

Sequence variants of *BIEC2-808543* near *LCORL* are associated with body composition in Thoroughbreds under training

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Ligand-dependent nuclear receptor compressor-like (LCORL) encodes a transcription factor, and its polymorphisms are associated with measures of skeletal frame size and adult height in several species. Recently, the single nucleotide polymorphism (SNP) BIEC2-808543 located upstream of LCORL was identified as a genetic diagnostic marker associated with withers height in Thoroughbreds. In this study, 322 Thoroughbreds-in-training were genotyped for BIEC2-808543 to evaluate the association between genotype and body composition traits, including body weight, withers height, the ratio of body weight to withers height, chest circumference, and cannon circumference. Of these, withers height and cannon circumference were significantly associated with LCORL genotypes throughout almost the entire training period in males and females. Animals with a C/T genotype had higher withers height (maximum differences of 1.8 cm and 2.1 cm in males and females, respectively) and cannon circumference (maximum differences of 0.65 cm and 0.48 cm in males and females, respectively) compared with animals with a T/T genotype. These results suggested that the regulation of LCORL expression influences the skeletal frame size in Thoroughbreds and thus, indirectly affects the body weight. Although LCORL and BIEC2-808543 would be useful for selective breeding in Thoroughbreds, the production of genetically modified animals and gene doping based on genetic information should be prohibited in order to maintain racing integrity.

Key words: *BIEC2-808543, cannon circumference, LCORL, Thoroughbred, withers height*

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The Thoroughbred is a breed that was developed in the early 18th century. The origin of Thoroughbreds can be traced back to a small number of Arab stallions and native British mares approximately 300 years ago [2, 4, 24]. Current Thoroughbreds have larger body sizes compared with their ancestors. For instance, the withers height of old racehorses, such as the Darley Arabian, was reported to be about 152 cm in old literature, whereas the withers height of current Thoroughbreds is about 160.5 cm [5]. Morphological characteristics of Thoroughbreds have been improved to

obtain superior performance by selective breeding [6, 13, 22, 23].

The *ligand-dependent nuclear receptor compressor-like (LCORL)* is thought to encode a transcription factor associated with measures of skeletal frame size and adult height in humans and several animals [8, 10, 11, 20, 21]. In horses, *LCORL* has been mapped on *Equus caballus* autosome 3 (ECA3), and it has been reported that *BIEC2-808543*, a single nucleotide polymorphism (SNP) located upstream of the gene, is significantly associated with withers height [14, 19]. It is known that height is a quantitative trait controlled by numerous genes in humans; however, the genetic basis of body size in Thoroughbreds is still understudied.

In this study, we investigated the relationships between sequence variants of *BIEC2-808543* and measured morphological characteristics, such as body weight, withers height, chest circumference, and cannon circumference, in male and female Thoroughbreds prior to the initiation of training

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and at monthly intervals during the first seven months of training. We hypothesized that identification of genes controlling body size in Thoroughbreds might be relatively simple, since the breeding population includes a limited number of sires and dams. We also assumed that *LCORL* might affect the body composition of Thoroughbreds.

Materials and Methods

Thoroughbreds

This study was performed according to the ethical standards in animal research. A total of 322 (165 males and 167 females) Thoroughbreds were used in this study. All horses were born between January and June of 2009–2014, and the majority of them were born in March and April. All horses were purchased at commercial livestock auctions at one year of age, introduced to the Hidaka Training and Research Center (Japan Racing Association, JRA) by the end of September, trained to be ridden (initial training) during October, and trained for racing (full-scale training) from November through April.

Body composition measurement

Body composition assessment was initially performed in September (approximately 18 months of age) and was subsequently performed at monthly intervals during the following six months of training. Measurements included body weight (kg), withers height (cm), chest circumference (cm), and cannon circumference (cm). The ratio of body weight to withers height (kg cm^{-1}) was calculated and defined as the “muscle content”, since the skeletal muscle mass of Thoroughbreds comprises over 55% of their total body mass [7], and thus, it can be used as an indicator of skeletal muscle mass.

SNP genotyping

Blood samples were collected from each animal and stored at -40°C . Genomic DNA was extracted using an MFX-2000 MagExtractor System (Toyobo, Osaka, Japan), according to the manufacturer’s protocol. Genotyping for *BIEC2-808543* was performed using a Taqman SNP Genotyping Assay with a 7500 Fast Real-Time PCR System (Applied Biosystems, Foster City, CA, U.S.A.), according to the manufacturer’s protocols. The primer and probe sequences used in this study are shown in Table 1.

