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*CORRESPONDENCE Xiaolan Zhang xlzhang_hebmu@163.com

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Construction of a prognostic model of colon cancer patients based on metabolismrelated lncRNAs

Chenyang Li, Qian Liu, Yiran Song, Wenxin Wang and Xiaolan Zhang*

The Department of Gastroenterology and Hepatology, The Second Hospital of Hebei Medical University, Shijiazhuang, China

Background: Many studies have shown that metabolism-related lncRNAs may play an important role in the pathogenesis of colon cancer. In this study, a prognostic model for colon cancer patients was constructed based on metabolism-related lncRNAs.

Methods: Both transcriptome data and clinical data of colon cancer patients were downloaded from the TCGA database, and metabolism-related genes were downloaded from the GSEA database. Through differential expression analysis and Pearson correlation analysis, long non-coding RNAs (IncRNAs) related to colon cancer metabolism were obtained. CRC patients were divided into training set and verification set at the ratio of 2:1. Based on the training set, univariate Cox regression analysis was utilized to determine the prognostic differential expression of metabolic-related lncRNAs. The Optimal lncRNAs were obtain by Lasso regression analysis, and a risk model was built to predict the prognosis of CRC patients. Meanwhile, patients were divided into high-risk and low-risk groups and a survival curve was drawn accordingly to determine whether the survival rate differs between the two groups. At the same time, subgroup analysis evaluated the predictive performance of the model. We combined clinical indicators with independent prognostic significance and risk scores to construct a nomogram. C index and the calibration curve, DCA clinical decision curve and ROC curve were obtained as well. The above results were all verified using the validation set. Finally, based on the CIBERSORT analysis method, the correlation between lncRNAs and 22 tumor-infiltrated lymphocytes was explored.

Results: By difference analysis, 2491 differential lncRNAs were obtained, of which 226 were metabolic-related lncRNAs. Based on Cox regression analysis and Lasso results, a multi-factor prognostic risk prediction model with 13 lncRNAs was constructed. Survival curve results suggested that patients with high scores and have a poorer prognosis than patients with low scores (P<0.05). The area under the ROC curve (AUC) for the 3-year survival and 5-year survival were 0.768 and 0.735, respectively. Cox regression analysis showed that age, distant metastasis and risk scores can be used as

independent prognostic factors. Then, a nomogram including age, distant metastasis and risk scores was built. The C index was 0.743, and the ROC curve was drawn to obtain the AUC of the 3-year survival and the 5-year survival, which were 0.802 and 0.832, respectively. The above results indicated that the nomogram has a good predictive effect. Enrichment analysis of KEGG pathway revealed that differential lncRNAs may be related to chemokines, amino acid and sugar metabolism, NOD-like receptor and Toll-like receptor activation as well as other pathways. Finally, the analysis results based on the CIBERSORT algorithm showed that the lncRNAs used to construct the model had a strong polarized correlation with B cells, CD8+T cells and M0 macrophages.

Conclusion: 13 metabolic-related lncRNAs affecting the prognosis of CRC were screened by bioinformatics methods, and a prognostic risk model was constructed, laying a solid foundation for the research of metabolic-related lncRNAs in CRC.

KEYWORDS

colon cancer, metabolism, IncNRA, LASSO, prognostic model

Background

Clorectal cancer (CRC) is one of the most common gastrointestinal malignancies in the world, whose morbidity and mortality are increasing year by year. In 2018, China's CRC accounted for the third in the national incidence of malignant tumors and the second in mortality, and the number of new cases and deaths was as high as 376,000 and 191,000, respectively (1). Genetics, living environment, diet and other factors are all risk factors resulting in CRC (2, 3). At present, with the development of medical diagnosis and treatment technology, tumor molecular target and biological therapy were set as the entry point, searching for target proteins and regulatory signal pathways related to tumor occurrence and development, so as to achieve better effects.

