breeding females, replacement heifers, and available bulls. The breeds of these animals were crosses of local breed with HF. None of the animals tested were vaccinated against brucellosis. For the Knowledge, Attitude and Practice (KAP) study, occupationally associated farm workers, willing to be interviewed, were included.

Study design and sample size

A cross-sectional study design was employed from November 2016 to May 2017 to address the objectives of the study. According to Addis Ababa city urban agriculture bureau there are about 880 registered smallholder, medium scale and large commercial dairy farms in Addis Ababa. Individual farm was considered as a herd and the primary sampling unit. The sample size for dairy farms was calculated using a 9.1% herd level seroprevalence of bovine brucellosis [23], 95% confidence interval (CI) and 5% required precision [41]. Hence, a total of 127 dairy farms were considered for this study and proportional allocation was made for each sub city based on the number of farms. In each sub city, herd-sampling frame was established in collaboration with sub city veterinary department and farms were selected randomly using computer generated random numbers. Before data collection, consent was made with the identified farms owners requesting their farms to be included in the study. Farms where the owners were not willing to participate in the study were replaced by other farms. All cattle above six months of age in the selected dairy farms were included and a total of 1550 animals were sampled for serological screening. For the KAP study farm workers from sero surveyed farms, who agreed to be interviewed, were included. Hence, 130 farm workers from 59 farms participated in the study.

Data collection

Data concerning farm workers KAPs towards the disease were collected by interviewing individuals using a pre-tested structured questionnaire. Verbal consent was obtained from the respondents and the objective of the survey explained to them before start of the interview. The interviews were conducted in local languages (Afaan Oromo or Amharic). The questionnaire focused on demographic characteristic of the interviewee, knowledge-attitude about the disease, handling and afterbirth/aborted fetus disposal practices, habit of raw animal product consumption and animal feeding and housing practices.

Blood samples (10 ml) from the jugular vein of each animal were collected, using sterile needles and plain vacutainer tubes labelled with individual animal identification number. The blood samples were centrifuged at $3000\times g$ for 10 min to obtain the serum within 12 h of collection. Sera were decanted into cryo-vials, identified and stored at $-20\,^{\circ}\mathrm{C}$ until screened for antibodies against natural *Brucella* exposure using serological analysis.

Rose Bengal plate test (RBPT)

All sera samples collected were initially screened by RBPT using RBPT antigen (Animal and Plant Health Agency, New Haw, Addlestone, Surrey, KT15 3NB, United Kingdom) according to OIE (2016) procedures. Sera and antigen were taken from refrigerator and left at room temperature for half an hour before the test to reach room temperature. Briefly, RBT antigen (30 μ l) was added onto a glass slide next to an equal amount of cattle sera. The antigen and test serum were mixed thoroughly in a plastic applicator, shaken for 4 min, and agglutination was read immediately. Any observed agglutination by the naked eye was considered to be a positive reaction.

Competitive ELISA

All RBPT positive sera were further tested using the COMPELISA 160 and 400, a competitive ELISA kit for the detection of antibodies against Brucella in serum samples (Animal and Plant Health Agency, New Haw, Addlestone, Surrey, KT15 3NB, United Kingdom) at Addis Ababa University, Aklilu Lemma Institute of Pathobiology (AAU-ALIPB). The test was performed according to the manufacturer's instructions. The test was conducted in 96-well polystyrene plates that are pre-coated with Brucella species lipopolysaccharide (LPS) antigen. 20 µl of each test serum was added to each well followed by 100 μl of prepared conjugate solution. The plates were then shaken vigorously for two minutes and incubated at room temperature for 30 min on rotary shaker, at 160 revs/min. Plates were washed 5 times and dried. Hydrogen peroxidase substrate and chromogen solution was developed for

 $10\,\text{min.}\ 100\,\mu\text{l}$ of o-Phenylenediamine dihydrochloride (OPD) solution was added to all wells and the plates were incubated at room temperature for 10 to $20\,\text{min.}\ \text{Micro}$ plate reader was switched on and the units allowed stabilizing for $10\,\text{min.}\ \text{The}$ reaction was then being stopped using stopping solution. Optical densities (OD) were read at $450\,\text{nm}$ using micro plate reader. The lack of color development indicated that the sample tested was positive. A positive/negative cut-off was calculated as 60% of the mean of the OD of the 4 conjugate control wells. Any test sample giving an OD equal to or below this value was regarded as being positive. An animal was considered positive if it tested seropositive on both RBPT and c-ELISA in serial interpretation.

