Distribution of pimonidazole and RSU 1069 in tumour and normal tissues

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Summary The tritium-labelled analogues of pimonidazole and RSU 1069 were injected into mice bearing the KHT murine sarcoma which has a hypoxic cell fraction of $\sim 10\%$. The distribution of activity at 24 h was recorded using autoradiography and measurement of tissue activity. Autoradiographs with both drugs showed high activity in particular cells within tumour, eye (melanin-associated cells), eyelid (Meibomian gland), liver (centrilobular area), skin (sebaceous gland and melanin), stomach (squamous area), footpad, oesophagus, labial gland, Zymbal's gland, preputial gland, parotid gland (intralobular ducts) and airway epithelium. These tissues had previously been identified as sites of binding of misonidazole. The measurement of total tissue radioactivity showed significantly higher activity in liver, eyelid (Meibomian gland), oesophageal lining, kidney and labial gland than was found in the tumour.

Misonidazole (1-(2-nitro-1-imidazolyl)-3-methoxy-2-propanol; MISO) sensitises hypoxic tumour cells to ionising radiation (Asquith et al., 1974). This radiosensitisation is associated with the electron affinity of MISO and is a fast (subsecond) free-radical process. MISO can also be involved in a separate, slower, enzyme dependent process in which, in the presence of the appropriate reductase(s), the nitro group is reduced yielding cytotoxic species (Varghese et al., 1976; Rauth, 1984). This second process is favoured by a hypoxic environment - such as occurs in parts of many tumours and this leads to locally enhanced retention. The concept of the preferential retention of reactive bioreduced metabolites in tumours has generated two lines of research. One is directed towards tumour therapy (bioreductive drug therapy). In this, drugs are being developed which, as with MISO, on reduction in the hypoxic milieu of the tumour, form cytotoxic metabolites(s) that bind to critical macromolecules such as DNA. The other line of research is in the field of imaging and spectroscopy. The parent compound is labelled with a radionuclide (Rasey et al., 1987), or with an element suitable for magnetic resonance imaging or MR spectroscopy (Raleigh et al., 1986; Maxwell et al., 1989a). The imaging compound is reduced and retained preferentially in the hypoxic milieu of the tumour and is then visualised when the differential between tumour and adjacent tissues is maximal. In most instances it has not yet been possible to identify the metabolite(s) responsible for cytotoxicity or imaging.

Preferential binding of MISO within tumours was first illustrated by Chapman et al. (1981). Carbon-14-labelled MISO was given to tumour-bearing mice which were killed after 3 h and autoradiographs (ARG) of the tumours were prepared. The ARGs showed a dense band of grains in the area between the viable tumour and its necrotic centre. It was assumed that this band of grains represented binding to hypoxic cells in the tumour. Recently, while repeating this work, we have observed similarly high levels of grain count in a number of apparently normoxic normal tissues in mice. These tissues are: those of sebaceous gland origin (Meibomian gland, skin sebaceous gland, Zymbal's gland, labial gland, vulval gland, perianal gland and preputial gland); stratified squamous epithelia (oesophagus, squamous stomach lining, footpad); lung (airway epithelium); liver and enamel organ of the incisor (Cobb et al., 1989; Cobb & Nolan, 1989). When many of these tissues are examined by histochemistry they stain strongly for reductase activity (Cobb et al., 1990a).