Tumor metabolism plays a key role in the occurrence and development of tumors, including sugar metabolism, nucleic acid metabolism, enzyme metabolism, and protein metabolism (4–7). Studies have shown that tumor metabolism has a significant correlation with tumor prognosis. Long non-coding RNAs (lncRNAs) are RNAs with a transcription length between 200 and 100 000 nt, which do not encode proteins themselves but participate in many physiological processes. Recent studies have shown that metabolism-related lncRNAs are related to the prognosis of a variety of cancers, including breast cancer and lung cancer (8, 9). However, the role of metabolism and related lncRNAs in CRC still remains unclear.

The study used bioinformatics methods to analyze the correlation between differential lncRNAs and metabolic genes, and obtain differentially expressed metabolic-related lncRNAs. Furthermore, we used LASSO regression analysis to construct a risk model to obtain metabolic-related lncRNAs that affect the prognosis of CRC, used single and multi-factor Cox analysis to construct a prognostic risk prediction nomogram model, and evaluated the correlation between genes and clinicopathological characteristicsin the model, providing a new idea for the prognosis of CRC.

Materials and methods

Data resources

The colon cancer-related data and metabolism-related data in this study were from the Cancer Genome Atlas (TCGA) (https://cancergenome.nih.gov) database and the Gene Set Enrichment Analysis database (GSEA, https://www.gseamsigdb.org/gsea/index.jsp), respectively.

Differential expression analysis

Limma package in the R software was used to perform differential analysis, and the differentially expressed

metabolism-related genes and lncRNA in colon cancer tissues and adjacent tissues were explored in TCGA, with | log2FC |>1 and the false discovery rate was set as FDR< 0.05 for screening criteria. Pearson correlation analysis was used to calculate the correlation coefficient (R2) between lncRNAs and metabolismrelated genes. The lncRNAs with R2>0.5 and p<0.05 were defined as metabolic-related lncRNAs.

Construction of the prognostic model

The colon cancer patients were divided into the training set and validation set at the ratio of 2:1. Based on the training set, the singlefactor Cox regression analysis of the metabolic-related lncRNAs was performed to obtain the lncRNAs related to the overall survival. P<0.05 was considered as statistically significant and was included in the LASSO regression analysis to determine the closely related IncRNAs. Multi-factor Cox regression generated risk coefficients and a risk regression model was then constructed. The risk scores of different patients were calculated. The prognostic risk score formula was constructed as follows: Risk score $\sum_{n=1}^{n} coefficie$ $nts \times Expression$ of metabolic – related lncRNAs(i) = . According to the median value of the risk score., the patients were divided into high-risk and low-risk groups, and the survival curve was used to compare the prognosis of the two groups for verification. The prediction effect of the area under the curve (AUC) evaluation model was analyzed by ROC curve analysis, all the above analyses were verified in the verification set.

Clinical subgroup analysis

The survival package in R was applied for the subgroup analysis of lncRNAs risk model combined with the clinical subgroup characteristics of patients, such as age, gender, TNM staging. The ability of the risk model to distinguish high and low risk patients in different subgroups was clarified.

Construction and evaluation of the nomogram

Univariate and multivariate Cox regression analysis were used to analyze risk scores and clinical factors including age, gender, tumor stage and TNM stage and screened independent prognostic factors. We built a nomogram based on the results of multivariate Cox regression including risk scores. The 3-year and 5-year OS for each patient were predicted based on the nomogram. At the same time, the C index, calibration curve and ROC curve were generated to evaluate the prediction effect of the model. The above results are all verified in the validation set to verify the stability of the results.

GSEA enrichment analysis

KEGG analysis was performed on the selected metabolicrelated prognostic lncRNAs based on the clusterProfiler package in R using GSEA software to identify genes with rich function and classify gene clusters (P<0.05).

Correlation analysis between prognostic IncRNAs and tumor immunity

CIBERSORT is a tool for deconvolving the expression matrix of human immune cell subtypes based on the principle of linear support vector regression. The CIBERSORT analysis method (10, 11) was used to clarify the level of infiltration of 22 immune cells in CRC patients, then based on correlation analysis, the correlation of 13 prognostic metabolism-related lncRNAs with immune cell infiltration was determined, and the mechanism of prognostic lncRNAs affecting the progression of CRC was explored.