Statistical analysis

Genetic association analysis for body weight, withers height, chest circumference, cannon circumference, and the ratio of body weight to withers height was performed using Student’s *t*-test. In a previous study, we observed differences in body composition between males and females [22].

Table 1. Primer and probe sequences used in this study

	Sequence
F-primer	CCAAATTTGCCTGGCTAGAGA
R-primer	TGTTCCCTGTGATTCTGCCTTT
MGB_VIC_C	CATCCAGCTTATTCTGTGA
MGB_FAM_T	CATCCAGTTTATTCTGTAC

Thus, phenotypic data and SNP associations were separately evaluated for each sex. In the present study, double blind experiments were designed to avoid errors arising from bias in phenotyping and SNP genotyping. All statistical analyses were performed using IBM SPSS Statistics 19 (IBM, Armonk, NY, U.S.A.).

Results

Growth trends in body composition

To avoid pedigree-based and age-based systematic errors, 59 males and 48 females were selected, excluding animals with full-sibling and half-sibling relationships and/or those born in January, February, May, or June. No significant differences were observed in age between the subpopulations.

Body weight, withers height, the ratio of body weight to withers height, and chest circumference increased with time in both males and females, although the changes (increase or decrease) were minimal prior to the initiation of full-scale training (Table 2, Fig. 1). Body weight, withers height, and chest circumference of both males and females significantly ($P<0.05$) increased in March compared with those in October (Table 2, Fig. 1). Body weight and the ratio of body weight to withers height decreased in November, when full-scale training started, and then increased from December through April (Table 2, Fig. 1).

Differences between males and females

Body weight and the ratio of body weight to withers height were significantly ($P<0.05$) different between males and females from December through March (Table 2, Fig. 1). Additionally, no significant differences were observed in withers height between males and females throughout the experimental period (Table 2, Fig. 1). Chest circumference was significantly ($P<0.05$) higher in females in September and November, whereas no differences were observed between males and females from December through March (Table 2, Fig. 1). Cannon circumference was significantly ($P<0.05$) different between males and females throughout the experimental period (Table 2, Fig. 1); the measured values were significantly ($P<0.05$) higher in males than in females.

Table 2. Body weight (kg), withers height (cm), ratio of body weight to withers height (kg cm^{-1}), chest circumference (cm), and cannon circumference (cm) (average values \pm standard error; SE) from September to March by gender

	Sep.	Oct.	Nov.	Dec.	Jan.	Feb.	Mar.
Ratio of body weight to withers height (kg cm^{-1})							
Male (59)	2.92 \pm 0.02	2.95 \pm 0.02	2.92 \pm 0.02	2.94 \pm 0.02	2.96 \pm 0.02	2.99 \pm 0.02	3.03 \pm 0.02
Female (48)	2.88 \pm 0.02	2.90 \pm 0.02	2.88 \pm 0.02	2.87 \pm 0.02	2.89 \pm 0.02	2.92 \pm 0.02	2.94 \pm 0.02
<i>P</i> -values	0.1631	0.0983	0.1754	0.0160*	0.0224*	0.0149*	0.0018*
Body weight (kg)							
Male (59)	448.5 \pm 3.0	455.0 \pm 3.2	453.1 \pm 3.2	459.1 \pm 3.2	464.3 \pm 3.2	473.6 \pm 3.1	482.7 \pm 3.0
Female (48)	443.4 \pm 3.3	448.2 \pm 3.2	447.5 \pm 3.3	447.6 \pm 3.4	453.0 \pm 3.4	462.0 \pm 3.5	467.1 \pm 3.4
<i>P</i> -values	0.2572	0.1386	0.2355	0.0170*	0.0170*	0.0143*	0.0007*
Withers height (cm)							
Male (59)	153.8 \pm 0.3	154.4 \pm 0.3	155.1 \pm 0.3	155.9 \pm 0.4	157.1 \pm 0.4	158.2 \pm 0.4	159.2 \pm 0.4
Female (48)	154.1 \pm 0.3	154.6 \pm 0.3	155.3 \pm 0.3	155.8 \pm 0.3	156.9 \pm 0.3	158.0 \pm 0.3	158.7 \pm 0.3
<i>P</i> -values	0.5261	0.7230	0.7076	0.8252	0.6340	0.7694	0.2954
Chest circumference (cm)							
Male (59)	174.3 \pm 0.4	175.7 \pm 0.5	176.3 \pm 0.5	177.3 \pm 0.4	178.7 \pm 0.4	179.5 \pm 0.4	180.8 \pm 0.4
Female (48)	176.8 \pm 0.5	177.0 \pm 0.5	177.8 \pm 0.5	178.1 \pm 0.4	178.9 \pm 0.4	179.3 \pm 0.4	180.9 \pm 0.4
<i>P</i> -values	0.0002*	0.0769	0.0339*	0.2187	0.8319	0.7280	0.8894
Cannon circumference (cm)							
Male (59)	19.54 \pm 0.08	19.58 \pm 0.08	19.56 \pm 0.08	19.60 \pm 0.08	19.63 \pm 0.08	19.67 \pm 0.08	19.74 \pm 0.07
Female (48)	19.18 \pm 0.07	19.13 \pm 0.07	19.16 \pm 0.06	19.17 \pm 0.06	19.18 \pm 0.06	19.20 \pm 0.06	19.24 \pm 0.06
<i>P</i> -values	0.0015*	<0.0001*	0.0002*	<0.0001*	<0.0001*	<0.0001*	<0.0001*