Because of the possible relevance of this high level of binding in normal tissues to research in tumour visualisation and bioreductive drug therapy we have extended this work to examine two analogues of misonidazole to see if the phenomenon is specific to MISO or can be observed more widely. Pimonidazole (α-[(2-nitro-1-imidazolyl)methyl]-1-piperidine-ethanol; RO 03-8799; PIMO) is a lipophilic MISO analogue in which the methoxy group has been replaced by a piperidine group (Smithen et al., 1980). It was thought that being weakly basic it might assist in tumour retention and produce a more rapid clearance in acid urine than the parent MISO. The theory that the basic side-chain could lead to improved concentration in the acidic milieu of tumours is supported by the work of Dische et al. (1986a,b) and Roberts et al. (1986) who have observed in patients a tumour concentration of approximately twice that of MISO. This compound has had a limited clinical trial (Saunders et al., 1984; Roberts et al., 1986). RSU 1069 (1-(2-nitro-1-imidazolyl)-3-(1-aziridinyl)-2-propanol) is a MISO analogue in which an aziridine group replaces the methoxy group in the N1 sidechain (Adams et al., 1984). It is thought to act as a monofunctional alkylating agent (aziridine group) under normoxic conditions and under hypoxic conditions as a bifunctional alkylating group through the aziridine group and the reduced nitro group (Stratford et al., 1986). As a hypoxic-cell cytotoxin it is 100 times more toxic to anoxic than to normoxic cells in vitro (Ahmed et al., 1986; Stratford et al., 1986). In a limited clinical trial it has been found to cause the side-effects of nausea and vomiting (Horwich et al., 1986).

In the present study we have recorded the distribution of ³H-PIMO and ³H-RSU 1069 in the tissues of tumour-bearing mice by whole organ measurement of the tritium, and by ARGs of the tissues.

Materials and methods

The ten animals used were male C3H/He mice aged 11-13 weeks. Their pathogen-free status was maintained by the use of positive pressure isolators. The mice had a subcutaneous KHT sarcoma growing on the rump. At the size of tumour used in the present study (6 mm diameter) the radiobiologically hypoxic cell fraction has been calculated as 10% (Stratford *et al.*, 1989).

Both PIMO and RSU 1069 were labelled with tritium on the two carbon of the side-chain (Webb & Threadgill, 1990). The relative specific activities were 18 and 17 MBq mg⁻¹ respectively. For each drug five mice were injected i.v. with a single injection in PBS (pH 7.4); 50 mg kg⁻¹ for PIMO and 40 mg kg⁻¹ for RSU 1069. The i.v. $LD_{50}/7d$ of cold PIMO and RSU 1069 are 580 and 160 mg per kg body weight respectively.

After 24 h all animals were killed by i.p. sodium barbiturate and samples of the following tissues taken from each animal and weighed: eyelids ($\sim 20\%$ of which is Meibomian gland), Zymbal's gland (ear canal: Pohl & Fouts, 1983), preputial gland, liver, oesophageal epithelium, footpads, KHT tumour, lung, muscle (quadriceps), kidney, brain (one

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whole side), spleen, parotid salivary gland, blood, labial gland (lingual surface of cheek). The labial gland is an aggregation of sebaceous glands associated with the hairs in the labial vestibule (Quay, 1965).

Tritiated water was removed from the tissue samples by three cycles of dehydration with ethanol (overnight in the first instance and subsequently for a minimum of 4 h) and rehydration with distilled water. It was expected that this would remove tritiated water and some small water-soluble molecules (Franko *et al.*, 1989). Blood was similarly dehydrated using three cycles of drying/rehydration *in vacuo* over P_20_5 . The samples were then prepared for liquid scintillation counting by dissolution in 1.0 ml of OptiSolve (LKB Scintillation Products (Pharmacia), Milton Keynes, UK) at 55°C for 16 h. Fifteen ml of OptiPhase (LKB Scintillation Products (Pharmacia), Milton Keynes, UK) was added and the samples counted on a Beckmann LS 5000 CE liquid scintillation counter (Beckman Instruments Inc., Fullerton, CA, USA) using the external standard generated H number to estimate the counting efficiency.