CRC tissue sample collection

A total of 12 pairs of CRC tissues and noncancerous adjacent tissues were collected from patients who had undergone surgical resection at the Second Hospital of Hebei Medical University (Shijiazhuang, China). All patients had signed informed consent. This study was approved by the Ethical Review Committee of the Second Hospital of Hebei Medical University and was conducted in accordance with accepted ethical guidelines.

RNA extraction and qRT-PCR analysis

Total tissue RNA was extracted using Trizol reagent (Vazyme, Nanjing, China) following the manufacturer's protocols. Then, RNA samples were reverse transcribed by Hiscript III Reverse Transcriptase kit (Vazyme, Nanjing, China) and corresponding RNA expression was evaluated by qRT-PCR with ChamQTM Universal SYBR qPCR Master Mix kit (Vazyme, Nanjing, China). GAPDH acted as the internal reference for normalization.

Statistical analysis

The analysis in this study was completed by R software version 3.6.2, in which the limma package was used for differential gene acquisition, cluster Profiler and org.Hs.eg.db package were used for functional enrichment analysis, and

survival package was used to perform Kaplan-Meie survival analysis.

Results

Differential expression analysis and correlation analysis to obtain metabolicrelated IncRNAs

The expression data of 473 colon cancer tissues and 41 normal colon tissues was downloaded from the TGCA database, and the metabolism-related genes were obtained from the FerrDb database. A total of 279 differentially expressed metabolism-related genes were obtained, of which 151 genes were down-regulated and 128 genes were up-regulated (Figure 1; Supplementary Materials Table 1). In addition, a total of 2491 differentially expressed lncRNAs were identified, of which 675 lncRNAs were downregulated and 1816 lncRNAs were up-regulated (Figure 1; Supplementary Materials Table 2). In addition, according to Pearson correlation analysis, 226 metabolic-related lncRNAs were obtained, and the standards were set as |R2|>0.5 and p<0.05 (Supplementary Materials Table 3).

Construction of prognostic risk prediction model

Single-factor Cox regression analysis was performed on the selected metabolic-related lncRNAs, and a total of 20 lncRNAs

related to prognosis were found. The 20 lncRNAs were included in the LASSO regression analysis to obtain the 13 most critical metabolic-related lncRNAs (Figure 2), and the multi-factor COX regression analysis was used to construct a prognostic risk prediction model based on the 13 metabolic-related lncRNAs (risk score $\sum_{i=1}^{i} (coef * exp) =$)(Supplementary Materials Table 4). According to the median risk score, patients were divided into high-risk groups (n=136) and low-risk groups (n=137). The survival curve showed that the OS of the low-risk group was significantly higher than that of the high-risk group (p<0.001, Figure 3). The scatter plot and risk curve revealed that compared with the high-risk group, the low-risk group has lower risk factors and mortality (Figure 3B). The heat map showed the expression levels of 13 lncRNAs between the high-risk group and the low-risk group (Figure 3). The AUC of the ROC curve for 3-year and 5-year survival were 0.768 and 0.735, respectively (Figure 3). Similar results were obtained in the validation set (Figure 3J).

Subgroup survival analysis

The results of subgroup analysis showed that the risk score model in the subgroups of age greater than 65, younger than 65, female, male, T1-2, T3-4, N1-2, M0, and stage III-IV, the OS of the low-risk group was significantly higher than that of the highrisk group, and the difference was statistically significant (P<0.05). However, there was no statistical difference in OS between the high-risk and low-risk groups in the N0, M1, and stage I-II-subgroups (Figure 4).