Complement fixation test (CFT)

All samples that were RBPT-positive were further subjected to complement fixation test as a confirmatory test at the National Veterinary Institute (NVI), Bishoftu, Ethiopia. The Brucella antigen and control sera (positive and negative) used during the test were produced by and Plant Health Agency, UK. standardization of the antigen was made at 1:20 working dilution (strength). The Brucella antigen, complement and 3% sensitized sheep red blood cells were added after the test sera were serially diluted (1:5, 1:10, 1:20, and 1:40) in microtitre plates. Then the plates were incubated at 37 °C for 30 min. The test was considered positive when the reading was as partial fixation (50% haemolysis) or complete fixation (no haemolysis) at 1:10 dilution. The validity of the test was considered when there was complete hemolysis in negative control serum and the positive control shows inhibition of hemolysis.

Case definition

Animals were considered positive to brucellosis when they tested positive on either RBPT/CFT or RBPT/c-ELISA tests in parallel interpretation. Similarly, a herd or farm was considered seropositive when at least one animal in a herd or farm tested positive. Since there is no history of vaccination against brucellosis in Ethiopia, seropositivity observed in this study was considered to be due to natural infection.

Data analysis

Data generated from the questionnaire survey and laboratory investigations were recorded and coded using a Microsoft Excel spreadsheet (Microsoft Corporation) and analyzed using STATA version 13.0 for Windows (Stata Corp. College Station, TX, USA). Descriptive statistics were used for demographic and farm characteristics as well as KAPs relating to bovine brucellosis. The sero-prevalence was calculated as the number of seropositive samples divided by the total number of samples tested.

Additional files

Additional file 1: Raw Data (XLSX 136 kb)

Additional file 2: Questionnaire prepared to assess the knowledge, attitude and practices (KAP) of farm workers towards brucellosis in Addis Ababa, Central Ethiopia. (DOCX 22 kb)

Abbreviations

AAU-ALIPB: Addis Ababa University, Aklilu Lemma Institute of Pathobiology; AW: Adrian Whatmore; AZ: Aboma Zewde; BM: Bedaso Mammo; cELISA: Competitive Enzyme Linked Immunosorbent Assay; CFT: Complement Fixation Test; CI: Confidence Interval; GA: Gobena Ameni; GH: Gizachew Hailegebreal; GZA: Gizat Almaw; HF: Holstein Friesian; iELISA: Indirect Enzyme Linked Immunosorbent Assay; JW: James Wood; KAP: Knowledge Attitude Practice; LPS: Lipopolysaccharide; NVI: National Veterinary Institute; RBPT: Rosbengal Plate Test; RFM: Retained Fetal Membranes; SAT: Serum Agglutination Tests; SB: Stefan Berg; TS: Teshale Sori; YZ: Yemiserach Zeleke; ZELS: Zoonoses and Emerging Livestock Systems

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Availability of data and materials

All data generated and analyzed during this study are included in this published article (and its Additional files 1 and 2).