The following tissues were taken for ARG: eyelids, eyeball, labial gland, Zymbal's gland, preputial gland, KHT tumour, lung (to include bronchi), liver, oesophagus, muscle (quadriceps), kidney, stomach (squamous and glandular), brain (sagittal section), footpads, spleen, parotid gland and rostral part of the nasal passages. The tissues were processed to $3-5 \mu m$ paraffin sections and dipped in K2 emulsion (Ilford Emulsion, Knutsford, Cheshire, UK) for ARG. Exposure periods of from 1 to 14 weeks were used. The ARGs were developed, stained with haematoxylin and eosin, and examined microscopically to identify the distribution and density of grains in the emulsion overlying the tissue.

Results

Tissue distribution studies

The following results expressed as per cent injected tritium per g wet weight are multiplied by 100 for ease of presentation. For PIMO and RSU 1069 respectively the values $(\pm \text{ s.d.})$ were: KHT tumour 7.8 (0.9), 33.0 (13.0); blood 1.2 (0.6), 33.6 (4.7); footpads 4.3 (1.1), 40.5 (4.1); parotid gland 3.2 (0.8), 21.6 (5.5); eyelid 20.5 (3.9), 171.9 (42.7); Zymbal's gland 3.6 (0.8), 22.8 (2.6); labial gland 5.5 (1.1), 62.8 (15.4); brain 1.3 (0.3), 18.5 (1.4); preputial gland 6.8 (2.9), 27.7 (11.8); liver 20.0 (5.4), 115.1 (25.1); spleen 3.5 (2.2), 43.3 (5.9); kidney 6.5 (1.4), 88.2 (9.9); oesophageal epithelium 34.2 (1.4), 99.4 (35.6); lung 3.1 (1.1), 32.2 (1.1); muscle 1.7 (0.3), 34.9 (3.5). The tissue measurements were then expressed as a ratio to the KHT tumour value and subjected to statistical analysis. All significant values together with a number of non-significant background tissue values are given in Table I.

 Table I
 Activity in various tissues 24 h after injection, expressed as a ratio to the tumour value

	RSU 1069	РІМО	MISO ^a
Liver	3.5 (0.72) ***	2.6 (0.34) ***	3.5 (0.41) ***
Meibomian gland ^b	5.2 (1.12) ***	2.6 (0.26) ***	1.3 (0.47) *
Oesophageal epithelium	3.1 (0.73) ***	4.37 (0.83) ***	9.8 (0.80) ***
Kidney	2.7 (0.51) **	0.83 (0.12)	1.2 (0.35)
Labial gland	1.9 (0.41) *	0.71 (0.07)	n.a.
Blood	1.0 (0.20)	0.16 (0.03)	0.49 (0.30)
Muscle	1.1 (0.20)	0.22 (0.02)	0.38 (0.08)
Spleen	1.3 (0.25)	0.45 (0.13)	0.40 (0.10)

*Values from Cobb *et al.* (1990*b*). These values are for ¹⁴C-labelled MISO injected into mice bearing the A 110 mouse lung tumour. n.a., not available; ^bThe eyelids, of which ~20% is Meibomian gland, were removed and the activity measured. ARGs showed that the Meibomian gland held much the greater part of the retained activity. The statistical significance of the tissue levels above tumour level was calculated using the Mann–Whitney test. *P < 0.05; **P < 0.01; ***P < 0.001. Each compound had been injected into a group of five mice. s.d. in parentheses.

Also in this table are measurements taken from a previous publication on MISO (Cobb et al., 1990b).

Autoradiography

With the exception of cells of macrophage lineage and melanin-containing cells (see below) the tissues at 24 h with noticeably high grain counts in PIMO- and RSU 1069-injected mice were the same as those in which high levels of MISO had previously been observed and measured (Cobb *et al.*, 1989; Cobb & Nolan, 1989). They fell into five distinct tissues or groups of tissues.

1. Sebaceous gland group. This group comprised the sebaceous gland of the hair follicles, the Meibomian gland of the eyelids, Zymbal's gland of the ear canal, the labial gland on the lingual surface of the cheek and the preputial gland. These are histologically similar glands which all exhibit holocrine secretion; that is, the entire mature cell is secreted. Basal cells proliferate and a percentage mature to form the glandular secretion. The maturation process involves a gross enlargement of the cells by lipids, and nuclear degeneration. The radioactivity at 24 h was observed to some extent over the maturing cells and more so those in the process of secretion through the duct(s) to the exterior.