FIGURE 1

The volcano map highlighted the differentially expressed metabolism-related genes and IncRNAs in colon cancer tissues and surrounding tissues. Red showed a considerable up-regulation, green represented a significant down-regulation, and black represented no significant variation in gene expression. (A) The volcano map and heat map demonstrated the differentially expressed metabolism-related genes (| log2FC |>1, FDR< 0.05). (B) The volcano map and heat map demonstrated differentially expressed lncRNAs (| log2FC |>1, FDR< 0.05)



FIGURE 2

Identify metabolic-related IncRNAs having prognostic significance for colon cancer. (A) COX forest plot demonstrated that 20 metabolic-related IncRNAs were substantially correlated with the OS of colon cancer (P< 0.05). (B, C) The adjustment parameters of the LASSO regression model, LASSO coefficient spectrum of prognostic-related IncRNAs, 10-fold cross-validation to filtrate candidate necroptosisrelated IncRNAs in LASSO regression analysis.

Clarify independent prognostic factors

Univariate Cox regression analysis showed that age, TNM stage, and risk score were related to the patient's prognosis (P<0.05) (Figure 5). However, results of multivariate Cox regression showed that only age, M staging and risk score were closely related to the prognosis of CRC patients and were independent prognostic factors (Figure 5).

Construction of the Nomogram

The Nomogram was constructed based on independent prognostic factors determined by the multivariate Cox regression results, such as age, M, and risk score (Figure 6). The C index of the nomogram in the training set were 0.823, the AUC of the 3-year and 5-year OS were 0.802 and 0.832, respectively (Figure 6). The results of the calibration curve (Figure 6), and the clinical decision curve (Figure 6) showed the model had good predictive performance and can bring benefits to patients. The above results were all verified in the internal validation set to verify the stability of the results (Figures 6C, E).

KEGG pathway enrichment analysis

The KEGG pathway enrichment analysis of the differentially expressed metabolic-related lncRNAs showed that differential lncRNAs were mainly related to the following pathways: chemokine pathways, amino acid and sugar metabolism, NOD-like receptors and Toll-like receptor activation (Figure 7).

Correlation analysis between prognostic lncRNAs and immune cell infiltration

The CIBERSORT-based analysis method clarified the infiltration level of 22 immune cells in colon cancer patients, and the results showed that lncRNAs used to construct the model had a strong correlation with the polarization of B cells, CD8+ T cells and M0 macrophages (Figure 8), indicating that these lncRNAs may regulate tumor growth and progression by affecting the tumor immune infiltration microenvironment.



Construction of the risk scoring model. (A, F) Kaplan-survival Meier's study of colon cancer patients in the training set and validation set indicated that the prognosis of the high-risk group was considerably poorer, while the low-risk group was better. (B, G) The survival rate and survival status of colon cancer patients in the training set and validation set. (C, H) The distribution of risk scores of 13 lncRNAs for colon cancer patients in the training and validation set. (D, I) Heat maps of 13 lncRNAs in the low-risk and high-risk group in the training set and validation set. (E, J) ROC curve analysis of colon cancer patients in the training set and suldation set. (E, J) ROC curve analysis of colon cancer patients in the training set and 0.735, respectively. The AUC values of the 3-year survival rates of the validation set were 0.768 and 0.735, respectively. The AUC values of the 3-year survival rates of the validation set were 0.768 and 0.735, respectively.

Validation of metabolic-related lncRNAs expression in tissue samples

The expression levels of selected metabolic-related lncRNAs were further evaluated and validated in tissues. As illustrated in Figure 9, the expression levels of LINC01703, AL139384.1, LINC00858, LINC01876, AC008760.1, AL590483.1, CASC9, AP001469.3, ZKSCAN2.DT, AP006621.4 and AC010973.2 were significantly higher in CRC tissue samples, while LINC01559 and LINC01133 were significantly lower in CRC tissue samples. The detailed sequence of primers used were listed in Supplementary Table 5. These results further confirmed the correctness of the above bioinformatics analyses.



groups in the age \leq 65 subgroup. (B) Survival curves of the high-risk and low-risk groups in the age>65 subgroup. (C) Survival curves of the high-risk and low-risk groups in the female subgroup. (D) Survival curves of the high-risk and low-risk groups in the female subgroup. (D) Survival curves of the high-risk and low-risk groups in the T1-2 subgroup. (F) Survival curves of the high-risk and low-risk groups in the T3-4 subgroup. (G) Survival curves of the high-risk and low-risk groups in the T3-4 subgroup. (G) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (H) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the N1-2 subgroup. (I) Survival curves of the high-risk and low-risk groups in the StageII-II subgroup. (L) Survival curves of the high-risk and low-risk groups in the stageII-II subgroup. (L) Survival curves of the high-risk and low-risk groups in the stageII-II subgroup.