Authors' contributions

BM and GH collected all the required data. GZA contributed to the collection of the data. BM and GH performed the laboratory work. AZ and YZ contributed to the lab work. JW, GA, SB, AW, BM and GH designed the study. BM and GH analyzed and interpreted the data. BM and GH drafted the manuscript. JW, GA, SB AW and TS critically and substantially revised the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Ethical clearance for this study was obtained from University of Cambridge, Department of Veterinary Medicine, Addis Ababa University, Aklilu Lema Institute of Pathobiology, Addis Ababa University, College of Veterinary Medicine and Agriculture (AAU-CVMA), Minutes of Animal Research Ethics and Review committee. Before conducting the research, participants were informed with the objectives of the study and written and signed consent was obtained from the farm owners or managers of the dairy farms to take samples from their cattle and farm workers to be included in the KAP survey.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- CSA. Federal Democratic Republic of Ethiopia. Central Statistical Agency, Agricultural Sample Survey Report on Livestock and Livestock Characteristics. 2015; Volume II, 2014/15. Addis Ababa, Ethiopia.
- Ibrahim N, Belihu K, Lobago F, Bekana M. Seroprevalence of bovine brucellosis and its risk factors in Jimma zone of Oromia region, South-Western Ethiopia. Trop Anim Health Prod. 2010;42:34–40.
- Schelling E, Diguimbaye C, Daoud S, Nicolet J, Boerlin P, Tanner M, Zinsstag J. Brucellosis and Q-fever sero prevalence of nomadic pastoralists and their livestock in Chad. Prev Vet Med. 2003;61:279–93.
- Diaz Aparicio E. Epidemiology of brucellosis in domestic animals caused by *Brucella melitensis*, *Brucella suis* and *Brucella abortus*. Rev Sci Tech. 2013;32(1):53–60.
- Radostits ED, Gay CC, Inchcliff KW. Veterinary Medicine Textbook of the disease of cattle, sheep, pigs, goats, and horses. 10th ed. Newyork: W. B. Sounders Company Itd; 2006. p. 963–94.
- OIE. BRUCELLOSIS. In: Manual of diagnostic tests and vaccines for terrestrial animals; 2016. Available at: http://www.oie.int/fileadmin/Home/eng/Health standards/tahm/ 2.01.04_BRUCELLOSIS.pdf. Accessed 2 Feb 2017.
- Debassa G, Tefera M, Addis M. Small ruminant brucellosis: serological survey in Yabello District, Ethiopia. Asia J Anim Sci. 2013;7(1):14–21.
- Roth F, Zinsstag J, Orkhon D, Chimed-Ochir G, Hutton G, Cosivi O, et al. Human health benefits from livestock vaccination for brucellosis: case study. Bull World Health Org. 2003;81:867–76.
- Musa MT, Eisa MZ, El Sanousi MA, Wahab EM, Perrett L. Brucellosis in camels (Camelus dromedarius) in Darfur, western Sudan. J Comp Pathol. 2008;13(8):151–5.
- Bechtol D, Carpenter LR, Mosites E, Smalley D, Dunn JR. Brucella melitensis infection following military duty in Iraq. Zoonoses Public Health. 2011;58(2):489–92.
- McDermott JJ, Arimi SM. Brucellosis in sub-Saharan Africa: epidemiology, control and impact. Vet Microbiol. 2002;90(1–4):111–34.
- Berhe G, Belihu K, Asfaw Y. Seroepidemiological investigation of bovine brucellosis in the extensive cattle production system of Tigray region of Ethiopia. Intern J Appl Res Vet Med. 2007;5:2.
- Jergefa T, Kelay B, Bekana M, Teshale S, Gustafson H, Kindahl H. Epidemiological study of bovine brucellosis in three agro-ecological areas of Central Oromiya, Ethiopia. Rev sci tech Off Int Epiz. 2009;28(3):933–43.
- Asmare K, Asfaw Y, Gelaye E, Ayelet G. Brucellosis in extensive management system of zebu cattle in Sidama zone, Southern Ethiopia. Afr J Agric Res. 2010;5:257–63.
- Degefu H, Mohamud M, Hailemelekot M, Yohannes M. Seroprevalence of bovine brucellosis in agro pastoral areas of Jijjiga zone of Somali National Regional State, eastern Ethiopia. Ethiop Vet J. 2011;15(1):37–47.
- Megersa B, Biffa D, Niguse F, Rufael T, Asmare K, Skjerve E. Cattle brucellosis in traditional livestock husbandry practice in southern and eastern Ethiopia, and its zoonotic implication. Acta Vet Scandinavica. 2011;53:24.
- Tesfaye G, Tsegaye W, Chanie M, Abinet F. Seroprevalence and associated risk factors of bovine brucellosis in Addis Ababa dairy farms. Trop Anim Health Prod. 2011;43:1001–5.