2. Keratinised stratified squamous epithelial cell group. This group comprised skin, oesophageal lining, the squamous area of the stomach lining and the footpads. The grains were concentrated over the stratum granulosum and the deeper parts of the keratinised layer (Figure 1). The highest grain density for both drugs (as for MISO) was in the oesophageal lining. The grain count in the skin of the eyelids, nares and footpads was patchy.

3. Lung. Activity was centred over the epithelium lining the airways. The grain density was clearly less in this tissue than in (1) and (2) above.

4. Liver. For both drugs, as for MISO, there was a high grain density in the cells surrounding the centrilobular veins (zone 3).

5. Parotid gland. As had previously been observed for MISO, PIMO and RSU 1069 showed a higher grain count over the epithelial cells lining the intralobular ducts than in the surrounding tissues.



Figure 1 Junction of glandular with keratinised stratified squamous area of stomach (ARG). The PIMO metabolite 24 h postinjection is localised only in the squamous area, predominantly in the upper cells of the stratum pigmentosum and the adjacent lowest layers of the keratinised cells (arrow heads). One such area is shown inset enlarged $\times 3.5$. At the bottom of the inset are the cells of the basal epithelium. They have a somewhat higher than background grain count; as do the cells of the next layer (stratum spinosum). The highest grain count is over the stratum granulosum and the first few layers of the keratinised cells. Thereafter the count falls away rapidly at the top of the inset. The bar represents 60 μ m.



Figure 2 Area of heavily pigmented skin to show binding of RSU 1069 to melanin (ARG). Twenty-four hours after the injection of ³H-RSU 1069 the ARG shows high activity over cells which are known to contain melanin (arrows). In this illustration only the grains in the ARG emulsion can be seen. The presence of melanin granules beneath required examination at high magnification when the black grains in the emulsion could be seen separately from the underlying pale golden brown melanin granules. The asterisk is in an area of heavy grain density over melanocytes and melanin-impregnated epidermal cells in the lower layers of the epidermis. The arrow head points to heavy grain density above a pigmented hair. The bar represents $60 \, \mu m$.

In addition to the above tissues, PIMO- and RSU 1069injected mice showed high grain density above melanincontaining cells (Figure 2). This was particularly clear in the skin of the eyelids and in the footpads. The melanin was in both melanocytes and adjacent melanin impregnated epithelial cells.

For RSU 1069 but not PIMO the macrophages in the liver (Kupffer cells), spleen and skin (tissue histiocytes) were associated with high grain density, indicative of the accumulation within these cells of a labelled metabolite and there was a high grain count over small cells in the brain which had a scattered distribution similar to that of microglia (macrophage lineage) (Merz *et al.*, 1987).

Discussion

The main points arising from the results are that with the two MISO analogues PIMO and RSU 1069 activity was retained in much the same tissues as had previously been observed for MISO; and that there was activity in some normal tissues significantly in excess of the subcutaneous tumour.

The tissue samples were dehydrated because there is good evidence that the tritium associated with MISO tissue levels at 24 h is to a significant extent due to tritiated water (Franko et al., 1989; Cobb et al., 1990b). In the absence of any other evidence we have concluded that the same might apply to the two analogues of MISO and that with the tritiated water removed we believed that the activity was likely to be associated predominantly with bound metabolites of the two drugs. This concept is supported by the close correlation of high tritium retention with high grain count in the target tissues: sebaceous gland group, stratified squamous epithelium (oesophagus) and liver. In a recent publication on MISO we have pointed out that these tissues have in common high levels of reductase and we have suggested that, despite the likely presence of significant amounts of oxygen, nitroimidazoles are reduced to reactive, binding, metabolite(s) in these tissues (Cobb et al., 1990a). The same may well apply to PIMO and RSU 1069.