Discussion

How to predict the prognosis of colon cancer is an urgent problem for gastrointestinal surgeons. With the development of clinical diagnosis and treatment, some prognostic factors have been discovered, including tumor size, tumor grade and stage. High-throughput biological technology has been widely used to predict cancer recurrence and tumor metastasis by detecting lncRNA or gene changes. In recent years, many studies have shown that changes in the expression patterns of metabolismrelated genes are closely related to the occurrence and prognosis of colon cancer [13]. In-depth study of metabolism-related



The predictive usefulness of clinicopathological features and risk ratings. (A) Univariate Cox regression of colon cancer patients. (B) Multivariate Cox regression of colon cancer patients.



validation set.

lncRNAs is expected to provide guidance for clinical decision-making.

Colon cancer is a common malignant tumor in the digestive system, which seriously affects patients' survival (12, 13). How to predict the prognosis of colon cancer is an urgent problem for gastrointestinal surgeons. With the development of clinical diagnosis and treatment, some prognostic factors have been discovered, including tumor size, tumor grade and stage. Highthroughput biological technology has been widely used to predict cancer and tumor metastasis by detecting lncRNAs or gene changes. In recent years, many studies have shown that changes in the expression patterns of metabolism-related genes are closely related to the occurrence and prognosis of colon cancer (14). To study metabolism-related lncRNAs in depth is expected to provide guidance for clinical decision-making.

This study identified 13 metabolic lncRNAs related to the prognosis of CRC patients through bioinformatics methods, namely LINC01703, LINC01559, AC083880.1, AC027796.4, AL139384.1, LINC00858, LINC01876, AC008760.1, AC006329.1 AL590483.1, AC073283.1, CASC9, AP001469.3,

CAPN10-DT, ZKSCAN2-DT, AP006621.4, AC074117.1, LINC01133, AL161729.4 and AC010973.2, thereby building a risk model for predicting prognosis. Combined with clinicopathological analysis, this model can be used as an independent predictor for the prognosis of lung cancer.

Among them, LINC01703 has been reported to be of great significance in the diagnosis of lung adenocarcinoma. Wang et al. found that LINC01703 enhanced the invasiveness of NSCLC cells by changing miR-605-3p/MACC1 (15). In addition, the bioinformatics analysis of the sequencing data of lung adenocarcinoma showed that LINC01703 and AC074117.1 can be used as potential diagnostic biomarkers for lung adenocarcinoma (16, 17). LINC01559 played an important role in gastrointestinal tumors such as pancreatic cancer, gastric cancer and liver cancer (18–21). AC083880.1 was also an important lncRNA, which was related to the activation of autophagy-related pathways in ovarian cancer (22). AC027796.4 was regarded as a potential biomarker for prostate cancer (23). As a ceRNA, LINC01133 regulated APC expression and Wnt/β-catenin pathway by releasing miR-106a-3p to inhibit gastric



GSEA pathway enrichment analysis of metabolism-related lncRNAs. (A) Chemokine signaling pathway. (B) Histidine metabolism. (C) NOD- like receptor signaling pathway. (D) Pentose phosphate pathway. (E) Peroxisome. (F) Toll like receptor signaling pathway.





cancer progression (23). LINC00858 regulated PAK2 signaling pathway through sponge adsorption of miR-4766-5p to promote the progression of colorectal cancer (24). Studies have confirmed that AC008760.1, AC006329.1, AL590483.1, CASC9 and AC010973.2 (25) can be potential biomarkers for colorectal cancer (25–28), while there are few studies about AL139384.1, LINC01876, AP001469.3, CAPN10-DT, ZKSCAN2-DT, AP006621.4, and AL161729.4 in solid tumors.