*Statistically significant *P*-values (<0.05) in a *t*-test.

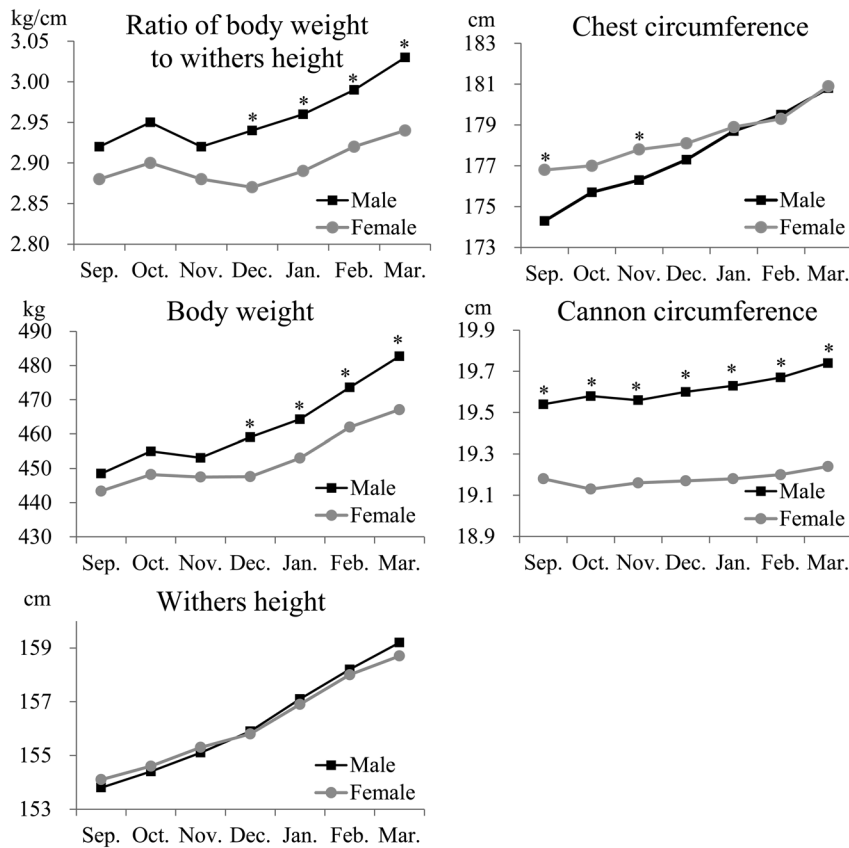


Fig. 1. Changes in the mean ratio of body weight to withers height, body weight, withers height, chest circumference, and cannon circumference from September to the following March in each sex. The black square (■) and gray circle (●) indicate male and female, respectively. An asterisk (*) indicates a statistical difference at $P < 0.05$ by sex.

Genotypic distribution

All the animals used in this study were successfully genotyped for *BIEC2-808543*. No significant differences were observed in genotypic distribution and allele frequency between males and females (Table 3). Since the C/C genotype had a very low frequency in the population, only the C/T and T/T genotypes were used for the genotype-phenotype association study.

Four subpopulations were assigned based on *BIEC2-808543* genotype and gender. Animals (n=200) with full and half-sibling relationships and/or those born in January, February, May, or June were excluded from the four subpopulations in order to eliminate pedigree-based and age-based systematic errors. Therefore, the four subpopulations included a total of 67 males and 55 females. These numbers were different from those in the growth trend analysis (59 males and 48 females) due to the classification by sex and genotype. No significant differences were observed in age among the four subpopulations. The four subpopulations were used for the association study.

