

POSTER PRESENTATION

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Partial characterization of two novel monoclonal antibodies for *Listeria* spp. Immunodetection

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From 5th Congress of the Brazilian Biotechnology Society (SBBIOTEC)
Florianópolis, Brazil. 10-14 November 2013

Background

The *Listeria* genus comprises ten different species, however, only two of them - *L. monocytogenes* and *L. ivanovii* - are pathogenic. Although being a well-known pathogen, its detection methodologies are still limited. The gold-standard technique involves enrichment, isolation, and biochemical characterization, which can take about 7 days [1]. In addition, since both pathogenic and non-pathogenic species are present in contaminated food, it is important to detect them both [2]. This way, new methods, such as the immunodetection using monoclonal antibodies (MAbs), have been developed to reduce the detection time of *Listeria* spp.. Recently, our group described a hybridoma clone secreting a MAb of the IgM isotype, called MAb-3F8, that reacts exclusively with an antigen of 30 kDa (P30) present in all *Listeria* spp. evaluated [3]. In the present work, we produced and initially characterized, by ELISA and Western blot, other two novel antibodies against P30 to be used in *Listeria* spp. immunodetection assays.

Materials and methods

A female BALB/c mouse was initially inoculated intraperitoneally with 100 µg of recombinant P30 (rP30) with complete Freund's Adjuvant (FA). Fifteen days after the first injection, a series of 13 doses of rP30 with incomplete FA were performed weekly. Then, the mouse splenocytes were obtained and further fused with Sp2/O cells. The resulting hybridomas were cultured and screened by ELISA using both rP30 and formalin-killed *L. monocytogenes* strain ATCC 19117 (Lm19117) as antigens. Thereby, two hybridomas were selected and used to produce ascites in mice [4]. MAbs isotyping kit (Sigma Aldrich) was used to determine the isotype of each MAb. For the ELISA, rP30 and formalin-killed Lm19117 and *L. innocua* (Linn) were used as antigens.

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For the Western blot (WB), Lm19117 cell-wall fraction [5] and rP30 were transferred to a PVDF membrane. In both assays, the ascites and anti-mouse/peroxidase (Sigma Aldrich) were primary and secondary antibodies, respectively. In addition, 3F8 (positive control) and a non-immunized mouse serum (negative control) were used. Absorbance results at least 3 times higher than the negatives were considered positives.

Results and conclusions

The isotyping assay determined that one MAb was IgG1 (MAb-G2A) and the other, IgM (MAb-M5A). In the ELISA, MAb-G2A reacted with rP30 ($OD_{450}=0.346$), but not with Lm19117 ($OD_{450}=0.020$) or Linn ($OD_{450}=0.018$). However, M5A reacted with rP30 ($OD_{450}=0.160$), Lm19117 ($OD_{450}=0.229$), and Linn ($OD_{450}=0.199$). Negatives for rP30, Lm, and Linn were 0.026, 0.048, and 0.062, respectively; while the positives were 0.418, 0.237, and 0.211, respectively. In the WB, the same pattern seem to occur with MAb-G2A, since it reacted only with rP30; while MAb-M5A presented no reaction. The fact that M5A had no reaction in WB, but showed promising results in ELISA indicate this MAb binds to a conformational epitope of P30 that is displayed on *Listeria* surface. Considering this, further studies will be conducted with a larger panel of bacteria to better characterize these MAbs and determine their sensibility and specificity in detecting *Listeria* spp. in food samples.

Acknowledgements

We would like to thanks CAPES, CNPq and FAPERGS for the financial support and scholarships.

Published: 1 October 2014

References

- Gasanov U, Hughes D, Hansbro PM: Methods for the isolation and identification of *Listeria* spp. and *Listeria monocytogenes*: a review. *FEMS microbiology reviews* 2005, 29:851-75.

2. Besse NG, Barre L, Buhariwalla C, Vignaud ML, Khamissi E, Decourseulles E, Nirsimloo M, Chelly M, Kalmokoff M: **The overgrowth of Listeria monocytogenes by other Listeria spp. in food samples undergoing enrichment cultivation has a nutritional basis.** *International journal of food microbiology* 2010, 136:345-351.
3. Mendonça M, Conrad NL, Conceição FR, Moreira AN, da Silva WP, Aleixo JAG, Bhunia AK: **Highly specific fiber optic immunosensor coupled with immunomagnetic separation for detection of low levels of Listeria monocytogenes and L. ivanovii.** *BMC microbiology* 2012, 12:275.
4. Misra SK, Milohanic E, Aké F, Mijakovic I, Deutscher J, Monnet V, Henry C: **Analysis of the serine/threonine/tyrosine phosphoproteome of the pathogenic bacterium Listeria monocytogenes reveals phosphorylated proteins related to virulence.** *Proteomics* 2011, 11:4155-4165.
5. Harlow E, Lane D: **Antibodies: A Laboratory Manual.** New York: Cold Spring Harbor; 1988.

doi:10.1186/1753-6561-8-S4-P64

Cite this article as: Moreira et al.: Partial characterization of two novel monoclonal antibodies for *Listeria* spp. Immunodetection. *BMC Proceedings* 2014 8(Suppl 4):P64.

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