Pathway enrichment analysis revealed that metabolismrelated lncRNAs were mainly related to pathways such as chemokine pathways, metabolic collaterals, NOD-like receptors and Toll-like receptor activation. Liu et al. (29) found that the activity of gastric cancer cells decreased after lncRNA and UCA-1 were knocked out, in which the chemokine pathway played an important role. Besides, Satoh et al. (30) found that LINC01876 can up-regulate chemokine (C-C motif) ligand 2, and promoted the proliferation of macrophages and myeloid-derived suppressor cells in hepatocellular carcinoma cell lines. While in colorectal cancer, CASC9 interacted with CPSF3 to regulate TGF- β signal transduction (31). LINC01133 inhibited epithelial-mesenchymal transition and metastasis of colorectal cancer by interacting with SRSF6 LINC01133 (32). At present, there are few research on signal pathways related to amino acid and sugar metabolism.

A large number of studies have shown that there is a significant correlation between tumor cell metabolism and immune cell infiltration in tumor tissues (33–35). In order to further reveal the role of 13 lncRNAs in colon cancer, we analyzed the correlation between 13 prognostic-related lncRNAs in the expression matrix of the colon cancer and 22 kinds of immune cell infiltration based on the CIBERSORT algorithm, whose results showed that these 13 lncRNAs showed a strong correlation with neutrophils, follicular helper T cells

and polarized M2 macrophages, suggesting that they may pose impact on the prognosis of colon cancer by affecting the tumor immune microenvironment. The specific regulatory mechanism remains to be studied in depth.

Conclusions

The relationship between metabolic-related lncRNAs and the prognosis of CRC was preliminarily explored through bioinformatics methods, a model based on 13 metabolicrelated lncRNAs was established to predict the prognosis of CRC, and the stability of the model in multiple data sets was fully verified. It provided a new direction for the study of metabolism and colon cancer. We still need further *in vitro* and *in vivo* studies to clarify the role and specific mechanisms of metabolism-related lncRNAs in the occurrence and development of colon cancer.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Author contributions

XZ designed the experiments. CL performed the analysis. CL and QL analyzed the TCGA data. CL, YS and WW wrote and reviewed the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2018) 686:394–424. doi: 10.3322/caac.21492

2. Arredondo EM, Dumbauld J, Milla M, et al. A promotor-led pilot study to increase colorectal cancer screening in latinos: The juntos contra el cancer program. *Health Promot Pract* (2021) 224:491–501. doi: 10.1177/1524839920912240

3. Connell LC, Mota JM, Braghiroli MI, Hoff PM. The rising incidence of younger patients with colorectal cancer: Questions about screening, biology, and treatment. *Curr Treat Options Oncol* (2017) 184:23. doi: 10.1007/s11864-017-0463-3

4. Kuo CY, Ann DK. When fats commit crimes: Fatty acid metabolism, cancer stemness and therapeutic resistance. *Cancer Commun (Lond)* (2018) 381:47. doi: 10.1186/s40880-018-0317-9

5. Alfarouk KO. Tumor metabolism, cancer cell transporters, and microenvironmental resistance. *J Enzyme Inhib Med Chem* (2016) 316:859–66. doi: 10.3109/14756366.2016.1140753

6. Alfarouk KO, Verduzco D, Rauch C, et al. Erratum: Glycolysis, tumor metabolism, cancer growth and dissemination. a new ph-based etiopathogenic perspective and therapeutic approach to an old cancer question. *Oncoscience* (2015) 24:317. doi: 10.18632/oncoscience.158

7. Lee WR, Ishikawa T, Umetani M. The interaction between metabolism, cancer and cardiovascular disease, connected by 27-hydroxycholesterol. *Clin Lipidol* (2014) 96:617–24. doi: 10.2217/clp.14.53