Genotype association with body composition in males

Significant ($P<0.05$) differences were observed in body weight in September, October, November, and February; in withers height from September to January; in chest circumference in September; and in cannon circumference from September to February between genotypes in males (Table 4, Fig. 2). The measured values were significantly ($P<0.05$) higher in the C/T genotype than in the T/T genotype throughout almost the entire experimental period. However, no significant differences were observed in the ratio of body weight to withers height between genotypes at any point during the experimental period.

Genotype association with body composition in females

Withers height and cannon circumference in females were significantly ($P<0.05$) higher in the C/T genotype than in the T/T genotype throughout the experimental period. Withers height tended to increase during the training period, whereas cannon circumference did not show any changes (Table 4, Fig. 2). No significant differences were observed in body weight, the ratio of body weight to withers height, and chest circumference between genotypes at any point during the experimental period, although the measured values tended to be higher in the C/T genotype than in the T/T genotype.

Discussion

To our knowledge, this is the first report of relationships between sequence variants of *BIEC2-808543* and measured morphological characteristics in Thoroughbreds under training. Our data demonstrated that *BIEC2-808543* was

Table 3. Genotypic frequency of *BIEC2-808543* near the *ligand-dependent nuclear receptor compressor-like (LCORL)* gene on *Equus caballus* autosome 3 (ECA3)

	C/C	T/C	T/T
Male	0	0.15	0.85
Female	0.01	0.18	0.81

significantly associated with withers height (Table 4, Fig. 2), and the results were consistent with those reported in previous studies on several horse breeds [14, 19]. Animals with a C/T genotype had higher withers heights (maximum differences of 1.8 cm in males; maximum differences of 2.1 cm in females) compared with animals with a T/T genotype. We also demonstrated that *BIEC2-808543* was associated with cannon circumference in Thoroughbreds under training. Animals with a C/T genotype had higher cannon circumferences (maximum differences of 0.65 cm in males; maximum difference of 0.48 in females) compared with animals with a T/T genotype. These findings suggested that *LCORL* on ECA3 was closely associated with the skeletal frame and body composition in Thoroughbreds. *BIEC2-808543* polymorphism disrupts a putative binding site of the transcription factor TFIID [15], which suggests that the influence on the skeletal frame is caused by a regulation mechanism of *LCORL* expression.

We also observed associations between genotypes and phenotypes when all 322 animals were used for analysis (data not shown). Although the C/C genotype was not evaluated in this study because of a low frequency (Table 3), one female horse with the C/C genotype showed a withers height of 162.0 cm in March. Therefore, it is expected that horses with a C/C genotype would have a much higher withers height and cannon circumference compared with those with a C/T or T/T genotype. Overall, our findings suggested that the C-allele at *BIEC2-808543* increased the skeletal frame size and that the T-allele decreased it.

In the present study, no statistical differences were observed in chest circumference throughout almost the entire experimental period, suggesting that *LCORL* does not have a strong effect on chest circumference in horses (Table 4, Fig. 2). The differences between genotypes in terms of the association with chest circumference seemed to decrease with growth and/or advanced training. The thoroughbreds used in this study trained almost every day; thus, *LCORL* was not able to strongly affect their chest circumference.

Our data showed that cannon circumference was affected by gender (Table 2, Fig. 1). In the four subpopulations, males with a C/T genotype had the highest cannon circumference, males with a T/T genotype and females with a C/T genotype had an intermediate cannon circumference,

Table 4. Body weight (kg), withers height (cm), ratio of body weight to withers height (kg cm^{-1}), chest circumference (cm), and cannon circumference (cm) (average values \pm standard error; SE) from September to March by genotype in males and females