The levels of radioactivity in the various tissues were measured at only one point in time (24 h) and it is possible that the ratio of one tissue level to another could change with time. However, the ARG grain density, which would reflect binding by reactive metabolites in the first few hours was high in tissues showing high tissue to tumour ratios at 24 h (Table I) thus indicating that the measurements of tissue activity at 24 h were probably not anomalous.

The high level of MISO retained by the liver has frequently been commented upon. Garrecht and Chapman (1983) injected ¹⁴C-MISO into mice bearing the EMT-6 tumour and reported high levels of adduct in liver and tumour as well as lower levels in all normal tissues sampled. The two drugs in the present report showed a similar high liver retention, with preferential localisation in the centrilobular zone (zone 3). Recently Maxwell *et al.* (1989*a,b*) and previously Van Os-Corby *et al.* (1987) have pointed to low oxygen tension being the likely cause of this retention for MISO. While this may be so there is also a higher reductase level in zone 3 which might contribute significantly to the localisation of all three drugs (Pette & Brandau, 1966; Cobb *et al.*, 1990*a*).

If it is true that high local reductase activity can outweigh the ability of oxygen to minimise the reduction of nitroimidazole by futile cycling there are clinical implications. For example, bioreductive cytotoxic drugs may be more effective in tumours which are not only hypoxic but have high reductase activity so that cytotoxic metabolites are produced in both hypoxic and normoxic regions. Also, where nitroimidazoles are used to identify hypoxic cells in pathological lesions by imaging techniques (e.g. NMR) they may also concentrate in normal tissues which have high levels of the appropriate reductase(s) and thus make interpretation more difficult.

The ARG grain localisation in part reflected the whole tissue measurements and supported the observation that some nitroimidazoles are able to form reactive, binding, metabolites in probably normoxic tissues with high reductase levels. It is by no means clear whether the high grain density can be regarded as a measure of cytotoxic potential in tissues. Cytotoxicity in the sebaceous gland group and the stratified squamous epithelial group would be difficult to recognise because the cells are normally discarded in a degenerate form from the skin. The position is different for the lung and liver where the cells observed to have a high grain count have a slow turn-over (replacement) rate. It is interesting to note that these same cells (airway epithelium and zone 3 of the liver) are damaged by CB 1954, a cytotoxin and radiosensitiser (Cobb, 1970) which has a similar electron affinity to MISO (Stratford et al., 1981). It is thought that the reduction of CB 1954 to a cytotoxin is dependent on the 2 electron transfer reductase DT-diaphorase (Knox et al., 1988).

In the present work it was noted that both PIMO and RSU 1069 were associated with melanin containing cells. This finding confirms previous observations that PIMO was associated with melanin in the eye in the mouse (Laurent et al., 1989), with B 16 melanoma cells in the mouse (Walling et al., 1989) and with xenografted human melanomas in nude mice (Lespinasse et al., 1989). When Dische (1987) combined PIMO with radiation in patients with malignant melanoma he observed a favourable response compared with the expected response to radiation alone. The connection between the high grain density over melanin-containing cells in normal mouse tissue and the apparent improvement in response to radiation by patients with malignant melanoma is not clear to us. Many drugs adsorb to, or react with, melanin, and occasionally this leads to cytotoxic effects (Lindquist et al., 1973). The presence of melanin in the uveal tract of the eye in pigmented mice and rats has not been observed to lead to toxic effect with PIMO or RSU 1069. In man the other critical tissue containing melanin is the brain (e.g. neuromelanin in the substantia nigra); however, melanin is not seen in the brain of mice or rats.

In conclusion, the accumulation of metabolites in normal tissues at levels greater than that for tumours is not restricted to MISO but occurs also in the analogues PIMO and RSU 1069.

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