8. Lin W, Zhou Q, Wang CQ, et al. Lncrnas regulate metabolism in cancer. Int J Biol Sci (2020) 167:1194–206. doi: 10.7150/ijbs.40769

9. Amelio I, Bernassola F, Candi E. Emerging roles of long non-coding rnas in breast cancer biology and management. *Semin Cancer Biol* (2021) 72:36–45. doi: 10.1016/j.semcancer.2020.06.019

10. Chen B, Khodadoust MS, Liu CL, Newman AM, Alizadeh AA. Profiling tumor infiltrating immune cells with cibersort. *Methods Mol Biol* (2018) 1711:243–59. doi: 10.1007/978-1-4939-7493-1_12

11. Newman AM, Liu CL, Green MR, et al. Robust enumeration of cell subsets from tissue expression profiles. *Nat Methods* (2015) 125:453–7. doi: 10.1038/ nmeth.3337

12. Mujagic Z, Masclee A, Keszthelyi D. Lymphangiomatosis of the colon. Clin Gastroenterol Hepatol (2020) 20:14–5. doi: 10.1016/j.cgh.2020.08.037

13. Tomkovich S, Dejea CM, Winglee K, et al. Human colon mucosal biofilms from healthy or colon cancer hosts are carcinogenic. *J Clin Invest* (2019) 1294:1699–712. doi: 10.1172/JCI124196

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fonc.2022.944476/full#supplementary-material

SUPPLEMENTARY TABLE 1

A total of 279 differentially expressed metabolism-related genes were obtained, of which 151 genes were down-regulated and 128 genes were up-regulated.

SUPPLEMENTARY TABLE 2

A total of 2491 differentially expressed lncRNAs were identified, of which 675 lncRNAs were down-regulated and 1816 lncRNAs were up-regulated.

SUPPLEMENTARY TABLE 3

According to Pearson correlation analysis, 226 metabolic-related lncRNAs were obtained, and the standards were set as |R2|>0.5 and p<0.05.

SUPPLEMENTARY TABLE 4

Multi-factor COX regression analysis was used to construct a prognostic risk prediction model based on the 13 metabolic-related lncRNAs.

SUPPLEMENTARY TABLE 5

Detailed sequences of primers used in qRT-PCR experiments.

14. Sirven MA, Venancio VP, Shankar S, et al. Ulcerative colitis results in differential metabolism of cranberry polyphenols by the colon microbiome *in vitro*. *Food Funct* (2021) 12:12751–64. doi: 10.1039/D1FO03047G

15. Wang Z, Zhang X, Zhang and X Jiang X. Long noncoding rna linc01703 exacerbates the malignant properties of non-small-cell lung cancer by upregulating macc1 in a microrna-605-3p-mediated manner. *Oncol Res* (2021) 28:913–27. doi: 10.3727/096504021X16310057751016

16. Wang Y, Fu and Z Wang J, et al. Screening key lncrnas for human lung adenocarcinoma based on machine learning and weighted gene co-expression network analysis. *Cancer biomark* (2019) 254:313–24. doi: 10.3233/CBM-190225

17. Li M, Yang and X Li B, et al. Identification of prognostic factors related to super enhancer-regulated cerna network in metastatic lung adenocarcinoma. *Int J Gen Med* (2021) 14:6261–75. doi: 10.2147/IJGM.S332317

18. Deng Z $\,$, Li and Y Shi X, et al. A novel autophagy-related incrnas signature for prognostic prediction and clinical value in patients with pancreatic cancer. Front Cell Dev Biol (2020) 8:606817. doi: 10.3389/fcell.2020.606817

19. Wang L, Bo X, Yi X, et al. Exosome-transferred linc01559 promotes the progression of gastric cancer *via* pi3k/akt signaling pathway. *Cell Death Dis* (2020) 119:723. doi: 10.1038/s41419-020-02810-5

20. Lou C, Zhao J, Gu Y, et al. Linc01559 accelerates pancreatic cancer cell proliferation and migration through yap-mediated pathway. *J Cell Physiol* (2020) 2354:3928–38. doi: 10.1002/jcp.29288