	Sep.	Oct.	Nov.	Dec.	Jan.	Feb.	Mar.
Male							
Ratio of body weight to withers height (kg cm^{-1})							
C/T (13)	2.98 \pm 0.04	3.02 \pm 0.04	2.98 \pm 0.05	3.00 \pm 0.04	2.99 \pm 0.04	3.05 \pm 0.04	3.07 \pm 0.03
T/T (54)	2.90 \pm 0.02	2.93 \pm 0.02	2.91 \pm 0.02	2.93 \pm 0.02	2.95 \pm 0.02	2.98 \pm 0.02	3.02 \pm 0.02
<i>P</i> -values	0.0582	0.0608	0.1099	0.1715	0.2955	0.1193	0.2389
Body weight (kg)							
C/T (13)	462.5 \pm 7.0	469.5 \pm 7.8	466.9 \pm 8.0	471.8 \pm 8.0	474.4 \pm 7.3	485.8 \pm 7.1	489.2 \pm 6.4
T/T (54)	445.2 \pm 3.1	451.4 \pm 3.1	450.1 \pm 3.2	456.4 \pm 3.3	461.6 \pm 3.2	470.1 \pm 3.1	480.0 \pm 3.1
<i>P</i> -values	0.0175*	0.0168*	0.0292*	0.0502	0.0901	0.0340*	0.2012
Withers height (cm)							
C/T (13)	155.1 \pm 0.6	155.7 \pm 0.6	156.5 \pm 0.6	157.2 \pm 0.7	158.5 \pm 0.6	159.4 \pm 0.6	159.4 \pm 0.8
T/T (54)	153.5 \pm 0.4	154.0 \pm 0.3	154.7 \pm 0.4	155.5 \pm 0.4	156.7 \pm 0.4	157.8 \pm 0.4	159.0 \pm 0.3
<i>P</i> -values	0.0434*	0.0368*	0.0270*	0.0382*	0.0351*	0.0594	0.6640
Chest circumference (cm)							
C/T (13)	176.2 \pm 0.8	177.3 \pm 1.2	177.7 \pm 1.2	178.5 \pm 1.1	179.5 \pm 0.9	180.2 \pm 0.9	180.2 \pm 0.6
T/T (54)	173.8 \pm 0.5	175.2 \pm 0.5	175.7 \pm 0.5	176.9 \pm 0.5	178.3 \pm 0.4	179.0 \pm 0.4	180.7 \pm 0.5
<i>P</i> -values	0.0277*	0.0797	0.1045	0.1430	0.2563	0.2272	0.5840
Cannon circumference (cm)							
C/T (13)	20.09 \pm 0.18	20.09 \pm 0.17	20.11 \pm 0.18	20.11 \pm 0.18	20.12 \pm 0.18	20.15 \pm 0.17	20.00 \pm 0.19
T/T (54)	19.44 \pm 0.08	19.48 \pm 0.08	19.47 \pm 0.08	19.51 \pm 0.08	19.54 \pm 0.08	19.59 \pm 0.07	19.70 \pm 0.07
<i>P</i> -values	0.0011*	0.0015*	0.0012*	0.0023*	0.0021*	0.0020*	0.0710
Female							
Ratio of body weight to withers height (kg cm^{-1})							
C/T (11)	2.91 \pm 0.04	2.93 \pm 0.04	2.92 \pm 0.04	2.88 \pm 0.04	2.89 \pm 0.04	2.93 \pm 0.04	2.96 \pm 0.04
T/T (44)	2.87 \pm 0.02	2.89 \pm 0.02	2.87 \pm 0.02	2.86 \pm 0.02	2.88 \pm 0.02	2.91 \pm 0.02	2.94 \pm 0.02
<i>P</i> -values	0.5163	0.4336	0.2508	0.6778	0.8515	0.7514	0.6298
Body weight (kg)							
C/T (11)	451.2 \pm 6.5	456.0 \pm 6.7	457.5 \pm 7.1	452.3 \pm 6.9	457.4 \pm 6.6	467.2 \pm 6.7	474.9 \pm 6.7
T/T (44)	441.5 \pm 3.6	445.6 \pm 3.5	443.6 \pm 3.4	444.6 \pm 3.6	449.8 \pm 3.7	458.5 \pm 3.7	465.4 \pm 3.6
<i>P</i> -values	0.2218	0.1846	0.0752	0.3440	0.3565	0.2961	0.2351
Withers height (cm)							
C/T (11)	155.2 \pm 0.7	155.5 \pm 0.7	156.5 \pm 0.9	156.8 \pm 0.9	158.4 \pm 0.7	159.6 \pm 0.8	160.2 \pm 0.8
T/T (44)	153.7 \pm 0.3	154.1 \pm 0.2	154.7 \pm 0.3	155.3 \pm 0.3	156.4 \pm 0.2	157.5 \pm 0.2	158.3 \pm 0.3
<i>P</i> -values	0.0131*	0.0176*	0.0154*	0.0347*	0.0013*	0.0018*	0.0059*
Chest circumference (cm)							
C/T (11)	178.5 \pm 0.7	178.3 \pm 0.9	179.2 \pm 1.0	179.0 \pm 0.9	179.8 \pm 1.1	180.6 \pm 0.9	181.4 \pm 0.8
T/T (44)	176.3 \pm 0.5	176.5 \pm 0.5	177.2 \pm 0.5	177.6 \pm 0.4	178.3 \pm 0.4	178.8 \pm 0.5	180.5 \pm 0.4
<i>P</i> -values	0.0505	0.1142	0.0744	0.2007	0.1226	0.0824	0.3515
Cannon circumference (cm)							
C/T (11)	19.58 \pm 0.13	19.50 \pm 0.11	19.55 \pm 0.10	19.56 \pm 0.10	19.55 \pm 0.10	19.53 \pm 0.11	19.51 \pm 0.12
T/T (44)	19.10 \pm 0.07	19.07 \pm 0.07	19.08 \pm 0.06	19.08 \pm 0.05	19.10 \pm 0.06	19.14 \pm 0.05	19.22 \pm 0.06
<i>P</i> -values	0.0020*	0.0038*	0.0004*	0.0002*	0.0006*	0.0021*	0.0344*