- Megersa B, Biffaf D, Abunna F, Regassa A, Godfroid J, Skjerve E.
 Seroepidemiological study of livestock brucellosis in a pastoral region. Epidemiol Infect. 2012;140:887–96. https://doi.org/10.1017/S0950268811001178.
- Yohannes M, Mersha T, Degefu H, Tolosa T, Woyesa MBB. Serological survey in Guto-Gida District, east Wollega zone, Ethiopia. Global Veterinaria. 2012;8(2):139–43.
- Adugna KE, Agga GE, Zewde G. Seroepidemiological survey of bovine brucellosis in cattle under a traditional production system in western Ethiopia. Rev Sci Tech. 2013;32(3):765–73.
- Gumi B, Firdessa R, Yamuah L, Sori T, Tolosa T, Aseffa A, Zinsstag J, Schelling E. Seroprevalence of brucellosis and Q-fever in southeast Ethiopian pastoral livestock. J Vet Sci Med Diagn. 2013;2:1.
- Tschopp R, Abera B, Sourou SY, Guerne-Bleich E, Aseffa A, Wubete A, Zinsstag J, Young D. Bovine tuberculosis and brucellosis prevalence in cattle from selected milk cooperatives in Arsi zone, Oromia region, Ethiopia. BMC Vet Res. 2013;9:163.
- Asmare K, Sibhat B, Molla B, Ayelet G, Shiferaw J, Martin AD, Skjerve E, Godfroid J. The status of bovine brucellosis in Ethiopia with special emphasis on exotic and cross bred cattle in dairy and breeding farms. Acta Trop. 2013;126:186–92.
- 24. Alemu F, Admasu P, Feyera T, Niguse A. Seroprevalence of bovine brucellosis in eastern Showa, Ethiopia. Acad J Anim Dis. 2014;3(3):27–32.
- Asgedom H, Damena D, Duguma R. Seroprevalence of bovine brucellosis and associated risk factors in and around Alage district, Ethiopia. Springer Plus. 2016;5:851. https://doi.org/10.1186/s40064-016-2547-0.
- Geresu MA, Ameni G, Kassa T, Tuli G, Arenas A, Gezahegne MK.
 Seropositivity and risk factors for *Brucella* in dairy cows in Asella and Bishoftu towns, Oromia regional state, Ethiopia. Afr J Microbiol Res. 2016; 10(7):203–13. https://doi.org/10.5897/AJMR2015.7707.
- Geresu MA, Ameni G, Wubete A, Arenas-Gamboa AM, Gezahegne MK. Isolation and Identification of *Brucella* Species from Dairy Cattle by Biochemical Tests: The First Report from Ethiopia. World Vet J. 2016;6(2):80–8.
- Asmare K, Krontveit IR, Ayelet G, Sibhat B, Godfroid J, Skjerve E. Metaanalysis of *Brucella* seroprevalence in dairy cattle of Ethiopia. Trop Anim Health Prod. 2014;46:1341–50. https://doi.org/10.1007/s11250-014-0669-3.
- Lindahl E, Sattorov N, Boqvist S, Magnusson UA. Study of knowledge, attitudes and practices relating to brucellosis among small-scale dairy farmers in an urban and Peri- urban area of Tajikistan. PLoS One. 2015;10(2):e0117318.
- Alem W, Solomon G. A retrospective sero-epidemiology study of bovine brucellosis in different production systems in Ethiopia. In: Proceeding of 16th annual conference; 2002. p. 53–7. June 5–6, Addis Ababa, Ethiopia.
- Belihu K. Analysis of dairy cattle breeding practices in selected areas of Ethiopia. PhD Thesis. Berlin: Humboldt University; 2002.
- Bashitu L, Afera B, Tuli G, Aklilu F. SeroPrevalence study of bovine brucellosis and its associated risk factors in Debrebirhan and ambo towns. J Adv Dairy Res. 2015;3:131. https://doi.org/10.4172/2329-888X.1000131.
- Rashid M. Reproductive wastage in cattle due to bovine brucellosis. In: Proceeding of the fourth livestock improvement conference. Addis Ababa: Ethiopian Institute of Agricultural Research; 1993. p. 270–2.
- 34. Pappas G, Papadimitriou P, Akritidis N, Christou L, Tsianos E. The new global map of human brucellosis. Lancet Infect Dis. 2006;6:91–9.
- Pieracci EG, Hall AJ, Gharpure R, Haile A, Walelign E, Deressa A, Bahiru G, Kibebe M, Walke H, Belay E. Prioritizing zoonotic diseases in Ethiopia using a one health approach. One Health. 2016;2:131–5.
- Godfroid J, Nielsen K, Saegerman C. Diagnosis of brucellosis in livestock and wildlife. Croat Med J. 2010;51:296–305. https://doi.org/10.3325/cmj.2010.51.296.
- Plackett P, Alton GG. A mechanism for prozone formation in the complement fixation test for bovine brucellosis. Aust Vet J. 1975;51:374.
- 38. Corbel MJ. Brucellosis in humans and animals. Geneva: Produced by the, WHO in collaboration with the, FAO and OIE; 2006.
- Fosgate GT, Carpenter TE, Chomel BB, Case JT, Debess EE, Reilly KF. Time-space clustering of human brucellosis, California, 1973–1992. Emerg Infect Dis. 2002; 8(7):672–8. https://doi.org/10.3201/eid0807.010351. PMID: 12095433.
- National Metrology Institute of Ethiopia. 2016. http://www.most.gov.et/ national-metrology-institute-of-ethiopia. Accessed Nov 9 2016.
- Thrusfield M. Sampling in veterinary epidemiology. 3rd ed. London: Black well Science Ltd.; 2007. p. 214–56.