21. Dong S, Fu Y, Yang K, et al. Linc01559 served as a potential oncogene and promoted resistance of hepatocellular carcinoma to oxaliplatin by directly sponging mir-6783-3p. *Anticancer Agents Med Chem* (2021) 212:278-86. doi: 10.2174/1871520620666200721122317

22. Meng C, Zhou JQ, Liao YS. Autophagy-related long non-coding rna signature for ovarian cancer. J Int Med Res (2020) 4811:1220770313. doi: 10.1177/0300060520970761

23. Wang Y, Wang Y, Feiyao D, et al. Bioinformatic analysis of immune-related lncrna based on tcga database in patients with prostate cancer. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi* (2021) 372:113–8. doi: 10.13423/j.cnki.cjcmi.009142

24. Zhan W, Liao X, Chen Z, et al. Linc00858 promotes colorectal cancer by sponging mir-4766-5p to regulate pak2. *Cell Biol Toxicol* (2020) 364:333–47. doi: 10.1007/s10565-019-09506-3

25. Cai HJ, Zhuang ZC, Wu Y, et al. Development and validation of a ferroptosis-related lncrnas prognosis signature in colon cancer. *Bosn J Basic Med Sci* (2021) 215:569–76. doi: 10.17305/bjbms.2020.5617

26. Wei J, Ge X, Tang Y, et al. An autophagy-related long noncoding rna signature contributes to poor prognosis in colorectal cancer. J Oncol (2020) 2020:4728947. doi: 10.1155/2020/4728947

27. Ghafouri-Fard S, Asadi M, Sohrabi B, et al. Down-regulation of a panel of immune-related lncrnas in breast cancer. *Pathol Res Pract* (2021) 224:153534. doi: 10.1016/j.prp.2021.153534

28. Sun Y, Peng P, He L, Gao X. Identification of lnc rnas related to prognosis of patients with colorectal cancer. *Technol Cancer Res Treat* (2020) 19:1079229768. doi: 10.1177/1533033820962120

29. Qin L, Jia Z, Xie D, Liu Z. Knockdown of long noncoding rna urothelial carcinoma-associated 1 inhibits cell viability, migration, and invasion by regulating microrna-182 in gastric carcinoma. *J Cell Biochem* (2018) 11912:10075–86. doi: 10.1002/jcb.27344

30. Fujisaka Y, Iwata T, Tamai K, et al. Long non-coding rna hotair up-regulates chemokine (c-c motif) ligand 2 and promotes proliferation of macrophages and myeloid-derived suppressor cells in hepatocellular carcinoma cell lines. *Oncol Lett* (2018) 151:509–14. doi: 10.3892/ol.2017.7322

31. Ding Y, Li X, Zhang Y, Zhang J. Long non-coding rna cancer susceptibility 9 (casc9) up-regulates the expression of erbb2 by inhibiting mir-193a-5p in colorectal cancer. *Cancer Manag Res* (2020) 12:1281–92. doi: 10.2147/CMAR.S234620

32. Kong J, Sun W, C Li, et al. Long non-coding rna linc01133 inhibits epithelial-mesenchymal transition and metastasis in colorectal cancer by interacting with srsf6. *Cancer Lett* (2016) 3802:476-84. doi: 10.1016/j.canlet.2016.07.015

33. Patel CH, Leone RD, Horton MR, Powell JD. Targeting metabolism to regulate immune responses in autoimmunity and cancer. *Nat Rev Drug Discovery* (2019) 189:669–88. doi: 10.1038/s41573-019-0032-5

34. Xia L, Oyang L, Lin J, et al. The cancer metabolic reprogramming and immune response. *Mol Cancer* (2021) 201:28. doi: 10.1186/s12943-021-01316-8

35. Ganeshan K, Chawla A. Metabolic regulation of immune responses. Annu Rev Immunol (2014) 32:609–34. doi: 10.1146/annurev-immunol-032713-120236