*Statistically significant *P*-values (<0.05) in a *t*-test.

and females with a T/T genotype had the lowest cannon circumference (Table 4, Fig. 2). Because it is generally considered that bone size is an important determinant of bone strength, cannon circumference may play an important role in bone fractures, which are a common cause of loss in Thoroughbreds [17]. Therefore, further studies are needed to investigate the association between bone fracture, *LCORL*, and sex in order to improve horse health and welfare.

Males with a C/T genotype at *LCORL* had relatively higher body weights and withers heights. Therefore, body weight was related to withers height in *LCORL* genotypes, but not to muscle content, since no significant differences were identified in the ratio of body weight to withers height (Table 4, Fig. 2). A similar tendency was also observed in females (Table 4, Fig. 2). In our previous study, racehorses with a C/C genotype at the *myostatin* (*MSTN*) gene demon-

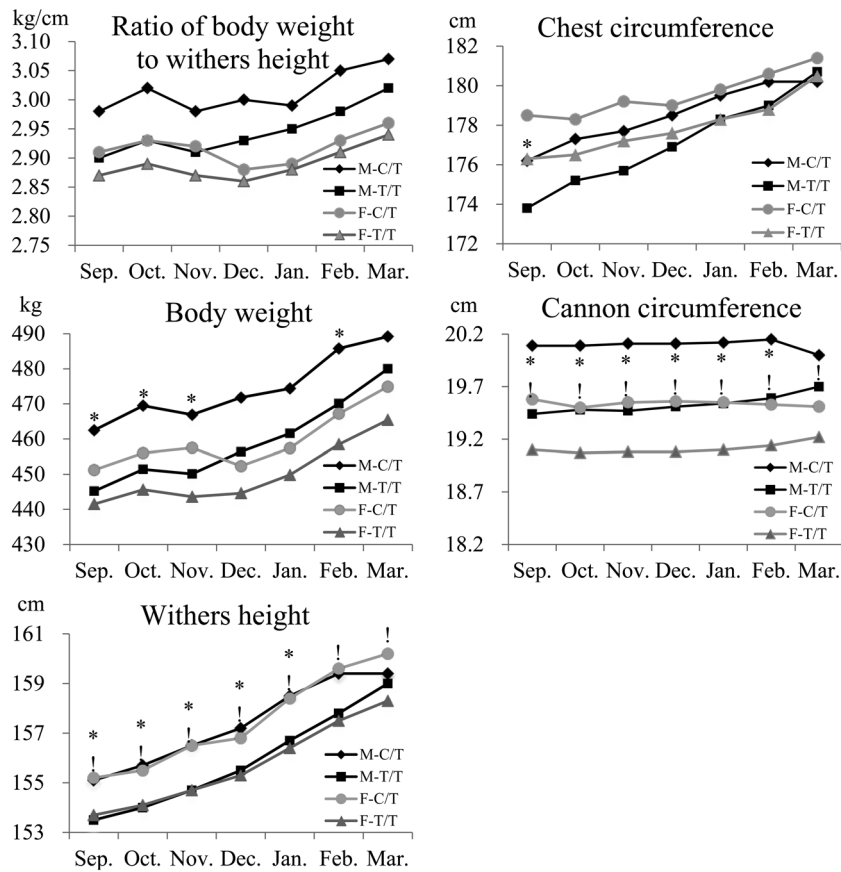


Fig. 2. Changes in the mean ratio of body weight to withers height, body weight, withers height, chest circumference, and cannon circumference from September to the following March expressed according to the genotype at *BIEC2-808543* in each sex. Black indicates male and gray indicates female, and the rhombus (◆), square (■), circle (●) and triangle (▲) indicate male-C/T, male-T/T, female-C/T and female-T/T, respectively. An asterisk (*) indicates a statistical difference at $P < 0.05$ in males. An exclamation mark (!) indicates a statistical difference at $P < 0.05$ in females.

strated a relatively higher body weights and skeletal muscle masses than racehorses with other genotypes [6, 13, 22, 23].

Based on genotypes at *LCORL* and *MSTN* and sex, we may be able to identify the ideal body composition for each racehorse. For instance, male racehorses with a C/C genotype at *LCORL* and a C/C genotype at *MSTN* may have large and heavy bodies, whereas female racehorses with a T/T genotype at *LCORL* and a T/T genotype at *MSTN* may have small and light bodies. Thus, animal care and training, such feeding and exercise for regulating body weight, could be applied based on individual genotypic information in a manner similar to that in personalized medicine in humans [1]. In addition, genotypic information could contribute to the development of effective breeding strategies for producing Thoroughbreds with favorable body traits. Because of the relatively low frequency of the C-allele at *LCORL*, withers height could be modified based on the

LCORL genotype, leading to improved racing performance.

Although selective breeding based on genetic information is effective for producing racehorses with desired traits, gene doping, which may be defined as the “abuse/misuse of gene therapy”, should be prohibited in order to maintain racing integrity [3]. The CRISPR/Cas system for genome editing [9] and the adeno-associated virus (AAV) vector [12, 18] for gene transfer have been developed for the production of genetically modified animals and gene therapy, respectively. In addition, the combination of both technologies allows for *in vivo* genome editing that directly modifies the genome in postnatal animals [16]. Therefore, these technologies raise concerns regarding the production of genetically modified animals and gene doping in racehorses. Since *LCORL* and *MSTN* influence body phenotype and/or racing performance, the genes should be monitored to protect the genomic and genetic integrity of racehorses.

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References

- Apellaniz-Ruiz, M., Gallego, C., Ruiz-Pinto, S., Carracedo, A., and Rodríguez-Antona, C. 2016. Human genetics: international projects and personalized medicine. *Drug. Metabol. Personal. Ther.* **31**: 3–8.
- Bower, M.A., Campana, M.G., Whitten, M., Edwards, C.J., Jones, H., Barrett, E., Cassidy, R., Nisbet, R.E., Hill, E.W., Howe, C.J., and Binns, M. 2011. The cosmopolitan maternal heritage of the Thoroughbred racehorse breed shows a significant contribution from British and Irish native mares. *Biol. Lett.* **7**: 316–320. [[Medline](#)] [[CrossRef](#)]
- Brzezińska, E., Domańska, D., and Jegier, A. 2014. Gene doping in sport - perspectives and risks. *Biol. Sport* **31**: 251–259. [[Medline](#)] [[CrossRef](#)]
- Cunningham, E.P., Dooley, J.J., Splan, R.K., and Bradley, D.G. 2001. Microsatellite diversity, pedigree relatedness and the contributions of founder lineages to thoroughbred horses. *Anim. Genet.* **32**: 360–364. [[Medline](#)] [[CrossRef](#)]
- Đermanović, V., Mitrović, S., Đordjević, N., and Novaković, M. 2010. Some significant exterior and reproductive properties of the English Thoroughbred horse population from the stud farm “Ljubicevo”–Serbia. *Bio-technol. Anim. Husb.* **26**: 75–82. [[CrossRef](#)]
- Fonseca, R.G., Kenny, D.A., Hill, E.W., and Katz, L.M. 2013. The relationship between body composition, training and race performance in a group of Thoroughbred flat racehorses. *Equine Vet. J.* **45**: 552–557. [[Medline](#)] [[CrossRef](#)]
- Gunn, H.M. 1987. Muscle, bone and fat productions and muscle distribution of thoroughbreds and quarter horses. *In: Equine Exercise Physiology 2: Proceedings of the Second International Conference on Equine Exercise Physiology; August 7–11 1986, San Diego.* http://www.iceep.org/pdf/iceep2/_1129101114_001.pdf (accessed on May 1, 2016).
- Hirschhorn, J.N., and Lettre, G. 2009. Progress in genome-wide association studies of human height. *Horm. Res.* **71**:(Suppl 2): 5–13. [[Medline](#)] [[CrossRef](#)]
- Hsu, P.D., Lander, E.S., and Zhang, F. 2014. Development and applications of CRISPR-Cas9 for genome engineering. *Cell* **157**: 1262–1278. [[Medline](#)] [[CrossRef](#)]
- Lettre, G. 2011. Recent progress in the study of the genetics of height. *Hum. Genet.* **129**: 465–472. [[Medline](#)] [[CrossRef](#)]
- Lindholm-Perry, A.K., Sexten, A.K., Kuehn, L.A., Smith, T.P., King, D.A., Shackelford, S.D., Wheeler, T.L., Ferrell, C.L., Jenkins, T.G., Snelling, W.M., and Freetly, H.C. 2011. Association, effects and validation of polymorphisms within the NCAPG - *LCORL* locus located on BTA6 with feed intake, gain, meat and carcass traits in beef cattle. *BMC Genet.* **12**: 103. [[Medline](#)] [[CrossRef](#)]
- Lisowski, L., Tay, S.S., and Alexander, I.E. 2015. Adeno-associated virus serotypes for gene therapeutics. *Curr. Opin. Pharmacol.* **24**: 59–67. [[Medline](#)] [[CrossRef](#)]
- Love, S., Wyse, C.A., Stirk, A.J., Stear, M.J., Calver, P., Voute, L.C., and Mellor, D.J. 2006. Prevalence, heritability and significance of musculoskeletal conformational traits in Thoroughbred yearlings. *Equine Vet. J.* **38**: 597–603. [[Medline](#)] [[CrossRef](#)]
- Makvandi-Nejad, S., Hoffman, G.E., Allen, J.J., Chu, E., Gu, E., Chandler, A.M., Loreda, A.I., Bellone, R.R., Mezey, J.G., Brooks, S.A., and Sutter, N.B. 2012. Four loci explain 83% of size variation in the horse. *PLoS One* **7**: e39929. [[Medline](#)] [[CrossRef](#)]
- Metzger, J., Schimpf, R., Philipp, U., and Distl, O. 2013. Expression levels of *LCORL* are associated with body size in horses. *PLoS One* **8**: e56497. [[Medline](#)] [[CrossRef](#)]
- Ran, F.A., Cong, L., Yan, W.X., Scott, D.A., Gootenberg, J.S., Kriz, A.J., Zetsche, B., Shalem, O., Wu, X., Makarova, K.S., Koonin, E.V., Sharp, P.A., and Zhang, F. 2015. In vivo genome editing using *Staphylococcus aureus* Cas9. *Nature* **520**: 186–191. [[Medline](#)] [[CrossRef](#)]
- Riggs, C.M. 2002. Fractures—a preventable hazard of racing thoroughbreds? *Vet. J.* **163**: 19–29. [[Medline](#)] [[CrossRef](#)]
- Salganik, M., Hirsch, M.L., and Samulski, R.J. 2015. Adeno-associated virus as a mammalian DNA vector. *Microbiol. Spectr.* **3**: 4. [[Medline](#)] [[CrossRef](#)]
- Signer-Hasler, H., Flury, C., Haase, B., Burger, D., Simianer, H., Leeb, T., and Rieder, S. 2012. A genome-wide association study reveals loci influencing height and other conformation traits in horses. *PLoS One* **7**: e37282. [[Medline](#)] [[CrossRef](#)]
- Soranzo, N., Rivadeneira, F., Chinappen-Horsley, U., Malkina, I., Richards, J.B., Hammond, N., Stolk, L., Nica, A., Inouye, M., Hofman, A., Stephens, J., Wheeler, E., Arp, P., Gwilliam, R., Jhamai, P.M., Potter, S., Chaney, A., Ghori, M.J., Ravindrarajah, R., Ermakov, S., Estrada, K., Pols, H.A., Williams, F.M., McArdle, W.L., van Meurs, J.B., Loos, R.J., Dermitzakis, E.T., Ahmadi, K.R., Hart, D.J., Ouwehand, W.H., Wareham, N.J., Barroso, I., Sandhu, M.S., Strachan, D.P., Livshits, G., Spector, T.D., Uitterlinden, A.G., and Deloukas, P. 2009. Meta-analysis of genome-wide scans for human adult stature identifies novel Loci and associations with measures of skeletal frame size. *PLoS Genet.* **5**: e1000445. [[Medline](#)] [[CrossRef](#)]
- Takasuga, A. 2016. *PLAG1* and *NCAPG-LCORL* in live-

- stock. *Anim. Sci. J.* **87**: 159–167. [[Medline](#)] [[CrossRef](#)]
22. Tozaki, T., Sato, F., Hill, E.W., Miyake, T., Endo, Y., Kakoi, H., Gawahara, H., Hirota, K., Nakano, Y., Nambo, Y., and Kurosawa, M. 2011. Sequence variants at the myostatin gene locus influence the body composition of Thoroughbred horses. *J. Vet. Med. Sci.* **73**: 1617–1624. [[Medline](#)] [[CrossRef](#)]
 23. Weller, R., Pfau, T., May, S.A., and Wilson, A.M. 2006. Variation in conformation in a cohort of National Hunt racehorses. *Equine Vet. J.* **38**: 616–621. [[Medline](#)] [[Cross-Ref](#)]
 24. Willet, P. 1991. *A History of the General Stud-Book*. Weatherbys, Northants.